

Session 4 – Quality assessment and read preprocessing

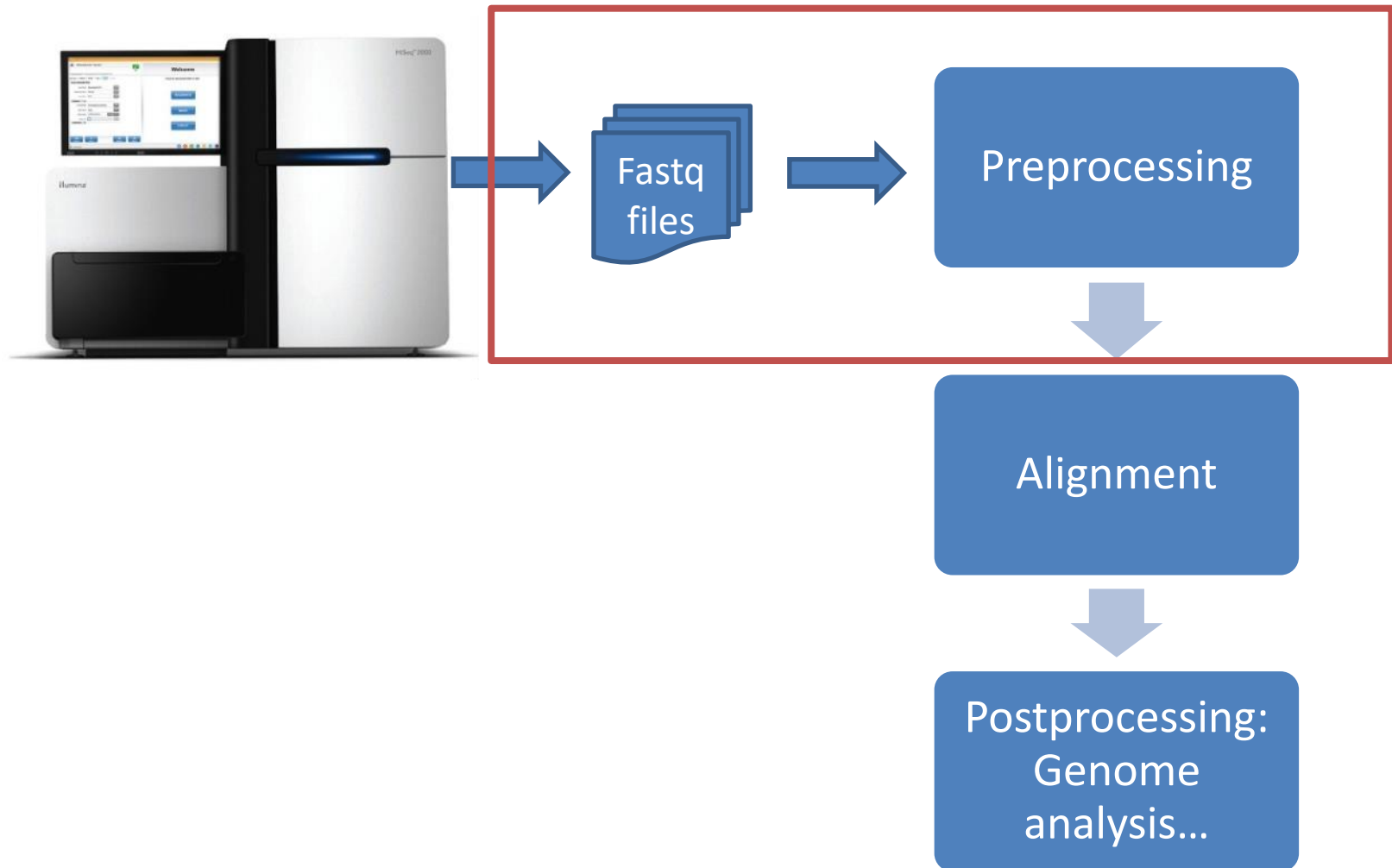
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Programa Formación Continua, ISCIII

Step in the process



Raw output files format

Illumina



.fastq



454 .sff



SOLiD

.fasta
.qual



Nanopore
FAST5



PacBio RSII
Bax.h5
fasta

FASTQ format

- Is a FASTA file with quality information
- Within HTS, FASTA contain genomes y FASTQ reads

>SEQ_ID|

```
AGCTTTTCATTCTGACTGCAACGGGCAATATGTCTCTGTGTGGATTAAAAAAAGAGTGTCTGATAGCAGC  
TTCTGAACTGGTTACCTGCCGTGAGTAAATTTAAATTTTATTGACTTAGGTCACTAAATACTTTAACC  
TATAGGCATAGCGCACAGACAGATAAAAATTACAGAGTACACAACATCCATGAAACGCATTAGCACCACC  
ATTACCACCACCATTACCATTACCACAGGTAACGGTGC GGGCTGACGCGTACAGGAAACACAGAAAAAAG
```

Sequence

@SEQ_ID

```
GATTTGGGGTTCAAAGCAGTATCGATCAAATAGTAAATCCATTTGTTCAACTCACAGTTT
```

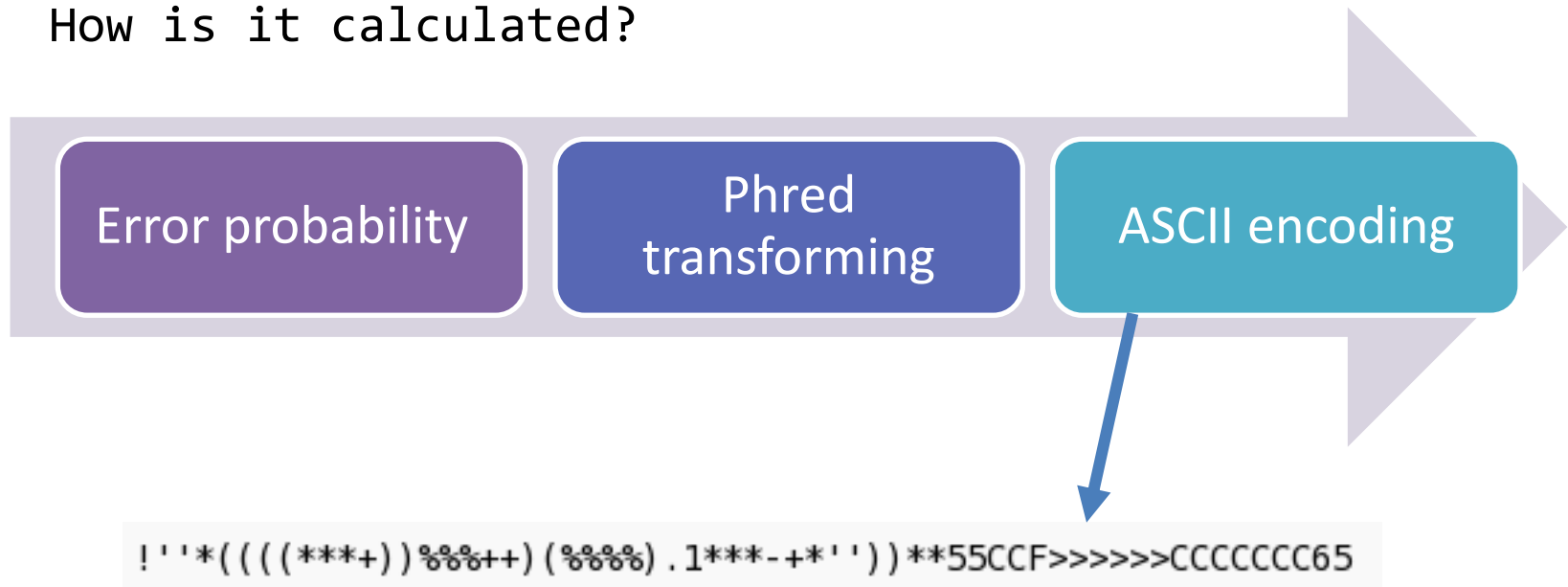
+

```
!''*(((((***+))%%%+))(%%%).1***-+*'))**55CCF>>>>>CCCCCCC65
```

Quality: must be 1 bit

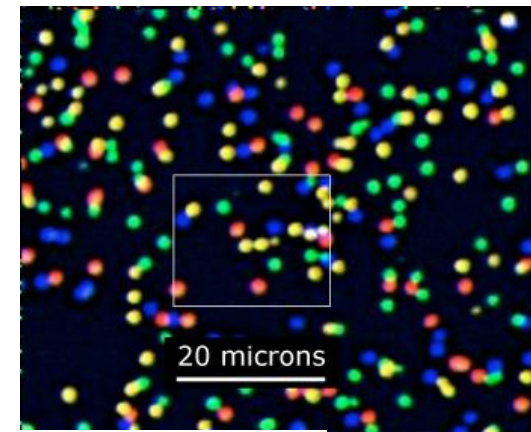
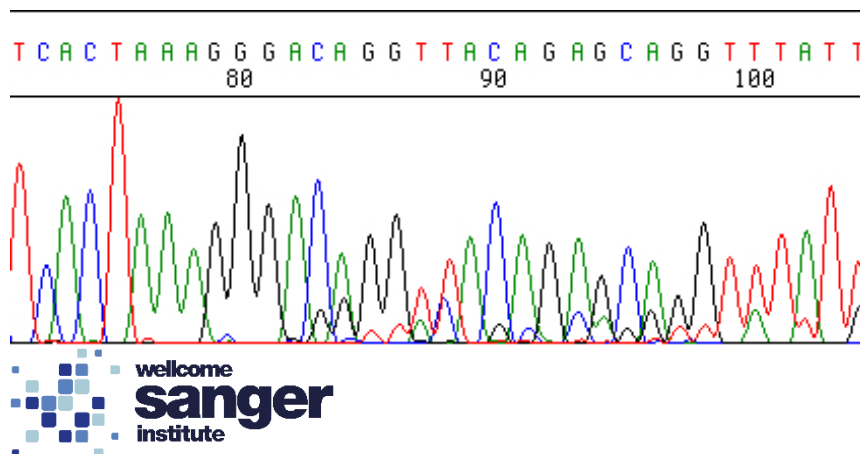
FASTQ format

- Each base has an assigned quality score
 - Sequencing quality scores measure the probability that a base is called incorrectly
- How is it calculated?



