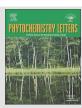
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Sceletorines A and B, two minor novel dimeric alkaloids of 1 Mesembryanthemum tortuosum (synonym Sceletium tortuosum)



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Mesembryanthemum tortuosun Sceletium tortuosum Sceletorine Alkaloids Absolute configuration Structure elucidation NMR

ABSTRACT5

Sceletorines A and B, two unique alkaloids, were isolated for the first time from the aerial parts of 7 Mesembryanthemum tortuosum. Their structures were determined by the extensive spectroscopic methods and their absolute configuration was established with ECD calculations. Plausible biosynthetic pathways for 1 and 2 are proposed and a relationship to the previously assumed artifact, channaine is also addressed.

1. Introduction 8

Mesembryanthemum tortuosum L. (synonym Sceletium tortuosum (L.) N.E.Br.), a succulent subshrub indigenous to South Africa, belongs to the family Mesembryanthemaceae (Smith et al., 1996). It is mainly distributed in the Karroid areas of the Cape region of South Africa and is locally referred to as "kanna" or "canna" (Khoi) and "kougoed" (Afrikaans) (Shikanga et al., 2012b). M. tortuosum is traditionally used to treat central nervous system (CNS) conditions, such as depression, stress, anxiety, and as a masticatory drug for the relief of thirst and hunger (Shikanga et al., 2012a,b). Recently, a ¹H NMR and UPLC-MS based metabolomic study, exploring chemotypic variation in S. tortuosum from two provinces in South Africa, was reported (Zhao et al., 2018). Alkaloids, commonly referred to as "mesembrine type alkaloids" (mesembrine, mesembrenone, mesembrenol and mesembranol), are recognized as the major chemical and bioactive constituents responsible for the psychoactive activities of the plant (Carpenter et al., 2016; Loria et al., 2014; Shikanga et al., 2012a, 2011, 2012b; Smith et al., 1996). As few alkaloids have been reported from this psychoac-

tive plant species so far, we decided to investigate it further for its 10 undiscovered phytoconstituents. The current phytochemical investigation of M. tortuosum resulted in the isolation and characterization of two novel alkaloids, sceletorine A (1) and sceletorine B (2), which are shown to have unusual carbon architecture (Fig. 1). Their structures and absolute configurations (ACs) were determined by the analysis of extensive spectroscopic data and electronic circular dichroism (ECD) calculations. Interestingly, compound 2 appears to be a precursor of channaine, which was reported as an artifact originating from the condensation of pair of N-desmethylmesembrenone molecules in 1978 by Jeffs et al. from S. strictum (Abou-Donia et al., 1978) and recently isolated and characterized on the basis of NMR from M. tortuosum (Veale et al., 2018). The possibility of compounds 1 and 2 to be the artifacts formed during acid-base extraction and/or isolation procedures was ruled out by observing their presence in the freshly prepared methanol extract of the plant by HPLC (see Supplementary file). Herein, the isolation, structure elucidation, stereochemical assignment, and plausible biosynthetic pathways of these alkaloids are presented.

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Fig. 1. Structures of compounds 1 and 2. 2

2. Results and discussion 3

Sceletorine A (1) was isolated as colorless solid. The molecular 4 formula of 1 was determined to be $C_{33}H_{40}N_2O_5$ by its ^{13}C NMR data and positive HRESIMS, which showed a [M+H]⁺ ion peak at m/z 545.3026 (calcd for $C_{33}H_{41}N_2O_5$, 545.3015). The ^{13}C NMR and HSQC spectra analysis revealed that 1 possessed 33 carbons, including five methyls, eight methylenes, nine methines, and 11 nonprotonated carbons, out of which 15 were olefinic/aromatic carbons. The resonance at δ_C 211.3 was due to a carbonyl group. The NMR resonances (Table 1) at δ_H 2.32 (3H, s), and δ_C 39.8 were ascribed to a methyl group on the nitrogen atom. The resonances at δ_H 7.11 (1 H), 7.04 (1 H), 6.94 (1 H), 6.86

