Webappendix for the article entitled Insights for quantifying the long-term benefit of immunotherapy using quantile regression

Bassirou Mboup, Christophe Le Tourneau, Aurélien Latouche

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This document makes an easier access to the supplementary material of the article entitled **Insights for** quantifying the long-term benefit of immunotherapy using quantile regression.

1) Importing the reconstructed data set

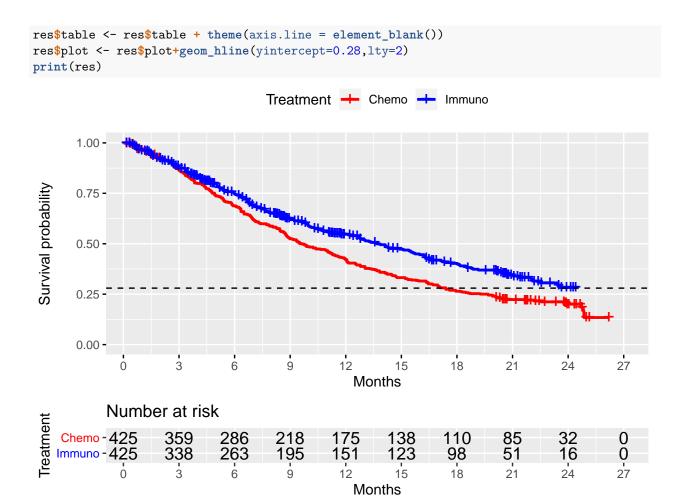
We use the algorithm of Guyot. al 2012 to reconstruct individual-level time-to-event data based on the published Kaplan–Meier curves of the randomized controlled trial (Rittmeyer et al. 2017).

The R code of the algorithm is available at https://www.mskcc.org/sites/default/files/node/137932/documents/2017-04-20-14-31-36/dataexample.zip

After reconstruction, we get in this dataset the following variables.

- time: vector of observed failure times e.g (death or censored).
- event: vector of indicator of status (1 for death and 0 for censoring).
- tmt.arm.number: vector of treatment indicator (binary with 1 for immunotherapy).
- treatment.type: the type of treatment (char) immonotherapy or chemotherapy

```
time event tmt.arm.number treatment.type
## 1 0.4059140
                   1
                                   1
                                       Atezolizumab
## 2 0.4059140
                                       Atezolizumab
## 3 0.4059140
                                       Atezolizumab
                                   1
## 4 0.5599768
                                   1
                                       Atezolizumab
## 5 0.5599768
                                   1
                                       Atezolizumab
## 6 0.5599768
                                   1
                                       Atezolizumab
## Kaplan Meier curves
fit_KM <- survfit(Surv(time, event)~tmt.arm.number,data=data_ICI_Rittmeyer)</pre>
res <- ggsurvplot(fit_KM,data=data_ICI_Rittmeyer,
           risk.table=TRUE,
           conf.int=FALSE,
           xlim=c(0.4,27),
           palette =c("red","blue"),
           xlab="Months",
           risk.table.y.text.col=T,
            break.time.by=3,
          ggtheme = theme_grey() ,
           legend.title="Treatment",
           legend.labs=c("Chemo", "Immuno")
```



2) Application of quantile regression for survival data

In the following R output, test of the coefficients compares this coefficients to 0 for a given quantile τ . For the coefficient $\beta_1(\tau)$, this test rejects or not the equality of the two treatment groups for a given quantile τ . This test is based of the Wald test and the variance of the coefficients is obtained using resampling bootstraps method.

Thus, we do not reject the hypothesis of equality of the two groups at 0.1 quantile (P.value=0.9470) illustrated by the overlap of the curves at this quantile.

We reject this hypothesis at 0.6 quantile (P.value $< 10^{-4}$).