Phred quality and error probability

- **Light intensity** is used to calculate the error probabilities
- Convert error probability into Phred score quality - Ewing B, Green P. (1998)
- Phred originated as an algorithmic approach that considered Sanger sequencing metrics, such as **peak resolution and shape**



illumina®

Phred quality and error probability

- Convert error probability into Phred score quality - in real time on Illumina platforms
- Q scores are defined as a property that is logarithmically related to the base calling error probabilities (P)
- Phred quality range between 0-40 for Sanger and Illumina 1.8+

$$Q = -10 \log_{10} P$$

Phred Quality Score	Probability of Incorrect Base Call	Base Call Accuracy
10	1 in 10	90%
20	1 in 100	99%
30	1 in 1,000	99.9%
40	1 in 10,000	99.99%
50	1 in 100,000	99.999%

Phred quality and error probability

- Convert Phred quality score into ASCII, a compact form, which uses only 1 byte per quality value

ASCII_BASE=33 Illumina, Ion Torrent, PacBio and Sanger

Q	P_error	ASCII	Q	P_error	ASCII	Q	P_error	ASCII	Q	P_error	ASCII
0	1.00000	33 !	11	0.07943	44 ,	22	0.00631	55 7	33	0.00050	66 B
1	0.79433	34 "	12	0.06310	45 -	23	0.00501	56 8	34	0.00040	67 C
2	0.63096	35 #	13	0.05012	46 .	24	0.00398	57 9	35	0.00032	68 D
3	0.50119	36 \$	14	0.03981	47 /	25	0.00316	58 :	36	0.00025	69 E
4	0.39811	37 %	15	0.03162	48 0	26	0.00251	59 ;	37	0.00020	70 F
5	0.31623	38 &	16	0.02512	49 1	27	0.00200	60 <	38	0.00016	71 G
6	0.25119	39 '	17	0.01995	50 2	28	0.00158	61 =	39	0.00013	72 H
7	0.19953	40 (18	0.01585	51 3	29	0.00126	62 >	40	0.00010	73 I
8	0.15849	41)	19	0.01259	52 4	30	0.00100	63 ?	41	0.00008	74 J
9	0.12589	42 *	20	0.01000	53 5	31	0.00079	64 @	42	0.00006	75 K
10	0.10000	43 +	21	0.00794	54 6	32	0.00063	65 A			

- Phred+33 (Sanger and current Illumina). 0 Phred quality correspond to decimal 33, which is the symbol !

ASCII_BASE=64 Old Illumina

Q	P_error	ASCII	Q	P_error	ASCII	Q	P_error	ASCII	Q	P_error	ASCII
0	1.00000	64 @	11	0.07943	75 K	22	0.00631	86 V	33	0.00050	97 a
1	0.79433	65 A	12	0.06310	76 L	23	0.00501	87 W	34	0.00040	98 b
2	0.63096	66 B	13	0.05012	77 M	24	0.00398	88 X	35	0.00032	99 c
3	0.50119	67 C	14	0.03981	78 N	25	0.00316	89 Y	36	0.00025	100 d
4	0.39811	68 D	15	0.03162	79 O	26	0.00251	90 Z	37	0.00020	101 e
5	0.31623	69 E	16	0.02512	80 P	27	0.00200	91 [38	0.00016	102 f
6	0.25119	70 F	17	0.01995	81 Q	28	0.00158	92 \	39	0.00013	103 g
7	0.19953	71 G	18	0.01585	82 R	29	0.00126	93]	40	0.00010	104 h
8	0.15849	72 H	19	0.01259	83 S	30	0.00100	94 ^	41	0.00008	105 i
9	0.12589	73 I	20	0.01000	84 T	31	0.00079	95 _	42	0.00006	106 j
10	0.10000	74 J	21	0.00794	85 U	32	0.00063	96 `			

- Phred+64 (Solexa and Illumina 1.3-1.5)

Phred quality and error probability

- Phred 33 example

```
@HWI-ST731_6:1:1101:1322:1938#1@0/1
NTGACAAAGGGCTAATATCCAGAATCTACAAAGAACTTAAACAAATGTATAAGAATAAAAGTATAGTGCTAACAAT
+
#1:BDDADFDFFDD@F>BGFIIIB@CFHHIICAGBC9CBCBGGIGCFF??>GGHFHIGGEGI<FECGDE=FHCHEG=
```

$P=0.0001 \longrightarrow Q=-10*\log_{10}(0.0001)=40 \longrightarrow \text{ASCII } 33+40=73 \longrightarrow \text{I}$

$P=0.001 \longrightarrow Q=-10*\log_{10}(0.001)=30 \longrightarrow \text{ASCII } 33+30=63 \longrightarrow ?$

Quality encoding: !"#\$%&'()*+,-./0123456789:;<=>?@ABCDEFGHI

Quality score: 0	10	20	30	40

FASTQ format

Illumina read header

@HWUSI-EAS100R:6:73:941:1973#0/1

HWUSI-EAS100R	the unique instrument name
6	flowcell lane
73	tile number within the flowcell lane
941	'x'-coordinate of the cluster within the tile
1973	'y'-coordinate of the cluster within the tile
#0	index number for a multiplexed sample (0 for no indexing)
/1	the member of a pair, /1 or /2 (<i>paired-end or mate-pair reads only</i>)

```
@HWUSI-EAS1752R:21:FC64JUKAAXX:3:1:2458:1027 1:N:0:ACAGTG
AGAAAAAACCTTGGANGGAAAAAATCAGACATTTTCTAGAGGTGGAAGGCAAACTGAACAAAGAAATAATTACA
+
DGGGEDHHHHGGGFE#CBACBCA<?HHHHBHHHHHHHHDHHHHEHEFEFGGGGGG/GGDDDGHFHGFCHFHEHEH8
@HWUSI-EAS1752R:21:FC64JUKAAXX:3:1:3082:1029 1:N:0:ACAGTG
GGTAATACAGACTGANATGATCAAAGGCTGCTGGAACAAACCTATTAAGATAAGCTTGGATCAAGCTTTCATT
+
B:B?BB/:=55177#55877<775EDD>E=B?BBBBGGGDDAG@G>GGGGGG@)EEEEBEG>GGGGGGGAAA?<D
@HWUSI-EAS1752R:21:FC64JUKAAXX:3:1:3185:1033 1:N:0:ACAGTG
TCTGGGACATTGCTCNTGGCTGGGAGTCACCTGTCTGGGACATTGCTCAGGCTGGGAGACACGTGTTGGAGGGAC
+
BC??A66; )74781<#7??;452.27'64(8,851DDG8GB?#####
@HWUSI-EAS1752R:21:FC64JUKAAXX:3:1:3268:1033 1:N:0:ACAGTG
ATTCAAATTAGAAGANAGTTGATCGTTCTTCATGATGCCCAAAATTTCACTGAGAAAACCTTTTTTAAGCCCCAC
+
IIIIIIIIIIFFFFE#ABACFEFFFIIGIIIFIHE@BIIIIIIIIHHIIFIIF>HHIHFIDIIIIIGFHIIEGH
@HWUSI-EAS1752R:21:FC64JUKAAXX:3:1:3400:1035 1:N:0:ACAGTG
TCCTGCTTTAGGAGANTCCTCATGCTCTGACAGGATGCTCTCTATGTGAGTTGAGCTGGTCTTCTCACTTTTATAG
+
IIIIHHIHIIGGEGG#AACA@=?BHHIIIIHHIHHIHHIHHIHHGHIHHGHIHHGEGGGHG@EFGGCEFAB
@HWUSI-EAS1752R:21:FC64JUKAAXX:3:1:3962:1033 1:N:0:ACAGTG
CCACCAACACAGTCTNCACCTTCTGTTGCTGGTGATAGATTTTGCACCTTCCATCTCCAGGTTTCAAATAGC
+
HHFHHDHDDH>C?CA#EEEE>?A?>HHDGHEGBGBCEEEEGHHF8HEHEEHECH,=>==EAE>BEBBAEACAB
@HWUSI-EAS1752R:21:FC64JUKAAXX:3:1:4491:1028 1:N:0:ACAGTG
AGAGAGAGAGAGAGANAGAGGACTCTGGAGATGCCGAAGCACAAGCCTGCAAGAGTCCAGCAAAGAAAAATAAAA
+
GADGGEGGEGBBB?B#@=@@72:64GGGFG>GGGBDG<DBG<DA??/?#####
```

