

# Package ‘CTD’

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**Title** CTD method for “connecting the dots” in weighted graphs

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**Description** An R package for pattern discovery in weighted graphs. Two use cases are achieved: 1) Given a weighted graph and a subset of its nodes; do the nodes show significant connectedness? 2) Given a weighted graph and two subsets of its nodes; do the subsets show significant similarity?

**Depends** R (>= 3.3.0),  
igraph,  
plotly,  
gplots,  
RColorBrewer

**License** MIT License

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**LazyData** true

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data.HMDBtoKEGG	<i>Convert HMDB IDs to KEGG compound IDs.</i>
-----------------	---

---

### Description

A function that converts HMDB IDs to KEGG compounds.

### Usage

```
data.HMDBtoKEGG(hmdb.ids)
```

### Arguments

hmdb.ids            - A character vector of HMDB IDs.

### Examples

```
kegg.ids = data.HMDBtoKEGG(hmdb.ids)
```

---

data.LabeltoKEGG	<i>Convert compound names to KEGG compound IDs.</i>
------------------	---

---

### Description

A function that converts HMDB IDs to KEGG compounds.

### Usage

```
data.LabeltoKEGG(compound.names)
```

### Arguments

compound.names    - A character vector of metabolite (compound) names.

### Examples

```
kegg.ids = data.HMDBtoKEGG(compound.names)
```

---

data.surrogateProfiles

*Surrogate profiles*


---

### Description

Fill in a data matrix rank, when your data is low n, high p. Fill in rank with surrogate profiles.

### Usage

```
data.surrogateProfiles(data, sd = 1, useMnDiseaseProfile = FALSE,
  addHealthyControls = TRUE, ref_data = NULL)
```

### Arguments

- data - Data matrix with observations as rows, features as columns.
- sd - The level of variability (standard deviation) around each feature's mean you want to add in surrogate profiles.
- useMnDiseaseProfile - Boolean. For disease cohorts not showing homogeneity, mean across disease profiles and generate disease surrogates around this mean.
- addHealthyControls - Boolean. Add healthy control profiles to data?

### Value

data\_mx - Data matrix with added surrogate profiles.

---

graph.diffuseP1

*Diffuse Probability P1 from a starting node.*


---

### Description

Recursively diffuse probability from a starting node based on the connectivity of the background knowledge graph, representing the likelihood that a variable will be most influenced by a perturbation in the starting node.

### Usage

```
graph.diffuseP1(p1, startNode, G, visitedNodes, graphNumber = 1,
  verbose = FALSE)
```

**Arguments**

p1	- The probability being dispersed from the starting node, startNode.
startNode	- The first variable drawn in the node ranking, from which p1 gets dispersed.
G	- A list of probabilities, with names of the list being the node names in the background knowledge graph.
visitedNodes	- The history of previous draws in the node ranking sequence.
graphNumber	- If testing against multiple background knowledge graphs, this is the index associated with the adjacency matrix that codes for G. Default value is 1.
verbose	- If debugging or tracking a diffusion event, verbose=TRUE will activate print statements. Default is FALSE.

**Value**

G - A list of returned probabilities after the diffusion of probability has truncated, with names of the list being the node names in the background knowledge graph.

**Examples**

```
# Read in any network via its adjacency matrix
tmp=matrix(1, nrow=100, ncol=100)
for (i in 1:100) {
  for (j in 1:100) {
    tmp[i, j]=rnorm(1, mean=0, sd=1)
  }
}
colnames(tmp)=sprintf("MolPheno%d", 1:100)
ig=graph.adjacency(tmp, mode="undirected", weighted=TRUE, add.colnames="name")
V(ig)$name=tolower(V(ig)$name)
adjacency_matrix=list(as.matrix(get.adjacency(ig, attr="weight"))) # Must have this declared as a GLOBAL variable
# Set other tuning parameters
p0=0.1 # 10% of probability distributed uniformly
p1=0.9 # 90% of probability diffused based on edge weights in networks
thresholdDiff=0.01
G=vector(mode="list", length=length(V(ig)$name))
names(G)=V(ig)$name
G=lapply(G, function(i) i[[1]]=0)
startNode=names(G)[1]
visitedNodes=G[1]
probs_afterCurrDraw=graph.diffuseP1(p1, startNode, G, visitedNodes, 1, TRUE)
```

---

graph.diffuseP1Movie    *Make a movie of the diffusion of probability, P1, from a starting node.*

---

**Description**

Recursively diffuse probability from a starting node based on the connectivity of the background knowledge graph, representing the likelihood that a variable will be most influenced by a perturbation in the starting node.

