

MiSeq Wet Lab SOP

Detailed Title:	16S rRNA Sequencing with the Illumina MiSeq: Library Generation, QC, & Sequencing
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1.0) Introduction and Workflow

1.1) *Introduction*

- The Illumina MiSeq Personal Sequencer can produce 2 x 250 paired-end reads and up to 8.5 Gb of data in a single run. Dual indexing of library samples allows up to 384 samples to be run simultaneously. The instrument is capable of producing in excess of 24 million reads, however, for low diversity runs about 12 million reads can be expected. A wide range of applications is possible including 16S analysis, metagenomics, genome sequencing, transcriptomics, and RNA sequencing.
- There are several steps in preparing samples for sequencing on the MiSeq. Broadly, these include library generation and indexing, quality control, normalization and pooling, quantification, sequencing, run quality assessment, and data export.

1.2) *16S Prep Workflow*

1. Extracted DNA should be arrayed in 96 well plate format. If possible, leave two wells on each plate open for controls.
2. PCR of 16S samples with Schloss lab indices. Each plate will contain a negative control (water) and a positive control (mock community)
3. A subset of 12-24 samples from each plate will undergo electrophoresis on a 1% agarose gel to ensure amplification proceeded normally.
4. Library clean up and normalization performed using the Invitrogen [SequalPrep](#) Plate Normalization Kit.
5. Samples from each plate are pooled into single wells (i.e. 1 well/plate, 4 wells total for a run with 4 plates).
6. (Optional) To assess the quality of the library, the investigator may choose to perform a Bioanalyzer trace using the [Agilent Technologies HS DNAkit](#) cat# 5067-4626.
7. (Optional) If the post-PCR gel or the Bioanalyzer trace suggests contaminant DNA from leftover indices/primer-dimer, an additional gel purification of the pooled plates is recommended. This often improves the quality of the sequencing run.
8. Each pooled plate is quantified using a KAPA Biosystems [Q-PCR kit](#) cat# KK4824.
9. Plates are pooled to equal concentration into a single well (i.e. 1 well per run)
10. The pooled library enters the Sequencing Workflow.

1.3) Sequencing Workflow

1. A Sample Plate is created for each plate using Illumina Experiment Manager. Sample Plates are then used to create a Sample Sheet. This sheet serves as the set of run parameters and indexing scheme used by the MiSeq for the run. The sample sheet is then transferred to the MiSeq via flash drive.
2. The reagent cartridge is thawed in a water bath per the [MiSeq User Guide](#).
3. Unless otherwise specified, dilution and loading will follow the steps outlined in the document: Preparing DNA Libraries for Sequencing on the MiSeq([15039740 C](#))
4. Any custom primers required (including those for 16S) will be mixed with the native primers on the reagent cartridge.
5. The technician will load the sample sheet, flow cell, reagent cartridge, PR2 bottle, and an empty waste bottle onto the MiSeq and start the run. A 500 cycle run takes approx. 44 hours.
6. The technician will monitor the run using Illumina Sequence Analysis Viewer.
7. The data will be stored on the Schloss Lab NAS drive and made available to the investigator following run completion. Unless otherwise requested, default output will be fastq files.
8. Technician will perform a post run wash and any required maintenance of instrument.

2.0) Safety and Waste Disposal

- The Schloss Lab Chemical Hygiene Plan should be followed at all times.
- Standard PPE (nitrile gloves, safety glasses, and lab coat) should be used at all times.
- Each reagent cartridge contains a small amount of formamide and must be disposed of in an appropriate container following the run. Liquid waste from a run must also be disposed of as hazardous due to the formamide content.
- All other safety concerns may be addressed to the lab chemical safety officer.