ASCII-coded (0-40):

- “!\"#\$%” lowest quality
- “FGHI” highest quality

Sequencing quality assessment

- To assess quality, software uses **Phred per-base quality** score is used
- Is the **first quality control step** after sequencing. There should be one after every step of the analysis
- After quality assessment user can know how **reliable** are their datasets
- QC will determine the next **filtering** step
- Filtering decisions will **impact** directly in **further analysis**
- Many other steps also use this quality as variable in their **algorithms**

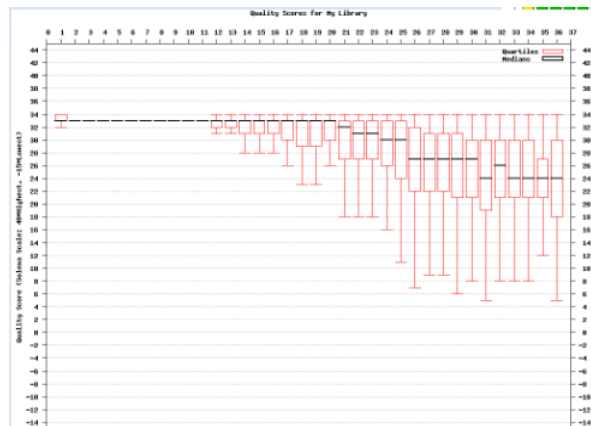
Sequencing quality assessment: Artifacts

HTS methods are bounded by their technical and theoretical limitations and sequencing errors cannot be completely eliminated (Hadigol M, Khiabani H. 2018)

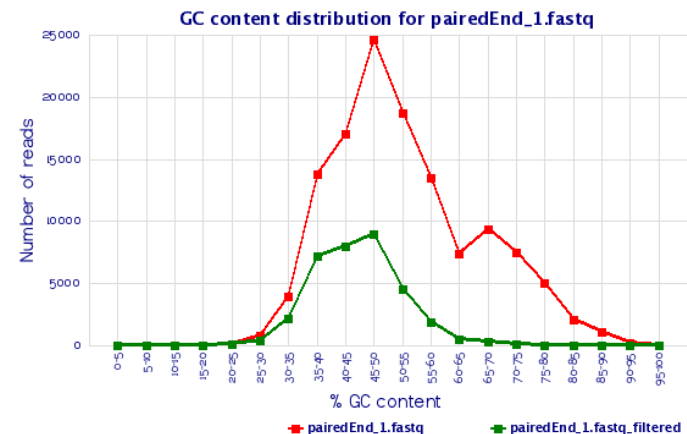
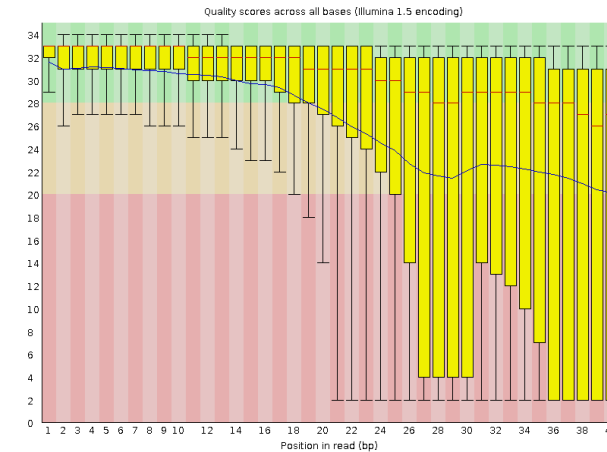
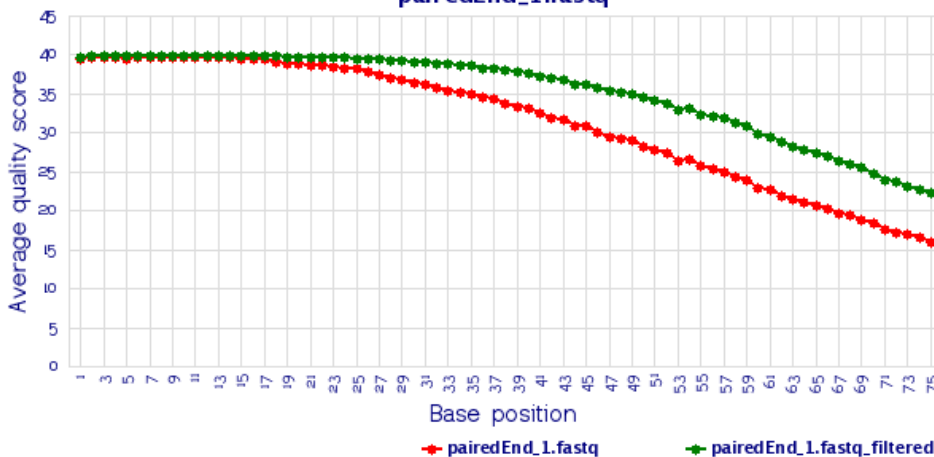
- **Artifacts in library preparation**
 - Remaining adapters
 - High rate of duplicates
 - GC regions bias
 - Polymerase error rate
 - DNA damage during breakdown
- **Artifacts during sequencing**
 - Low quality in sequence ends(Phasing: cluster loose sync)
 - Complication in certain regions:
 - Repetitions
 - Homopolymers
 - High CG content

Sequencing quality assessment

- FastQC, fastx-toolkit, sfftools, NGSQCToolkit, etc...

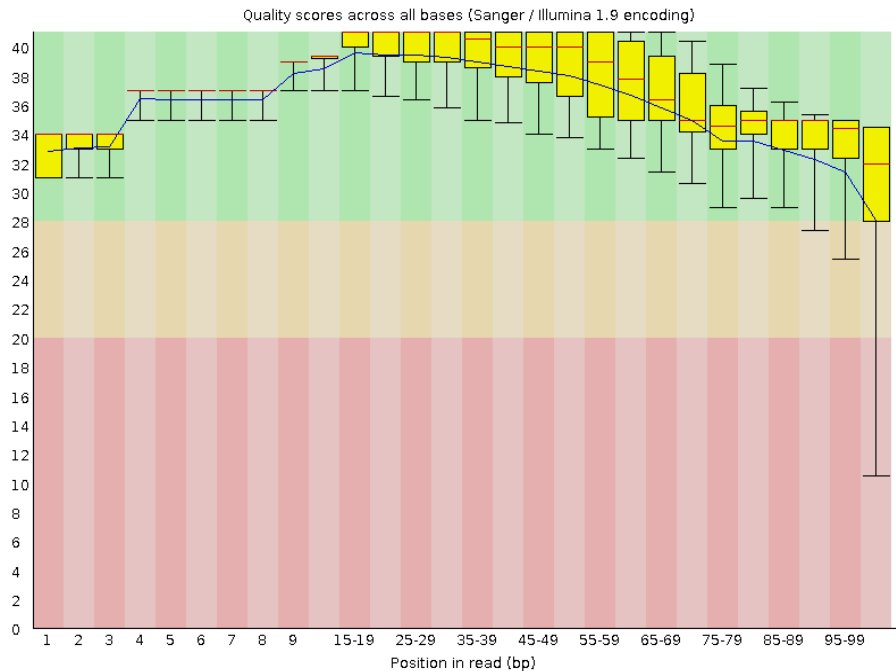


pairedEnd_1.fastq

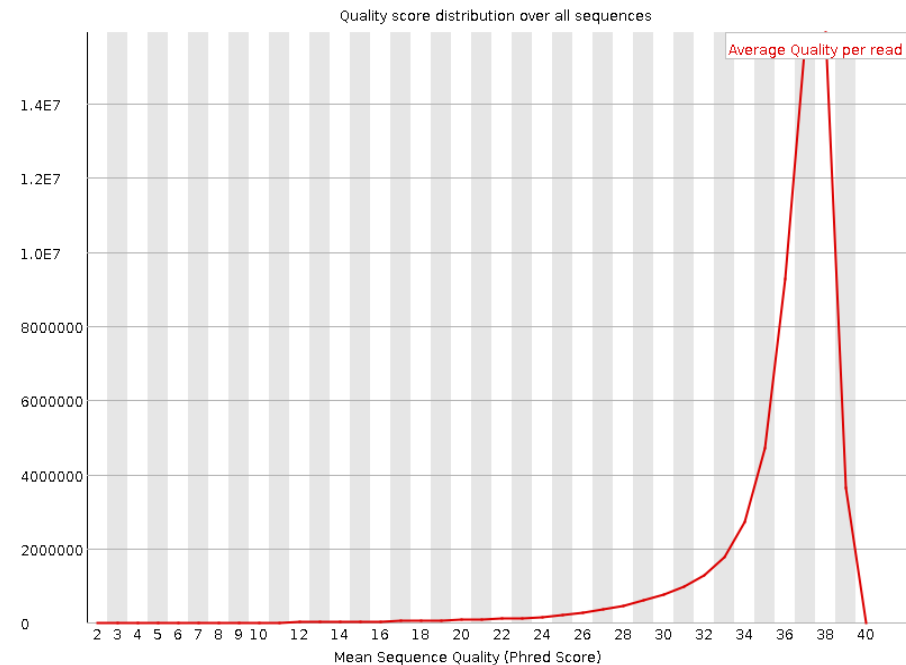


Sequencing quality assessment: FastQC

Per base sequence quality



Per sequence quality scores



<https://www.bioinformatics.babraham.ac.uk/projects/fastqc/>

Sequencing quality assessment: fastp

• Fastp fastp report

Summary

General

fastp version:	0.20.1 (https://github.com/OpenGene/fastp)
sequencing:	paired end (149 cycles + 149 cycles)
mean length before filtering:	116bp, 116bp
mean length after filtering:	117bp, 117bp
duplication rate:	1.704150%
Insert size peak:	95
Detected read1 adapter:	CACCTAAGTTGGCGTATACGCGTAATATATCTGGGTTTTCTACAAAATCATACCACTCT
Detected read2 adapter:	CACCTAAGTTGGCGTATACGCGTAATATATCTGGGTTTTCTACAAAATCATACCACTCT