**Usage**

```
graph.diffuseP1Movie(p1, startNode, G, visitedNodes, ig,
  recursion_level = 1, output_dir = getwd())
```

**Arguments**

p1	- The probability being dispersed from the starting node, startNode.
startNode	- The first variable drawn in the node ranking, from which p1 gets dispersed.
G	- A list of probabilities, with names of the list being the node names in the background knowledge graph.
visitedNodes	- A character vector of node names, storing the history of previous draws in the node ranking.
graphNumber	- If testing against multiple background knowledge graphs, this is the index associated with the adjacency matrix that codes for G. Default value is 1.

**Value**

G - A list of returned probabilities after the diffusion of probability has truncated, with names of the list being the node names in the background knowledge graph.

**Examples**

```
# 7 node example graph illustrating diffusion of probability based on network connectivity
# from Thistlethwaite et al., 2019.
adj_mat = rbind(c(0,2,1,0,0,0,0), # A
               c(2,0,1,0,0,0,0), # B
               c(1,0,0,1,0,0,0), # C
               c(0,0,1,0,2,0,0), # D
               c(0,0,0,2,0,2,1), # E
               c(0,0,0,1,2,0,1), # F
               c(0,0,0,0,1,1,0)  # G
               )
rownames(adj_mat) = c("A", "B", "C", "D", "E", "F", "G")
colnames(adj_mat) = c("A", "B", "C", "D", "E", "F", "G")
adjacency_matrix = list(adj_mat)
ig = graph.adjacency(as.matrix(adj_mat), mode="undirected", weighted=TRUE)
G=vector(mode="list", length=7)
G[1:length(G)] = 0
names(G) = c("A", "B", "C", "D", "E", "F", "G")
startNode = "A"
visitedNodes = startNode
# Diffuse 100% of probability from startNode "A"
p1 = 1.0
# Probability diffusion truncates at
thresholdDiff=0.01
coords = layout.fruchterman.reingold(ig)
V(ig)$x = coords[,1]
V(ig)$y = coords[,2]
# Global variable imgNum
imgNum=1
G_new = graph.diffuseP1Movie(p1, startNode, G, visitedNodes, ig, 1, getwd())
```

---

<code>graph.naivePruning</code>	<i>Pruning edges from disease differential network that also occur in reference-only network.</i>
---------------------------------	---

---

### Description

Recursively diffuse probability from a starting node based on the connectivity of the background knowledge graph, representing the likelihood that a variable will be most influenced by a perturbation in the starting node.

### Usage

```
graph.naivePruning(ig_dis, ig_ref)
```

### Arguments

<code>ig_dis</code>	- The igraph object associated with the disease+reference trained differential interaction network.
<code>ig_ref</code>	- The igraph object associated with the reference-only trained interaction network.

### Value

`ig_pruned` - The pruned igraph object of the disease+reference differential interaction network, with reference edges subtracted.

### Examples

```
ig_pruned=graph.naivePruning(ig_dis, ig_ref)
```

---

<code>mle.getEncodingLength</code>	<i>Minimum encoding length (MLE)</i>
------------------------------------	--------------------------------------

---

### Description

This function calculates the minimum encoding length associated with a subset of variables given a background knowledge graph.

### Usage

```
mle.getEncodingLength(bs, pvals, ptID, G)
```

### Arguments

<code>bs</code>	- A list of bitstrings associated with a given patient's perturbed variables.
<code>pvals</code>	- The matrix that gives the perturbation strength significance for all variables (columns) for each patient (rows)
<code>ptID</code>	- The row name in <code>data.pvals</code> corresponding to the patient you specifically want encoding information for.
<code>G</code>	- A list of probabilities with list names being the node names of the background graph.

**Value**

df - a data.frame object, for every bitstring provided in bs input parameter, a row is returned with the following data: the patientID; the bitstring evaluated where T denotes a hit and 0 denotes a miss; the subsetSize, or the number of hits in the bitstring; the individual p-values associated with the variable's perturbations, delimited by '/'; the combined p-value of all variables in the set using Fisher's method; Shannon's entropy, IS.null; the minimum encoding length IS.alt; and IS.null-IS.alt, the d.score.

