

Lecture 9



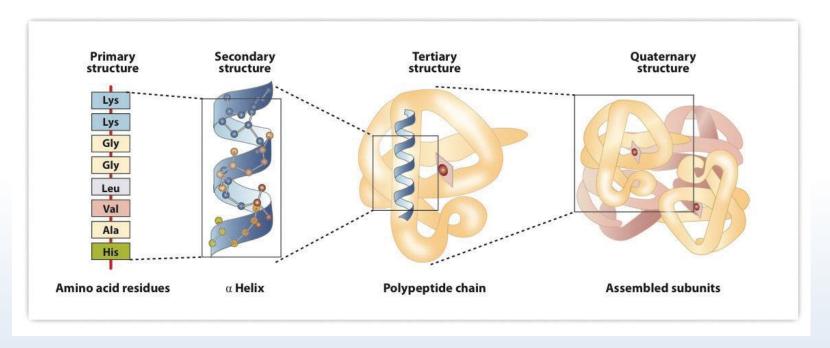


QC Testing
Protein Estimation Assays
Slab Gel Electrophoresis
Capillary Electrophoresis
HPLC
Immunoassays and Bioassays
PCR



What makes a protein?

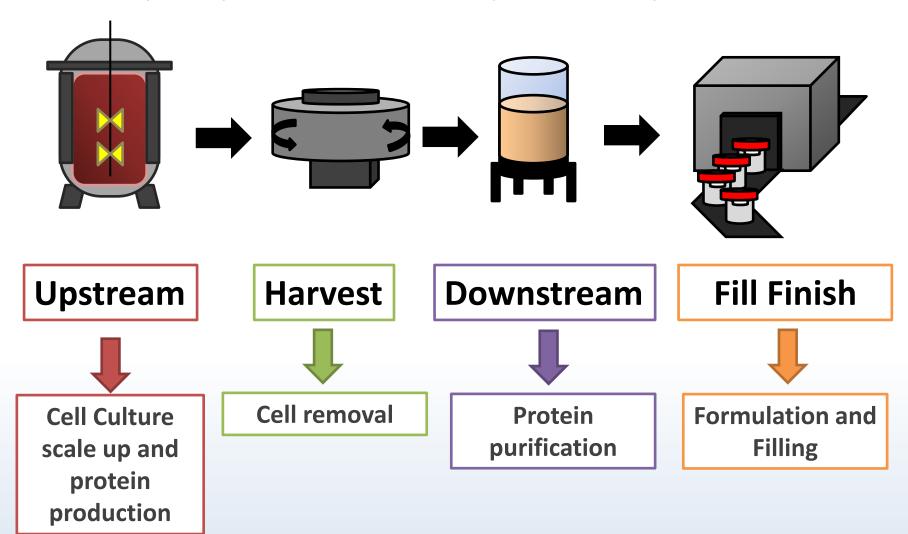
- The amino acids are arranged in a specific sequence which is dictated by the gene for that protein
- The sequence is extremely important for the protein to take its biologically active shape





Bioprocessing: Complex and Unique

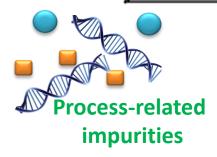
Many complex cell culture and purification procedures.



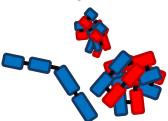


ICH Topic Q 6 B

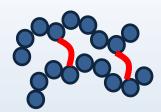
Specifications: Test Procedures and Acceptance Criteria for Biotechnological/Biological Products



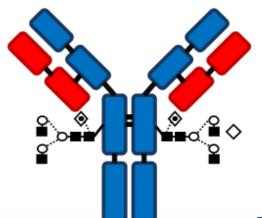
Product-related impurities



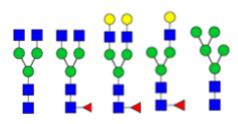
Disulfide bridges

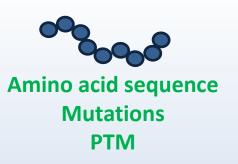


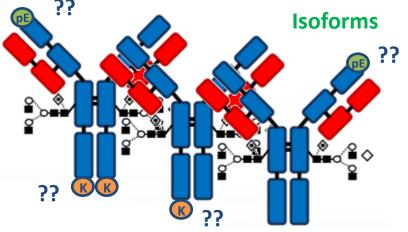
Intact mass
Higher Order Structure
Potency
Binding properties



