



Sceletium tortuosum demonstrates *in vitro* anti-HIV and free radical scavenging activity



P. Kapewangolo^{a,*}, T. Tawha^a, T. Nawinda^a, M. Knott^b, R. Hans^a

^a Department of Chemistry and Biochemistry, Faculty of Science, University of Namibia, Namibia

^b School of Pharmacy, University of Namibia, Namibia

ARTICLE INFO

Article history:

Received 29 April 2016

Received in revised form 10 June 2016

Accepted 24 June 2016

Available online 9 July 2016

Edited by AR Ndhala

Keywords:

Sceletium tortuosum

Anti-HIV

Radical scavenging

Phytochemicals

ABSTRACT

Crude extracts of commercially available *Sceletium tortuosum* (Aizoaceae) were investigated for *in vitro* anti-HIV and free radical scavenging activity for the first time. In addition, the chemical profile of *S. tortuosum* was determined using qualitative and quantitative phytochemical analysis. Crude extracts were prepared from dried plant parts of *S. tortuosum*. Ethanol and ethyl acetate extracts from the plants were tested for inhibitory activity against HIV-1 enzymes; protease (PR), reverse transcriptase (RT) and integrase (IN). *S. tortuosum* revealed the presence of anthraquinones, terpenes, polyphenols, anthocyanin, tannins, alkaloids, glycosides, carbohydrates and coumarins. The total phenolic, flavonoid and tannin contents were 171.89 ± 0.05 mg GAE/g, 678.51 ± 0.01 mg QE/g and 903.15 ± 0.02 mg TAE/g extract, respectively. Ethanol and ethyl acetate extracts of *S. tortuosum* inhibited HIV-1 RT and PR respectively. HIV-1 RT inhibition testing had IC_{50} values of <50 and 121.7 ± 2.5 μ g/mL for ethanol and ethyl acetate extracts, respectively. In addition, both extracts also inhibited HIV-1 PR with IC_{50} values < 100 μ g/mL. Results of the antiradical activity indicate a dose-dependent ability of the extracts to scavenge DPPH radicals. For this experiment, IC_{50} values of 49.0 ± 0.2 and 64.7 ± 3.1 μ g/mL were obtained from the ethanolic and ethyl acetate extracts, respectively. This suggests that *S. tortuosum* might be a useful potential source of new lead compounds in the development of new anti-HIV and radical scavenging compounds.

© 2016 SAAB. Published by Elsevier B.V. All rights reserved.

1. Introduction

Medicinal plants constitute an effective source of both traditional and modern medicine. About 80% of rural population depends on herbal medicine for primary health care (WHO, 2005). Over the years, the World Health Organization has advocated that countries should devote more attention to traditional medicine with a view to identify and explore aspects that provide safe and effective remedies for ailments of both microbial and non-microbial origin (WHO, 2005).

Southern Africa has a remarkable diversity of plants with a long tradition of medicinal use. A number of these plants reportedly contain compounds with anti-HIV and antioxidant properties (Van Wyk and Gericke, 2000; Van Wyk, 2011). Due to drug toxicity and the emergence of drug resistant viral strains (Clavel and Hance, 2004), there is an ongoing search for improved treatment against HIV/AIDS.

Sceletium tortuosum is widely distributed throughout Southern Africa and belongs to the family Aizoaceae, sub-family Mesembryanthemaceae. It is commonly referred to as Koegoed (Afrikaans), Kanna (Khoi) and

Sceletium (English) (Smith et al., 1996). *S. tortuosum* powders are commercially available as capsules known as “kanna” (Harvey et al., 2011; Shikanga et al., 2012) and are used to treat psychiatric disorders such as depression and anxiety (Gericke and Viljoen, 2008). Recent studies have highlighted a potential link between oxidative stress in the pathogenesis of neuropsychiatric disorders (Bakunina et al., 2015; Moylan et al., 2014; Smaga et al., 2015). Oxidative stress is caused by the accumulation of free radicals in the body which can be neutralized or scavenged by either endogenous or exogenous antioxidants. Some plant species belonging to the Aizoaceae family, for example *Carpobrotus edulis*, are used traditionally in the Eastern Cape, South Africa to boost the immune system of persons with HIV/AIDS as well as treat associated opportunistic infections (Omoruyi et al., 2012a; Vuuren and Naidoo, 2010). Omoruyi et al. (2012b) reported the antioxidant properties of *C. edulis* and linked the free radical scavenging activity of the extracts to the traditional success of the plant in managing aspects of HIV/AIDS.