**Examples**

```
# Identify the most significant subset per patient, given the background graph
data_mx.pvals = t(apply(data_mx, c(1,2), function(i) 2*pnorm(abs(i), lower.tail = FALSE)))
for (pt in 1:ncol(data_mx)) {
  ptID = colnames(data_mx)[pt]
  res = mle.getEncodingLength(ptBSbyK[[ptID]], data_mx.pvals, ptID, G)
  res = res[order(res[, "d.score"], decreasing=TRUE),]
  print(res)
}
```

---

mle.getMinPtDistance	<i>Metabolite set enrichment analysis (MSEA) using pathway knowledge curated by Metabolon</i>
----------------------	---

---

**Description**

A function that returns the pathway enrichment score for all perturbed metabolites in a patient's full metabolomic profile.

**Usage**

```
mle.getMinPtDistance(allSimMatrices)
```

**Arguments**

allSimMatrices - A list of all similarity matrices, across all k for a given graph, or across many graphs.

**Examples**

```
# Get patient distances
data_mx.pvals = apply(data_mx, c(1,2), function(i) 2*pnorm(abs(i), lower.tail = FALSE))
res = list()
t = list(ncd=matrix(NA, nrow=ncol(data_mx), ncol=ncol(data_mx)))
rownames(t$ncd) = colnames(data_mx)
colnames(t$ncd) = colnames(data_mx)
for (i in 1:kmx) {
  res[[i]] = t
}
for (pt in 1:ncol(data_mx)) {
  print(pt)
  ptID = colnames(data_mx)[pt]
  for (pt2 in pt:ncol(data_mx)) {
    ptID2 = colnames(data_mx)[pt2]
```

```

    tmp = mle.getPtSim(ptBSbyK[[ptID]], ptID, ptBSbyK[[ptID2]], ptID2, data_mx, ranks)
    for (k in 1:kmx) {
      res[[k]]$ncd[ptID, ptID2] = tmp$NCD[k]
      res[[k]]$ncd[ptID2, ptID] = tmp$NCD[k]
    }
  }
}
patientSimilarity = mle.getMinPtDistance(res)

```

---

mle.getPtSim	<i>Patient similarity using mutual information MLE metric of patients' most modular, perturbed subsets.</i>
--------------	---

---

## Description

This function calculates the universal distance between patients, using a mutual information metric, where self-information comes from the minimum encoding length of each patient's encoded modular perturbations in the background knowledge graph.

## Usage

```
mle.getPtSim(p1.optBS, ptID, p2.optBS, ptID2, data_mx, ranks)
```

## Arguments

p1.optBS	- The optimal bitstring associated with patient 1.
ptID	- The identifier associated with patient 1's sample.
p2.optBS	- The optimal bitstring associated with patient 2.
data_mx	- The matrix that gives the perturbation strength (z-scores) for all variables (columns) for each patient (rows).
ptID2	- The identifier associated with patient 2's sample.

## Value

patientSim - a similarity matrix, where row and columns are patient identifiers.

## Examples

```

# Get patient distances
data_mx.pvals = apply(data_mx, c(1,2), function(i) 2*pnorm(abs(i), lower.tail = FALSE))
res = list()
tt = list(ncd=matrix(NA, nrow=ncol(data_mx), ncol=ncol(data_mx)))
rownames(tt$ncd) = colnames(data_mx)
colnames(tt$ncd) = colnames(data_mx)
for (i in 1:kmx) {
  res[[i]] = tt
}
for (pt in 1:ncol(data_mx)) {
  print(pt)
  ptID = colnames(data_mx)[pt]
  for (pt2 in pt:ncol(data_mx)) {
    ptID2 = colnames(data_mx)[pt2]

```



```

    for (k in 1:kmx) {
      tmp = mle.getPtSim(ptBSbyK[[ptID]][k], ptID, ptBSbyK[[ptID2]][k], ptID2, data_mx, ranks)
      res[[k]]$ncd[ptID, ptID2] = tmp$NCD
      res[[k]]$ncd[ptID2, ptID] = tmp$NCD
    }
  }
}

```

---

mle.kraftMcMillan	<i>Apply the Kraft-McMillian Inequality using a specific encoding algorithm.</i>
-------------------	--

---

## Description

A power analysis of the encoding algorithm using to encode subsets of S in G.