(3 H), 3.89 (s, 3 H), 3.93 (s, 9 H) and at $\delta_{\rm C}$ 149.6, 148.9, 147.9, 147.7, 5 139.0, 135.8, 118.8, 118.7, 111.5, 111.1, 110.3, 110.0, 56.2, 56.0, 56.0, 55.9 in ¹H or ¹³C NMR spectra suggested the presence of two 3, 4-dimethoxyphenyl groups, the typical aryl moiety for mesembrine type alkaloids (Bastida et al., 1989; Chavan et al., 2004), which was further supported by the detailed HMBC correlations (Fig. 2). The aforementioned resonances implied that 1 could be a dimeric alkaloid, which was confirmed by the HMBC correlations of H-7′ ($\delta_{\rm H}$ 3.86) with C-4, C-5, C-6; H-7a′ ($\delta_{\rm H}$ 2.76, dd, J = 6.9, 15.8 Hz) with C-5; H-2′ ($\delta_{\rm H}$ 3.82 and 3.68, brd, J = 18.6 Hz) with C-6; and H-4 ($\delta_{\rm H}$ 5.95, s) to C-7′. Determination of the relative configuration at each stereogenic center of 1 was achieved with NOESY experiment in benzene- d_6 . The correlations

Table 1 1 H * and 13 C NMR spectroscopic data of 1 and 2 (δ in ppm).

No.	1			2	
	$\delta_{\rm C}^{\ a}$, type	$\delta_{ m H}^{\ \ a}$ mult. (J in Hz)	$\delta_{ m H}^{\;\; m b}$ mult. (J in Hz)	$\delta_{ m C}{}^{ m c}$	$\delta_{ m H}^{\ \ c}$ mult. (J in Hz)
N-Me	39.8, CH ₃	2.32 s	2.25 s	40.0, CH ₃	2.28 s
2	56.5, CH ₂	2.51, 3.23 brt (8.0)	2.30 m, 3.05 brt (8.0)	53.5, CH ₂	2.30 m, 3.16 td (4.8, 9.0)
3	40.1, CH ₂	2.45, 2.08 ddd (8.0, 10.2, 13.9)	2.04, 2.20	38.6, CH ₂	2.01
3a	51.0, C			48.2, C	
4	142.1, CH	5.95 s	6.23 s	38.0, CH ₂	2.09, 2.58 d (15.0)
5	134.5, C			122.3, C	
6	164.8, C			138.9, C	
7	36.2, CH ₂	2.51	2.62 brd (13.5)	29.9, CH ₂	2.16
			2.79 dd (3.0, 13.5)		2.23 dd (6.3, 18.6)
7a	73.3, CH	2.52	2.53	68.4, CH	2.66 d (6.3)
8	139.0, C			139.6, C	
9	110.3, CH	6.86	6.96 d (2.3)	111.4, CH	6.99 brs
10	148.9, C			149.8, C	
10-OMe	56.0, CH ₃	3.93	3.64 s	55.1, CH ₃	3.86 s
11	147.7, C			148.1, C	
11-OMe	55.9, CH ₃	3.89 s	3.49 s	55.3, CH ₃	3.79 s
12	111.1, CH	6.86	6.66 d (8.5)	112.0, CH	6.91
13	118.8, CH	6.86	6.93	118.7, CH	6.91
2′	47.9, CH ₂	3.68 brd (18.6)	3.74 m	54.0, CH ₂	2.86
		3.82	3.97 dd (6.4, 17.8)		2.89
3′	45.5, CH ₂	1.84	1.64 dd (6.4, 15.0)	40.9, CH ₂	1.75 ddd (2.5, 8.3, 12.6)
			1.88 dd (10.0, 15.0)		1.93
3a'	43.3, C			46.2, C	
4′	38.3, CH ₂	2.15, 2.28 brd (13.2)	2.02, 2.20	48.0, CH	2.46 brs
5′	31.8, CH ₂	2.16, 2.41	2.09	40.8, CH ₂	1.95, 2.15
6′	211.3, C			207.3, C	
7′	42.2, CH	3.86	3.55	39.9, CH ₂	2.53 d (18.9)
				· -	2.72 dd (8.3, 18.9)
7a′	42.6, CH ₂	2.49, 2.76 dd (6.9, 15.8)	2.47	62.4, CH	3.84 dd (2.4, 8.3)
8′	135.8, C			137.4, C	
9′	110.0, CH	7.04 brs	6.78 d (2.3)	111.3, CH	6.92 d (2.4)
10′	149.6, C			149.1, C	
10'-OMe	56.2, CH ₃	3.93	3.41 s	55.2, CH ₃	3.80 s
11'	147.9, C			147.8, C	
11'-OMe	56.0, CH ₃	3.93	3.47 s	55.5, CH ₃	3.79 s
12'	111.5, CH	6.94 d (8.4)	6.63 d (8.4)	110.5, CH	6.91
13'	118.7, CH	7.11 brd (8.4)	6.70 dd (2.3, 8.0)	117.9, CH	6.85 dd (2.4, 8.4)
	110.7, 011		2.70 da (2.0, 0.0)	117.5, 011	2.00 da (2.1, 0.1)