HIV reportedly induces oxidative stress by disturbing cellular antioxidant defences and by initiating oxidative reactions. For this reason oxidative stress is regarded as one of the mechanisms that contributes to the progression of HIV to AIDS (Gil et al., 2003; Kashou and Agarwal, 2011; Mallery et al., 2004). Advanced cases of HIV infection leaves individuals prone to opportunistic infections because of progressive

* Corresponding author at: Department of Chemistry and Biochemistry, Faculty of Science, University of Namibia, P/Bag 13301, Windhoek, Namibia. Tel.: +264 61 2063384; fax: +264 61 2063791.

E-mail address: pkapewangolo@unam.na (P. Kapewangolo).

immunodeficiency caused by the virus. Therefore, naturally occurring antioxidants could play a role as inexpensive therapeutic agents that could reduce the HIV viral load of affected individuals. As a result, investigating both the anti-HIV and radical scavenging properties of such plants is recommended.

2. Material and methods

2.1. Plant cultivation

The dried whole plant parts were kindly donated by Farm Vredelus in July 2014. Farm Vredelus is located in Mariental, Namibia and specializes in rare desert plants with well-known medicinal and healing properties. The farm exports various medicinal plant products including *S. tortuosum* capsules. *S. tortuosum* identification (M1362) was undertaken by Silke Rugheimer at the National Herbarium of Namibia. *S. tortuosum* seedlings were grown in shade net tunnels and subsequently planted out. Plant coordinates: S 23° 56' 30" E 18° 19' 12". *S. tortuosum* was manually harvested and the roots were removed, dried and milled. Drying of the plant samples was done in a shade; air dried manner away from direct sunlight. The seed was produced in 2005 and certified organic by Namibia Organic Association (NOA) and Ecologic Certification (ECOCERT) for EU and USA.

2.2. Plant extraction

Powdered plant parts (111.2 g) were steeped in ethanol (1 L) at room temperature for 48 h. The filtrate was concentrated under vacuum and one half of the residue was re-dissolved in ethyl acetate to exclude highly polymeric tannins which are regarded as non-specific enzyme inhibitors (Kapewangolo et al., 2013). The resulting extracts were dried and stored at room temperature until further use.

2.3. Qualitative phytochemical screening

The major secondary metabolite classes such as alkaloids, anthraquinones, coumarins, terpenes, saponins, phenols, flavonoids, emodins and tannins were screened for using reported methods (Mir et al., 2013; Ugochukwu et al., 2013).

2.4. Quantitative phytochemical analysis

Total phenolic, flavonoid and tannin contents present in *S. tortuosum* ethanol extract were determined and quantified following previously described standard procedures (Gracelin et al., 2013; Marinova et al., 2005).

2.4.1. Total phenolic content (TPC)

The TPC was determined by using the Folin–Ciocalteu assay (Gracelin et al., 2013). In brief, 1 mL of extract (1 mg/mL) in double distilled water (ddH₂O) was mixed with 0.5 mL of 2 M Folin–Ciocalteu reagent and 1.5 mL of 20% Na₂CO₃. The volume was made up to 8 mL with ddH₂O. After incubation for 2 h at room temperature, the UV absorbance against the prepared reagent blank was taken at 765 nm. The TPC was determined from extrapolation of the calibration curve ($R^2 = 0.997$) which was made by preparing various gallic acid solutions (0.3, 0.6, 1.3, 2.5, 5 mg/mL). The TPC was expressed as milligrammes (mg) of gallic acid equivalents (GAE) per g of extract (mg GAE/g extract).