## Usage

```
mle.kraftMcMillan(G, k, multiNode = FALSE)
```

## Arguments

G	- A character vector of all node names in the background knowledge graph.
k	- The size of the node name subsets of G.
multiNode	- Boolean, indicating whether to use the multi-node diffusion encoding algorithm (TRUE) or the single-node diffusion encoding algorithm (FALSE). Default is FALSE.

## Value

IA - a list of bitlengths associated with all outcomes in the N choose K outcome space, with the names of the list elements the node names of the encoded nodes

## Examples

```

G = list(A=0, B=0, C=0, D=0, E=0, F=0, G=0)
names(G) = tolower(names(G))
adj_mat = rbind(c(0,2,1,0,0,0,0), #A's neighbors
               c(2,0,1,0,0,0,0), #B's neighbors
               c(1,1,0,1,0,0,0), #C's neighbors
               c(0,0,1,0,2,1,0), #D's neighbors
               c(0,0,0,2,0,2,1), #E's neighbors
               c(0,0,0,1,2,0,1), #F's neighbors
               c(0,0,0,0,1,1,0)  #G's neighbors
               )
rownames(adj_mat) = names(G)
colnames(adj_mat) = names(G)
adjacency_matrix = list(adj_mat)
IA = mle.kraftMcMillian(G, 2)
# Power to find effects is
sum(2^unlist(IA))

```

---

```
multiNode.getNodeRanks
```

*Generate the "adaptive walk" node rankings, starting from a given perturbed variable*

---

### Description

This function calculates the node rankings starting from a given perturbed variable in a subset of variables in the background knowledge graph.

### Usage

```
multiNode.getNodeRanks(S, G, num.misses = NULL)
```

### Arguments

**S** - A character vector of the node names for the subset of nodes you want to encode.

### Value

**ranks** - A list of character vectors of node names in the order they were drawn by the probability diffusion algorithm, from each starting node in S.

### Examples

```
# Read in any network via its adjacency matrix
tmp = matrix(1, nrow=100, ncol=100)
for (i in 1:100) {
  for (j in 1:100) {
    tmp[i, j] = rnorm(1, mean=0, sd=1)
  }
}
colnames(tmp) = sprintf("Compound%d", 1:100)
ig = graph.adjacency(tmp, mode="undirected", weighted=TRUE, add.colnames="name")
V(ig)$name = tolower(V(ig)$name)
# Get node rankings for graph
G = vector(mode="list", length=length(V(ig)$name))
names(G) = V(ig)$name
S = names(G)[1:3]
ranks = multiNode.getNodeRanks(S, G)
```

---

```
multiNode.getNodeRanksMovie
```

*Capture the movement of the adaptive walk of the diffusion probability method.*

---

### Description

Make a movie of the adaptive walk the diffusion probability method makes in search of a given patient's perturbed variables.

**Usage**

```
multiNode.getNodeRanksMovie(subset.nodes, ig, output_filepath,
                             movie = TRUE, zoomIn = FALSE)
```

**Arguments**

subset.nodes     - The subset of variables, S, in a background graph, G.

ig                - The igraph object associated with the background knowledge graph.

output\_filepath     - The local directory at which you want still images to be saved.

movie             - If you want to make a movie, set to TRUE. This will produce a set of still images that you can stream together to make a movie. Default is TRUE. Alternatively (movie=FALSE), you could use this function to get the node labels returned for each node ranking starting with a perturbed variable.

zoomIn            - Boolean. Delete nodes outside of node subset's order 1 neighborhood?. Default is FALSE.

**Value**

ranksByStartNode - a list object of node rankings Each element is based on a different startNode. Images are also generated in the output\_directory specified.

