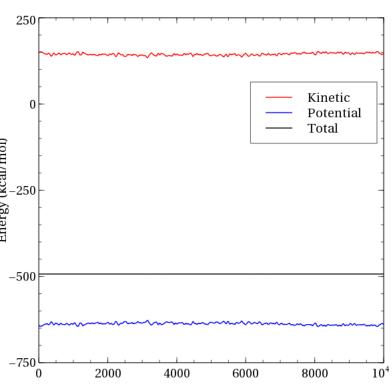
Molecular Dynamics

Analysis of MD trajectory (Part 1)

P. Satpati

Why Temperature changes of MD simulation?

Total Energy (U) = Kinetic Energy + Potential Energy = CONSTANT **Positive**



https://en.wikibooks.org/wiki/Molecul ar_Simulation/Molecular_Dynamics

1. At the start of simulation

Maxwell-Boltzmann distribution Temperature is fixed (say, 300K) Kinetic energy is fixed.

Usually negative

If the initial structure is bad => Starting Potential Energy (Less negative)

2. During simulation Structure will be better => Potential Energy is more negative Energy conservation => Kinetic energy should go up → Temperature will increase

- >THERMOSTAT fix "Temperature.
- > But if you use thermostat energy not longer is conserved.

MD trajectory

Time-step =
$$\Delta t = 1 \text{ fs} = 10^{-15} \text{ sec}$$

Total Simulation time/ Trajectory = $1 ns = 10^{-9} sec$

If you want to save frame/snapshot after every time-step = 10^6 fames (1 ns trajectory)

Every Fame = position and velocities of 'N' particles of the system

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Every particle = 3 positions (x, y, z) + 3 velocities (v_x, v_y, y_z)

Single frame = 3N positions + 3N velocities = 6N information's

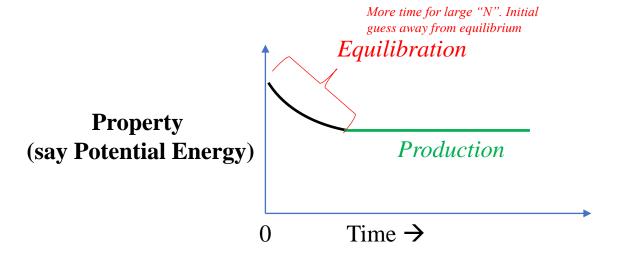
1 ns Trajectory (frames at every 1fs) = 10^6 x 6N information's

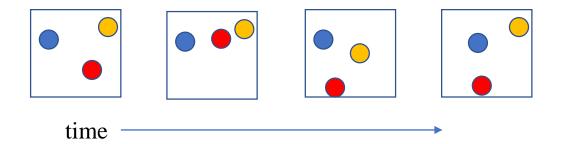
= 10^6 x 6 x 10^6 information's

= 6 x 10^{12} information's
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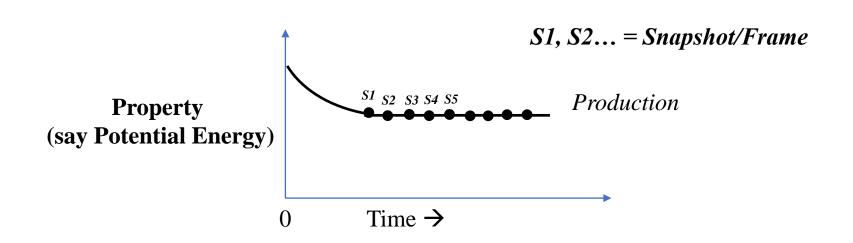
Trajectory file (Too much information) -> Storage Issue + Post-processing issue

Monitor the MD trajectory



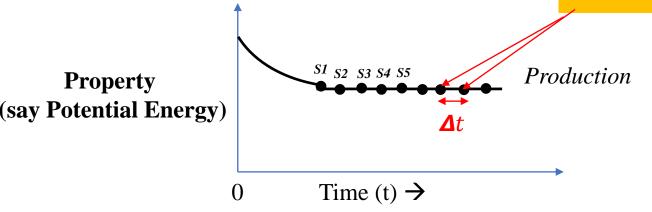


AVERAGE PROPERTIES
WILL BE CALCULATED
FROM THE SNAPS (S1, S2...)



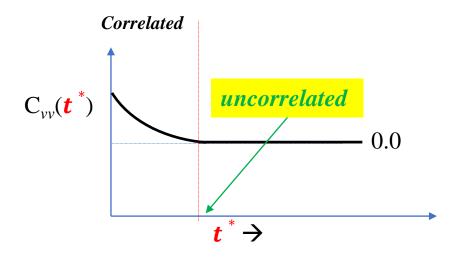
Do we need all the Snaps?

- . We need uncorrelated Snaps($\Delta t = t^{\uparrow}$?)
- 2. Simulation Time $\gg \Delta t$



How to get uncorrelated snaps (t^*) ?

