# Comparison between the new method and the old method

#### 2024-04-24

## Load Dataset

```
##this file tests the proteomics example
library("IHWpaper")
## Loading required package: IHW
##
## Attaching package: 'IHW'
## The following object is masked _by_ '.GlobalEnv':
##
      ihw
library("IHW")
library(tidyverse)
## Warning: package 'ggplot2' was built under R version 4.2.3
## Warning: package 'tidyr' was built under R version 4.2.3
## Warning: package 'readr' was built under R version 4.2.3
## Warning: package 'dplyr' was built under R version 4.2.3
## Warning: package 'stringr' was built under R version 4.2.3
                                             ----- tidyverse 2.0.0 --
## -- Attaching core tidyverse packages ----
## v dplyr
           1.1.4
                       v readr
                                    2.1.5
## v forcats 1.0.0
                       v stringr
                                   1.5.1
## v ggplot2 3.5.0
                     v tibble
                                  3.2.1
## v lubridate 1.9.3
                        v tidyr
                                    1.3.1
## v purrr
              1.0.2
## -- Conflicts ----- tidyverse_conflicts() --
## x ggplot2::alpha() masks IHW::alpha()
## x dplyr::filter() masks stats::filter()
                       masks CVXR::id()
## x dplyr::id()
## x purrr::is_vector() masks CVXR::is_vector()
## x dplyr::lag() masks stats::lag()
## i Use the conflicted package (<a href="http://conflicted.r-lib.org/">http://conflicted.r-lib.org/</a>) to force all conflicts to become error
proteomics file <- system.file("extdata/real data",</pre>
                              "science_signaling.csv", package = "IHWpaper")
proteomics_df <- read.csv(proteomics_file, stringsAsFactors = F)</pre>
proteomics_df$pvalue <- rank(proteomics_df$p1, ties.method="first")*proteomics_df$p1/nrow(proteomics_df
```

## Run the three methods

N <- length(num bins)

 $num_bins \leftarrow seq(from = 10, to = 50, by = 10)$ 

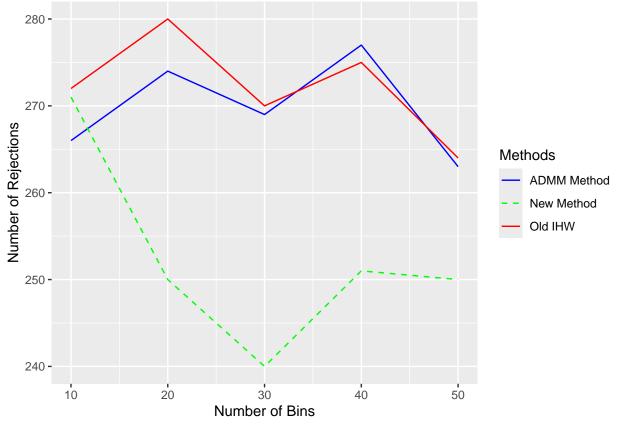
```
time.new <- rep(NA, N)
time.old <- rep(NA, N)
time.cvxr <- rep(NA, N)
rej_new <- rep(NA, N)</pre>
rej_old <- rep(NA, N)</pre>
rej_cvxr <- rep(NA, N)</pre>
for(i in 1:N){
  time.new[i] <- system.time(ihw_res_new <- ihw(proteomics_df$pvalue, proteomics_df$X..peptides, alpha
  rej_new[i] <- IHW:::rejections(ihw_res_new)</pre>
  time.old[i] <- system.time(ihw_res_old <- IHWold:::ihwOld(proteomics_df$pvalue, proteomics_df$X..pept
  rej_old[i] <- IHW:::rejections(ihw_res_old)</pre>
  print(paste0("Ending iteration ", i))
}
## We recommend that you supply (many) more than 1000 p-values for meaningful data-driven hypothesis we
## [1] "Ending iteration 1"
## We recommend that you supply (many) more than 200 p-values for meaningful data-driven hypothesis wei
## We recommend that you supply (many) more than 1000 p-values for meaningful data-driven hypothesis we
## [1] "Ending iteration 2"
## We recommend that you supply (many) more than 200 p-values for meaningful data-driven hypothesis wei
## We recommend that you supply (many) more than 1000 p-values for meaningful data-driven hypothesis we
## [1] "Ending iteration 3"
## We recommend that you supply (many) more than 200 p-values for meaningful data-driven hypothesis wei
## We recommend that you supply (many) more than 1000 p-values for meaningful data-driven hypothesis we
## [1] "Ending iteration 4"
## We recommend that you supply (many) more than 200 p-values for meaningful data-driven hypothesis wei
## We recommend that you supply (many) more than 1000 p-values for meaningful data-driven hypothesis we
## [1] "Ending iteration 5"
```