*Multiplicity not clear for some signals due to overlapping. Data were recorded in aCDCl₃, bC₆D₆, and cC₃D₆O.

Fig. 2. Key COSY, HMBC, and NOESY correlations of 1. 2

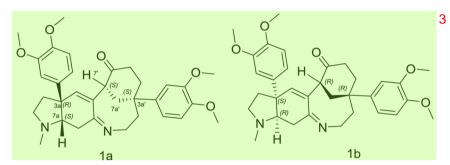


Fig. 3. Possible configurations of 1. 4

between H-7a with H-9/H-13 are in agreement with mesembrine al-5 kaloid type *cis* ring fusion. Whereas, NOESY correlations of H-7′ with H-9′/H-13′ and H-4 (olefinic) with H-7a′ and H-9/H-13 indicated that both aryl groups, H-7a, and H-7′ placed on same side of the plane, implying that the absolute configuration (AC) of compound 1 might be either 3a*R*,7a*S*,3a′*S*,7′*S*- or 3a*S*,7a*R*,3a′*R*,7′*R*- (Fig. 3).

Electronic circular dichroism (ECD) spectra were calculated to as-6 sign the exact AC of compound 1 (Ding et al., 2007, 2010; Li et al., 2015; Yang et al., 2014). Since the ECDs of enantiomers are completely opposite, only configuration 1a was employed for the conformational random search with an energy window of 130 kJ/mol by using the OPLS 2005 force field in MacroModel software package of Schrodinger suite, yielding 320 conformers. Thirty-two conformers within an energy cut-off of 20 kJ/mol were submitted to the geometry optimization and harmonic vibrational frequency computation at the B3LYP/6-311+ +G** level in the gas phase. Twenty-one conformers were relocated and conformational analysis was performed using the electronic and zero-point energies obtained above (Table S1 and Fig. 4). The ECDs of individual conformers were calculated using time-dependent density functional theory (TDDFT) at the B3LYP-SCRF(COSMO)/6-311+ $+G^{**}/B3LYP/6-311 + +G^{**}$ level in MeOH. The weighted ECD of 1a convincingly matches the experimental curve of 1 (Fig. 5). Exampled by the predominant conformer 1a7 which possesses 35.5%, the experimentally observed positive Cotton Effect (CE) at 288 nm is mainly contributed by the electronic transition at 286 nm, whereas the negative CE at 241 nm is attributed to the transition states at 264, 257, and 234 nm, respectively, and that at 211 corresponds to the excitations at 217, 207, 204, 203 and 201 nm, respectively. Therefore, the AC of compound 1 was unambiguously assigned as 3aR,7aS,3a'S,7'S (1a in Fig. 3).

Sceletorine B (2) was obtained as a colorless solid. Its molecular 7 formula, $C_{33}H_{40}N_2O_5$, was established by the positive HR-ESI-MS (m/z 545.3073 [M+H]⁺, calcd. for $C_{33}H_{41}N_2O_5$, 545.3015) and ¹³C NMR data. Analysis of ¹H NMR, ¹³C NMR and HSQC spectra revealed that the 33 carbons in this compound are comprised of five methyls, eight methylenes, nine methines (six olefinic/aromatic and three aliphatic) and 11 nonprotonated carbons (one non-conjugated carbonyl, eight