Glycan profile









Vial inspection

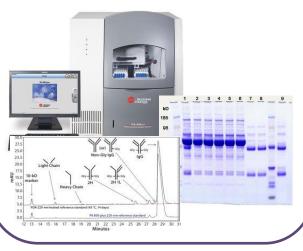


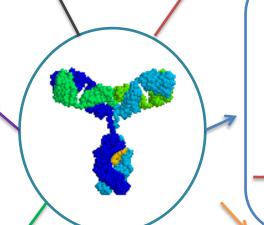




pH, conductivity, osmolality

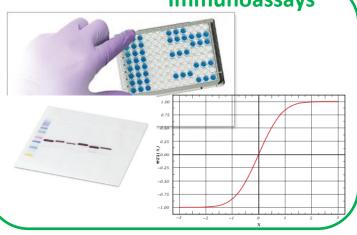
Electrophoresis

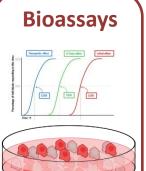






Immunoassays





Microbial testing

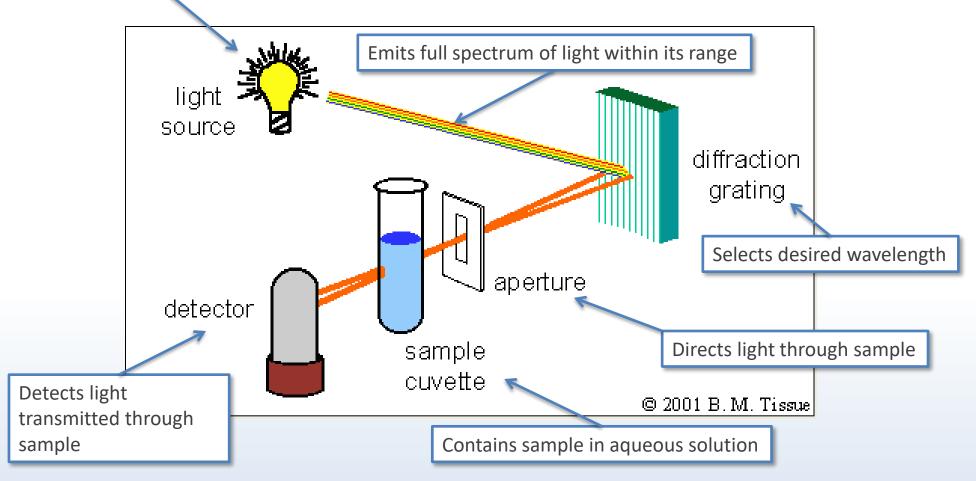




QC Testing **Protein Estimation Assays** Slab Gel Electrophoresis **Capillary Electrophoresis HPLC** Immunoassays and Bioassays PCR

Measuring Protein Concentration

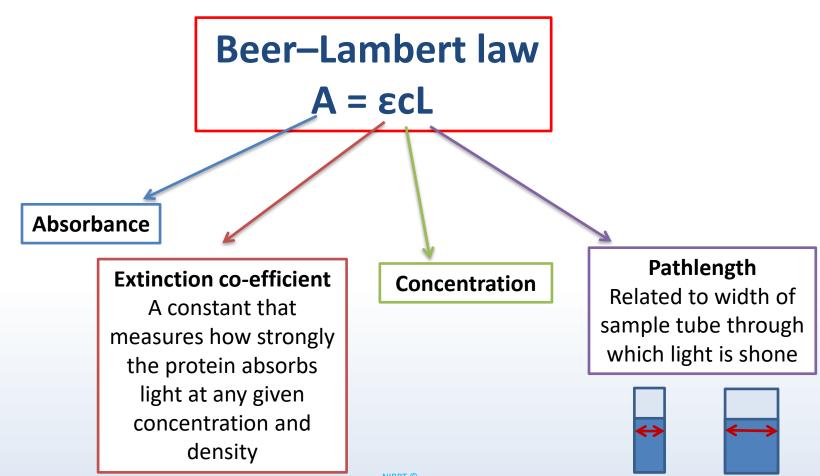
Tungsten lamp = 340 - 1100nm range (Visible) Deuterium lamp = 168 - 360nm range (UV)





