## ### GENE EXPRESSION ANALYSIS

#### # BACKGROUND FUNCTIONS

```
read.pc1 <- function(filename,na.type = "",make.rownames = TRUE,Nrows=</pre>
-1, Comment.char="",...) {
     x.df <- read.table(paste(filename, "pcl", sep="."), header=TRUE, sep="\t",</pre>
     quote="\"", as. is = rep(T, 3),
na.strings=na.type,skip=0,nrows=Nrows,comment.char=Comment.char,...);
     v <- x.df[[1]];
     ifelse(make.rownames,rownames(x.df)<-v,rownames(x.df)<-(1:\{length(v)\}));
return(x.df)};
write.pcl <- function(df,dataname,fileaddress="") {</pre>
      dir.address <- paste(fileaddress,dataname,".pcl",sep="");</pre>
      X <- write.table(df,file=dir.address,</pre>
append=FALSE, quote=FALSE, sep="\t", eol="\n", na="", row.names=FALSE, col.names=TR
UE);
return(X)};
11vec <- function(vec, na.rm = TRUE) {junk1 <- sum(abs(vec), na.rm =
na.rm);return(junk1)};
# MODIFIED SURVIVAL FUNCTIONS:
# group = FACTOR of the different classes of patients (for distinct KM curves
# survival.time & censoring.status = vectors
# survival.type = string for the title e.g. "metastasis" or "death" etc...
# p-value is obtained using log-rank test
m.plotsurvival <- function (group, survival.time, censoring.status, mark.time
= TRUE, my.colors,my.title = "",time.units = "",survival.type =
"",class.names = as.character(1:length(unique(group))),pv.legend = 0.25,
groups.legend = 0.2, x.legend = 0.1)
    require(survival)
    n.class <- length(unique(group))</pre>
    junk <- survfit(Surv(survival.time, censoring.status) ~ as.factor(group))</pre>
    junk2 <- coxph(Surv(survival.time, censoring.status) ~ as.factor(group))</pre>
    pv \leftarrow 1 - pchisq(2 * (junk2$loglik[2] - junk2$loglik[1]),
        df = n.class - 1)
    plot(junk, mark.time = mark.time, col = my.colors, xlab =
paste("Time",time.units), ylab = paste("Probability of
survival",survival.type),main = paste("KM survival",my.title))
    legend(x.legend * max(survival.time, na.rm = TRUE), groups.legend, col =
my.colors,
```

```
lty = rep(1, n.class), legend = class.names, bty = "n")
    text(x.legend * max(survival.time, na.rm = TRUE), pv.legend,
paste("pvalue=", as.character(round(pv,
        4))))
    return()
}
m.PVAL.survival <- function (group, survival.time, censoring.status)</pre>
{
    require(survival)
    n.class <- length(unique(group))</pre>
    junk2 <- coxph(Surv(survival.time, censoring.status) ~ as.factor(group))</pre>
    pv <- 1 - pchisq(2 * (junk2$loglik[2] - junk2$loglik[1]), df = n.class -</pre>
1)
    return(pv)
}
m.coxphCOEF.survival <- function (group, survival.time, censoring.status)</pre>
    require(survival)
    n.class <- length(unique(group))</pre>
    junk2 <- coxph(Surv(survival.time, censoring.status) ~ as.factor(group))</pre>
      junk3 <- junk2$coef;</pre>
    return(junk3)
}
sign0.mat <- function(mat, thr){junk0 <- mat; junk0[abs(junk0) < thr] <- NA;</pre>
junk1 = junk0>0; junk2 = 2*junk1 - 1; junk2[is.na(junk2)] <- 0;</pre>
return(junk2)};
  NKI ANALYSIS ON DSGA TRANSFORMED DATA.
# PERFORM THE DSGA ANALYSIS, ON THE BREAST CANCER DATA
(NKI tumor and BCN normal) AND PLACE THE OUTPUT IN A
DIRECTORY CALLED:
                 work/DSGA.NKI
      MAKE "work" the working directory.