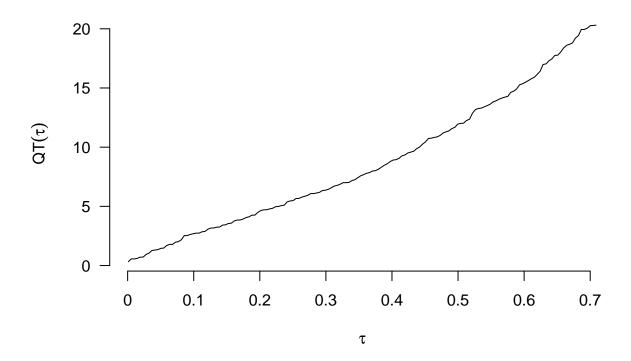
```
set.seed(123456)
x <- c(0.1, 0.2, 0.3, 0.4,0.5,0.6)
Rq <- crq(Surv(time,event)~tmt.arm.number,data=data_ICI_Rittmeyer,method="Pen")
result <- summary(Rq,taus=x)

result

##
## tau: [1] 0.1
##
## Coefficients:
## Value Lower Bd Upper Bd Std Error T Value Pr(>|t|)
```

```
## (Intercept)
                  2.70243 1.75658 2.81841 0.27088
                                                      9.97646 0.00000
## tmt.arm.number -0.01844 -0.13442 0.95376 0.27760 -0.06642 0.94704
## tau: [1] 0.2
##
## Coefficients:
                         Lower Bd Upper Bd Std Error T Value Pr(>|t|)
##
                 Value
## (Intercept)
                 4.15397 3.30234 6.26304 0.75529
                                                     5.49982 0.00000
## tmt.arm.number 0.88255 0.05166 2.45479 0.61305
                                                     1.43960 0.14998
##
## tau: [1] 0.3
##
## Coefficients:
##
                 Value
                          Lower Bd Upper Bd Std Error T Value Pr(>|t|)
                  5.86294 5.75690 6.26086 0.12857 45.60287 0.00000
## (Intercept)
## tmt.arm.number 1.02312 0.55678 2.04722 0.38022
                                                       2.69086 0.00713
##
## tau: [1] 0.4
##
## Coefficients:
##
                          Lower Bd Upper Bd Std Error T Value Pr(>|t|)
                 Value
                  7.80078 4.60256 9.11294 1.15063
## (Intercept)
                                                       6.77959 0.00000
## tmt.arm.number 2.12340 -0.49612 4.36664 1.24052
                                                       1.71169 0.08695
## tau: [1] 0.5
## Coefficients:
##
                          Lower Bd Upper Bd Std Error T Value Pr(>|t|)
                 Value
                  9.78031 8.72650 10.71357 0.50691 19.29381 0.00000
## (Intercept)
## tmt.arm.number 4.35360 2.26212 6.58893 1.10380
                                                       3.94420 0.00008
## tau: [1] 0.6
##
## Coefficients:
                          Lower Bd Upper Bd Std Error T Value Pr(>|t|)
##
                 Value
## (Intercept)
                 12.69400 10.75741 13.09069 0.59524 21.32596 0.00000
## tmt.arm.number 5.46031 3.04456 6.91123 0.98641
                                                       5.53551 0.00000
## jack
## jack
## jack
## jack
## jack
## jack
# Quantile function
tau <- Rq\sol["tau",][1:160]
q<- Rq$sol["Qhat",][1:160]</pre>
plot(tau,q,type="l",xlab = expression(tau),ylab = expression(QT(tau)),
     main="Quantile function",axes=FALSE)
axis(1,at=seq(from=0,to=0.7,by=0.1),labels=seq(from=0,to=0.7,by=0.1),las=1)
axis(2,at=seq(from=0,to=20,by=5),labels=seq(from=0,to=20,by=5),las=2)
```