Before filtering

total reads:	1.296756 M
total bases:	151.424921 M
Q20 bases:	143.112834 M (94.510754%)
Q30 bases:	137.905419 M (91.071812%)
GC content:	40.410939%

After filtering

total reads:	854.250000 K
total bases:	100.537720 M
Q20 bases:	99.598139 M (99.065444%)
Q30 bases:	97.968091 M (97.444115%)
GC content:	39.665634%

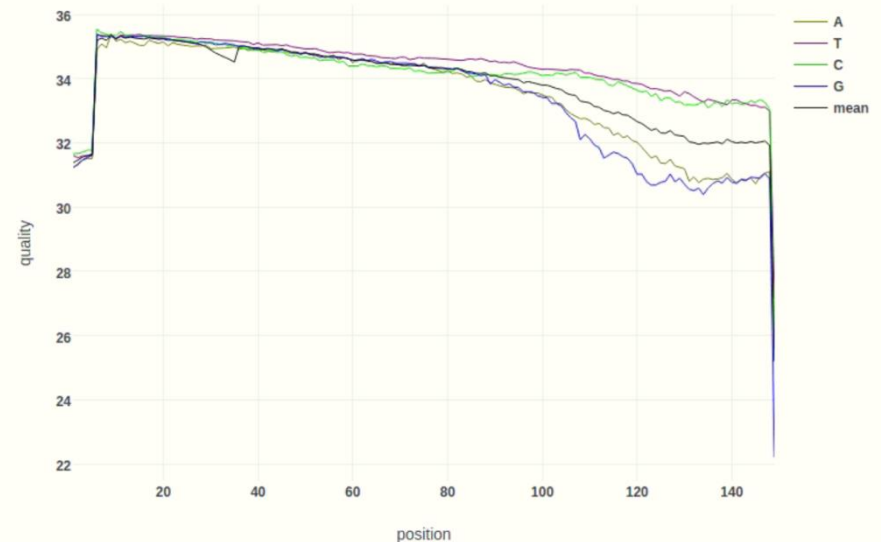
Filtering result

reads passed filters:	854.250000 K (65.875924%)
reads with low quality:	352.272000 K (27.165635%)
reads with too many N:	84 (0.006478%)
reads too short:	90.150000 K (6.951963%)

Before filtering

Before filtering: read1: quality

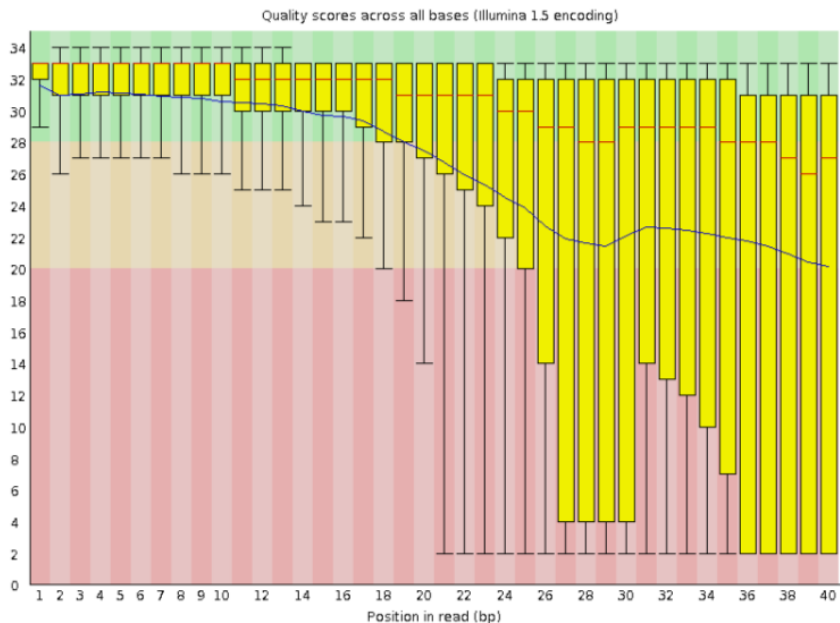
Value of each position will be shown on mouse over.



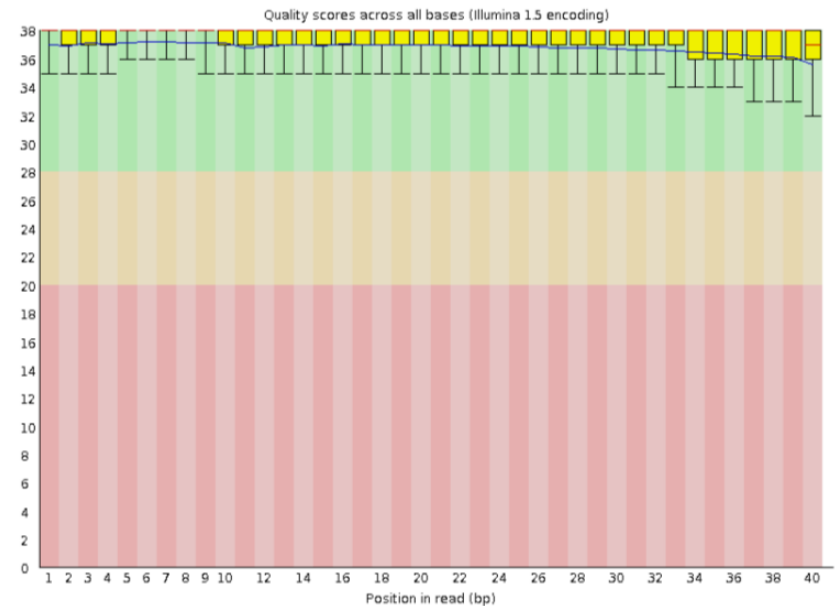
FastQC: Per base sequence quality

- Overview of the range of quality values across all bases at each position in the FastQ file
- **Median**, **inter-quartile range (25-75%)**, **10-90% points**, **mean quality**