**Examples**

```
# Read in any network via its adjacency matrix
tmp = matrix(1, nrow=100, ncol=100)
for (i in 1:100) {
  for (j in 1:100) {
    tmp[i, j] = rnorm(1, mean=0, sd=1)
  }
}
colnames(tmp) = sprintf("Compound%d", 1:100)
ig = graph.adjacency(tmp, mode="undirected", weighted=TRUE, add.colnames="name")
V(ig)$name = tolower(V(ig)$name)
# Set other tuning parameters
p0=0.1 # 10% of probability distributed uniformly
p1=0.9 # 90% of probability diffused based on edge weights in networks
thresholdDiff=0.01
G = vector(mode="list", length=length(V(ig)$name))
names(G) = V(ig)$name
subset.nodes = names(G)[sample(1:length(G), 3)]
multiNode.getNodeRanksMovie(subset.nodes, ig, output_filepath = getwd(), movie=TRUE)
```

---

multiNode.getPtBSbyK     *Generate patient-specific bitstrings from adaptive network walk.*

---

**Description**

This function calculates the bitstrings (1 is a hit; 0 is a miss) associated with the adaptive network walk made by the diffusion algorithm trying to find the variables in the encoded subset, given the background knowledge graph.

**Usage**

```
multiNode.getPtBSbyK(S, ranks)
```

**Arguments**

**S** - A character vector of node names describing the node subset to be encoded.

**ranks** - The list of node ranks calculated over all possible nodes, starting with each node in subset of interest.

**Value**

pt.byK - a list of bitstrings, with the names of the list elements the node names of the encoded nodes

**Examples**

```
# Get bitstrings associated with each patient's top kmx variable subsets
kmx = 15
ptBSbyK = list()
for (pt in 1:ncol(data_mx)) {
  S = data_mx[order(abs(data_mx[,pt]), decreasing=TRUE),pt][1:kmx]
  ptBSbyK[[ptID]] = multiNode.getPtBSbyK(S, ranks)
}
```

---

```
pathway.ListMaps_metabolon
```

*Get All Metabolites In Metabolon's Pathway Knowledgebase*

---

**Description**

Get All Metabolites In Metabolon's Pathway Knowledgebase

**Usage**

```
pathway.ListMaps_metabolon()
```

**Value**

pwys - List of pathway maps curated by Metabolon's Metabolync.

**Examples**

```
pwys = pathway.ListMaps_metabolon()
print(pwys)
```

---

```
pathway.ListMetabolites_metabolon
```

*Get All Metabolites In Metabolon's Pathway Knowledgebase*

---

### Description

Get All Metabolites In Metabolon's Pathway Knowledgebase

### Usage

```
pathway.ListMetabolites_metabolon()
```

### Value

mets - a character vector of unique metabolites and enzymes found in at least 1 pathway in Metabolon's pathway knowledgebase.

### Examples

```
mets = pathway.ListMetabolites_metabolon()
print(mets)
```

---

```
plot.getPathwayIgraph  plot.getPathwayIgraph
```

---

### Description

```
plot.getPathwayIgraph
```

### Usage

```
plot.getPathwayIgraph(input, Pathway.Name)
```

### Arguments

input	- A list object of parameters (esp. from R shiny app). Required parameters are ptIDs, diagClass and pathwayMapId.
Pathway.Name	- The name of the pathway map for which you want the topological information.

### Value

template.ig - Igraph object of selected pathway map.

**Examples**

```
data(Miller2015)
# Input is supplied by R shiny app, but you can hard code parameters as a list object, too, to test functionality
input = list()
input$ptIDs = colnames(Miller2015)[4]
input$diagClass = "paa"
input$pathwayMapId = "All"
ig = plot.getPathwayIgraph(input, Miller2015)
# Returns a blank template for selected pathway.
plot.igraph(ig, edge.arrow.size = 0.01)
```

---

plot.hmSim	<i>Generate heatmap plot of patient similarity matrix.</i>
------------	--

---

**Description**

This function plots a heatmap of a patient similarity matrix.

**Usage**

```
plot.hmSim(patientSim, path=getwd(), diagnoses=NULL)
```

**Arguments**

patientSim	- The patient similarity matrix.
path	- The filepath to a directory in which you want to store the .png file.
diagnoses	- A character vector of diagnostic labels associated with the rownames of patientSim.

**Examples**

```
plot.hmSim(patientSim, path=getwd(), diagnoses)
```

---

plot.knnSim	<i>Visualize the confusion matrix using nearest neighbor as classification model.</i>
-------------	---

---

**Description**

Visualize the confusion matrix using nearest neighbor as classification model.

**Usage**

```
plot.knnSim(patientSim, diagnoses, diag)
```

**Arguments**

patientSim	- The patient similarity matrix.
diagnoses	- A character vector of diagnostic labels associated with the rownames of patientSim. The names of this vector are patient IDs, and values are diagnostic labels.
diag	- The diagnosis associated with positive controls in your data.