2.4.2. Total flavonoid content (TFC)

The TFC was determined following a modified version of the method reported by Marinova et al. (2005). Briefly, 1 mL of sample (1 mg/mL) or a standard solution of quercetin (0.3, 0.6, 1.3, 2.5, 5 mg/mL) was added to 4 mL of ddH₂O and 0.3 mL of 5% NaNO₂. After 5 min, 0.3 mL 10% AlCl₃ and 2 mL 1 M NaOH were added to the mixture. The total volume was made up to 10 mL with ddH₂O. The solution was mixed well and the

UV absorbance was measured against the reagent blank at 510 nm. The total flavonoids were expressed as mg of quercetin equivalents per g of extract (mg QE/g extract) using the gallic acid calibration curve ($R^2 = 0.990$).

2.4.3. Total tannin content (TTC)

The TTC of *S. tortuosum* was determined according to a method previously described by Gracelin et al. (2013). In brief, 5 mL of the filtered sample (10 mg/mL) was mixed with 2 mL of 0.1 M FeCl₃ in 0.1 M HCl and 0.008 M K₄[Fe(CN)₆]. UV absorbance at 120 nm was read after 10 min of incubation at room temperature. A standard calibration curve was prepared using various concentrations of tannic acid (0.3, 0.6, 1.3, 2.5, 5 mg/mL). TTC was expressed as mg tannic acid equivalents per g of extract (mg TAE/g extract) using the tannic acid calibration curve ($R^2 = 0.999$).

2.5. Anti-HIV assays

2.5.1. HIV-1 protease assay

The HIV-1 protease (PR) inhibition assay was conducted using the FRET-based Sensolyte HIV-1 protease assay kit (Anaspec). Briefly, HIV-1 PR and the fluorogenic substrate were mixed with the extracts to a final reaction volume of 100 μ L in a 96 well black assay plate (BRAND). The fluorescence intensity was measured at an excitation wavelength of 340 nm and an emission wavelength of 490 nm using a SpectraMax M2 plate reader. Acetyl pepstatin (AP) was used as a known inhibitor of HIV-1 PR. Other assay controls included a blank treatment with no extracts and, background controls with just extracts and buffer. *S. tortuosum* was tested at final concentrations of 25, 50, 100, 150, 200 and 250 μ g/mL. The IC₅₀ values were determined using GraphPad prism software (GraphPad Software Inc.).

2.5.2. HIV-1 reverse transcriptase colorimetric assay

The *in vitro* HIV-1 reverse transcriptase (RT) activity of the extracts was evaluated using a non-radioactive HIV-RT colorimetric assay kit from Roche Diagnostics. The protocol outlined in the kit was followed using 4 ng of enzyme per well. Extracts were tested at known concentrations (25, 50, 100, 150, 200, 250 μ g/mL). The extract concentrations were incubated with RT for 1 h at 37 °C. Subsequent hourly incubations involved the addition of an antibody conjugated to peroxidase, which binds to the digoxigenin-labelled DNA. In the final step, the peroxidase substrate solution [2,2'-azino-bis-(3-ethylbenzthiazoline-6-sulfonic acid)] was cleaved by the peroxidase enzyme, producing a coloured reaction product. Doxorubicin was used as a positive control (Kuethe et al., 2010; Mbaveng et al., 2011). The absorbance of the samples was read at 405 nm using a SpectraMax M2 plate reader. IC₅₀ values were computed using GraphPad prism software.

2.5.3. HIV-1 Integrase colorimetric assay

A non-radioactive HIV-1 integrase assay kit (XpressBio) was used to measure the inhibitory effects of *S. tortuosum* extracts against HIV-1 integrase (Jiang et al., 2010). Streptavidin-coated 96-well plates were coated with a double-stranded HIV-1 LTRU5 donor substrate (DS) DNA containing an end-labelled biotin by manufacturer. Full-length recombinant HIV-1 integrase protein was then loaded onto the oligo substrate. Extracts (0.2, 0.4, 0.8, 1 and 2 mg/mL) were added to the enzyme reaction and then a different double stranded target substrate (TS) DNA containing a 3'-end modification was added to the reaction mixture. The HRP-labelled antibody was directed against the TS 3'-end modification and the absorbance due to the HRP antibody-TMB peroxidase substrate reaction was measured at 450 nm. Sodium azide was included as a positive control.