Beer-Lambert Law

Absorbance is related to concentration by:





Protein concentration assays

Methods:

• Absorbance at 280nm — No dye or chemicals

- Biuret
- Lowry

BCA

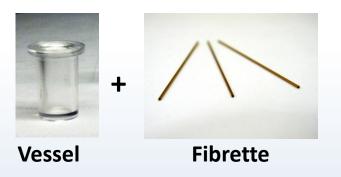
→ Copper-based colorimetric

Method	Estimated working range
A280nm	0.2 – 2mg/ml
Coomassie (Bradford)	$100 - 1500 \mu g/m I$ (Bio-Rad website)
Modified Lowry Assay	1 - 1500µg/ml (Pierce website)
BCA	$20 - 2000 \mu g/m I$ (Pierce website)

SoloVPE

- SoloVPE is a slope spectrophotometer developed by C-Technologies
- It uses UV-Vis measurement platform and variable pathlength technology
- Composed of Agilent Cary 60 spectrophotometer and fibre optic coupler
- Fibre optic cable carries light from spec to the fibre platform to a piece of optical fibre called a "Fibrette"
- Light passes through fibrette into the sample and various pathlength measurements taken.

To process a sample you need:



F Protein +
Sample





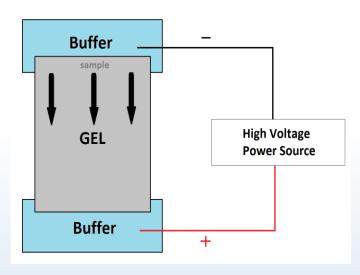
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Electrophoresis

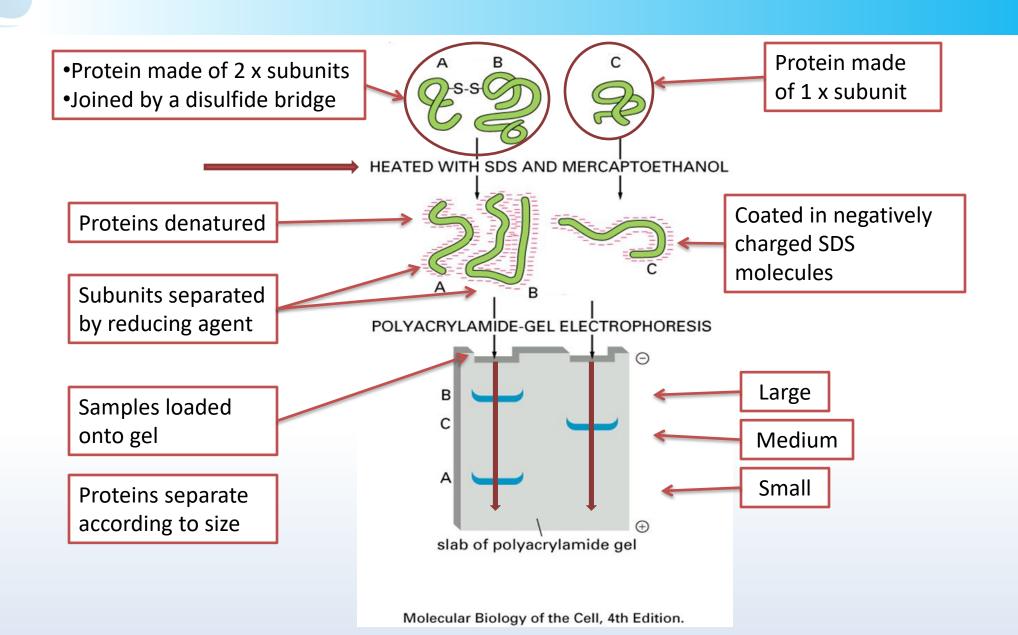
 Usually performed by placing sample in a non-charged gel through which an electrical current is applied