#### Plot the results

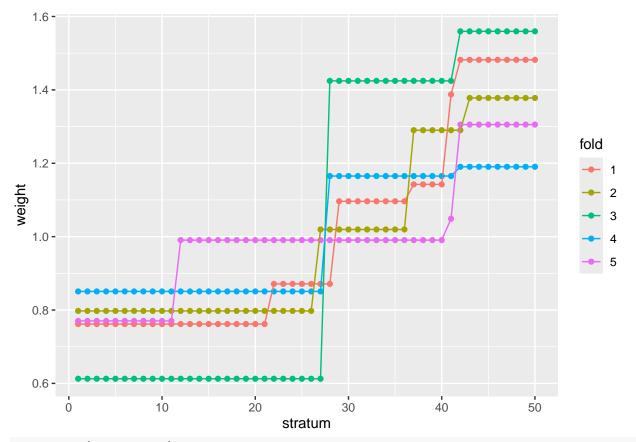
```
load("~/Library/Mobile Documents/com~apple~CloudDocs/Desktop/UchicagoStudy/Nikolaos Research/IHW_New/CV.
load("~/Library/Mobile Documents/com~apple~CloudDocs/Desktop/UchicagoStudy/Nikolaos Research/IHW_New/CV.
load("~/Library/Mobile Documents/com~apple~CloudDocs/Desktop/UchicagoStudy/Nikolaos Research/IHW_New/CV.
data <- data.frame(num_bins = num_bins, rejection_new = rej_new[1:5], rejection_admm = rej_admm[1:5], r
#we first plot runtime versus bins
# Create the line plot</pre>
```

```
p1 <- ggplot(data) +
  geom_line(aes(x = num_bins, y = log(time_admm), col = "ADMM Method", linetype = "ADMM Method")) +
  geom_line(aes(x = num_bins, y = log(time_old), col = "Old IHW", linetype = "Old IHW")) +
  geom_line(aes(x = num_bins, y = log(time_new), col = "New Method", linetype = "New Method")) +
  labs(x = "Number of Bins", y = "Runtime (log sec)") +
scale_color_manual(values = c("ADMM Method" = "blue", "Old IHW" = "red", "New Method" = "green"), name
  scale_linetype_manual(values = c("ADMM Method" = "solid", "Old IHW" = "solid", "New Method" = "solid"
#we then plot rejections versus bins
# Create the line plot
p2 <- ggplot(data) +
  geom_line(aes(x = num_bins, y = rejection_admm, col = "ADMM Method", linetype = "ADMM Method")) +
  geom_line(aes(x = num_bins, y = rejection_old, col = "Old IHW", linetype = "Old IHW")) +
  geom_line(aes(x = num_bins, y = rejection_new, col = "New Method", linetype = "New Method")) +
  labs(x = "Number of Bins", y = "Number of Rejections") +
scale_color_manual(values = c("ADMM Method" = "blue", "Old IHW" = "red", "New Method" = "green"), name
  scale_linetype_manual(values = c("ADMM Method" = "solid", "Old IHW" = "solid", "New Method" = "dashed
р1
   2 -
   1 -
Runtime (log sec)
   0 -
                                                                        Methods
                                                                             ADMM Method
                                                                             New Method
                                                                             Old IHW
   -2 -
   -3 -
                                    30
       10
                      20
                                                   40
                                                                 50
                              Number of Bins
```