olefinic/aromatic and two aliphatic). Analysis of the $^1\text{H}/^{13}\text{C}$ NMR data 8 (Table 1) demonstrated that 2 contained a non-conjugated carbonyl group (δ_{C} 207.3), *N*-methyl [$\delta_{\text{H}}/\delta_{\text{C}}$ 2.28 (3H, s)/40.0], four methoxy groups [$\delta_{\text{H}}/\delta_{\text{C}}$ 3.79 (6H, s)/55.3, 55.5, 3.80 (3H, s)/55.2, and 3.86 (3H, s)/55.1], and two 1,3,4-trisubstituted phenyl groups [$\delta_{\text{H}}/\delta_{\text{C}}$ 6.99/111.4, 6.91/112.0, 6.91/118.7, 6.92/111.3, 6.91/110.5, and 6.85/117.9 and δ_{C} 139.6, 149.8, 148.1. 137.4, 149.1, 147.84]. The aforesaid data suggested that 2 is also a dimeric alkaloid, which was further supported by the HMBC correlations of H-2' with C-6, H-4' with C-5/C-6, H-5' with C-5, and H-7a' with C-6, as well as supported by the $^1\text{H}-^1\text{H}$ COSY correlations of H-2/H-3, H-7/H-7a, H-2'/H-3', H-4'/H-5', and H-7'/H-7a' (Fig. 6). Thus, the planar structure of 2 was determined to be that shown in Fig. 1.

The relative configuration at the stereogenic centers in 2 was assigned by analysis of the NOESY correlation (Fig. 6). The correlations of H-7a with H-9/H-13 in the NOESY spectrum revealed that H-7a and phenyl group at 3a are on the same side, similar to mesembrine type alkaloids. The presence of similar correlations of H-4' with H-9'/H-13', and H-7a' with H-9'/H-13' and no correlation between H-4' and H-7a' verified that these two protons occupy equatorial-equatorial positions flanked by a cyclopentane and dimethoxy aryl group as shown in Fig. 6. Similar to compound 1, ECD spectra were calculated to facilitate the AC assignment of compound 2. Since configurations 2a and 2c are enantiomeric to 2b and 2d, respectively, only the ECDs of 2a and 2c were calculated using the previous protocols. Fifty-two of 242 conformers of 2a and 48 of 264 conformers of 2c within an energy cut-off of 10 kJ/ mol were included in the ECD computations. Fifty-two conformers of 2a and 42 conformers of 2c were relocated at the B3LYP/6-311++G* level in the gas phase, both with very broad distribution (Tables S2 and S3). The weighted ECD spectrum of configuration 2a overall matches the experimental ECD curve, whereas that of 2c does not, even though noticeable redshift was found in the simulated ECD spectrum of 2a for the experimental CEs at 235 and 253 nm (Fig. 7). The experimentally negative CEs at 205 and 235 nm were reproduced at 202 and 254 nm, respectively, whereas those positive CEs at 217 and 253 nm correspond to the simulated ones at 227 and 279 nm, respectively. Therefore, the AC of compound 2 was assigned as 3aS,7aS,3a'R,4'R,7a'S (2a) (Fig. 8).

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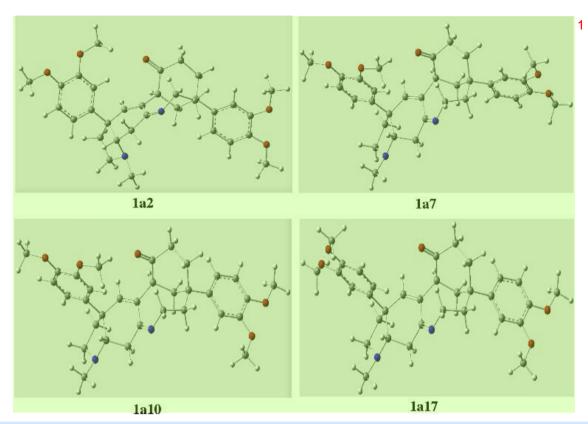


Fig. 4. Optimized geometries of predominant conformers of 3aR,7aS,3a'S,7'S- configuration of compound 1 at the B3LYP/6-311 + + G** level in the gas phase.