 The electrical charge and mass of the biomolecule affect its mobility through the gel during electrophoresis

- Can separate based on:
 - Size
 - Isoelectric point
 - Size:charge ratio

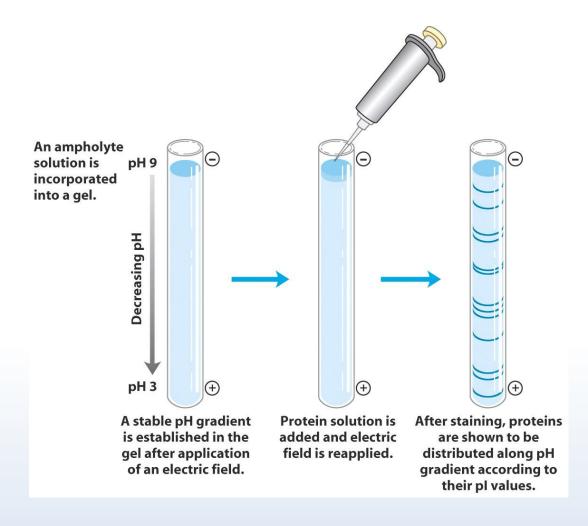


Performing SDS PAGE

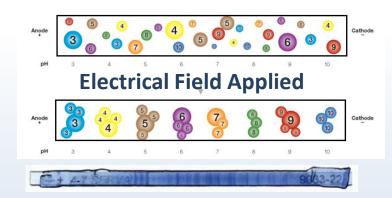


Isoelectric Focusing (IEF)

Protein mixture is loaded and electrophoresed until the proteins in the sample reach the pH that is equal to their pI.



- When the protein in the pH gradient reaches a zone in which the net surface charge is zero, it will no longer move.
- At this point the protein becomes "focused" and a band is formed in the gel.



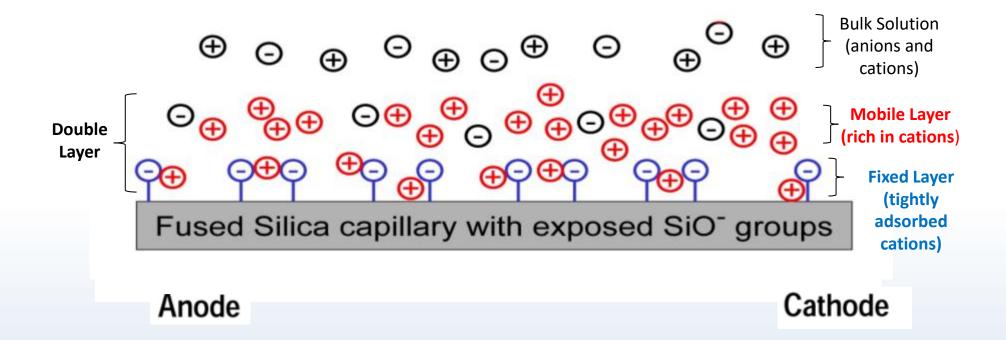
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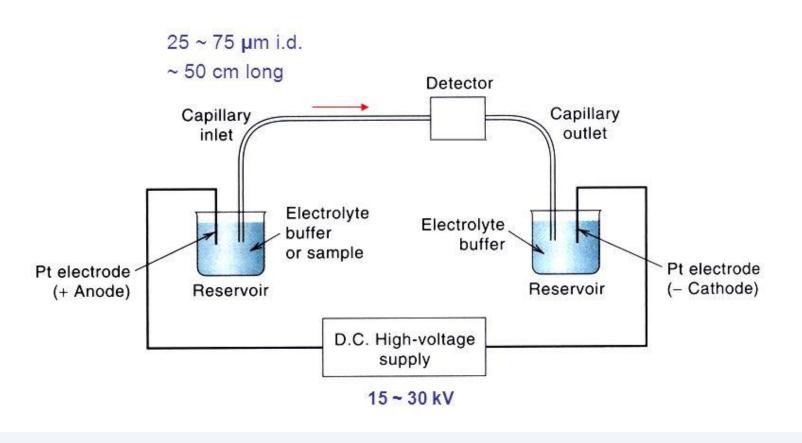