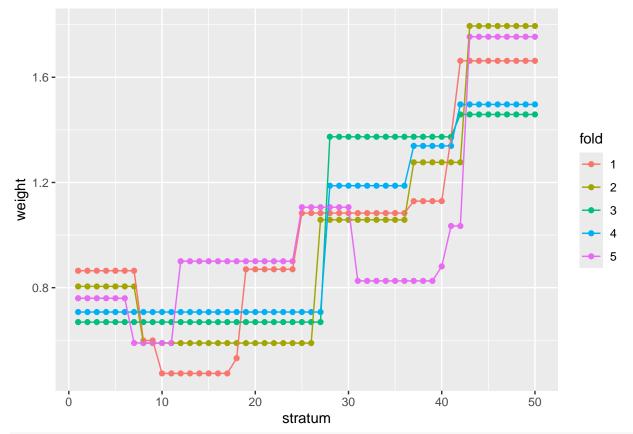
p2



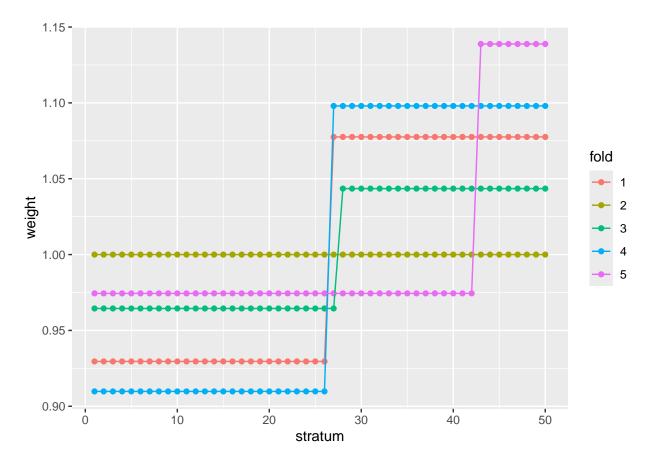
#plot weights: num bins = 50
IHW:::plot(ihw\_res\_admm)



IHW:::plot(ihw\_res\_old)



IHW:::plot(ihw\_res\_new)



## 2nd example

```
library("IHWpaper")
library("IHW")
bottomly <- analyze_dataset("bottomly")</pre>
##
     Note: levels of factors in the design contain characters other than
     letters, numbers, '_' and '.'. It is recommended (but not required) to use
##
##
     only letters, numbers, and delimiters \verb|'_'| or \verb|'.'|, as these are safe characters
##
     for column names in R. [This is a message, not a warning or an error]
## estimating size factors
##
     Note: levels of factors in the design contain characters other than
     letters, numbers, \verb|'_'| and \verb|'.'| . It is recommended (but not required) to use
##
     only letters, numbers, and delimiters '\_' or '.', as these are safe characters
##
     for column names in R. [This is a message, not a warning or an error]
## estimating dispersions
## gene-wise dispersion estimates
## mean-dispersion relationship
##
     Note: levels of factors in the design contain characters other than
     letters, numbers, '_' and '.'. It is recommended (but not required) to use
##
     only letters, numbers, and delimiters '_' or '.', as these are safe characters
##
     for column names in R. [This is a message, not a warning or an error]
## final dispersion estimates
```

```
Note: levels of factors in the design contain characters other than
    letters, numbers, '_' and '.'. It is recommended (but not required) to use
##
##
     only letters, numbers, and delimiters '_' or '.', as these are safe characters
     for column names in R. [This is a message, not a warning or an error]
##
## fitting model and testing
    Note: levels of factors in the design contain characters other than
     letters, numbers, '_' and '.'. It is recommended (but not required) to use
##
##
     only letters, numbers, and delimiters '_' or '.', as these are safe characters
     for column names in R. [This is a message, not a warning or an error]
## -- replacing outliers and refitting for 4 genes
## -- DESeq argument 'minReplicatesForReplace' = 7
## -- original counts are preserved in counts(dds)
## estimating dispersions
##
    Note: levels of factors in the design contain characters other than
     letters, numbers, '_' and '.'. It is recommended (but not required) to use
##
     only letters, numbers, and delimiters '_' or '.', as these are safe characters
     for column names in R. [This is a message, not a warning or an error]
## fitting model and testing
##
    Note: levels of factors in the design contain characters other than
##
     letters, numbers, '_' and '.'. It is recommended (but not required) to use
##
     only letters, numbers, and delimiters '_' or '.', as these are safe characters
     for column names in R. [This is a message, not a warning or an error]
num bins \leftarrow seq(from = 10, to = 50, by = 10)
N <- length(num_bins)</pre>
```