```

```
# read non-DSGA-transformed gene expression data for
NKI

torg.pcl <- read.pcl("DSGA.NKI/NKI.normMesh");
norg.pcl <- read.pcl("DSGA.NKI/BCN.normMesh");

torg.mat <- as.matrix(torg.pcl[-(1:3)][-1,]);
norg.mat <- as.matrix(norg.pcl[-(1:3)][-1,]);</pre>
```

```
tumor.names <- colnames(torg.mat);</pre>
# read DSGA-transformed gene expression data for NKI
tdc.pcl <- read.pcl("DSGA.NKI/NKI.Tdis");</pre>
ndc.pcl <- read.pcl("DSGA.NKI/BCN.L1out");</pre>
tnc.pcl <- read.pcl("DSGA.NKI/NKI.Tnorm");</pre>
nnc.pcl <- read.pcl("DSGA.NKI/BCN.Nnorm");</pre>
tdc.mat <- as.matrix(tdc.pcl[-(1:3)][-1,]);
ndc.mat <- as.matrix(ndc.pcl[-(1:3)][-1,]);
tnc.mat <- as.matrix(tnc.pcl[-(1:3)][-1,]);</pre>
nnc.mat <- as.matrix(nnc.pcl[-(1:3)][-1,]);</pre>
# read basement-membrane (BM) gene list with Uniquene
cluster ID from current build and with NKI build 219
bm.df <-
read.table("List 2 BM protein.modifying enzymes list.tx
t", header=TRUE, sep="\t", quote="\"", as.is =
rep(T,3), na.strings=c("NA", "na", ""), skip=0);
v.hs <- as.vector(bm.df[["Hs.ID 219.NKI"]], mode =</pre>
"character");
bm.all.df <- bm.df[!is.na(v.hs),];</pre>
bm.true.df <- bm.all.df[bm.all.df[["BMgene.TF"]],];</pre>
id.vec <- rownames(tdc.mat);</pre>
# identify id's for ESR1 and ERBB2 for tumor placement
into molecular subtypes
esr1.id <- "Hs.208124"
erbb2.id <- "Hs.446352";
# tumor subtypes (ER-status and ERBB2 or her2 status)
are based on DSGA-transformed data: Disease component
for nki
esr.tdc.vec <- tdc.mat[esr1.id,];</pre>
```

```
erbb2.tdc.vec <- tdc.mat[erbb2.id,];</pre>
```

# # NKI stratification based on ESR1 status and her2 status in Disease component

```
hist(esr.tdc.vec, breaks=100, col="red", border="red3",
main="Histogram of all tumors ESR1\nNKI Disease
component", xlab="ESR1 Dc all tumors");
abline(v = -4, lty = "dashed", col = "red4");
legend(x = -4.4, y = 10, legend = "-4", bty = "n");
legend(x = -6.5, y = 9, legend = "ER negative", bty =
"n");
legend(x = -4.3, y = 9, legend = "ER positive", bty =
"n");
esr1.cut = -4
hist(erbb2.tdc.vec, breaks=100, col="purple",
border="purple3", main="Histogram of all tumors
her2\nNKI Disease component", xlab="her2 Dc all
tumors");
abline(v = 0.5, lty = "dashed", col = "purple4");
legend(x = 0, y = 19.5, legend = "0.5", bty = "n");
legend(x = -2.5, y = 17, legend = "her2 negative", bty
= "n");
legend(x = 0.5, y = 17, legend = "her2 positive", bty =
"n");
```

# tumor stratification based on Disease component of

her2.cut = 0.5

DSGA-transformed NKI data

```
basal.names <- tumor.names[{esr.tdc.vec < esr1.cut} &
{erbb2.tdc.vec < her2.cut}];
length(basal.names)</pre>
```

```
#[1] 49
her2.names <- tumor.names[{erbb2.tdc.vec > her2.cut}];
length(her2.names)
#[1] 52
luminal.A.names <- tumor.names[{esr.tdc.vec > esr1.cut}
& {erbb2.tdc.vec < her2.cut}];
length(luminal.A.names)
#[1] 194
luminal.B.names <- tumor.names[{esr.tdc.vec > esr1.cut}
& {erbb2.tdc.vec > her2.cut}];
length(luminal.B.names)
#[1] 30
her2.erN.names <- tumor.names[{esr.tdc.vec < esr1.cut}</pre>
& {erbb2.tdc.vec > her2.cut}];
length(her2.erN.names)
#[1] 22
erP.names <- tumor.names[{esr.tdc.vec > esr1.cut}];
length(erP.names)
#[1] 224
erN.names <- tumor.names[{esr.tdc.vec < esr1.cut}];</pre>
length(erN.names)
#[1] 71
# extract Basement Membrane genes only
rownames(bm.true.df) <-
as.vector(bm.true.df[["Hs.ID_219.NKI"]], mode =
"character");
V.hs <- rownames(bm.true.df);</pre>
V.gene <- as.vector(bm.true.df[["gene.symbol"]], mode =</pre>
```

```
"character");
names(V.hs) <- V.gene;

tdc.T.mat <- tdc.mat[V.hs,];
ndc.T.mat <- ndc.mat[V.hs,];
tnc.T.mat <- tnc.mat[V.hs,];
nnc.T.mat <- nnc.mat[V.hs,];
torg.T.mat <- torg.mat[V.hs,];
norg.T.mat <- norg.mat[V.hs,];</pre>
```

# # separate BM data matrices by breast cancer type:

```
tdc.basal.mat <- tdc.T.mat[,paste(basal.names,"Dis",sep</pre>
= ".")];
tdc.luminal.A.mat <-
tdc.T.mat[,paste(luminal.A.names, "Dis", sep = ".")];
tdc.luminal.B.mat <-
tdc.T.mat[,paste(luminal.B.names, "Dis", sep = ".")];
tdc.her2.mat <- tdc.T.mat[,paste(her2.names,"Dis",sep =</pre>
".")1;
tdc.her2.erN.mat <-
tdc.T.mat[,paste(her2.erN.names,"Dis",sep = ".")];
tdc.erP.mat <- tdc.T.mat[,paste(erP.names,"Dis",sep =</pre>
".")1;
tdc.erN.mat <- tdc.T.mat[,paste(erN.names,"Dis",sep =</pre>
".")1;
tnc.basal.mat <-</pre>
tnc.T.mat[,paste(basal.names, "Norm", sep = ".")];
tnc.luminal.A.mat <-</pre>
tnc.T.mat[,paste(luminal.A.names, "Norm", sep = ".")];
tnc.luminal.B.mat <-</pre>
tnc.T.mat[,paste(luminal.B.names, "Norm", sep = ".")];
tnc.her2.mat <- tnc.T.mat[,paste(her2.names, "Norm", sep</pre>
= ".")];
tnc.her2.erN.mat <-</pre>
```

```
tnc.T.mat[,paste(her2.erN.names,"Norm",sep = ".")];
tnc.erP.mat <- tnc.T.mat[,paste(erP.names,"Norm",sep =
".")];
tnc.erN.mat <- tnc.T.mat[,paste(erN.names,"Norm",sep =
".")];

torg.basal.mat <- torg.T.mat[,basal.names];
torg.luminal.A.mat <- torg.T.mat[,luminal.A.names];
torg.luminal.B.mat <- torg.T.mat[,luminal.B.names];
torg.her2.mat <- torg.T.mat[,her2.names];
torg.her2.erN.mat <- torg.T.mat[,her2.erN.names];
torg.erP.mat <- torg.T.mat[,erP.names];
torg.erN.mat <- torg.T.mat[,erP.names];</pre>
```

#### # read in clinical data

```
clin.df <-
read.table("NKI.Clinical Data Supplement.txt", header=TR
UE,sep="\t",quote="\"");
rownames(clin.df) <- paste("SAMPLE",clin.df[["ID"]],sep</pre>
= ".")