This first layer of cations (fixed layer) is not of sufficient density to totally neutralise the negative charges of the silanoate, so a second outer layer of cations forms (mobile layer)





CE Instrumentation

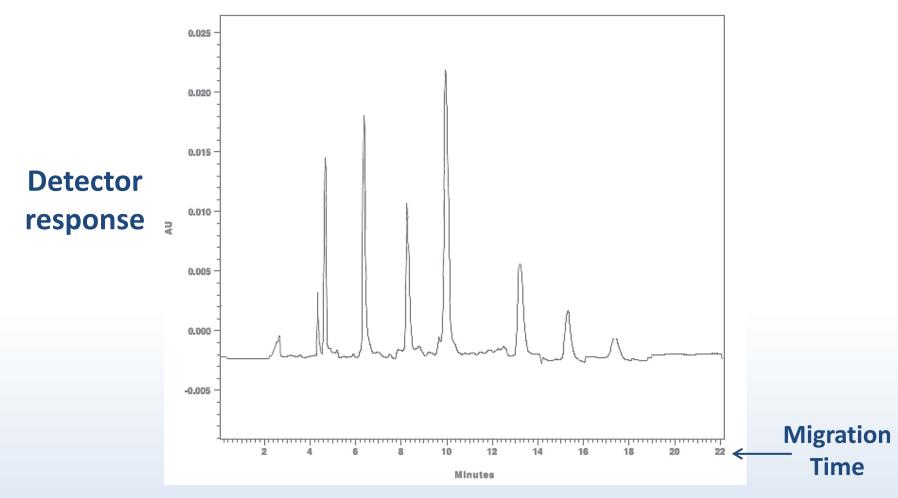


G. D. Christian, Analytical Chemistry, 6th ed., John Wiley, 2004, p. 632



CE Results

• Results are shown as an **electropherogram** (similar to a chromatogram in HPLC): time v absorbance/fluorescence units





CE separation techniques

 Several separation techniques can be used depending on the capillary and the electrolytes used

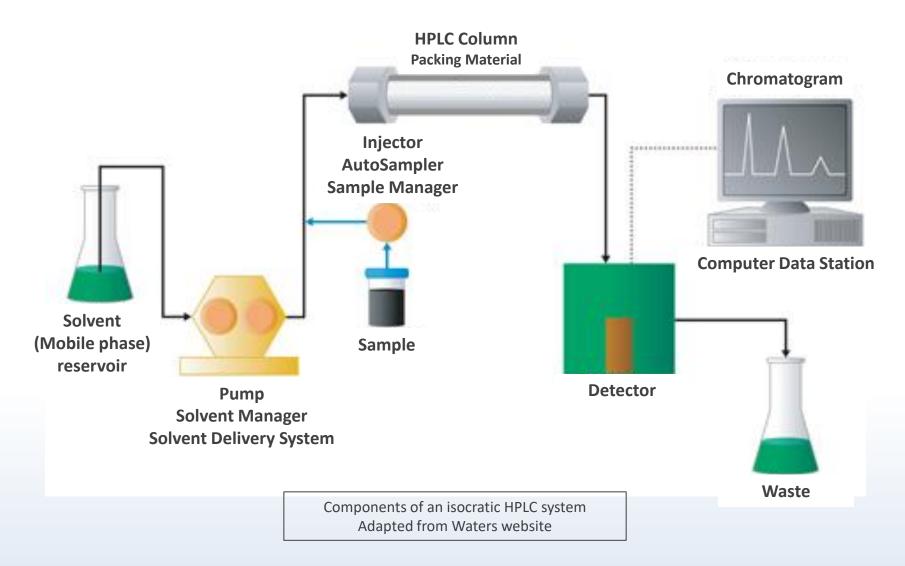
- Capillary Zone Electrophoresis (CZE): simplest form of CE, separation is by charge-to-mass ratio
- Capillary Gel Electrophoresis (CGE): separation occurs in a gel in the capillary e.g.
 polyacrylamide gel. SDS can be used to coat protein in negative charge so
 separation is by size
- Capillary iso-electric focusing (CIEF): separation is by <u>charge</u> & occurs along a pH gradient so proteins migrate depending on their isoelectric point