3.0) Consumables

Reagent/Kit	Catalog #	Price
MiSeq® Reagent Kit v2 (500 cycle)	MS-102-2003	\$910.80
PhiX Control Kit v3	FC-110-3001	\$132.00
16s Index Primers (40 total)	IDT	\$1180.00
16s Sequencing Primers (Read 1, Index, Read 2)	IDT	\$41.71
AccuPrime™ Pfx SuperMix (200rxns)	12344-040	\$331.28
Library Quantification Kit - Illumina/Universal	KK4824	\$575.00
SequalPrep™ Normalization Plate (96) Kit	A10510-01	\$453.60
(optional) Agilent High Sensitivity DNA Kit	5067-4626	\$434.70
(optional) QIAquick Gel Extraction Kit (50 rxns)	28704	\$111.00
epT.I.P.S. Motion 1-50 uL Reloads 24 racks of 96 tips	30014421	\$190.00
Tip One Refill Wafers 200uL elongated graduated	NC9549602	\$39.24
Tip One Refill Wafers 1000uL graduated	1111-2721	\$25.25
Fisher 1N NaOH 1L	SS266-1	\$46.32
TWIN.TEC 96 Well Plate Skirted Blue	E951020460	\$99.23

Reagent/Kit	Catalog #	Price
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4.0) Run Costs

For 384 sample run	PCR and Indexing	Cleanup Pooling & Normalization	Library QC	Sequencing	Totals	Total Cost
16S Reagents	\$588	\$265	\$138	\$941	\$1932	\$2,232
16S Man hours	4	3	4	4	15	

5.0) Detailed Method(s)

5.1) Published Protocols

- The following methods and references are used in the workflows above.
 - [MiSeq User Guide](#) Rev. L
 - Preparing DNA Libraries for Sequencing on the MiSeq([15039740 C](#))
 - Kapa Biosystems Q-PCR Library [Quantification Kit](#) Illumina
 - Accuprime Pfx [Super Mix](#)
 - Agilent High Sensitivity DNA Kit Guide
 - SequalPrep Normalization Plate (96) Kit

5.2) Initial Set up

1. Reconstitute indexed primers and sequencing primers to 100 uM. See Appendix D for primer design.
2. Prepare 100ul 10 uM aliquots of indexed primers. Do not dilute sequencing primers.
3. Array aliquots of indexed primers into four 96 well plates using the following scheme:
 - a. A701 – A712 with A501 – A508
 - b. A701 – A712 with B501 – B508
 - c. B701 – B712 with B501 – B508
 - d. B701 – B712 with A501 – A508

Note: These primer plates can be stored at -20C and used for subsequent runs.

4. Extract template DNA and array in 96 well format leaving two wells open. (One for a negative water control and another for the positive Mock Community control)

5.3) PCR

Note: These steps may be performed using an epMotion or similar automated pipetting system.

1. Dispense 17 ul of Accuprime Pfx Supermix into each well of a new 96 well plate.

2. Using a multichannel pipette, transfer 1 ul of template DNA per well to the corresponding well on the PCR plate.
3. Using a multichannel pipette, transfer 2 ul of each paired set of index primers from the primer plate to the corresponding well on the PCR plate. Be sure to follow the layout chosen in the sample sheet.
4. Add 1 ul of PCR grade dH₂O to the negative control well, and 1 ul of Mock Community at a 1:3 dilution to the positive control well.
5. Repeat for up to four 96 well plates. Seal plates, vortex briefly and spin down contents.
6. Place in thermocycler.

Use the following program:

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95C 2:00
——— 30 cycles ———
95C 00:20
55C 00:15
72C 5:00
—————
72C 10:00
4C Hold

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Gel Electrophoresis

1. A random row of 12 should be selected from each PCR plate and run on a gel to confirm success of the PCR.
2. Use 2 ul of sample, 4 ul of loading dye in a 1% agarose gel.
3. Run at 100v for 30 minutes alongside a standard ladder.
4. Photograph gel under UV. Check to be sure there is a band for every well.

Cleanup, Normalization, and Pooling

Use the SequalPrep Normalization Plate Kit

1. Transfer 18 ul of PCR product from PCR plate to corresponding well on the normalization plate.
2. Add 18 ul of Binding Buffer. Mix by pipetting, sealing, vortexing, and spinning briefly.
3. Incubate at room temperature for 60 minutes. Note: can incubate overnight if needed. Extra time does not improve results.
4. Aspirate the liquid from the wells. Do not scrape the sides.
5. Add 50 ul of Wash Buffer and pipette up and down twice, then aspirate immediately. Ensure there is no residual wash buffer in any wells.
6. Add 20 ul of Elution Buffer. Mix by pipetting up and down 5 times. Seal, vortex, and spin briefly.
7. Incubate at room temperature for 5 minutes.
8. Create a pool from each plate. Take 5 ul of each well to pool. The use of an empty 96 well plate may facilitate the use of multichannel pipettes.