# Run the three methods

```
time.new <- rep(NA, N)
time.old <- rep(NA, N)
#time.cuxr <- rep(NA, N)

rej_new <- rep(NA, N)
rej_old <- rep(NA, N)

rej_cuxr <- rep(NA, N)

for(i in 1:N){
    time.new[i] <- system.time(ihw_res_new <- ihw(bottomly$pvalue, bottomly$baseMean, 0.1, nbins = num_bi:
    rej_new[i] <- IHW:::rejections(ihw_res_new)

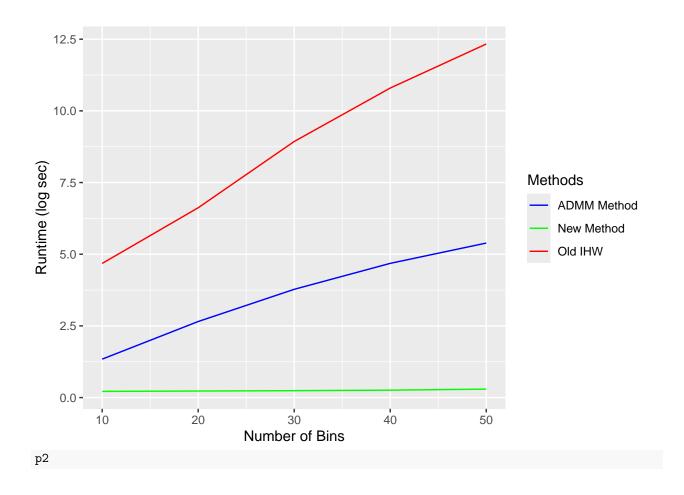
time.old[i] <- system.time(ihw_res_old <- IHWold:::ihwOld(bottomly$pvalue, bottomly$baseMean, 0.1, nb
    rej_old[i] <- IHW:::rejections(ihw_res_old)

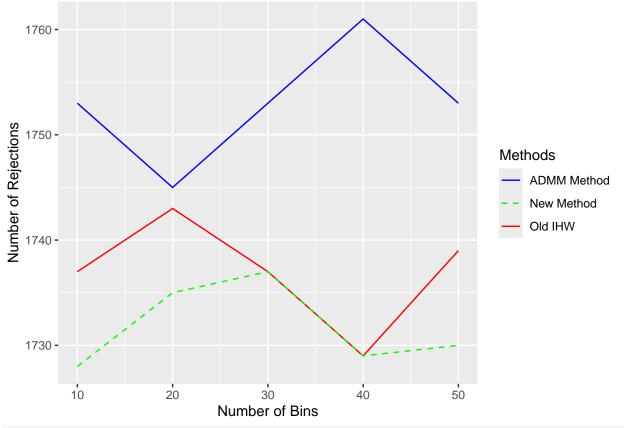
#time.cuxr[i] <- system.time(ihw_res_cuxr <- IHW:::ihw(bottomly$pvalue, bottomly$baseMean, 0.1, nbins
#rej_cuxr[i] <- IHW:::rejections(ihw_res_cuxr)

print(paste0("Ending iteration ", i))
}</pre>
```