2.5.4. Antiradical activity: DPPH assay method

The free radical scavenging activity of *S. tortuosum* extracts was evaluated using a modified method previously described by Kapewangolo

et al. (2013). Briefly, extracts dissolved in ethanol were mixed with a 90 μ M 2,2-diphenyl-1-picrylhydrazyl (DPPH) ethanol solution to give final extract concentrations of 7.81, 15.63, 31.25, 62.5, 125, 250, 500 and 1000 μ g/mL. After 30 min incubation at room temperature, sample absorbance was measured at 520 nm (SpectraMax M2). Absorbance values obtained were converted into percentage antiradical activity (Kute et al., 2010; Marxen et al., 2007). Ascorbic acid was used as a standard control. IC₅₀ values were then calculated using GraphPad Prism software.

3. Results and discussion

Phytochemical results revealed that *S. tortuosum* extracts contained anthraquinones, terpenes, polyphenols, anthocyanin, tannins, alkaloids, glycosides, carbohydrates and coumarins. The amounts of total phenolic, flavonoid and tannin from ethanol extract were found to be 171.89 \pm 0.05 mg GAE/g, 678.51 \pm 0.01 mg QE/g and 903.15 \pm 0.02 mg TAE/g extract, respectively. The relative amounts of tannin and flavonoid components were observed to be higher than that of the phenolic components as shown in Table 1. Phytochemical content was only determined for the ethanol extract of *S. tortuosum* due to the type of extraction undertaken in this study; extraction of dried plant parts in ethanol followed by subsequent extraction of the ethanol filtrate with ethyl acetate to exclude highly polar tannins which are non-specific enzyme inhibitors (Klos et al., 2009). Minimal literature was available on the phytochemical analysis of *S. tortuosum*, with the exception of alkaloids from *S. tortuosum*. In the literature, the phytochemical investigation of *C. edulis*, a plant also belonging to the same family Aizoaceae, demonstrated a high tannin content in the ethanolic extract which is in agreement with data presented in this study (Omoriyi et al., 2012b). However, the relative amount of total phenolic content for *C. edulis* in the literature was higher than that reported for *S. tortuosum* in this study. In addition, Omoriyi et al. (2012b) reported lower flavonoid content for *C. edulis* ethanolic extract. A literature search revealed minimal information on the phytochemical investigation of plants belonging to the Aizoaceae family, thus, encouraging extensive research to be conducted on these plants. The high tannin and flavonoid contents observed here for *S. tortuosum* can be attributed to the good radical scavenging activity detected. The effect of tannins and flavonoids on the protection of cells from oxidative-induced damage/death has been reported (Panat et al., 2016; Zielińska-Przyjemska et al., 2015) and the relatively high content of these phytochemicals recorded for *S. tortuosum* is noteworthy.

The activity of *S. tortuosum* extracts against HIV-1 RT, IN and PR varied. IC₅₀ values were obtained for both the ethanolic and ethyl acetate extracts (Table 2). Both extracts possessed anti-RT and anti-PR activities. The ethanolic extract inhibited HIV-1 RT by 59.9% at 100 μ g/mL (IC₅₀ < 50 μ g/mL) and at the same concentration HIV-1 PR was inhibited by 91.1%. The ethyl acetate extract inhibited HIV-1 RT by 56.7% at 200 μ g/mL (IC₅₀ = 121.7 \pm 2.5 μ g/mL) and inhibition of HIV-1 PR was 100% at 100 μ g/mL. Doxorubicin, used as a positive control for HIV-1 RT, showed an inhibition of 90.1% at 25 μ g/mL, while the known inhibitor for HIV-1 PR, acetyl pepstatin, gave an inhibition of 92.6% at 10 μ g/mL. Both extracts demonstrated weak activity against HIV-1 integrase with less than 20% inhibition at the highest concentration tested (400 μ g/mL). This data suggests that *S. tortuosum* contains

Table 2

HIV-1 enzyme inhibitory activity of *S. tortuosum* extracts and positive controls.

Sample	IC ₅₀ (μ g/mL) \pm SD		
	HIV-1 RT	HIV-1 PR	HIV-1 IN
Ethanol extract	<50	<100	>400
Ethyl acetate extract	121.7 \pm 2.5	<100	>400
Acetyl pepstatin ^a	–	<10	–
Doxorubicin ^b	<25	–	–
Sodium azide ^c	–	–	<10

^a Positive control for HIV-1 protease (PR).