9. Freeze the remaining sample for later use.

Library QC

1. Prepare the following dilutions of each pooled library in PCR grade H₂O:
 - a. 1:1
 - b. 1:10
 - c. 1:1000 (dilute in several steps for better results)
 - d. 1:2000
 - e. 1:4000
2. Agilent Bioanalyzer Trace
 - a. Prepare Gel-Dye mix if not already prepared.
 - b. Let reagents equilibrate to room temperature.
 - c. Turn Bioanalyzer on and load 350 ul of dH₂O onto electrode cleanser and place in analyzer for 5 minutes.
 - d. Open a high sensitivity chip and place on the priming station. Base plate should be a position “C” and syringe clip should be at lowest position.
 - e. Load 9.0 ul of gel-dye mix to position 12 marked with a large “G”. Ensure the syringe plunger is at 1.0 ml and close the station. Press plunger until it is held by clip.
 - f. Wait for exactly 60 seconds then release the plunger clip. Wait an additional 5 seconds, then slowly pull the plunger back to the 1.0 ml position.
 - g. Open the priming station. Pipette 9.0 ul of gel-dye mix into the other wells marked “G” in positions 4,8, and 16.
 - h. Pipette 5.0 ul of marker to all wells excluding the right column. (No marker positions 4,8,12, and 16)
 - i. Load 1 ul of ladder into position 15 marked by the ladder symbol.
 - j. Pipette 1 ul of each of dilutions a – b above. Top row Plate 1 Pool 1:1 x 1, 1:10 x 2. Second row Plate 2 pool 1:1 x 1, 1:10 x 2. Third row Plate 3 Pool 1:1 x 1, 1:10 x 2. Bottom row Plate 4 Pool 1:1 x 1, 1:10 x 2.
 - k. Place chip in the designated vortex for 1 minute, then transfer chip to the Bioanalyzer.
 - l. Open the 2100 Expert software and select the HS DNA Assay. Enter sample names/dilutions for each of the test wells. Click Start.
 - m. Print .pdf when run finishes.
3. Kapa Q-PCR Library Quantification
 - a. Before Q-PCR reaction setup, add 1 ml Primer Premix (10X) to the 5 ml bottle of KAPA SYBR® FAST Q-PCR Master Mix (2X) and mix by vortexing for 10 sec. Record the date of Primer Premix addition on the KAPA SYBR® FAST Q-PCR Master Mix bottle.
 - b. Reaction can be either 10 ul or 20 ul. A 10 ul reaction volume is recommended.
 - c. Prepare a 96 well Q-PCR plate compatible with the real time thermocycler. There are six standards. Each should be run in triplicate. Each pool at each dilution should be run in triplicate.
 - d. For 10 ul reaction volume dispense 6 ul of master mix into each well needed.
 - e. Pipette 4 ul of standards and library dilutions into appropriate wells. Mix by pipetting. Vortex and spin optional.