**Value**

p - a plotly scatter plot colored by provided diagnostic labels.

**Examples**

```
# if you have diagnostic labels associated with the colnames(data_mx), send them using diagnoses parameter
p = plot.knnSim(patientSim, diagnoses, diag="diseased")
p
```

---

plot.mdsSim	<i>View patient clusters using multi-dimensional scaling.</i>
-------------	---

---

**Description**

This function plots the provided patient similarity matrix in a lower dimensional space using multi-dimensional scaling, which is well suited for similarity metrics.

**Usage**

```
plot.mdsSim(patientSim, diagnoses, k, diag)
```

**Arguments**

patientSim	- The patient similarity matrix.
diagnoses	- A character vector of diagnostic labels associated with the rownames of patientSim.
k	- The number of dimension you want to plot your data using multi-dimensional scaling.
diag	- The diagnosis associated with positive controls in your data.

**Value**

p - a plotly scatter plot colored by provided diagnostic labels.

**Examples**

```
# if you have diagnostic labels associated with the colnames(data_mx), send them using diagnoses parameter
p = plot.mdsSim(patientSim, diagnoses, k=2, diag="diseased")
p
p = plot.mdsSim(patientSim, diagnoses, k=3, diag="diseased")
p
```

---

plot.pathwayMap	<i>Generate pathway map with patient perturbation data superimposed.</i>
-----------------	--

---

### Description

Generate pathway map with patient perturbation data superimposed.

### Usage

```
plot.pathwayMap(Pathway, ptID, pt.zscore, zscore.threshold, scale, out.path, SVG=TRUE)
```

### Arguments

Pathway	- The name of the pathway map you want to plot patient data on.
ptID	- An identifier string associated with the patient.
pt.zscore	- A named vector of metabolites with corresponding z-scores.
zscore.threshold	- Plot all z-scores > or < this threshold.
scale	- Integer associated with increase in node size.
out.path	- The directory in which you want to store image files.
SVG	- Save as SVG or PNG? If SVG is TRUE, then an SVG image is saved. If FALSE, a PNG is saved.

### Examples

```
Pathway = pathway.ListMaps_metabolon()
data(Miller2015)
Miller2015 = Miller2015[,grep("IEM", colnames(Miller2015))]
ptID = colnames(Miller2015)[1]
pt.zscore = Miller2015[,1]
plot.pathwayMap(Pathway[1], ptID, pt.zscore, zscore.threshold, scale=1, out.path=getwd(), SVG=TRUE)
```

---

singleNode.getNodeRanksMovie
------------------------------

*Capture the movement of the fixed, single-node walk of the diffusion probability method.*

---

### Description

Make a movie of the fixed, single-node walk the diffusion probability method makes in search of a given patient's perturbed variables.

### Usage

```
singleNode.getNodeRanksMovie(subset.nodes, ig, output_filepath,
  movie = TRUE, zoomIn = FALSE)
```



**Arguments**

- subset.nodes     - The subset of variables, S, in a background graph, G.
- ig                - The igraph object associated with the background knowledge graph.
- output\_filepath     - The local directory at which you want still images to be saved.
- movie            - If you want to make a movie, set to TRUE. This will produce a set of still images that you can stream together to make a movie. Default is TRUE. Alternatively (movie=FALSE), you could use this function to get the node labels returned for each node ranking starting with a perturbed variable.
- zoomIn           - Boolean. Delete nodes outside of node subset's order 1 neighborhood?. Default is FALSE.

**Value**

ranksByStartNode - a list object of node rankings Each element is based on a different startNode. Images are also generated in the output\_directory specified.

**Examples**

```
# Read in any network via its adjacency matrix
tmp = matrix(1, nrow=100, ncol=100)
for (i in 1:100) {
  for (j in 1:100) {
    tmp[i, j] = rnorm(1, mean=0, sd=1)
  }
}
colnames(tmp) = sprintf("Compound%d", 1:100)
ig = graph.adjacency(tmp, mode="undirected", weighted=TRUE, add.colnames="name")
V(ig)$name = tolower(V(ig)$name)
adjacency_matrix = list(tmp) # MUST BE GLOBAL VARIABLE
# Set other tuning parameters
p0=0.1 # 10% of probability distributed uniformly
p1=0.9 # 90% of probability diffused based on edge weights in networks
thresholdDiff=0.01
G = vector(mode="list", length=length(V(ig)$name))
names(G) = V(ig)$name
subset.nodes = names(G)[sample(1:length(G), 3)]
singleNode.getNodeRanksMovie(subset.nodes, ig, output_filepath = getwd(), movie=TRUE)
```

---

singleNode.getNodeRanksN

*Generate the fixed, single-node diffusion node rankings, starting from a given perturbed variable.*

---

**Description**

This function calculates the node rankings starting from a given perturbed variable in a subset of variables in the background knowledge graph.