surv.time.death <-</pre>
as.vector(clin.df[["survival.death."]], mode =
"numeric");
names(surv.time.death) <- rownames(clin.df);</pre>
censor.death <- as.vector(clin.df[["event death"]],</pre>
mode = "numeric");
names(censor.death) <- rownames(clin.df);</pre>
surv.time.met <-</pre>
as.vector(clin.df[["Follow up time or metastasis"]],
mode = "numeric");
names(surv.time.met) <- rownames(clin.df);</pre>
censor.met <- as.vector(clin.df[["event metastasis"]],</pre>
mode = "numeric");
names(censor.met) <- rownames(clin.df);</pre>
```

```
list.names <- list(basal.names, luminal.A.names,</pre>
luminal.B.names, her2.names, her2.erN.names, erP.names,
erN.names);
names(list.names) <- c("Basal", "Lum.A", "Lum.B",</pre>
"Her2", "Her2.ESRneg", "ERpos", "ERneg");
add.dis <- function(vec){return(paste(vec, "Dis", sep =
"."))};
add.norm <- function(vec){return(paste(vec, "Norm", sep</pre>
= "."))};
list.dc.names <- lapply(list.names,add.dis);</pre>
names(list.dc.names) <- paste(names(list.names), "Dc",</pre>
sep = ".");
list.nc.names <- lapply(list.names,add.norm);</pre>
names(list.nc.names) <- paste(names(list.names), "Nc",</pre>
sep = ".");
list.org.names <- list.names;</pre>
names(list.org.names) <- names(list.names);</pre>
library(survival)
pval.death.Dc.mat <- matrix(data = NA, nrow =</pre>
length(V.gene), ncol = 7, dimnames =
list(V.gene,names(list.names)));
pval.death.Nc.mat <- matrix(data = NA, nrow =</pre>
length(V.gene), ncol = 7, dimnames =
list(V.gene,names(list.names)));
pval.death.Org.mat <- matrix(data = NA, nrow =</pre>
length(V.gene), ncol = 7, dimnames =
list(V.gene,names(list.names)));
coef.death.Dc.mat <- matrix(data = NA, nrow =</pre>
```

```
length(V.gene), ncol = 7, dimnames =
list(V.gene,names(list.names)));
coef.death.Nc.mat <- matrix(data = NA, nrow =</pre>
length(V.gene), ncol = 7, dimnames =
list(V.gene,names(list.names)));
coef.death.Org.mat <- matrix(data = NA, nrow =</pre>
length(V.gene), ncol = 7, dimnames =
list(V.gene,names(list.names)));
pval.met.Dc.mat <- matrix(data = NA, nrow =</pre>
length(V.gene), ncol = 7, dimnames =
list(V.gene,names(list.names)));
pval.met.Nc.mat <- matrix(data = NA, nrow =</pre>
length(V.gene), ncol = 7, dimnames =
list(V.gene,names(list.names)));
pval.met.Org.mat <- matrix(data = NA, nrow =</pre>
length(V.gene), ncol = 7, dimnames =
list(V.gene,names(list.names)));
coef.met.Dc.mat <- matrix(data = NA, nrow =</pre>
length(V.gene), ncol = 7, dimnames =
list(V.gene,names(list.names)));
coef.met.Nc.mat <- matrix(data = NA, nrow =</pre>
length(V.gene), ncol = 7, dimnames =
list(V.gene,names(list.names)));
coef.met.Org.mat <- matrix(data = NA, nrow =</pre>
length(V.gene), ncol = 7, dimnames =
list(V.gene,names(list.names)));
for(i in (1:length(V.gene))){
    for(j in (1:7)){
         G.dc.vec <- tdc.T.mat[i,list.dc.names[[j]]];</pre>
    q.G.dc.33.67 <- quantile(G.dc.vec, probs = c(0,
0.33, 0.67, 1));
group3.dc.33.67 \leftarrow cut(G.dc.vec, breaks = q.G.dc.33.67,
labels = (1:3), include.lowest = TRUE);
v.G.dc.33.67.tmp <- as.vector(group3.dc.33.67, mode =
"character")
```

```
group.dc.33.67.f <- as.factor(v.G.dc.33.67.tmp[!