Answer: Autocorrelation function



Velocity

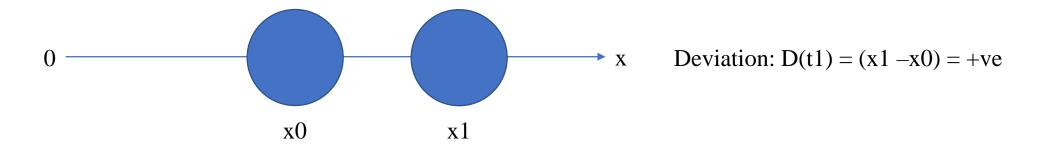
$$C_{vv}(t^*) = \langle v(t+t^*), v(t) \rangle = \langle \frac{1}{N} \sum_{i=1}^{N} v_i(t+t^*), v(t) \rangle$$

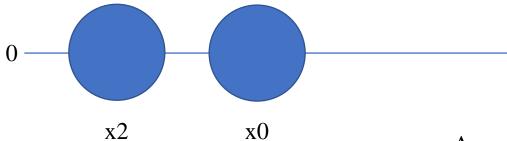
autocorrelation

function

Use uncorrelated snaps for estimating average properties (Biomolecular simulations typically: $t^* > 1$ ps / 10^{-12} sec)

Compare two structures.





Deviation: D(t2) = (x2 - x0) = -ve

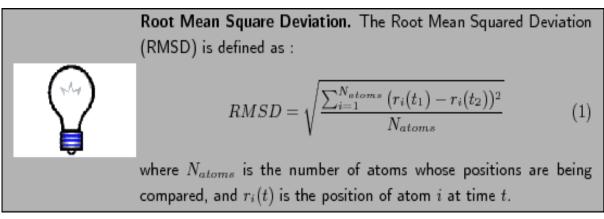
Average deviation= [D(t1)+D(t2)+...]/total no of snaps = 0.0

How to compare estimate the difference?

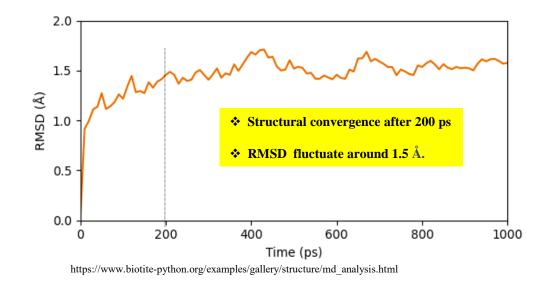
Deviation: $D^*(t1) = \sqrt{(x1-x0)2} = Always positive number$

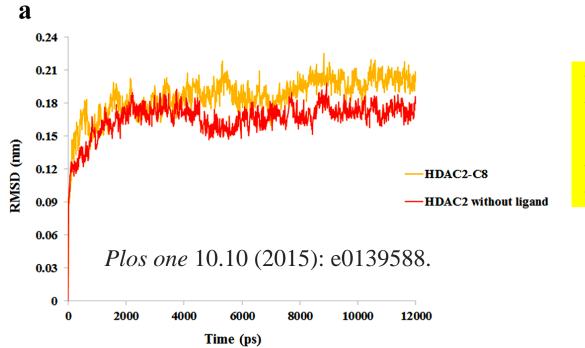
Average deviation for N-number of particles:
$$RMSD = \sqrt{\frac{1}{N}\sum_{i=1}^{N}(x_i(t) - x_i(0))^2}$$

Root Mean Square deviation (RMSD)



https://www.ks.uiuc.edu/Training/Tutorials/vmd/tutorial-html/node5.html



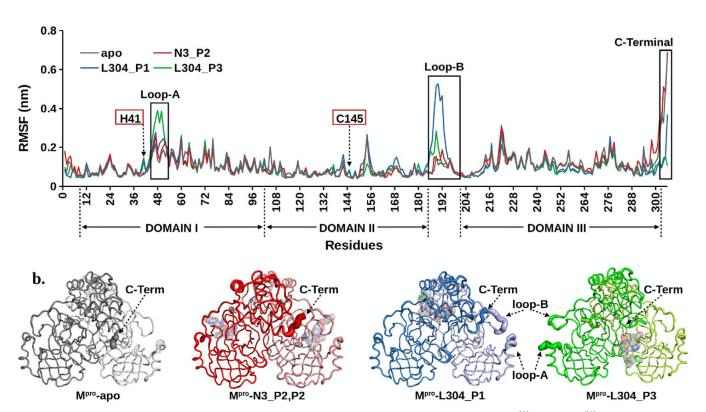


- Measure of structural Similarity between two structures
- Indicate structural convergence

Root Mean Square fluctuation (RMSF)

$$RMSF = \sqrt{\frac{1}{T}\sum_{t_j=1}^{T} (r_i(tj) - \widetilde{r}_i)^2}$$

 $\widetilde{r_i}$ = Average position of particle "i" from trajectory 'T' $r_i(t_j)$ = position of particle "i" at time/snap ' t_j ' T = Trajectory/snapshots



- Porcupine plot
- > Flexibility as a function of residue number
- ➤ Flexibility = RMSF = averaged over MD snaps

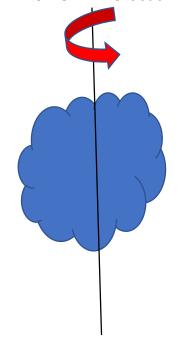
Measure of "FLEXIBILITY" at different region of the protein during a MD trajectory segment