**Usage**

```
singleNode.getNodeRanksN(n, G, S = NULL, misses.thresh = NULL)
```

**Arguments**

- n                   - The index (out of a vector of node names) of the node ranking you want to calculate.
- G                   - A list of probabilities with list names being the node names of the background graph.
- S                   - A character vector of node names in the subset you want the network walker to find.
- misses.thresh      - The number of "misses" the network walker will tolerate before switched to fixed length codes for remaining nodes to be found.

**Value**

current\_node\_set - A character vector of node names in the order they were drawn by the probability diffusion algorithm.

**Examples**

```
# Get node rankings for graph
ranks = list()
for (n in 1:length(G)) {
  print(sprintf("Generating node rankings starting with node %s", names(G)[n]))
  ranks[[n]] = singleNode.getNodeRanksN(n, G)
}
names(ranks) = names(G)
```

---

singleNode.getPtBSbyK   *Generate patient-specific bitstrings from the fixed, single-node network walker.*

---

**Description**

This function calculates the bitstrings (1 is a hit; 0 is a miss) associated with the fixed, single-node network walker trying to find the variables in the encoded subset, given the background knowledge graph.

**Usage**

```
singleNode.getPtBSbyK(S, ranks, num.misses = NULL)
```

**Arguments**

- S                   - A character vector of node names describing the node subset to be encoded.
- ranks               - The list of node ranks calculated over all possible nodes, starting with each node in subset of interest.
- num.misses          - The number of misses tolerated by the network walker before path truncation occurs.

**Value**

pt.byK - a list of bitstrings, with the names of the list elements the node names of the encoded nodes

**Examples**

```
# Load in your profiling data (rows are compounds, columns are samples)
data_mx = read.table("your_profiling_data.txt", sep="\t", header=TRUE)
# Get bitstrings associated with each patient's top kmx variable subsets
kmx = 15
ptBSbyK = list()
for (pt in 1:ncol(data_mx)) {
  S = data_mx[order(abs(data_mx[,pt]), decreasing=TRUE),pt][1:kmx]
  ptBSbyK[[ptID]] = singleNode.getPtBSbyK(S, ranks)
}
```

---

stats.entropyFunction *Entropy of a bit-string*


---

**Description**

The entropy of a bitstring (ex: 1010111000) is calculated.

**Usage**

```
stats.entropyFunction(bitString)
```

**Arguments**

x                      - A vector of 0's and 1's.

**Value**

e - a floating point percentage, between 0 and 1.

**Examples**

```
stats.entropyFunction(c(1,0,0,0,1,0,0,0,0,0,0,0,0))
> 0.6193822
stats.entropyFunction(c(1,1,1,1,1,1,1,1,0,0,0,0,0,0,0))
> 1
stats.entropyFunction(c(1,1,1,1,1,1,1,1,1,1,1,1,1,1,1))
> 0
```

---

stats.fishersMethod *Fisher's Combined P-value*


---

**Description**

Fisher's combined p-value, used to combine the results of individual statistical tests into an overall hypothesis.

**Usage**

```
stats.fishersMethod(x)
```

**Arguments**

x - A vector of p-values (floating point numbers).

**Value**

a floating point number, a combined p-value using Fisher's method.

**Examples**

```
stats.fishersMethod(c(0.2,0.1,0.3))
> 0.1152162
```

---

```
stats.getMSEA_Metabolon
```

*Metabolite set enrichment analysis (MSEA) using pathway knowledge  
curated by Metabolon*

---

**Description**

A function that returns the pathway enrichment score for all perturbed metabolites in a patient's full metabolomic profile.