✗ Per base sequence quality



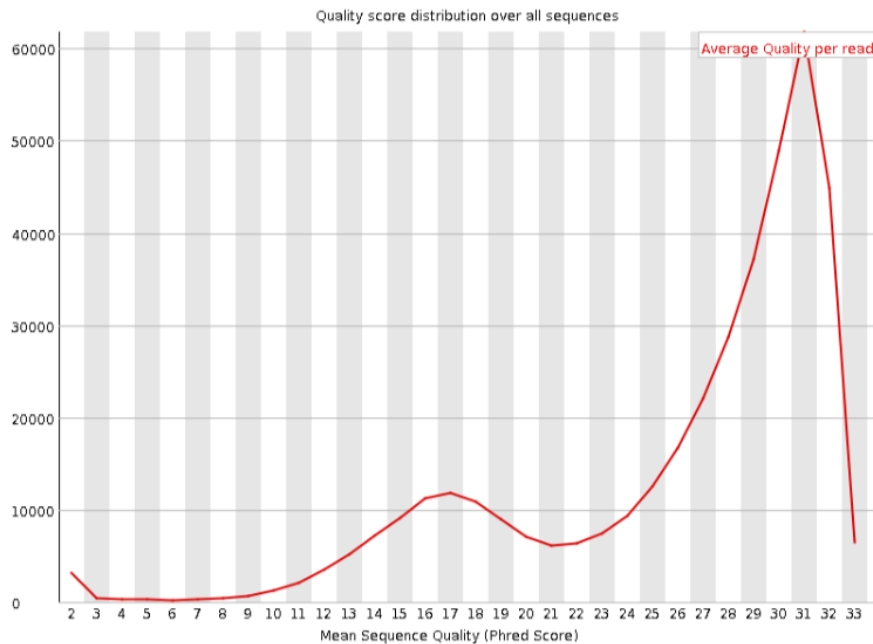
✓ Per base sequence quality



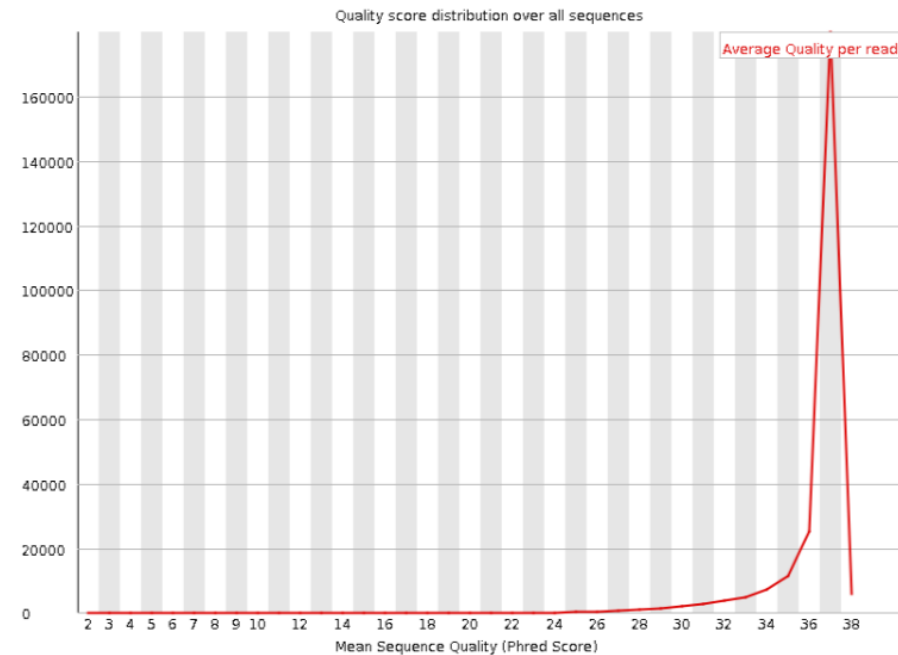
FastQC: Per sequence quality score

- Number of sequences with the same mean quality

✓ Per sequence quality scores



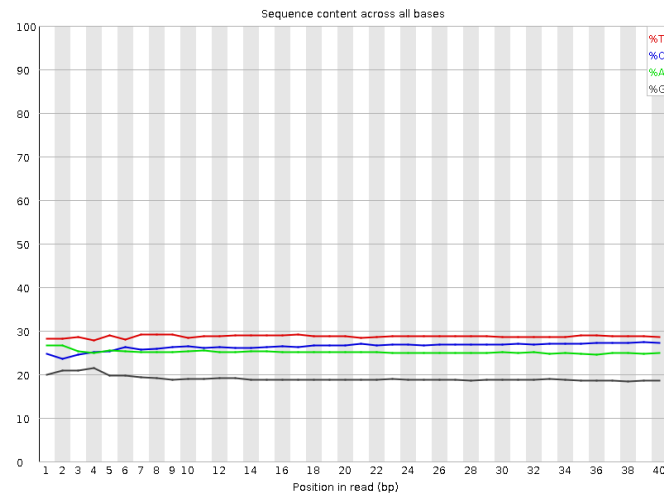
✓ Per sequence quality scores



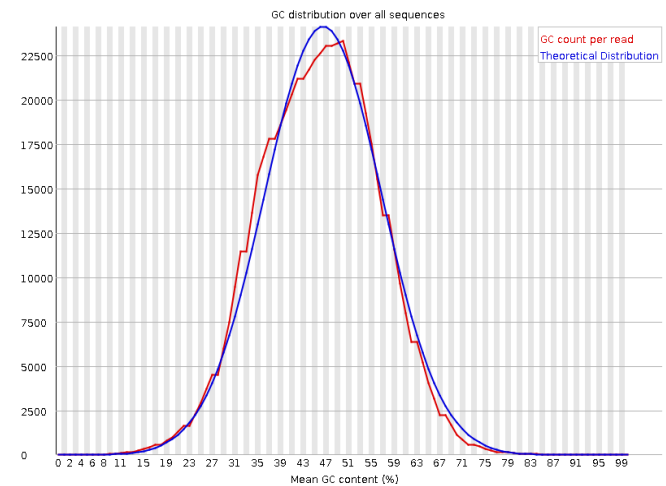
FastQC: Nucleotide related errors

- How expected nucleotide distribution deviates from expected
 - Per base sequence content
 - Per base GC content
 - Per sequence GC content
 - Per base N content

❗ Per base sequence content



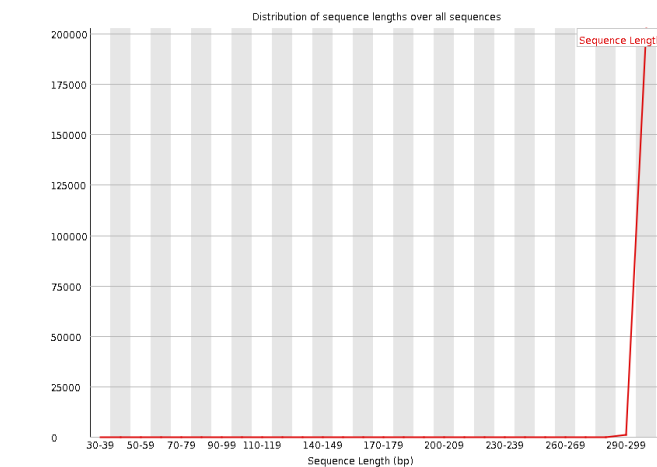
✅ Per sequence GC content



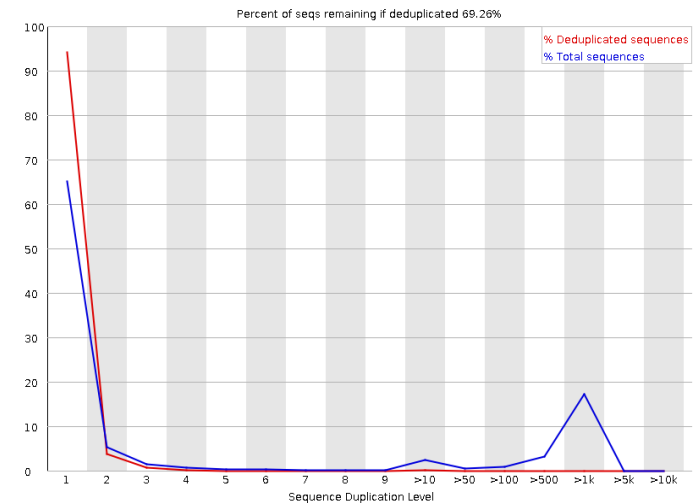
FastQC: Sequence related errors

- How expected nucleotide distribution deviates from expected
 - Sequence Length Distribution - Fragments
 - Sequence Duplication Levels
 - Overrepresented sequences
 - Adapter Content