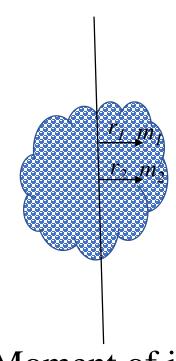
Figure 6. Residue-wise RMSF (averaged from the last 50 ns of the 150 ns trajectory) of protein-heavy atoms. (a) M^{pro}-apo and M^{pro}-holo. The C-terminal region, Loop-A and Loop-B are shown in the rectangular box. H41, C154 (catalytic dyad) is highlighted in the red box. (b) RMSF-based putty representations of M^{pro} protein is shown for visual analysis of protein flexibility. N3 and L304 are represented with van der Waals spheres.

Ghosh S. et al. Journal of Biomolecular Structure & Dynamics. 2021 Aug:1-12. DOI: 10.1080/07391102.2021.1967786. PMID: 34424151.

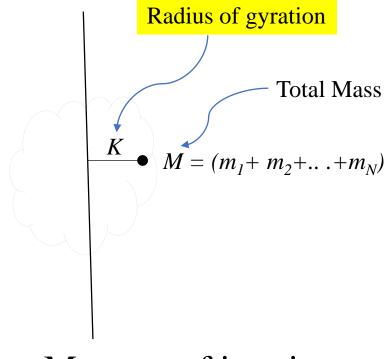
Radius of gyration

Axis of rotation





Moment of inertia
$$= \sum_{i=1}^{N} m_i r_i^2$$



Moment of inertia
$$= MK^2$$

$$MK^2 = \sum_{i=1}^{N} m_i r_i^2$$

Radius of gyration (From MD simulation)

A radius of gyration in general is the distance from the center of mass of a body at which the whole mass could be concentrated without changing

Radius of gyration as an indicator of protein structure compactness

•Physical Chemistry Research, 2016. 5(2):205-219 DOI: 10.22036/pcr.2017.40480

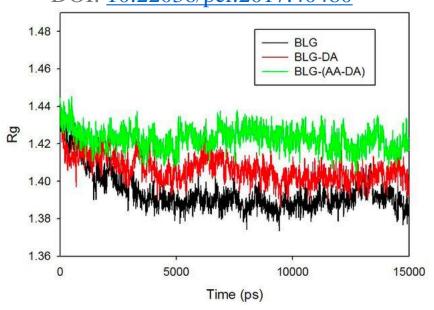
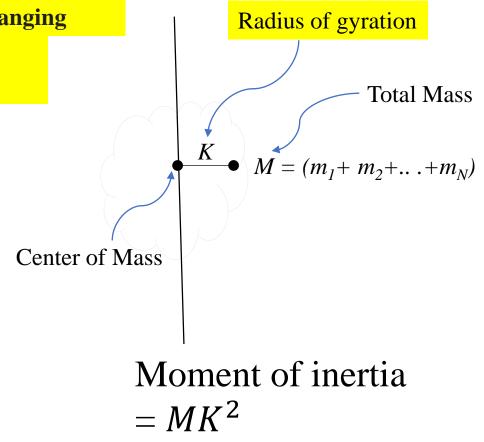


Fig. 4. RMSF of BLG residues with respect to their time-averaged positions for free and bound BLG. RMSF reductions in regions that directly in contact with ligands are shown with arrows.



Molecular BioSystems 11.3 (2015): 958-968.

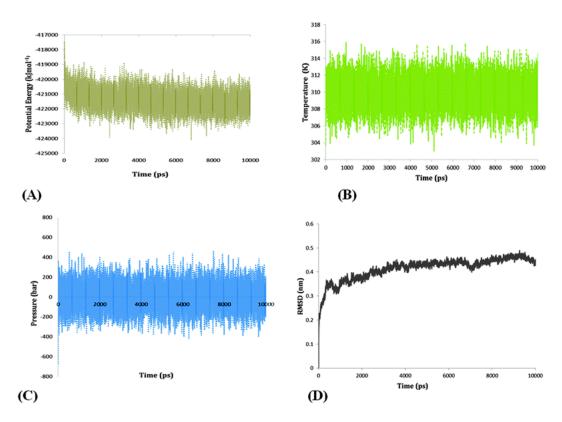


Fig. 5 Molecular dynamics simulation (A) potential energy (B) temperature 310 K (C) pressure of 1 bar (D) RMSD of the protein with respect to time and the protein structure.

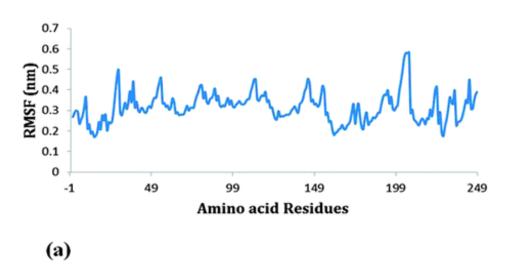


Fig. 6 (a) Root mean square fluctuation



Next: Analysis of MD trajectory (Part 2)