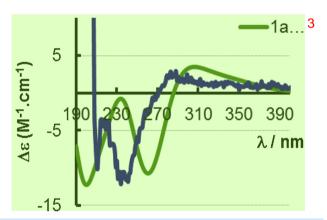


Fig. 5. Experimental (recorded in methanol) and weighted ECD spectra of 1.4

A plausible biogenetic route towards the unique dimer (1) forma-5 tion may be initiated by oxidative keto-enamine coupling of mesembrine with a N-desmethylmesembrine unit(Guo et al., 2012) to yield a tetracyclic intermediate. A proton rearrangement followed by an allylic oxidation and a subsequent elimination of a water molecule results in compound 1 (Scheme 1). Similar to compound 1, the pentacyclic compound 2 could be envisioned from intramolecular Michael addition of (desmethyl)mesembrine enolate to the N-demethylmesembrenone (Guo et al., 2012), via enamine/iminium intermediate. Further, possible P450 mediated allylic oxidation (when R=H), followed by a rearrangement of the hydroxyl group could yield amino-enone, aminol or hemiacetal, channaine (Scheme 1). Based on the results along with the proposed biogenesis for these dimeric alkaloids, it was speculated that the possible intermolecular condensation of monomeric mesembrine alkaloids as well as cytochrome P450 mediated secondary oxidations are plausible in M. tortuosum. Moreover, the optical rotation of these two dimeric alkaloids, conserved a stereopreference at the

bicyclic bridge and their presence in freshly prepared extract indicate 6 that these compounds are not artifacts of extraction/isolation procedures. Indeed, they are natural secondary metabolites of mesembrine-type mono alkaloids present in *M. tortuosum*. Nonetheless, the current findings highlight the complex chemistry of *M. tortuosum* wherein the major mono alkaloids are used as building blocks for the formation of novel, unique scaffolds.

3. Experimental section 7

3.1. General experimental procedures 8

Optical rotations were measured on an autopol IV polarimeter. CD 9 spectra were recorded on an Olis DSM20 instrument. UV spectra were obtained on Varian Cary 50 Bio UV-vis spectrophotometer. IR spectra were detected on an Agilent Cary 630 FTIR spectrophotometer. 1D and 2D NMR spectra were recorded on Bruker AVIII-400, AVIII-500 and AVIII-600 MHz spectrometers. Chemical shifts (δ) were expressed in ppm with reference to the solvent signals. HRESIMS spectra were determined on API QSTAR time-of-flight spectrometer and on Waters Autospec Premier P776 mass spectrometer. Semi-preparative HPLC analysis was conducted using an Agilent 1100 HPLC system equipped with a degasser, quaternary pump, auto sampler, column oven, and UV-Diode detector controlled by Chemstation software using RP-C18 column (250 × 10 mm; particle size 10 µm; Luna). HPLC grade acetonitrile and water were used as eluting solvents. Preparative TLC was performed on Sigma-Aldrich and Analtech uniplate silica gel plates $(20 \text{ cm} \times 20 \text{ cm}, 500 \mu\text{m})$, and column chromatography was carried out using silica gel (230-400 mesh, Silicycle Co., Ltd., Canada). Fractions were monitored by TLC (aluminum-backed cards, pre-coated with silica gel F254 (200 µm, 60 Å, Merck)) and spots were visualized by Dragendorff's and vanillin staining solutions.

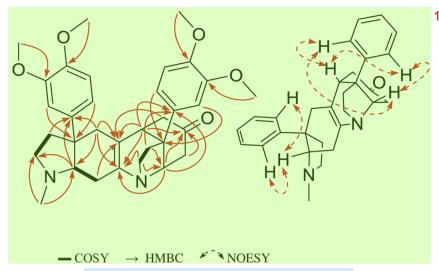


Fig. 6. Key ¹H-¹H COSY, HMBC, and NOESY correlations of 2.2

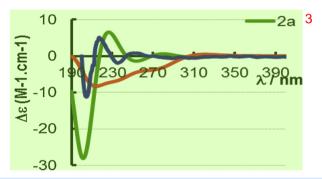


Fig. 7. Experimental (recorded in methanol) and weighted ECD spectra of 2. 4

3.2. Plant material 5

The aerial parts of *M. tortuosum* were collected (2012) from culti-6 vated material, Farm Vredelus, Uhlenhorst (Namibia). Retention samples are retained in the Department of Pharmaceutical Sciences,

Tshwane University of Technology (Pretoria, South Africa). The sample 7 (#13120) was also deposited at the National Center for Natural Products Research, University of Mississippi, USA.