Sequence Length Distribution



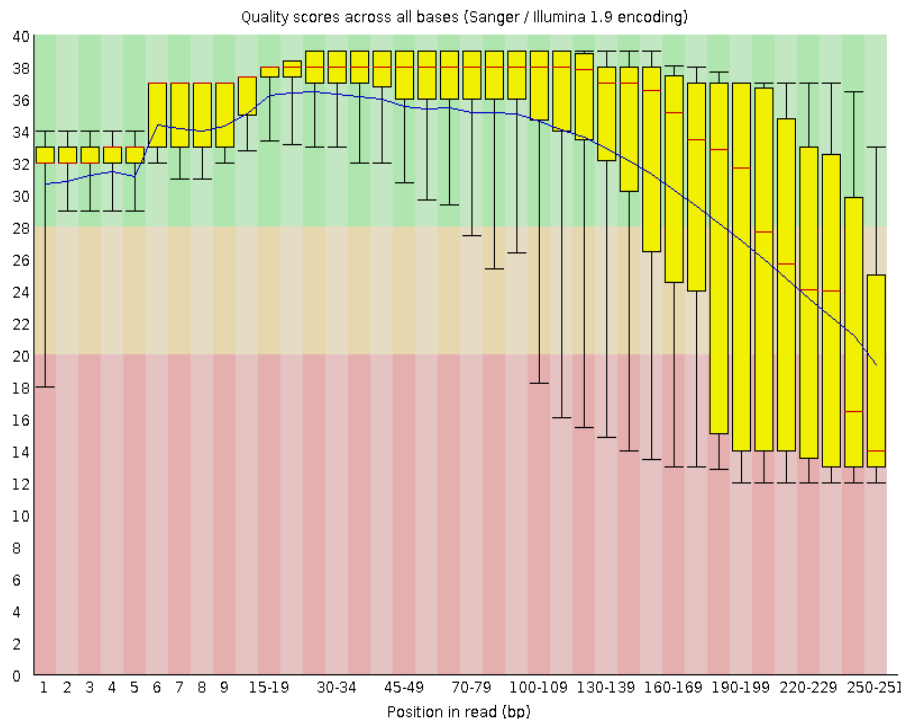
Sequence Duplication Levels



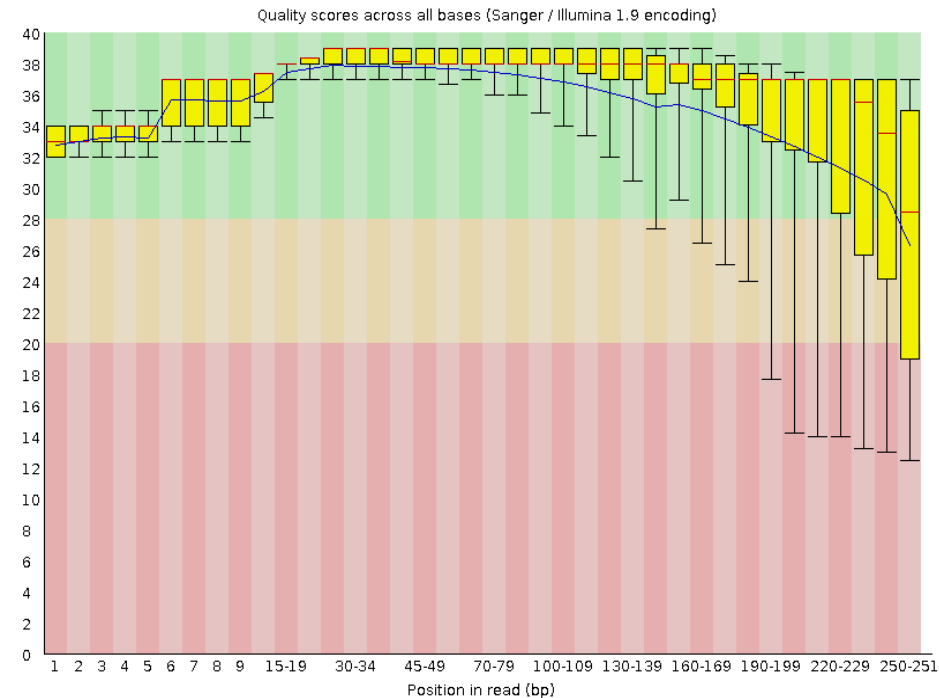
FastQC: Per base sequence quality

- Miseq assymetry

✗ Per base sequence quality



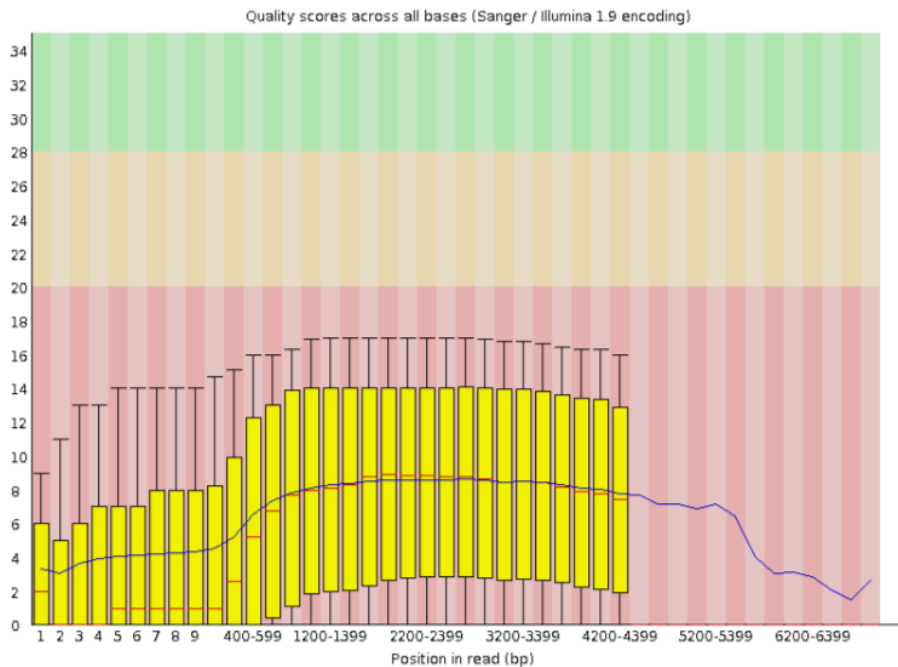
✓ Per base sequence quality



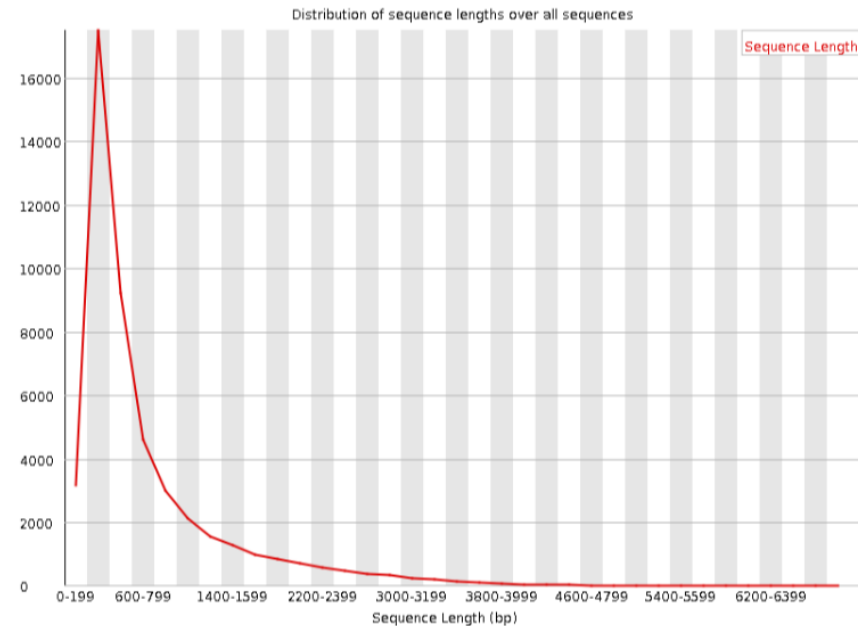
FastQC: Per base sequence quality

- SMRT PacBio

✖ Per base sequence quality



⚠ Sequence Length Distribution



Sequence filtering

- Remove residual adapters
 - Depending on used library
- Filtering parameters
 - Quality filtering
 - Overall mean quality
 - Local mean quality
 - Sequence end
 - Sliding window
 - Size filtering
 - Overall sequence size
 - Remaining sequence size after filtering