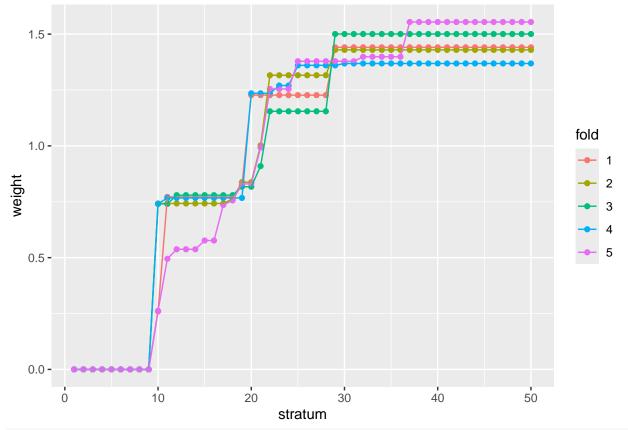
```
## [1] "Ending iteration 1"
## We recommend that you supply (many) more than 1000 p-values for meaningful data-driven hypothesis we
## [1] "Ending iteration 2"
## We recommend that you supply (many) more than 1000 p-values for meaningful data-driven hypothesis we
## [1] "Ending iteration 3"
## We recommend that you supply (many) more than 1000 p-values for meaningful data-driven hypothesis we
## [1] "Ending iteration 4"
## We recommend that you supply (many) more than 1000 p-values for meaningful data-driven hypothesis we
## [1] "Ending iteration 5"
load("~/Library/Mobile Documents/com~apple~CloudDocs/Desktop/UchicagoStudy/Nikolaos Research/IHW_New/CV
load("~/Library/Mobile Documents/com~apple~CloudDocs/Desktop/UchicagoStudy/Nikolaos Research/IHW_New/CV
load("~/Library/Mobile Documents/com~apple~CloudDocs/Desktop/UchicagoStudy/Nikolaos Research/IHW_New/CV
data <- data.frame(num_bins = num_bins, rejection_new = rej_new[1:5], rejection_admm = rej_admm[1:5], r
library(tidyverse)
#we first plot runtime versus bins
# Create the line plot
p1 <- ggplot(data) +
  geom_line(aes(x = num_bins, y = time_admm, col = "ADMM Method", linetype = "ADMM Method")) +
  geom_line(aes(x = num_bins, y = time_old, col = "Old IHW", linetype = "Old IHW")) +
  geom_line(aes(x = num_bins, y = time_new, col = "New Method", linetype = "New Method")) +
  labs(x = "Number of Bins", y = "Runtime (log sec)") +
scale_color_manual(values = c("ADMM Method" = "blue", "Old IHW" = "red", "New Method" = "green"), name
  scale_linetype_manual(values = c("ADMM Method" = "solid", "Old IHW" = "solid", "New Method" = "solid"
#we then plot rejections versus bins
# Create the line plot
p2 <- ggplot(data) +
  geom_line(aes(x = num_bins, y = rejection_admm, col = "ADMM Method", linetype = "ADMM Method")) +
  geom_line(aes(x = num_bins, y = rejection_old, col = "Old IHW", linetype = "Old IHW")) +
  geom_line(aes(x = num_bins, y = rejection_new, col = "New Method", linetype = "New Method")) +
  labs(x = "Number of Bins", y = "Number of Rejections") +
scale_color_manual(values = c("ADMM Method" = "blue", "Old IHW" = "red", "New Method" = "green"), name
  scale_linetype_manual(values = c("ADMM Method" = "solid", "Old IHW" = "solid", "New Method" = "dashed
```

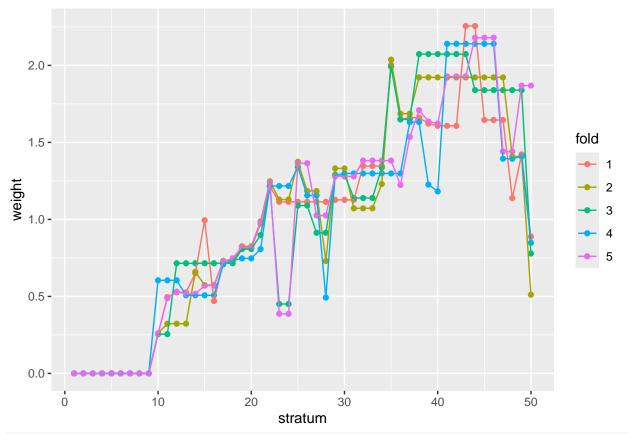
p1





#plot weights: num bins = 50
IHW:::plot(ihw\_res\_admm)





IHW:::plot(ihw\_res\_new)