- f. Place plate in thermocycler. Start control software
 - g. Program the following cycle
 - i. Initial Activation 95 °C 5 minutes
 - ii. 35 cycles
 - 1. Denaturation 95 °C 30 seconds
 - 2. Annealing 60 °C 45 seconds
 - 3. If library fragment size exceeds 700bp, extend annealing step to 90 seconds.
 - iii. Perform melt curve to check for primer/adaptor dimer
 - h. Assign wells and group replicates.
 - i. Enter values for standards
 - i. Std. 1 20pM
 - ii. Std. 2 2pM
 - iii. Std. 3 0.2pM
 - iv. Std. 4 0.02pM
 - v. Std. 5 0.002pM
 - vi. Std. 6 0.0002pM
 - vii. Note: The concentrations provided here are for the DNA Standards as supplied in the kit, and are NOT the concentrations in the reactions. Provided that the volume of template added to each reaction is the same for Standards and for library samples (i.e. 4 ul in each case), there is no need to account for these volumes when calculating the concentrations of library samples, nor should one need to calculate the concentration of template in the reaction.
 - j. Run program
 - k. To calculate library concentration use the following formula:
 - i. Average x (452/Avg fragment length from bioanalyzer) x dilution factor
 - ii. Use the average of the triplicate data points corresponding to the most concentrated library DNA dilution that falls within the dynamic range of the DNA Standards to calculate the concentration of the undiluted library.
 - iii. Do not include outliers in calculation. If there is more than one outlier in a group, the assay must be repeated.
4. Create normalized pools from each plate by diluting to the concentration of the least concentrated plate. Create a single final pool by adding equal amounts of each post qpcr normalized pool. Final pool must be >10ul in total volume. 40-80ul is ideal.

Sequencing

1. Remove a 500 cycle reagent cartridge from the -20 °C freezer. Place in room temperature water bath for one hour. Place HT1 buffer tube in 4 °C fridge.
2. Copy sample sheet to sample sheet folder on MiSeq. Rename sample sheet to match barcode of reagent cartridge.
3. When the reagent cartridge has thawed, dry bottom with paper towel. Invert the cartridge repeatedly to check each well is thawed. This also serves to mix the reagents. Place in hood.
4. Thaw library, PhiX, and sequencing primers. Check to make sure HT1 is thawed.
5. Place 3 ul of the 100 uM Read 1 Sequencing Primer(s) into a clean PCR tube. Repeat in separate tubes for the Index Primer(s) and Read 2 Sequencing Primer(s).

6. Using a 1000 ul pipette tip, break the foil over wells 12, 13, 14, and 17.
7. Use an extra long 100 ul tip with the pipettor set on 75 ul to transfer the 3 ul of Read 1 Sequencing Primer to the bottom of well 12 and pipette 10X to mix. Repeat this process spiking the Index Primer into well 13 and the Read 2 Sequencing Primer into well 14.
8. Prepare a fresh dilution of 0.2N NaOH.
9. To a 1.5ml tube add 10 ul of library, and 10 ul of 0.2N NaOH. To a separate tube add 2 ul PhiX, 3 ul PCR grade water, and 5 ul of 0.2N NaOH. Vortex both tubes to mix and spin for 1 minute at 400rcf. Note: NaOH concentration on the flow cell must remain under 0.001N. Adjusting the concentration of the NaOH used to denature the DNA to 0.1N may be necessary if library concentration is 1nM or below.¹
10. Allow the tubes to incubate at room temperature for 5 minutes. Immediately add 980 ul of HT1 to the library tube, and 990 ul HT1 to the PhiX tube.
11. Use HT1 to dilute both the library and PhiX to 4pM for a v2 kit. Can load up to 8pM for a v3 kit. For a 10% PhiX run, combine 900 ul of 4.0pM Library and 100 ul PhiX in a final tube. Vortex. Load 600 ul of this solution into well 17 on the reagent cartridge. See example below:
 - a. $(1.45 \text{ nM library} \times 10 \text{ ul}) + (0.2\text{N NaOH} \times 10 \text{ ul}) + 980 \text{ ul HT1} = 14.5\text{pM Lib, } 0.002\text{N NaOH}$
 - b. $(14.5\text{pM lib} \times 275.86 \text{ ul}) + 724.14 \text{ ul HT1} = 4.0\text{pM lib, } 0.00055\text{N NaOH}$
 - c. $[(10\text{nM PhiX} \times 2 \text{ ul}) + 3 \text{ ul H}_2\text{O}] + (0.2\text{N NaOH} \times 5 \text{ ul}) + 990 \text{ ul HT1} = 20\text{pM PhiX, } 0.001\text{N NaOH}$
 - d. $(20\text{pM PhiX} \times 200 \text{ ul}) + 800 \text{ ul HT1} = 4.0\text{pM PhiX, } 0.0002\text{N NaOH}$
 - e. $(4.0\text{pM Lib} \times 900 \text{ ul}) + (4.0\text{pM PhiX} \times 100 \text{ ul}) = \text{solution loaded}$
 - f. Solution loaded is 4.0pM overall with a 3.6pM Library concentration, 0.4pM PhiX concentration, and 0.000515N NaOH
13. Set reagent cartridge aside. Unbox flow cell and PR2 bottle.
14. Thoroughly rinse the flow cell with Milli-Q water. Carefully dry by blotting with lint free wipes (Kimwipes). Give special attention to the edges and points of intersection between the glass and plastic.
15. Wet a new wipe with 100% alcohol and wipe the glass on both sides avoiding the rubber intake ports.
16. Visually inspect the flow cell to ensure there are no blemishes, particles, or fibers on the glass.
17. Follow on screen instructions and load the flow cell, reagent cartridge, and PR2 bottle. Empty and replace the waste bottle.
18. Ensure the machine recognizes the correct sample sheet and the run parameters are correct.
19. Wait for the MiSeq to perform its pre-run checks, and press start.