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HPLC Equipment



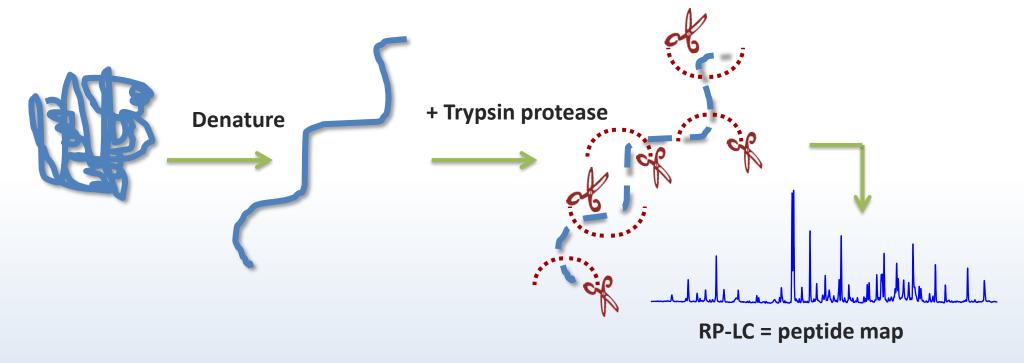


Key Analytical methods Chromatography

Isolation Method	Feature	
 Normal phase chromatography (NP-HPLC) Hydrophilicity 		
 Reversed phase chromatography (RP-HPLC) 	Hydrophobicity	
 Affinity chromatography 	Biorecognition	
 Size exclusion chromatography (SEC) 	Size	
 Ion exchange chromatography (IEX) 	Charge	
 Hydrophilic interaction chromatography (HILIC) 	Hydrophilicity	
	l	

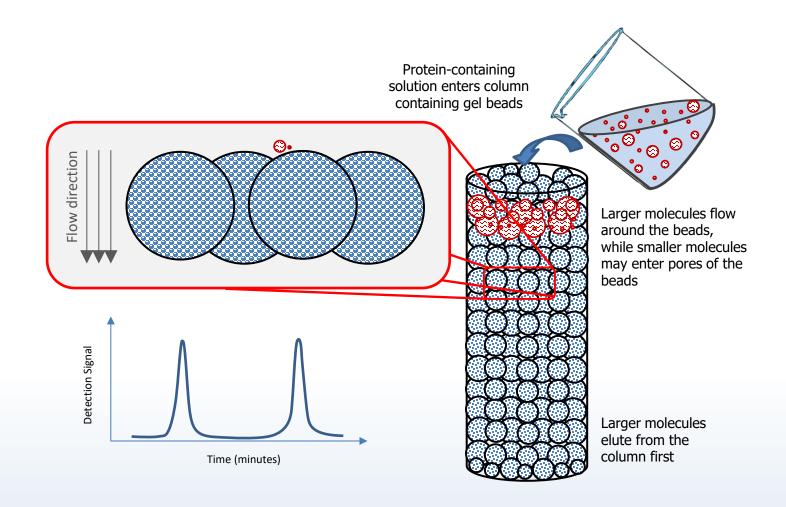


- **Analytical: Peptide mapping**
- Peptide mapping involves digesting a protein with a protease such as trypsin to create specific **peptides**
- Used to produce a unique 'fingerprint' of an individual protein when analysed by RP-LC





Size Exclusion Chromatography

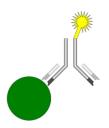




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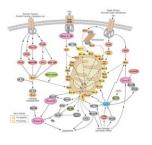
Biological Characterisation