3.3. Extraction and isolation 8

Powdered aerial parts of *M. tortuosum* (1.5 kg) were extracted with 9 methanol (4 L×24 h × 4). The combined extracts were concentrated under reduced pressure to afford methanol extracts (256.1 g). The major part of extract (225.5 g) was suspended in 5% HCl in water (1.3 L) and extracted with EtOAc (1.3 L × 3). The water soluble part was then basified (pH = 9–10) with NH₄OH solution and again extracted with EtOAc (1.7 L × 5) to afford alkaloid extract (40.8 g). The major part (32.2 g) of the alkaloid extract was chromatographed on a silica gel column, eluted with CHCl₃ – CH₃OH–NH₄OH (from 100:0:0 to 0:100:0.3), to yield 62 fractions. Fraction 28–32 (3.1 g) was separated over a silica gel column, eluted with CHCl₃, CHCl₃ – CH₃OH–NH₄OH (300:7.5:1) and (2000:100:7), to afford 37 subfractions. Compound 1 (1.7 mg) was obtained from subfractions 12–15 by preparative TLC.

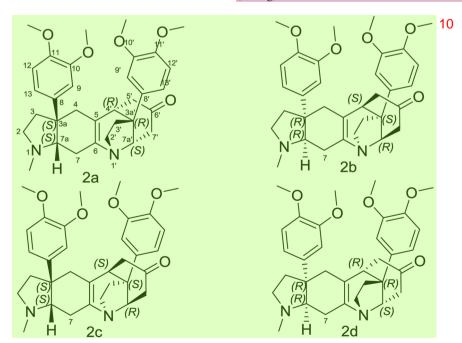


Fig. 8. Possible configurations of compound 2. 11

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Scheme 1. Plausible biogenetic pathway for 1, channaine and its precursor 2. 2

Compound **2** (3.5 mg) was isolated from subfraction 21 by semi preparative reversed phase HPLC. The HPLC analysis was carried out on RP-C18 column (250 \times 10 mm; particle size 10 μ m) with column oven temperature set at 25 °C and using gradients of eluent A (water with 0.1% acetic acid) and B (acetonitrile with 0.1% acetic acid). The gradient condition was as follows: 0–2 min (5% B), 2–32 min (5% B \rightarrow 40% B), 32–33 min (40% B \rightarrow 100% B), 33–37 min (100% B). The flow rate

of the solvent was 4.0 mL/min and the injection volume was 100 μ L. All 4 the analysis was carried out at wavelengths of 257 nm.

3.3.1. Sceletorine A (1) 5

 $C_{33}H_{40}N_2O_5$; Colorless solid; $[\alpha]_D^{26}+320$ (c 0.05, CH₃OH); UV 6 (CH₃OH) λ_{max} (log ϵ) 279.0 (3.59) nm; IR (neat) ν_{max} 2940, 1740, 1694, 1631, 1595, 1522, 1466, 13.72, 1260, 1154, 1059, 1031, 919,

858, 808, 774 cm $^{-1}$; HRESIMS m/z 545.3026 [M+H] $^{+}$ (calcd for 1 $C_{33}H_{41}N_2O_5$ 545.3015); 1 H NMR and 13 C NMR data: see Table 1.

3.3.2. Sceletorine B (2) 2

 $C_{33}H_{40}N_2O_5$; colorless solid; $[\alpha]_D^{26}+60$ (c 0.02, CH₃OH); UV 3 (CH₃OH) λ_{max} (log ϵ) 279.1 (3.30) nm; IR (neat) ν_{max} 2951, 1706, 1695, 1634, 1522, 1455, 1422, 1388, 1260, 1154, 1031, 830, 769 cm⁻¹; HRESIMS: m/z 545.3073 [M+H]⁺ (calcd for $C_{33}H_{41}N_2O_5$, 545.3015); ¹H NMR and ¹³C NMR data: see Table 1.

Acknowledgments 4

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Appendix A. Supplementary data 6

Supplementary material related to this article can be found, in the 7 online version, at doi:https://doi.org/10.1016/j.phytol.2019.03.013.

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