{v.G.dc.33.67.tmp %in% c(2)}]);
         G.nc.vec <- tnc.T.mat[i,list.nc.names[[j]]];</pre>
    q.G.nc.33.67 <- quantile(G.nc.vec, probs = c(0,
0.33, 0.67, 1);
qroup3.nc.33.67 < - cut(G.nc.vec, breaks = q.G.nc.33.67,
labels = (1:3), include.lowest = TRUE);
v.G.nc.33.67.tmp < - as.vector(qroup3.nc.33.67, mode =
"character")
group.nc.33.67.f <- as.factor(v.G.nc.33.67.tmp[!
\{v.G.nc.33.67.tmp %in% c(2)\}\};
         G.org.vec <- torg.T.mat[i,list.names[[j]]];</pre>
    q.G.org.33.67 <- quantile(G.org.vec, probs = c(0,
0.33, 0.67, 1);
group3.org.33.67 <- cut(G.org.vec, breaks =</pre>
q.G.org.33.67, labels = (1:3), include.lowest = TRUE);
v.G.org.33.67.tmp <- as.vector(group3.org.33.67, mode =
"character")
group.org.33.67.f <- as.factor(v.G.org.33.67.tmp[!</pre>
{v.G.org.33.67.tmp %in% c(2)}]);
surv.time.death.G <- surv.time.death[list.names[[j]]];</pre>
SURV.time.death.dc 3367 <- surv.time.death.G[!
{v.G.dc.33.67.tmp %in% c(2)}];
SURV.time.death.nc 3367 <- surv.time.death.G[!
{v.G.nc.33.67.tmp %in% c(2)}];
SURV.time.death.org 3367 <- surv.time.death.G[!
{v.G.org.33.67.tmp %in% c(2)}];
censor.death.G <- censor.death[list.names[[j]]]</pre>
CENSOR.death.dc 3367 <- censor.death.G[!
{v.G.dc.33.67.tmp %in% c(2)}];
CENSOR.death.nc 3367 <- censor.death.G[!
{v.G.nc.33.67.tmp %in% c(2)}];
CENSOR.death.org 3367 <- censor.death.G[!
{v.G.org.33.67.tmp %in% c(2)}];
```

```
pval.death.Dc.mat[i,j] <- m.PVAL.survival(group =</pre>
group.dc.33.67.f, survival.time =
SURV.time.death.dc 3367, censoring.status =
CENSOR.death.dc 3367)
pval.death.Nc.mat[i,j] <- m.PVAL.survival(group =</pre>
group.nc.33.67.f, survival.time =
SURV.time.death.nc 3367, censoring.status =
CENSOR.death.nc 3367)
pval.death.Org.mat[i,j] <- m.PVAL.survival(group =</pre>
group.org.33.67.f, survival.time =
SURV.time.death.org 3367, censoring.status =
CENSOR.death.org 3367)
coef.death.Dc.mat[i,j] <- m.coxphCOEF.survival(group =</pre>
group.dc.33.67.f, survival.time =
SURV.time.death.dc 3367, censoring.status =
CENSOR.death.dc 3367)
coef.death.Nc.mat[i,j] <- m.coxphCOEF.survival(group =</pre>
group.nc.33.67.f, survival.time =
SURV.time.death.nc 3367, censoring.status =
CENSOR.death.nc 3367)
coef.death.Org.mat[i,j] <- m.coxphCOEF.survival(group =</pre>
group.org.33.67.f, survival.time =
SURV.time.death.org 3367, censoring.status =
CENSOR.death.org 3367)
surv.time.met.G <- surv.time.met[list.names[[j]]];</pre>
SURV.time.met.dc 3367 <- surv.time.met.G[!
{v.G.dc.33.67.tmp %in% c(2)}];
SURV.time.met.nc 3367 <- surv.time.met.G[!
{v.G.nc.33.67.tmp %in% c(2)}];
SURV.time.met.org 3367 <- surv.time.met.G[!
{v.G.org.33.67.tmp %in% c(2)}];
censor.met.G <- censor.met[list.names[[j]]]</pre>
CENSOR.met.dc 3367 <- censor.met.G[!{v.G.dc.33.67.tmp
%in% c(2)}];
CENSOR.met.nc 3367 <- censor.met.G[!{v.G.nc.33.67.tmp
```

```
%in% c(2)}];
CENSOR.met.org 3367 <- censor.met.G[!{v.G.org.33.67.tmp
%in% c(2)}];
pval.met.Dc.mat[i,j] <- m.PVAL.survival(group =</pre>
group.dc.33.67.f, survival.time =
SURV.time.met.dc 3367, censoring.status =
CENSOR.met.dc 3367)
pval.met.Nc.mat[i,j] <- m.PVAL.survival(group =</pre>
group.nc.33.67.f, survival.time =
SURV.time.met.nc 3367, censoring.status =
CENSOR.met.nc 3367)
pval.met.Org.mat[i,j] <- m.PVAL.survival(group =</pre>
group.org.33.67.f, survival.time =
SURV.time.met.org 3367, censoring.status =
CENSOR.met.org 3367)
coef.met.Dc.mat[i,j] <- m.coxphCOEF.survival(group =</pre>
group.dc.33.67.f, survival.time =
SURV.time.met.dc 3367, censoring.status =
CENSOR.met.dc 3367)
coef.met.Nc.mat[i,j] <- m.coxphCOEF.survival(group =</pre>
group.nc.33.67.f, survival.time =
SURV.time.met.nc_3367, censoring.status =
CENSOR.met.nc 3367)
coef.met.Org.mat[i,j] <- m.coxphCOEF.survival(group =</pre>
group.org.33.67.f, survival.time =
SURV.time.met.org_ 3367, censoring.status =
CENSOR.met.org 3367)
}};
# Figure 1b & 1c
# dir.create("extras")
```

i = 23; # NTN4

```
j = 6; # ER positive
```

```
G.nc.vec <- tnc.T.mat[i,list.nc.names[[j]]];</pre>
    q.G.nc.33.67 <- quantile(G.nc.vec, probs = c(0,
0.33, 0.67, 1);
group3.nc.33.67 \leftarrow cut(G.nc.vec, breaks = g.G.nc.33.67,
labels = (1:3), include.lowest = TRUE);
v.G.nc.33.67.tmp < - as.vector(qroup3.nc.33.67, mode =
"character")
group.nc.33.67.f \le as.factor(v.G.nc.33.67.tmp[!]