Run Monitoring

1. The run should be monitored periodically using Illumina Sequence Analysis Viewer.
2. Ideal parameters for a 90% 16S run:

¹For extremely low concentration libraries we have adapted a method published by Quail et al. <http://www.nature.com/nmeth/journal/v5/n12/full/nmeth.1270.html>

Libraries as low as 0.1nM do not allow for sufficient dilution to reduce NaOH to 0.001N. It must be neutralized using 200mM Tris pH 7.0. Example: 40 ul 0.1nM library + 40 ul 0.1N NaOH. Incubate for 5 min. Add 80 ul 200mM Tris. Then add 840 ul HT1. This results in a 4.0pM library.

- a. Cluster density 700-800k/mm² for v2 kits
- b. Cluster density 1000-1100k/mm² for v3 kits
- c. >85% clusters passing filter
- d. 10% aligned (amount of PhiX)
- e. No spikes in corrected intensity plot
- f. All indices identified following index reads
- g. Final >Q30 score of >70%

Final Steps

1. Perform post run wash.
2. Dispose of liquid waste in appropriate hazardous jug and reagent cartridge in hazardous bucket.
3. When MiSeq Reporter finishes, copy the fastq files from the analysis folder to the run folder on the NAS drive.
4. Perform maintenance wash if required.
5. Confirm with investigator that data are of sufficient quality and quantity.

Appendix A: Adding An Assay To Illumina Experiment Manager

5. Using Illumina Experiment Manager, create a sample plate for each 96 well plate of template. Choose indexes that correspond to one of the four index pair plates above. See Appendix C for instruction on creating a custom assay in IEM.
6. Using Illumina Experiment Manager, create a sample sheet for the run. Ensure that index choices are compatible with one another and there is sufficient diversity in the index reads so as to activate both light channels every cycle. Note: Primer plate A has insufficient diversity to be run alone. If sequencing 96 or fewer samples, choose plate B, C, or D.
7. The MiSeq requires base diversity on every cycle. 16S is a low diversity library. With MiSeq software v2.2, 16S libraries can be loaded with 5% PhiX. Additionally, other high diversity samples such as metagenomes can be run simultaneously. This requires manually editing the sample sheet. Older software versions required “hardcoding” the matrix and phasing/pre-phasing values. See Appendix B.

Introduction

Illumina Experiment Manager is used to generate sample plates and sheets. A new assay must be added to the system to efficiently prepare sample sheets for 16S sequencing. Not only does this eliminate the need to manually assemble a sample sheet for 16S runs, but allows the user to retain use of the IEM index analysis feature. This ensures the indices selected for a particular run have sufficient diversity on a cycle-by-cycle basis and will successfully demultiplex.