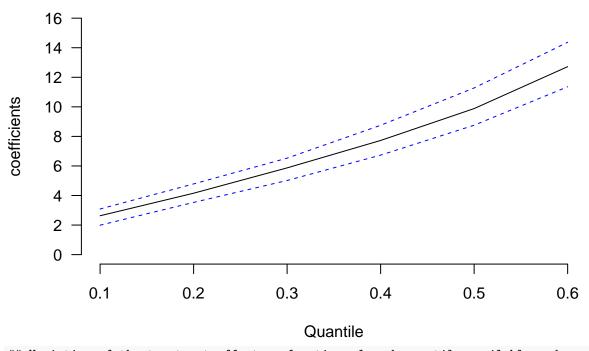
Quantile function



3) Resampling for confidence interval of coefficients

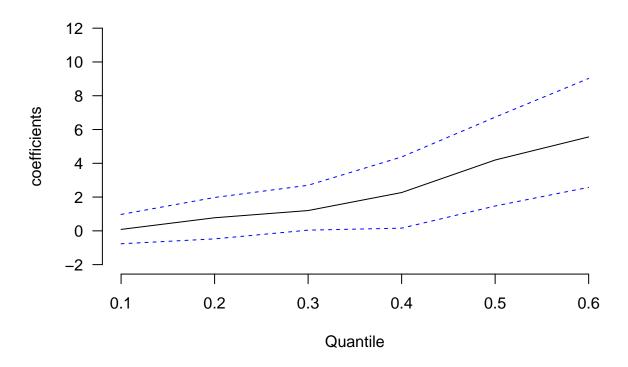
```
load("Mat.RData")
load("Moy.RData")
load("Mat1.RData")
load("Moy1.RData")
b <- matrix(data=NA,ncol=6,nrow = 1000)
for(i in 1:6){
b[,i] <-Mat1[,i][order(Mat1[,i])]</pre>
}
infO <- NA
for(i in 1:6){
inf0[i] <- b[,i][25]
sup0 <- NA
for(i in 1:6){
 \sup 0[i] \leftarrow b[,i][975]
}
plot(x,Moy1,type="l",ylim=c(0,16),ylab="coefficients",xlab = "Quantile",main="Intercept",axes = FALSE)
axis(1,at=seq(from=0.1,to=0.6,by=0.1),labels=seq(from=0.1,to=0.6,by=0.1),las=1)
axis(2,at=seq(from=0,to=16,by=2),labels=seq(from=0,to=16,by=2),las=2)
lines(x,sup0,col="blue",lty=2)
lines(x,inf0,col="blue",lty=2)
```

Intercept



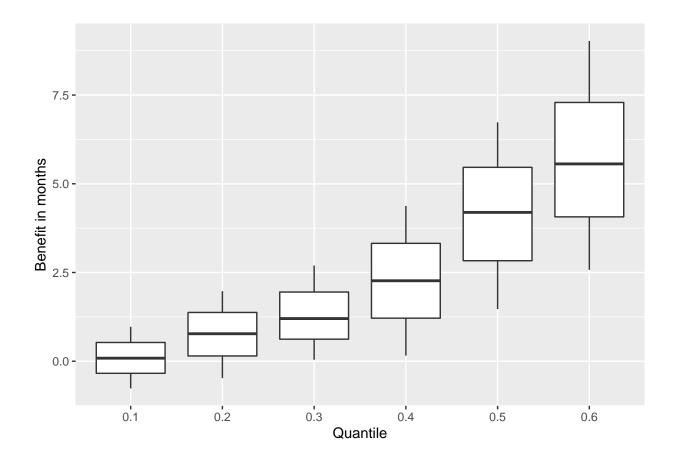
```
## Variation of the treatment effect as function of each quantile available and confidence
# intervals
b <- matrix(data=NA,ncol=6,nrow = 1000)
for(i in 1:6){
b[,i] <-Mat[,i][order(Mat[,i])]</pre>
}
inf <- NA
for(i in 1:6){
inf[i] <- b[,i][25]
sup <- NA
for(i in 1:6){
  \sup[i] \leftarrow b[,i][975]
}
plot(x,Moy,type="l",ylim=c(-2,12),ylab="coefficients",xlab = "Quantile",main="Treatment effect",axes = F
axis(1,at=seq(from=0.1,to=0.6,by=0.1),labels=seq(from=0.1,to=0.6,by=0.1),las=1)
axis(2,at=seq(from=-2,to=12,by=2),labels=seq(from=-2,to=12,by=2),las=2)
lines(x,sup,col="blue",lty=2)
lines(x,inf,col="blue",lty=2)
```