{v.G.nc.33.67.tmp %in% c(2)}]);
surv.time.death.G <- surv.time.death[list.names[[j]]];</pre>
SURV.time.death.nc 3367 <- surv.time.death.G[!
{v.G.nc.33.67.tmp %in% c(2)}];
censor.death.G <- censor.death[list.names[[j]]]</pre>
CENSOR.death.nc 3367 <- censor.death.G[!</pre>
{v.G.nc.33.67.tmp %in% c(2)}];
m.plotsurvival(group = group.nc.33.67.f, survival.time
= SURV.time.death.nc 3367,
censoring.status = CENSOR.death.nc 3367, my.colors =
c("blue", "red"), survival.type = "death", class.names
= c("low NTN4", "high NTN4"),
my.title = paste(names(list.nc.names)[[j]], " group\nNKI
33% 67% separation of", V.gene[[i]], "levels"));
group.nc.33.67.vec <- as.vector(group.nc.33.67.f, mode</pre>
= "character");
ERp.survival.grouplo.NTN4 <-</pre>
SURV.time.death.nc 3367[group.nc.33.67.vec %in%
c("1")];
ERp.survival.grouphi.NTN4 <-</pre>
SURV.time.death.nc 3367[group.nc.33.67.vec %in%
c("3")];
```

```
ERp.censor.grouplo.NTN4 <-</pre>
CENSOR.death.nc 3367[group.nc.33.67.vec %in% c("1")];
ERp.censor.grouphi.NTN4 <-</pre>
CENSOR.death.nc 3367[group.nc.33.67.vec %in% c("3")];
km.ERp.df <- data.frame("Sample" =</pre>
c(names(ERp.survival.grouplo.NTN4), names(ERp.survival.g
rouphi.NTN4)),
"group" = c(rep("lo.NTN4",
length(ERp.survival.grouplo.NTN4)), rep("hi.NTN4",
length(ERp.survival.grouphi.NTN4))),
"time" = c(ERp.survival.grouplo.NTN4,
ERp.survival.grouphi.NTN4),
"censor" = c(ERp.censor.grouplo.NTN4,
ERp.censor.grouphi.NTN4));
# write.table(km.ERp.df,file="extras/figure1b.txt",
append=FALSE, quote=FALSE, sep="\t", eol="\n", na="", row.na
mes=FALSE,col.names=TRUE);
j = 7; # ER negative
         G.nc.vec <- tnc.T.mat[i,list.nc.names[[j]]];</pre>
    q.G.nc.33.67 < - quantile(G.nc.vec, probs = c(0,
0.33, 0.67, 1);
group3.nc.33.67 \leftarrow cut(G.nc.vec, breaks = q.G.nc.33.67,
labels = (1:3), include.lowest = TRUE);
v.G.nc.33.67.tmp <- as.vector(group3.nc.33.67, mode =
"character")
group.nc.33.67.f \le as.factor(v.G.nc.33.67.tmp[!]
{v.G.nc.33.67.tmp %in% c(2)}]);
surv.time.death.G <- surv.time.death[list.names[[j]]];</pre>
SURV.time.death.nc 3367 <- surv.time.death.G[!
{v.G.nc.33.67.tmp %in% c(2)}];
```

```
censor.death.G <- censor.death[list.names[[j]]]</pre>
CENSOR.death.nc 3367 <- censor.death.G[!