Immunoassay

An immunoassay is a quantitative or qualitative analytical method that relies on the binding of an antibody to an antigen



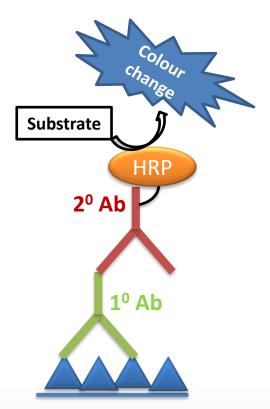


Bioassay

A bioassay is a quantitative analytical method that measures a defined biological effect in a living system, such as an intact cell.

Antibodies used in immunoassays

- The antibody that detects the antigen is called the primary antibody (1°)
- For detection of this interaction, a second antibody is required (the secondary antibody, 2°)
- The 2⁰ Ab is designed such that its binding partner is the 1⁰ Ab
- The 2⁰ Ab is labelled (conjugated) with an enzyme
 - E.g. horseradish peroxidase (HRP)
- Adding the substrate of the enzyme leads to a measurable response (e.g. colour change)
- Can also use a fluorophore (non enzymatic detection)





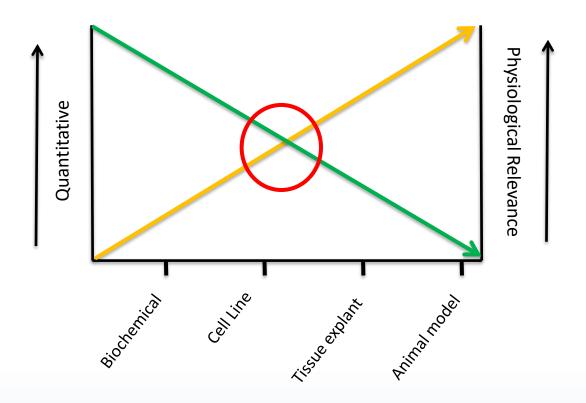


Western Blotting -v- ELISA

Western Blotting	ELISA
Immobilised antigen on membrane	Immobilised antigen on plastic plate
Qualitative and Semi-quantitative	Qualitative and Quantitative
Detection of protein of interest	Detection of protein of interest
Provides information on purity and MW of protein	Does not provide information on purity or MW of protein



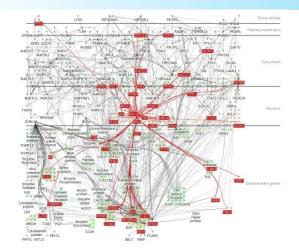
Types of Potency Assays



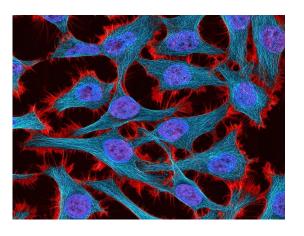
Cell based bioassays can be both quantitative and physiologically relevant and represent a good balance between the two extremes



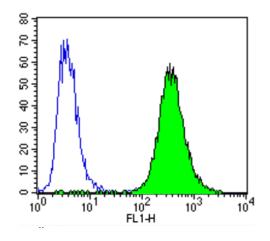
The Problem with Bioassays



1. Functional bioassays rely on the translation of complicated cellular biochemistry into an analytical read out.



2. Functional bioassays require the use of cultured or primary cells. How does in vitro reflect in vivo??



3. Analytical component dependent upon the performance of cells and biochemistry, does it answer the question posed?

Informative cell based functional bioassays require careful experimental design, optimization, validation and operation to generate consistent and reliable data.