**Usage**

```
stats.getMSEA_Metabolon(abs_filename_dataset, abs_filename_classes,
  pathway_knowledgebase = "Metabolon", output_dir = getwd(),
  expt_name = "msea_results")
```

**Arguments**

abs\_filename\_dataset

- Relative or absolute path to relevant .gct file. A .gct file contains profiling data, rows are compounds and columns are sample IDs.

abs\_filename\_classes

- Relative or absolute path to relevant .cls file. A .cls file contains a mapping of class labels to columns in the .gct file.

output\_dir

- The path associated with the folder in which MSEA results will be saved.

expt\_name

- A name to be associated with the experiment you are analyzing. This name will be used in filestems of results rendered in output\_dir.

pathway.knowledgebase

- The filename of the .gmt file associated with the pathway knowledge desired. Currently only "Metabolon" is offered, though "KEGG", "WikiPathways", "SM-PDB" and/or "Reactome" can be added in future versions.

## Examples

```
data(Miller2015)
Miller2015 = Miller2015[,grep("IEM", colnames(Miller2015))]
# Generate a .cls file for your data.
diagnoses = gsub("[[:digit:]]", "", colnames(Miller2015))
diag.ind = diagnoses
diag.ind[which(diag.ind!="Argininemia")] = 0
diag.ind[which(diag.ind=="Argininemia")] = 1
diag.ind = as.numeric(diag.ind)
# Manually add the following text to 1st line of .cls,
  where num_samples is the length of diag.ind: #num_samples 1 2
# Manually add the following text to 2nd line of .cls: #disease control
write.table(diag.ind, file=system.file("extdata/MSEA_Datasets/Miller2015_arg.cls", package="CTD"),
  sep=" ", quote=FALSE, row.names = FALSE, col.names = FALSE)

# Create a .gct file.
data_mx = Miller2015
data_mx = data_mx[, order(diags.ind)]
data_mx = cbind(rep(NA, nrow(data_mx)), data_mx)
colnames(data_mx)[1] = "DESCRIPTION"
write.table(data_mx, file=system.file("extdata/MSEA_Datasets/Miller2015.gct", package="CTD"),
  sep="\t", quote=FALSE, row.names = TRUE)

# Generate a .gmt file.
population = names(met.profile)
paths.hsa = list.dirs(path="../inst/extdata", full.names = FALSE)
paths.hsa = paths.hsa[-which(paths.hsa %in% c("", "RData", "allPathways"))]
sink(system.file("extdata/Pathway_GMTs/Metabolon.gmt", package="CTD"))
for (p in 1:length(paths.hsa)) {
  load(sprintf("../inst/extdata/RData/%s.RData", paths.hsa[p]))
  pathway.compounds = V(ig)$label[which(V(ig)$shape=="circle")]
  pathCompIDs = unique(tolower(pathway.compounds[which(pathway.compounds %in% population)]))
  print(sprintf("%s\t%s", paths.hsa[p], paste(pathCompIDs, collapse=" ")), quote=FALSE)
}
sink()
print("test")
abs_filename_dataset = system.file("extdata/MSEA_Datasets/Miller2015.gct", package="CTD")
abs_filename_classes = system.file("extdata/MSEA_Datasets/Miller2015_arg.cls", package="CTD")
pathway.data = stats.getMSEA_Metabolon(abs_filename_dataset, abs_filename_classes, pathway_knowledgebase = '
  output_dir = getwd(), expt_name="msea_results")
```

---

stats.getORA\_Metabolon

*Metabolite set enrichment analysis (MSEA) (using a hypergeometric test) using pathway knowledge curated by Metabolon*

---

## Description

A function that returns the pathway enrichment score for all perturbed metabolites in a patient's full metabolomic profile.

## Usage

```
stats.getORA_Metabolon(met.profile, threshold = 3, type = "zscore",
  gene.profile = NULL)
```

**Arguments**

met.profile	- A character vector of a patient's metabolomic profile, including KEGG IDs and the associated z-score or p-value describing the level of the metabolite compared to controls.
threshold	- A cutoff to select metabolites with a zscore > threshold or < -1*threshold.
type	- Either "p-value" or "z-score".
gene.profile	- Default set to NULL, meaning the default enrichment analysis only considers metabolites. However, if you have gene data, too, set this parameter to a character vector of the gene names with found variants in the patient's record. Gene IDs must be converted to Entrez Identifiers.

**Examples**

```
pathway.data = stats.getORA_Metabolon(met.profile, threshold=3, "z-score", NULL)
```

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