Treatment effect



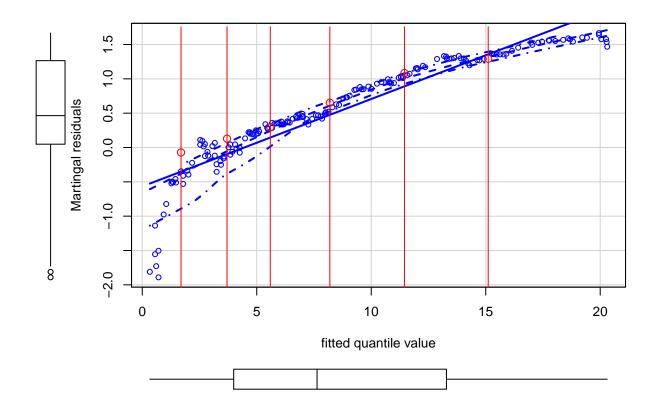
4) Benefit of treatment for each quantile

```
datafr <- rbind.data.frame(Moy,inf,sup)
names(datafr) <- c( "0.1", "0.2", "0.3", "0.4", "0.5","0.6")
p <- ggplot(stack(datafr), aes(x=factor(ind,levels=names(datafr)), y=values))+geom_boxplot()
p+labs(x="Quantile",y="Benefit in months")</pre>
```



5) Visual check of the linearity assumption

```
x <- c(0.1,0.2,0.3,0.4,0.5,0.6)
Rq <- crq(Surv(time,event)~tmt.arm.number,data=data_ICI_Rittmeyer,method="Pen")
tau <- Rq$sol["tau",][1:160]
obs <- unname(quantile(data_ICI_Rittmeyer$time,tau))
obss <- unname(quantile(data_ICI_Rittmeyer$time,x))
q<- Rq$sol["Qhat",][1:160]
q1 <- Rq$sol["Qhat",][c(24,46,69,91,114,136)]
residu <- obs-q
residus <- obs-q1
scatterplot(q,log(abs(residu)),xlab="fitted quantile value",ylab="Martingal residuals")
points(q1,log(abs(residus)),col='red')
for(i in 1:6){
   abline(v=q1[i],col="red")
}</pre>
```



```
set.seed(4567)
load("ImmunoRit.RData")
library(knitr)
library(riskRegression)
## riskRegression version 2020.02.05
db <- ImmunoRit
cox1<-coxph(Surv(time,event)~Treat, data=db,x=TRUE)</pre>
db2<-mutate(db,Treat=0)</pre>
pr<-predictCox(cox1,times=db2$time,newdata=db2,type="cumhazard")</pre>
cumhaz<-pr$cumhazard[1,]</pre>
beta1 <- log(0.69)
lamda <- cumhaz
beta2 < -log(0.72)
beta3 <- log(0.36)
genereTime <- function(x,lamda,trt,uni){</pre>
  return(-log(uni)/(lamda*exp(beta2*trt+beta1*x+beta3*x*trt)))
ttt <- ImmunoRit$Treat</pre>
```

```
n=850
PDL1 <- rep(NA,n)
for(i in 1:n) {
 x <- -1
  while(x<0){</pre>
    uni <- runif(1,0,1)
    x <- uniroot((function(x)(genereTime(x,lamda[i],ttt[i],uni)-ImmunoRit$time[i])),lower=-10,upper=10,
 }
 PDL1[i] \leftarrow x
}
summary(PDL1)
       Min. 1st Qu.
                       Median
                                   Mean 3rd Qu.
## 0.00724 1.06234 2.55940 4.24953 6.35912 24.56969
ImmunoRit$PDL1 <- PDL1</pre>
Rq_1 <- crq(Surv(time,event)~Treat*PDL1,data=ImmunoRit,method="PengHuang")</pre>
result <- summary(Rq_1,tau=c(0.1,0.20,0.30,0.40,0.50,0.6))
kable(result[[6]][2]$coefficients)
```

	Value	Lower Bd	Upper Bd	Std Error	T Value	$\Pr(> t)$
(Intercept)	1.429052	-0.3058327	2.564273	0.7321832	1.951769	0.0509657
Treat	1.734772	0.4067428	3.314709	0.7418417	2.338466	0.0193631
PDL1	1.921049	1.5126417	2.325287	0.2073113	9.266495	0.0000000
Treat:PDL1	6.553311	5.5314831	7.335916	0.4603229	14.236335	0.0000000