{v.G.nc.33.67.tmp %in% c(2)}];
m.plotsurvival(group = group.nc.33.67.f, survival.time
= SURV.time.death.nc 3367,
censoring.status = CENSOR.death.nc 3367, my.colors =
c("blue", "red"), survival.type = "death", class.names
= c("low NTN4", "high NTN4"),
my.title = paste(names(list.nc.names)[[j]], " group\nNKI
33% 67% separation of", V.gene[[i]], "levels"));
group.nc.33.67.vec <- as.vector(group.nc.33.67.f, mode</pre>
= "character");
ERn.survival.grouplo.NTN4 <-
SURV.time.death.nc 3367[group.nc.33.67.vec %in%
c("1")];
ERn.survival.grouphi.NTN4 <-</pre>
SURV.time.death.nc 3367[group.nc.33.67.vec %in%
c("3")];
ERn.censor.grouplo.NTN4 <-
CENSOR.death.nc 3367[group.nc.33.67.vec %in% c("1")];
ERn.censor.grouphi.NTN4 <-
CENSOR.death.nc 3367[group.nc.33.67.vec %in% c("3")];
km.ERn.df <- data.frame("Sample" =</pre>
c(names(ERn.survival.grouplo.NTN4), names(ERn.survival.g
rouphi.NTN4)),
"group" = c(rep("lo.NTN4",
length(ERn.survival.grouplo.NTN4)), rep("hi.NTN4",
length(ERn.survival.grouphi.NTN4))),
"time" = c(ERn.survival.grouplo.NTN4,
ERn.survival.grouphi.NTN4),
"censor" = c(ERn.censor.grouplo.NTN4,
ERn.censor.grouphi.NTN4));
```

```
# write.table(km.ERn.df,file="extras/figure1c.txt",
append=FALSE, quote=FALSE, sep="\t", eol="\n", na="", row.na
mes=FALSE,col.names=TRUE);
# compute statistics and consensus statistics
# take sign of the coefficient matrix, with anything
whose abs.val < 0.1 \longrightarrow 0
sign0.death.Dc.1.mat <- sign0.mat(mat =
coef.death.Dc.mat, thr = 0.01);
sign0.death.Nc.1.mat <- sign0.mat(mat =
coef.death.Nc.mat, thr = 0.01);
sign0.death.Org.1.mat <- sign0.mat(mat =</pre>
coef.death.Nc.mat, thr = 0.01);
sign0.met.Dc.1.mat <- sign0.mat(mat = coef.met.Dc.mat,
thr = 0.01);
sign0.met.Nc.1.mat <- sign0.mat(mat = coef.met.Nc.mat,
thr = 0.01);
sign0.met.Org.1.mat <- sign0.mat(mat = coef.met.Nc.mat,
thr = 0.01);
# log10, then turn 0 anything with pval > threshold
signO.death.Dc.mat <- signO.death.Dc.1.mat *
log10(pval.death.Dc.mat);
signO.death.Nc.mat <- signO.death.Nc.1.mat *
log10(pval.death.Nc.mat);
signO.death.Org.mat <- signO.death.Org.1.mat *</pre>
log10(pval.death.Org.mat);
sign0.met.Dc.mat <- sign0.met.Dc.1.mat *
```

log10(pval.met.Dc.mat);

```
sign0.met.Nc.mat <- sign0.met.Nc.1.mat *
log10(pval.met.Nc.mat);
sign0.met.Org.mat <- sign0.met.Org.1.mat *</pre>
log10(pval.met.Org.mat);
signed.sep.mean.Nc.vec <- apply(X =
cbind(sign0.death.Nc.mat, sign0.met.Nc.mat),
MARGIN=1,FUN=mean);
signed.sep.mean.Dc.vec <- apply(X =
cbind(sign0.death.Dc.mat,sign0.met.Dc.mat),
MARGIN=1,FUN=mean);
signed.sep.mean.Org.vec <- apply(X =
cbind(sign0.death.Org.mat, sign0.met.Org.mat),
MARGIN=1,FUN=mean);
signed.sep.death.mean.Nc.vec <- apply(X =
signO.death.Nc.mat, MARGIN=1,FUN=mean);
#ol.separation.vec gives L1-norm of log10 p-values for
all molecular subtypes & death & Disease comp. & Normal
comp & Original data
ol.separation.death.Nc.vec <-
apply(log10(pval.death.Nc.mat), 1, l1vec);
    names(o1.separation.death.Nc.vec) <- V.gene;</pre>
ol.separation.death.Dc.vec <-
apply(log10(pval.death.Dc.mat), 1, l1vec);
    names(o1.separation.death.Dc.vec) <- V.gene;</pre>
ol.separation.death.Org.vec <-
apply(log10(pval.death.Org.mat), 1, l1vec);
    names(o1.separation.death.Org.vec) <- V.gene;</pre>
ol.separation.met.Nc.vec <-
apply(log10(pval.met.Nc.mat), 1, l1vec);
    names(o1.separation.met.Nc.vec) <- V.gene;</pre>
```

```
ol.separation.met.Dc.vec <-
apply(log10(pval.met.Dc.mat), 1, l1vec);
    names(ol.separation.met.Dc.vec) <- V.gene;

ol.separation.met.Org.vec <-
apply(log10(pval.met.Org.mat), 1, l1vec);
    names(ol.separation.met.Org.vec) <- V.gene;

ol.separation.Nc.vec <- ol.separation.death.Nc.vec +
ol.separation.met.Nc.vec;

ol.separation.Dc.vec <- ol.separation.death.Dc.vec +
ol.separation.met.Dc.vec;

ol.separation.org.vec <- ol.separation.death.Org.vec +
ol.separation.org.vec <- ol.separation.death.Org.vec +
ol.separation.org.vec;</pre>
```

# # extended data figure 1a

```
plot(x =
c(signed.sep.mean.Nc.vec, signed.sep.mean.Dc.vec, signed.
sep.mean.Org.vec, rep(-2.8, 3)),
y = c(o1.separation.Nc.vec,
o1.separation.Dc.vec, o1.separation.Org.vec, 10,8,6),
main = "Comparison of overall association (L1) vs.
consensus signed association (mean)\nNormal comp. vs.