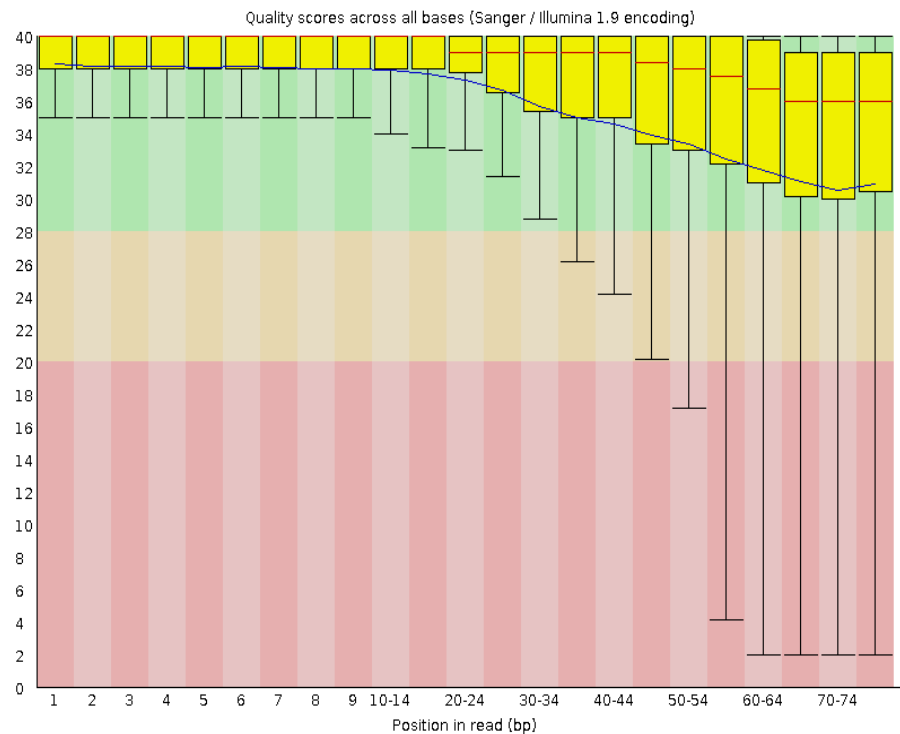


Sequence filtering

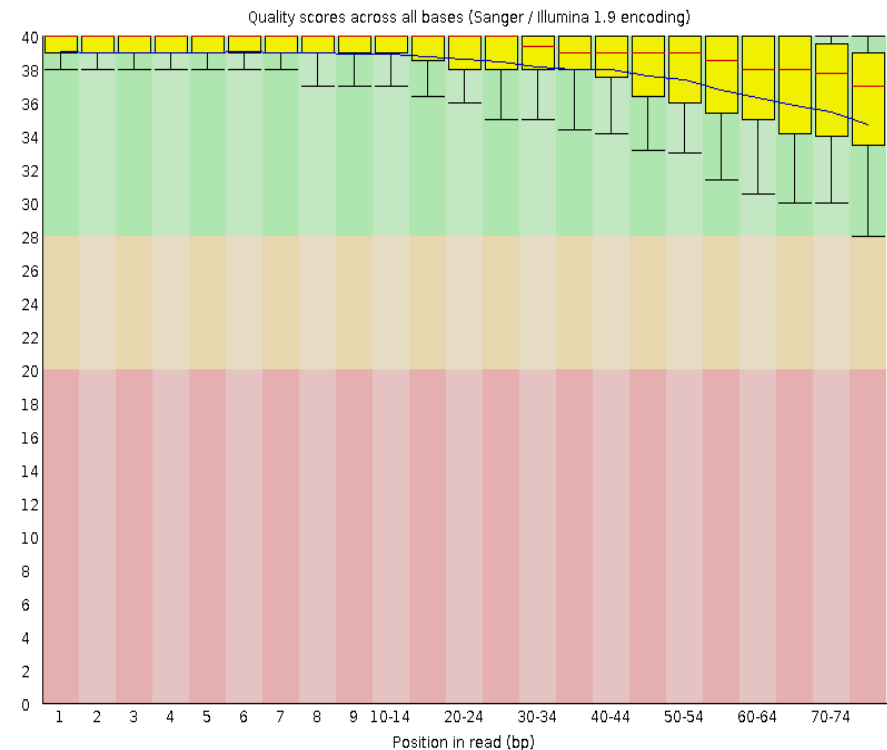
- Example of quality filtering



Per base sequence quality



Per base sequence quality



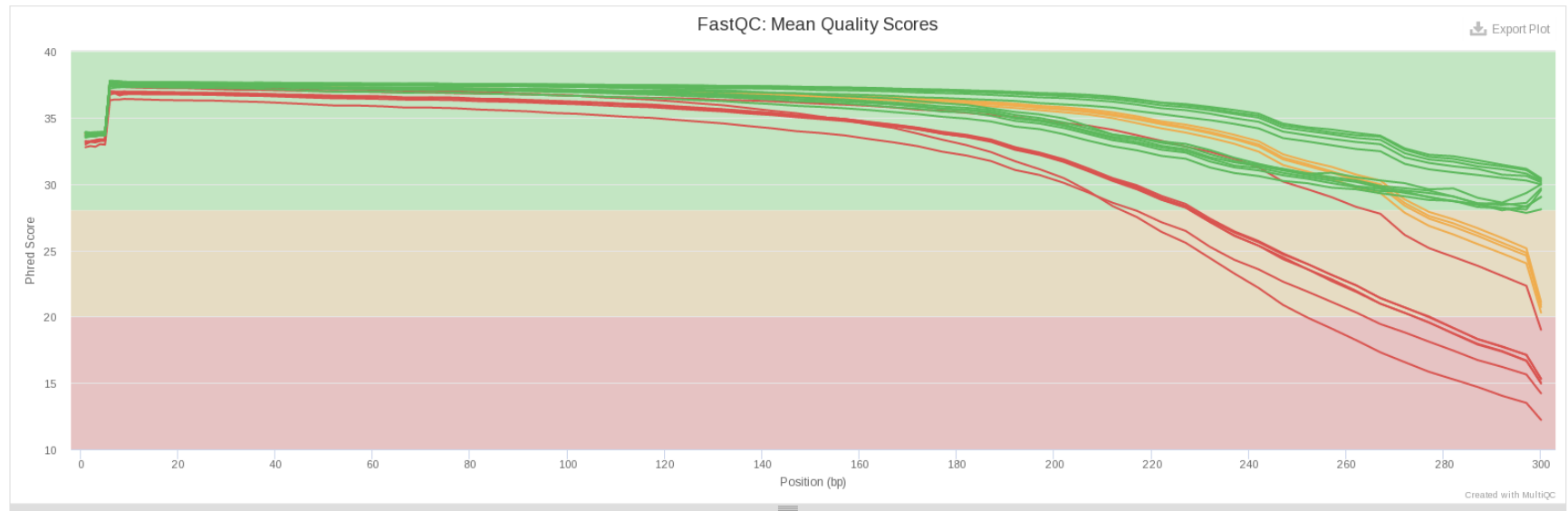
Sequence filtering: stats with MultiQC

Sequence Quality Histograms

11 4 7

The mean quality value across each base position in the read. See the [FastQC help](#).