^b Positive control for HIV-1 reverse transcriptase (RT).

^c Positive control for HIV-1 integrase (IN): concentration in %.

compounds that specifically inhibit HIV-1 RT and PR. However, as only crude extracts were tested, it should be noted that the synergistic therapeutic actions of various phytochemicals (Ma et al., 2009) could also be responsible for the bioactivity observed here.

The results of the antiradical activity indicated in Fig. 1 showed a dose-dependent response. IC₅₀ values of 49.0 \pm 0.2 and 64.7 \pm 3.1 μ g/mL were obtained for both the ethanol and ethyl acetate extracts, respectively. The free radical scavenging activity of *S. tortuosum* observed in the present study could be attributed to the presence of potential antioxidant compounds including coumarins, flavonoids, phenols and alkaloids (Gupta and Sharma, 2006). The radical scavenging activity of *S. tortuosum* is an indication that it is a potential natural antioxidant. *S. tortuosum* is traditionally used to treat depression and anxiety (Gerike and Viljoen, 2008). There are recent reports on the potential role of oxidative stress in the pathogenesis of psychiatric diseases (Bakunina et al., 2015; Smaga et al., 2015) and the antiradical potential of *S. tortuosum* could be attributed to the popular use of this commercial plant in managing depression and anxiety.

Oxidative stress is also largely linked to the activation of HIV-1 leading to the development of AIDS (Allard et al., 1998; Gil del Valle et al., 2013). Therefore, having an agent with both anti-HIV and antioxidant properties is of therapeutic importance. Currently, there exist no reports on the HIV inhibitory properties of *S. tortuosum*, making this *in vitro* HIV study a first for this plant.

4. Conclusion

The *in vitro* anti-HIV data presented here provides evidence that *S. tortuosum* contains compounds with inhibitory effects against both HIV-1 RT and PR. In addition, the plant also appears to possess compounds with antiradical properties. The free radical scavenging results obtained in this study justify the traditional success of *S. tortuosum* in

Table 1

Total phenolic (TPC), total flavonoid (TFC) and total tannin (TTC) contents in ethanol extract of *S. tortuosum* dried plant parts.

Phytochemical detected	<i>S. tortuosum</i> sample
TPC (mg GAE/g)	171.89 \pm 0.05
TFC (mg QE/g)	678.51 \pm 0.01
TTC (mg TAE/g)	903.15 \pm 0.02

All analyses are the mean of triplicate measurements \pm standard deviation.

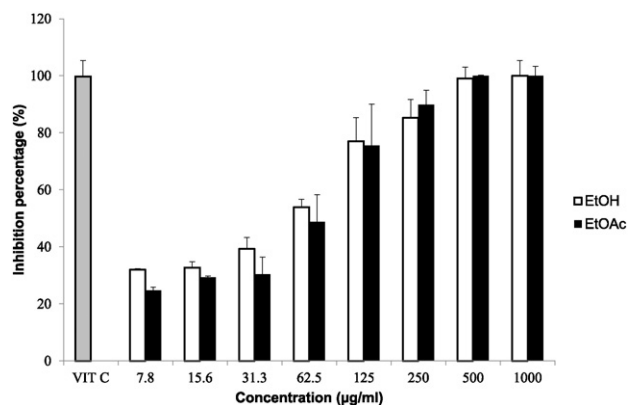


Fig. 1. Free radical scavenging activity of the crude extracts from *S. tortuosum*. The data represents the mean of multiple replicates. Vitamin C was used as a standard control with an inhibition of 99.8% at 50 μ g/mL (IC₅₀ < 1.6 μ g/mL). *S. tortuosum* ethanol (EtOH) and ethyl acetate (EtOAc) extracts demonstrated good antiradical activity with IC₅₀ values of 49.0 \pm 0.2 and 64.7 \pm 3.1 μ g/mL, respectively.

managing depression and anxiety. The anti-HIV results obtained substantiate the use of selected Aizoaceae plants in managing HIV/AIDS. Further research will include the isolation and identification of these active compounds (using bio-assayed fractionated guidance) as well as determining their exact mechanisms of action.