Procedure

1. Open C:\program files\Illumina\Illumina Experiment Manager\Sample Prep Kits
2. Copy the file Nextera.txt, and rename the file Schloss.txt
3. Open in a text editor
4. Under [Name] change to Schloss

5. Under [PlateExtension] change to Schloss
6. Under [I7], clear the Nextera indices and paste in the SA701 through SB712 index names, and the REVERSE COMPLIMENT of the primer index sequences.
7. Under [I5], remove the Nextera indices and paste in the SA501 through SB512 index names and index sequences with no alteration (NOT the reverse compliment).
8. Under [DefaultLayout_SingleIndex] and [DefaultLayout_DualIndex] each well of a 96 well plate is listed with a corresponding Nextera index name. These must be replaced with Schloss index names. It is recommended that a text editor with column select capability be used to leave the index name number unchanged (i.e. 701, 702, etc.), while replacing the character "N" for Nextera with "SA" for Schloss A.
9. Save and close.
10. Return to the IEM folder and open the Applications folder. Open and edit the file GenerateFASTQ.txt.
11. Add a line at to the bottom with the text "Schloss"

Appendix B: Primer design

Overall design considerations

- The sequencing primers must have a melting temperature near 65°C. This can be achieved by altering the pad sequence
- The index sequences must balance the number of bases at each position. The index sequences listed here have a 25% ATGC composition at each site. If you are going to cherry pick indices from the list, make sure that you have even representation.

Generic PCR primer design:

AATGATACGGCGACCAACCGAGATCTACAC <i5><pad><link><16Sf> VX.N5??

CAAGCAGAAGACGGCATAACGAGAT <i7><pad><link><16Sr> VX.N7??

Generic read 1 primer design

<pad><link><16Sf> VX.read1

Generic read 2 primer design

<pad><link><16Sr> VX.read2

Generic index read primer design

Reverse complement of (<pad><link><16Sr>) VX.p7_index

The listed sequences in the generic design, above, are the adapter sequences to allow annealing of the amplicons to the flow cell. The i5 and i7 sequences are the 8-nt index sequences. The pad is a 10-nt sequence to boost the sequencing primer melting temperatures. The link is a 2-nt sequence that is anti-complementary to the known sequences. The 16Sf and 16Sr are the gene specific primer sequences. Primers are purchased from IDT with no special purification. This system should work for any other region of the 16S rRNA gene or any other gene. The only thing to change would be the 16Sf/16Sr sequences and confirm that when combined with the pad sequence that the melting temperature is near 65°C.

16Sf

V3: CCTACGGGAGGCAGCAG

V4: GTGCCAGCMGCCGCGGTAA

16Sr

V4: GGAATCHVGGGTWTCTAAT

V5: CCCGTCAATTCMTTTRAGT

Link:

V4f: GT

V4r: CC

V3f: GG

V5r: GG

Pad:

Forward: TATGGTAATT

Reverse: AGTCAGTCAG

i5

SA501 ATCGTACG

SA502 ACTATCTG

SA503 TAGCGAGT

SA504 CTGCGTGT

SA505 TCATCGAG

SA506 CGTGAGTG

SA507 GGATATCT

SA508 GACACCGT

SB501 CTACTATA

SB502 CGTTACTA

SB503 AGAGTCAC

SB504 TACGAGAC

SB505 ACGTCTCG

SB506 TCGACGAG

SB507 GATCGTGT

SB508 GTCAGATA

SC501 ACGACGTG

SC502 ATATACAC

SC503 CGTCGCTA

SC504 CTAGAGCT

SC505 GCTCTAGT

SC506 GCACTGA

SC507 TGCGTACG

SC508 TAGTGTAG
SD501 AAGCAGCA
SD502 ACGCGTGA
SD503 CGATCTAC
SD504 TGCCTCAC
SD505 GTCTAGTG
SD506 CTAGTATG
SD507 GATAGCGT
SD508 TCTACACT

i7

SA701 AACTCTCG
SA702 ACTATGTC
SA703 AGTAGCGT
SA704 CAGTGAGT
SA705 CGTACTCA
SA706 CTACGCAG
SA707 GGAGACTA
SA708 GTCGCTCG
SA709 GTCGTAGT
SA710 TAGCAGAC
SA711 TCATAGAC
SA712 TCGCTATA
SB701 AAGTCGAG
SB702 ATACTTCG
SB703 AGCTGCTA
SB704 CATAGAGA
SB705 CGTAGATC
SB706 CTCGTTAC
SB707 GCGCACGT
SB708 GGTACTAT
SB709 GTATACGC
SB710 TACGAGCA
SB711 TCAGCGTT
SB712 TCGCTACG
SC701 ACCTACTG
SC702 AGCGCTAT