Cell Based Bioassays (mAB examples)

Each Bioassay is unique

as it depends on the

drug and its MOA

- Cell signalling assay
- Proliferation assay
- Cytotoxicity assay
- Antibody dependant cell-mediated cytotoxicity (ADCC)
- Complement dependent cytotoxicity (CDC)

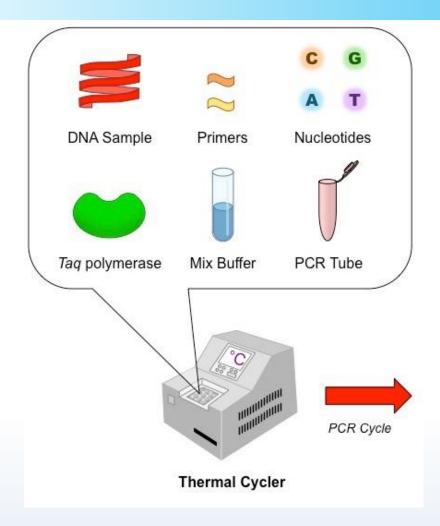
If the biopharmaceutical product has more than one biological activity, it will be necessary to measure all **functions**



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PCR Components







Thermal Cycling

Taq polymerase primers,

extends the synthesizing new strands of DNA.

Template Denaturation (96°C):

Causes melting of the DNA template by disrupting the hydrogen bonds between complementary bases, yielding single-stranded DNA molecules.

Primer **Extension** (72°C)



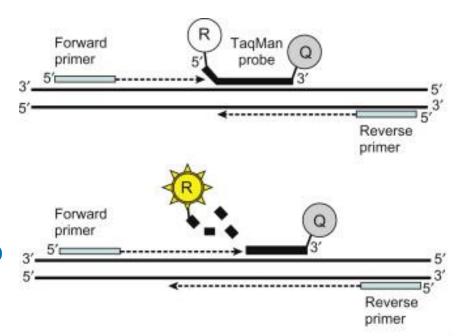
30-40x

Primer **Annealing** (55 - 65°C)

Primers bind to their complementary sequences on the singlestranded template DNA.



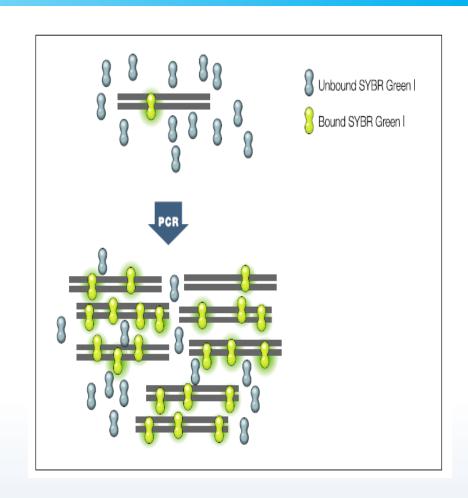
- Sequence-specific probe that anneals between the forward and reverse primers.
- Labelled with a reporter (R) and a quencher (Q) dye.
 - > R is usually a green dye (e.g. fluorescein)
 - Q is typically a red dye (e.g. TAMRA)
- When the probe is intact **Q** interferes with **R** little to no fluorescence.
- As the PCR primer extends, the probe is cleaved from the DNA strand
- Cleavage separates R and Q increase in fluorescence.





SYBR Green I

- SYBR Green I is a DNA dye that binds non-discriminately to double-stranded DNA (dsDNA).
- SYBR Green I exhibits minimal fluorescence when it is free in solution, but its fluorescence increases dramatically (up to 1000-fold) upon binding to dsDNA (new DNA)
- As more double stranded amplicons are produced,
 SYBR Green dye signal will increase respectively.





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LAQ Exam Format

- Open from 12pm on Wednesday 16th April (closes on Friday 18th April at 12pm)
- Time allowed = 1 hour 30min
- You will be given a choice of 4 questions and you only need to answer 3
- DO NOT attempt all 4 questions as I will only correct 3 per student



Example long answer questions for Final Exam

- An analyst wishes to use an ELISA method of detect for the presence of HCPs in a drug substance test sample. Discuss in your own words how this assay should be carried out. Include the following information in your answer:
 - 1. A description of the reagents and instrumentation required
 - 2. Describe how the assay is working



 All questions will focus on a specific technique but will draw on your knowledge of protein chemistry and general quality testing

 Answers should be written in your own language. Do not copy from other sources. Answers that are copied will receive zero marks

BRT ©



Thank You