Disease comp. vs. Original data, death, met",
xlab = "consensus signed association (mean)", ylab =
"overall association L1", pch =
c(rep(19,35),rep(20,70),19,20,20),
col = c(rep("black",35), rep("red", 35), rep("blue",
35), "black", "red", "blue"))

legend(x = -2.8, y = 11, legend = "Normal component",
cex = 0.7,bty = "n");
```

```
legend(x = -2.8, y = 9, legend = "Disease component", cex = 0.7, bty = "n"); legend(x = -2.8, y = 7, legend = "Original data, no DSGA", cex = 0.7, bty = "n");
```

```
e_figurela.df <- data.frame("GENE" =
rownames(e_figurela.mat),
    "consensus.signed.association" =
c(signed.sep.mean.Nc.vec,signed.sep.mean.Dc.vec,signed.
sep.mean.Org.vec),
    "overall.association.L1" = c(o1.separation.Nc.vec,
o1.separation.Dc.vec,o1.separation.Org.vec));

# write.table(e_figurela.df,file="extras/
Efigurela.txt",
# append=FALSE,quote=FALSE,sep="\t",eol="\n",na="",row.na
mes=FALSE,col.names=TRUE);</pre>
```

# # Figure 1a

```
signed.sep.Nc.Dc.vec <- apply(X =
cbind(sign0.death.Nc.mat,sign0.death.Dc.mat,sign0.met.N
c.mat,sign0.met.Dc.mat), MARGIN=1,FUN=mean);
ol.separation.Nc.Dc.vec <- ol.separation.death.Nc.vec +
ol.separation.met.Nc.vec + ol.separation.death.Dc.vec +
ol.separation.met.Dc.vec

plot(x = signed.sep.Nc.Dc.vec, y =
ol.separation.Nc.Dc.vec,
main = "Comparison of overall association (L1) vs.
consensus signed association (mean)\ncombined Normal
comp. Disease comp. death, met",
xlab = "consensus signed association (mean)", ylab =</pre>
```

```
"overall association L1", pch = 19,
col = "gray");
figure1a.df <- data.frame("GENE" =
names(signed.sep.Nc.Dc.vec),
 "consensus.signed.association NcDc.met.death" =
signed.sep.Nc.Dc.vec,
 "overall.association.L1 NcDc.met.death" =
ol.separation.Nc.Dc.vec);
# write.table(figure1a.df,file="extras/figure1a.txt",
append=FALSE, quote=FALSE, sep="\t", eol="\n", na="", row.na
mes=FALSE,col.names=TRUE);
# COMPUTING CORRELATIONS OF BASEMENT MEMBRANE GENES TO
NTN4 IN BREAST, AND COMPARING TO OVARIAN CANCER
# compute correlation to NTN4 of all BM genes r.value
and p.value
ntnc.T.mat <- cbind(nnc.T.mat, tnc.T.mat);</pre>
# correlation plot between NKI.Nc basement membrane
genes correlation to NTN4 and OC basement membrane
correlation to NTN4
get.cor.p <- function(x,y){j = cor.test(x,y,</pre>
alternative = "two.sided", method = "pearson"); CC =
j$estimate; PP = j$p.value;
J \leftarrow c(CC,PP); names(J) = c("cor", "p.val");
return(J)};
ntn4.id <- "Hs.201034";
```

```
cor.NTN4.ntnc <- t(apply(ntnc.T.mat,MARGIN = 1, FUN =</pre>
get.cor.p, y = ntnc.T.mat[ntn4.id,]));
rownames(cor.NTN4.ntnc) <- names(V.hs);</pre>
cor2ntn4.bc.oc.df <-</pre>
read.table("SupplementaryTable.5 N31 BM.mechanics.txt",
header=TRUE, sep="\t", quote="\"", na.strings=c("NA"),
skip=0);
rownames(cor2ntn4.bc.oc.df) <-
as.vector(cor2ntn4.bc.oc.df[[1]], mode = "character");
cor2ntn4.bc.oc.mat <- as.matrix(cor2ntn4.bc.oc.df[-</pre>
c(1));
tf.vec <- {!is.na(cor2ntn4.bc.oc.mat[,1])} & {!
is.na(cor2ntn4.bc.oc.mat[,3])}
small.bc.oc.mat <- cor2ntn4.bc.oc.mat[tf.vec,]</pre>
library("GmicR");
library("WGCNA")
```

### # EXTENDED DATA FIGURE 9 d

```
verboseScatterplot(x = small.bc.oc.mat[,3], y =
small.bc.oc.mat[,1], xlab = "OC cor to NTN4", ylab =
"NKI.Nc cor to NTN4", abline = TRUE,
xlim = range(-1,1), ylim = range(-1,1), pch = 20, col =
"blue", main = "Correlation of BM genes to NTN4 in
\novarian cancer vs. breast cancer normal component\n",
cex.axis = 1, cex.lab = 1, cex.main = 1);
```

```
e_figure_9d.df <- data.frame("Gene" =
rownames(small.bc.oc.mat), "cor.NKI.ntNc" =
small.bc.oc.mat[,1], "cor.OC" = small.bc.oc.mat[,3]);</pre>
```

```
# write.table(e figure 9d.df,file="extras/
Efigure9d.txt",
append=FALSE, quote=FALSE, sep="\t", eol="\n", na="", row.na
mes=FALSE,col.names=TRUE);
     #. DONE NKI
  RENAL CANCER ANALYSIS OF NTN4 ASSOCIATION
WITH SURVIVAL
# RENAL CANCER ANALYSIS ON DSGA TRANSFORMED DATA.
# PERFORM THE DSGA ANALYSIS, ON THE RENAL CANCER DATA
(RTum tumor data & RNorm normal data) AND PLACE THE
OUTPUT IN A DIRECTORY CALLED:
             work/DSGA.Renal
    MAKE "work" the working directory.