SC703 AGTCTAGA
SC704 CATGAGGA
SC705 CTAGCTCG
SC706 CTCTAGAG
SC707 GAGCTCAT
SC708 GGTATGCT
SC709 GTATGACG
SC710 TAGACTGA
SC711 TCACGATG
SC712 TCGAGCTC
SD701 ACCTAGTA
SD702 ACGTACGT
SD703 ATATCGCG
SD704 CACGATAG
SD705 CGTATCGC
SD706 CTGCGACT
SD707 GCTGTAAC
SD708 GGACGTTA
SD709 GGTCGTAG
SD710 TAAGTCTC
SD711 TACACAGT
SD712 TTGACGCA

Primers used to amplify 384 samples using the V4 region:

v4.SA501 AATGATACGGCGACCACCGAGATCTACACATCGTACGTATGGTAATTGTGTGCCAGCMGC-
CGCGGTAA

v4.SA502 AATGATACGGCGACCACCGAGATCTACACACTATCTGTATGGTAATTGTGTGCCAGCMGC-
CGCGGTAA

v4.SA503 AATGATACGGCGACCACCGAGATCTACACTAGCGAGTTATGGTAATTGTGTGCCAGCMGC-
CGCGGTAA

v4.SA504 AATGATACGGCGACCACCGAGATCTACACCTGCGTGTTATGGTAATTGTGTGCCAGCMGC-
CGCGGTAA

v4.SA505 AATGATACGGCGACCACCGAGATCTACACTCATCGAGTATGGTAATTGTGTGCCAGCMGC-
CGCGGTAA

v4.SA506 AATGATACGGCGACCACCGAGATCTACACCGTGAGTGTATGGTAATTGTGTGCCAGCMGC-
CGCGGTAA

v4.SA507 AATGATACGGCGACCACCGAGATCTACACGGATATCTTATGGTAATTGTGTGCCAGCMGC-
CGCGGTAA

v4.SA508 AATGATACGGCGACCACCGAGATCTACACGACACCGTTATGGTAATTGTGTGCCAGCMGC-
CGCGGTAA

v4.SB501 AATGATACGGCGACCACCGAGATCTACACCTACTATATATGGTAATTGTGTGCCAGCMGC-
CGCGGTAA

v4.SB502 AATGATACGGCGACCACCGAGATCTACACCGTTACTATATGGTAATTGTGTGCCAGCMGC-
CGCGGTAA

v4.SB503 AATGATACGGCGACCACCGAGATCTACACAGAGTCACTATGGTAATTGTGTGCCAGCMGC-
CGCGGTAA

v4.SB504 AATGATACGGCGACCACCGAGATCTACACTACGAGACTATGGTAATTGTGTGCCAGCMGC-
CGCGGTAA

v4.SB505 AATGATACGGCGACCACCGAGATCTACACACGTCTCGTATGGTAATTGTGTGCCAGCMGC-
CGCGGTAA

v4.SB506 AATGATACGGCGACCACCGAGATCTACACTCGACGAGTATGGTAATTGTGTGCCAGCMGC-
CGCGGTAA

v4.SB507 AATGATACGGCGACCACCGAGATCTACACGATCGTGTTATGGTAATTGTGTGCCAGCMGC-
CGCGGTAA

v4.SB508 AATGATACGGCGACCACCGAGATCTACACGTCAGATATATGGTAATTGTGTGCCAGCMGC-
CGCGGTAA

v4.SA701 CAAGCAGAAGACGGCATAACGAGATAACTCTCGAGTCAGTCAGCCGGACTACHVGGGTWTC-
TAAT

v4.SA702 CAAGCAGAAGACGGCATAACGAGATACTATGTCAGTCAGTCAGCCGGACTACHVGGGTWTC-
TAAT