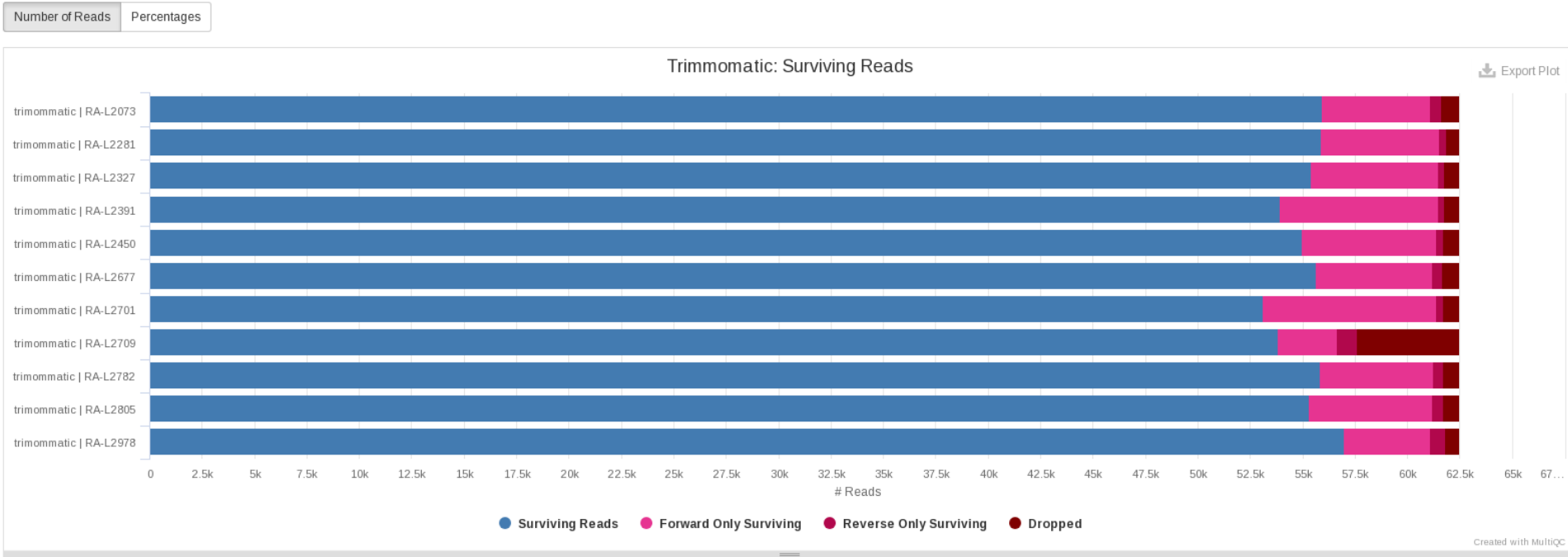
Y-Limits: ☐ off



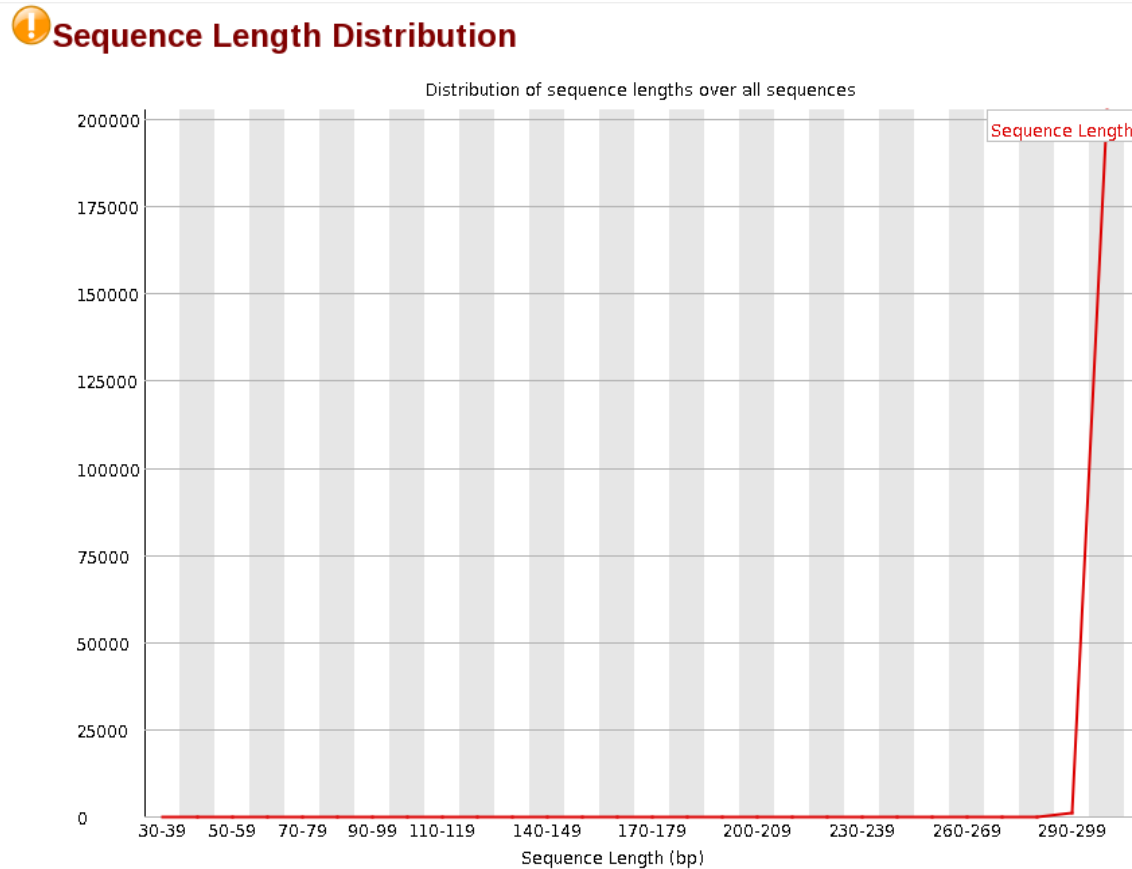
Sequence filtering: stats with MultiQC

Trimmomatic

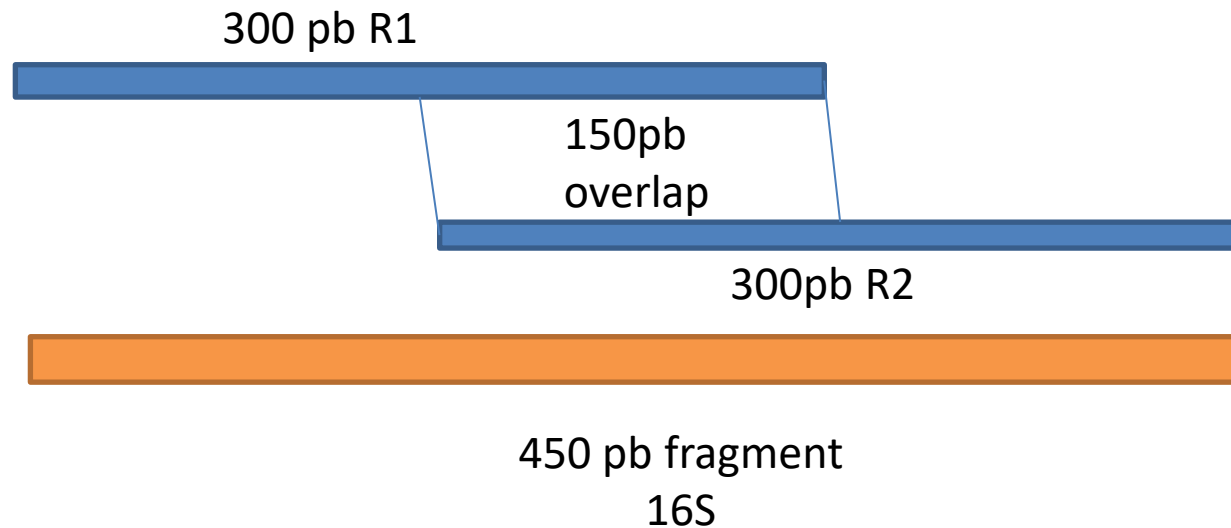
Trimmomatic is a flexible read trimming tool for Illumina NGS data.



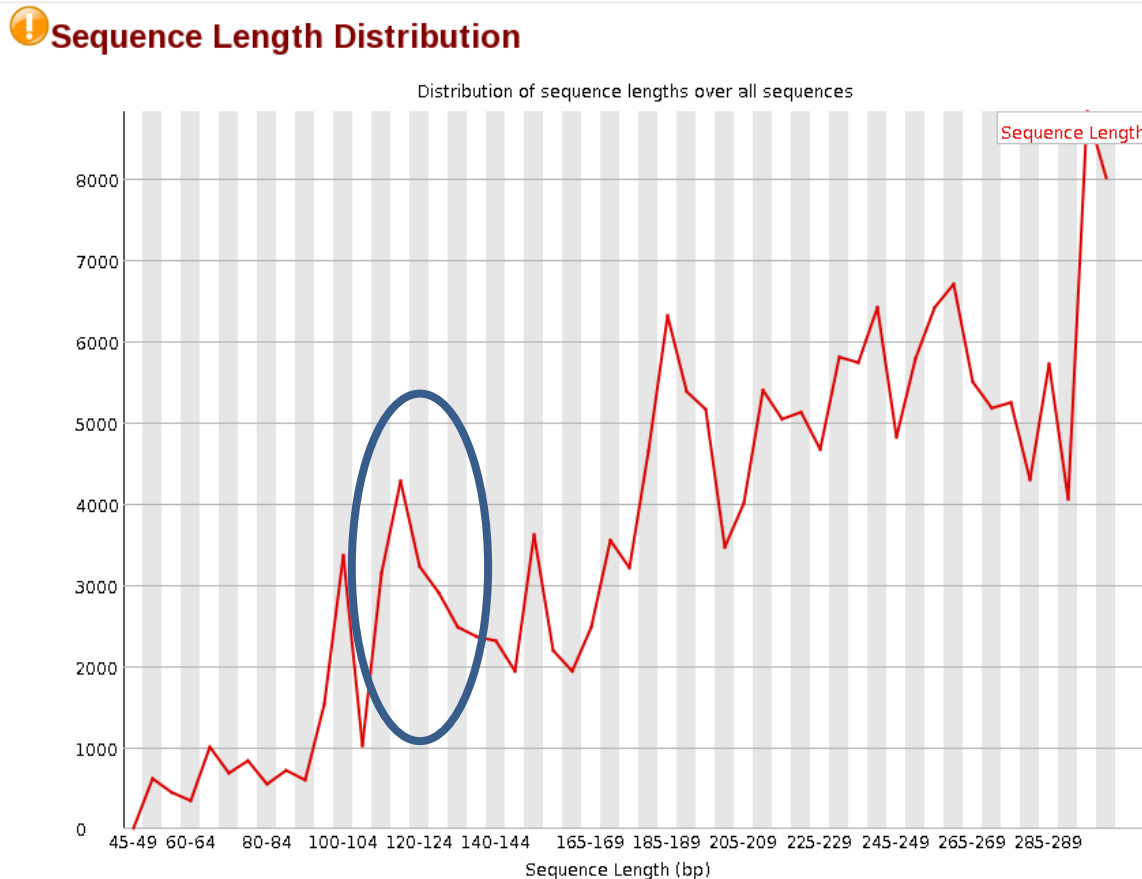
Quality filtering in metagenomic samples



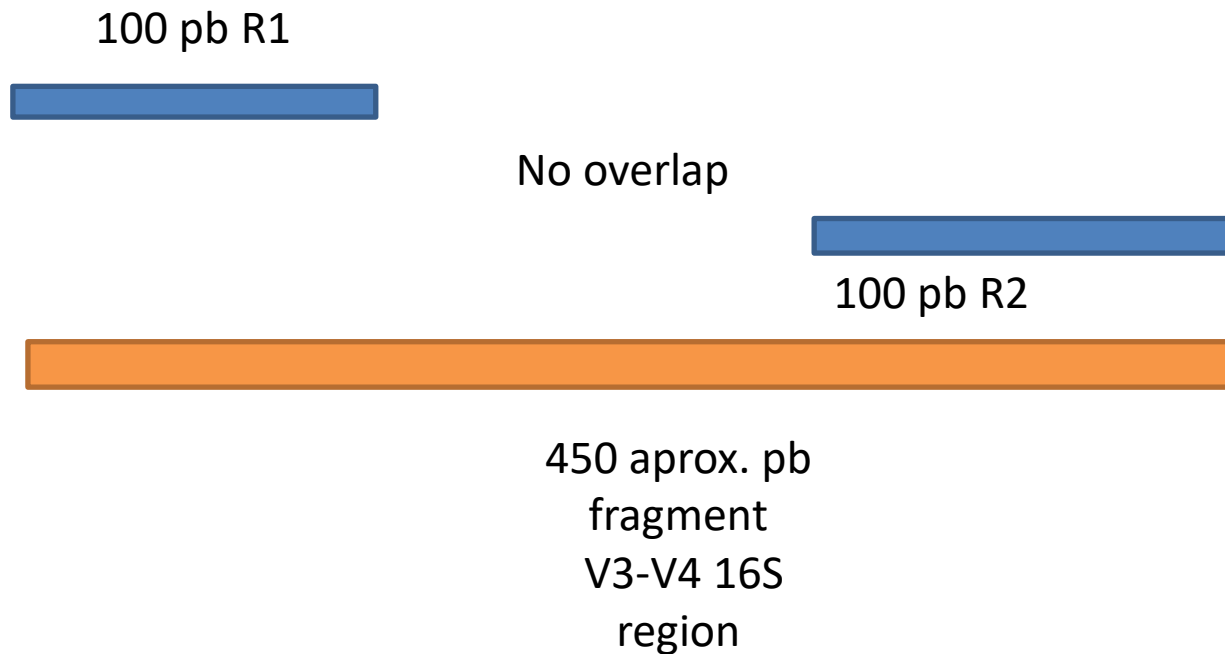
Quality filtering in metagenomic samples



Quality filtering in metagenomic samples



Quality filtering in metagenomic samples



Questions?