# read DSGA-transformed gene expression data for NKI
tdc.pcl <- read.pcl("DSGA.Renal/RTum.Tdis");</pre>
ndc.pcl <- read.pcl("DSGA.Renal/RNorm.Llout");</pre>
tnc.pcl <- read.pcl("DSGA.Renal/RTum.Tnorm");</pre>
nnc.pcl <- read.pcl("DSGA.Renal/RNorm.Nnorm");</pre>
tnc.mat <- as.matrix(tnc.pcl[-(1:3)]);</pre>
tumor.names <- sub( pattern = ".Norm", replacement =</pre>
```

```
"", x = colnames(tnc.mat))
NTN4.id <- "IMAGE:143661"
# read in clinical data
clin.df <-
read.table("cRCC.clinical.information.clean.txt", header
=TRUE, sep="\t", quote="\"");
rownames(clin.df) <- as.vector(clin.df[[1]], mode =
"character");
Clin.df <- clin.df[tumor.names,];</pre>
surv.time <- as.vector(Clin.df[["SurvivalMonths"]],</pre>
mode = "numeric");
names(surv.time) <- rownames(Clin.df);</pre>
censor.death <- as.vector(Clin.df[["Event.Death"]],</pre>
mode = "numeric");
names(censor.death) <- rownames(Clin.df);</pre>
censor.met <- as.vector(Clin.df[["EventRecur"]], mode =</pre>
"numeric");
names(censor.met) <- rownames(Clin.df);</pre>
library(survival)
         G.nc.vec <- tnc.mat[NTN4.id,];</pre>
    q.G.nc.33.67 < - quantile(G.nc.vec, probs = c(0,
0.33, 0.67, 1);
group3.nc.33.67 < - cut(G.nc.vec, breaks = q.G.nc.33.67,
labels = (1:3), include.lowest = TRUE);
v.G.nc.33.67.tmp <- as.vector(group3.nc.33.67, mode =
"character")
group.nc.33.67.f <- as.factor(v.G.nc.33.67.tmp[!
{v.G.nc.33.67.tmp %in% c(2)}]);
SURV.time 3367 <- surv.time[!{v.G.nc.33.67.tmp %in%
```

```
c(2)}];
CENSOR.death.nc 3367 <- censor.death[!{v.G.nc.33.67.tmp
%in% c(2)}];
CENSOR.met.nc 3367 <- censor.met[!{v.G.nc.33.67.tmp
%in% c(2)}];
m.plotsurvival(group = group.nc.33.67.f, survival.time
= SURV.time 3367,
censoring.status = CENSOR.death.nc 3367, my.colors =
c("blue", "red"), survival.type = "death", class.names
= c("low NTN4", "high NTN4"),
my.title = paste("Renal cancer death\nNKI 33% 67%
separation of", "NTN4 in Normal component", "levels"));
m.plotsurvival(group = group.nc.33.67.f, survival.time
= SURV.time 3367,
censoring.status = CENSOR.met.nc 3367, my.colors =
c("blue", "red"), survival.type = "met", class.names =
c("low NTN4", "high NTN4"),
my.title = paste("Renal cancer metastasis\nNKI 33% 67%
separation of", "NTN4 in Normal component", "levels"));
group.nc.33.67.vec <- as.vector(group.nc.33.67.f, mode</pre>
= "character");
Renal.survival.grouplo.NTN4 <-
SURV.time 3367[group.nc.33.67.vec %in% c("1")];
Renal.survival.grouphi.NTN4 <-
SURV.time 3367[group.nc.33.67.vec %in% c("3")];
Renal.censor.death.grouplo.NTN4 <-
CENSOR.death.nc 3367[group.nc.33.67.vec %in% c("1")];
Renal.censor.death.grouphi.NTN4 <-
CENSOR.death.nc 3367[group.nc.33.67.vec %in% c("3")];
Renal.censor.met.grouplo.NTN4 <-</pre>
CENSOR.met.nc 3367[group.nc.33.67.vec %in% c("1")];
Renal.censor.met.grouphi.NTN4 <-</pre>
```

```
CENSOR.met.nc 3367[group.nc.33.67.vec %in% c("3")];
km.Renal.df <- data.frame("Sample" =</pre>
c(names(Renal.survival.grouplo.NTN4), names(Renal.surviv
al.grouphi.NTN4)),
"group" = c(rep("lo.NTN4",
length(Renal.survival.grouplo.NTN4)), rep("hi.NTN4",
length(Renal.survival.grouphi.NTN4))),
"time" = c(Renal.survival.grouplo.NTN4,
Renal.survival.grouphi.NTN4),
"censor.death" = c(Renal.censor.death.grouplo.NTN4,
Renal.censor.death.grouphi.NTN4),
"censor.met" = c(Renal.censor.met.grouplo.NTN4,
Renal.censor.met.grouphi.NTN4));
# write.table(km.Renal.df,file="extras/
Efigure1.bc.txt",
append=FALSE, quote=FALSE, sep="\t", eol="\n", na="", row.na
mes=FALSE,col.names=TRUE);
```

## #. DONE RenalCancer