v4.SA703 CAAGCAGAAGACGGCATAACGAGATAGTAGCGTAGTCAGTCAGCCGGACTACHVGGGTWTC-
TAAT

v4.SA704 CAAGCAGAAGACGGCATAACGAGATCAGTGAGTAGTCAGTCAGCCGGACTACHVGGGTWTC-
TAAT

v4.SA705 CAAGCAGAAGACGGCATAACGAGATCGTACTCAAGTCAGTCAGCCGGACTACHVGGGTWTC-
TAAT

v4.SA706 CAAGCAGAAGACGGCATAACGAGATCTACGCAGAGTCAGTCAGCCGGACTACHVGGGTWTC-
TAAT

v4.SA707 CAAGCAGAAGACGGCATAACGAGATGGAGACTAAGTCAGTCAGCCGGACTACHVGGGTWTC-
TAAT

v4.SA708 CAAGCAGAAGACGGCATAACGAGATGTCGCTCGAGTCAGTCAGCCGGACTACHVGGGTWTC-
TAAT

v4.SA709 CAAGCAGAAGACGGCATAACGAGATGTCGTAGTAGTCAGTCAGCCGGACTACHVGGGTWTC-
TAAT

v4.SA710 CAAGCAGAAGACGGCATAACGAGATTAGCAGACAGTCAGTCAGCCGGACTACHVGGGTWTC-
TAAT

v4.SA711 CAAGCAGAAGACGGCATAACGAGATTCATAGACAGTCAGTCAGCCGGACTACHVGGGTWTC-
TAAT

v4.SA712 CAAGCAGAAGACGGCATAACGAGATTCGCTATAAGTCAGTCAGCCGGACTACHVGGGTWTC-
TAAT

v4.SB701 CAAGCAGAAGACGGCATAACGAGATAAGTCGAGAGTCAGTCAGCCGGACTACHVGGGTWTC-
TAAT

v4.SB702 CAAGCAGAAGACGGCATAACGAGATATACTTCGAGTCAGTCAGCCGGACTACHVGGGTWTC-
TAAT

v4.SB703 CAAGCAGAAGACGGCATACGAGATAGCTGCTAAGTCAGTCAGCCGGACTACHVGGGTWTC-TAAT

v4.SB704 CAAGCAGAAGACGGCATACGAGATCATAGAGAAGTCAGTCAGCCGGACTACHVGGGTWTC-TAAT

v4.SB705 CAAGCAGAAGACGGCATACGAGATCGTAGATCAGTCAGTCAGCCGGACTACHVGGGTWTC-TAAT

v4.SB706 CAAGCAGAAGACGGCATACGAGATCTCGTTACAGTCAGTCAGCCGGACTACHVGGGTWTC-TAAT

v4.SB707 CAAGCAGAAGACGGCATACGAGATGCGCACGTAGTCAGTCAGCCGGACTACHVGGGTWTC-TAAT

v4.SB708 CAAGCAGAAGACGGCATACGAGATGGTACTATAGTCAGTCAGCCGGACTACHVGGGTWTC-TAAT

v4.SB709 CAAGCAGAAGACGGCATACGAGATGTATACGCAGTCAGTCAGCCGGACTACHVGGGTWTC-TAAT

v4.SB710 CAAGCAGAAGACGGCATACGAGATTACGAGCAAGTCAGTCAGCCGGACTACHVGGGTWTC-TAAT

v4.SB711 CAAGCAGAAGACGGCATACGAGATTCAGCGTTAGTCAGTCAGCCGGACTACHVGGGTWTC-TAAT

v4.SB712 CAAGCAGAAGACGGCATACGAGATTCGCTACGAGTCAGTCAGCCGGACTACHVGGGTWTC-TAAT

Read 1 primer for V4 region

TATGGTAATTGTGTGCCAGCMGCCGCGGTAA

Read 2 primer for V4 region

AGTCAGTCAGCCGGACTACHVGGGTWTC-TAAT

Index primer for V4 region

ATTAGAWACCCBDGTAGTCCGGCTGACTGACT