Acknowledgements

Authors acknowledge Farm Vredelus in Mariental (Namibia) for the donation of *S. tortuosum*. This work was supported by the University of Namibia's Research and Publications Office [Grant number URPC/2014/184].

References

- Allard, J.P., Aghdassi, E., Chau, J., Salit, I., Walmsley, S., 1998. Oxidative stress and plasma antioxidant micronutrients in humans with HIV infection. *The American Journal of Clinical Nutrition* 67, 143–147.
- Bakunina, N., Pariente, C.M., Zunszain, P.A., 2015. Immune mechanisms linked to depression via oxidative stress and neuroprogression. *Immunology* 144, 365–373.
- Clavel, F., Hance, A.J., 2004. HIV drug resistance. *The New England Journal of Medicine* 350, 1023–1035.
- Gericke, N., Viljoen, A.M., 2008. *Scelletium*—a review update. *Journal of Ethnopharmacology* 119, 653–663.
- Gil del Valle, L., Hernández, R.G., Ávila, J.P., 2013. Oxidative stress associated to disease progression and toxicity during antiretroviral therapy in human immunodeficiency virus infection. *Journal Virology Microbiology* 2013. <http://dx.doi.org/10.5171/2013.279685>.
- Gil, L., Mart, G., González, I., Tarinas, A., Álvarez, A., Giuliani, A., Molina, R., Tápanes, R., Pérez, J., Sonia, O., 2003. Contribution to characterization of oxidative stress in HIV/AIDS patients. *Pharmacological Research* 47, 217–224.
- Gracelin, S.D.H., John, G.A., Jeya, P.B., Kumar, R., 2013. Qualitative and quantitative analysis of phytochemicals in five *Pteris* species. *International Journal of Pharmacy and Pharmaceutical Sciences* 5, 4–6.
- Gupta, V.K., Sharma, S.K., 2006. Plants as natural antioxidants. *Natural Product Radiance* 5, 326–334.
- Harvey, A.L., Young, L.C., Viljoen, A.M., Gericke, N.P., 2011. Pharmacological actions of the South African medicinal and functional food plant *Scelletium tortuosum* and its principal alkaloids. *Journal of Ethnopharmacology* 137, 1124–1129.
- Jiang, F., Chen, W., Yi, K., Wu, Z., Si, Y., Han, W., Zhao, Y., 2010. The evaluation of catechins that contain a galloyl moiety as potential HIV-1 integrase inhibitors. *Clinical Immunology* 137, 347–356.
- Kapewangolo, P., Hussein, A.A., Meyer, D., 2013. Inhibition of HIV-1 enzymes, antioxidant and anti-inflammatory activities of *Plectranthus barbatus*. *Journal of Ethnopharmacology* 149, 184–190.
- Kashou, A.H., Agarwal, A., 2011. Oxidants and antioxidants in the pathogenesis of HIV/AIDS. *Open Reproductive Science Journal* 3, 154–161.
- Klos, M., van de Venter, M., Milne, P.J., Traore, H.N., Meyer, D., Oosthuizen, V., 2009. In vitro anti-HIV activity of five selected South African medicinal plant extracts. *Journal of Ethnopharmacology* 124, 182–188.
- Kuete, V., Metuno, R., Keilah, P.L., Tshikalange, E.T., Ngadjui, B.T., 2010. Evaluation of the genus *Treculia* for antimycobacterial, anti-reverse transcriptase, radical scavenging and antitumor activities. *South African Journal Botany* 76, 530–535.
- Ma, X.H., Zheng, C.J., Han, L.Y., Xie, B., Jia, J., Cao, Z.W., Li, Y.X., Chen, Y.Z., 2009. Synergistic therapeutic actions of herbal ingredients and their mechanisms from molecular interaction and network perspectives. *Drug Discovery Today* 14, 579–588.
- Mallery, S.R., Pei, P., Landwehr, D.J., Clark, C.M., Bradburn, J.E., Ness, G.M., Robertson, F.M., 2004. Implications for oxidative and nitrate stress in the pathogenesis of AIDS-related Kaposi's sarcoma. *Carcinogenesis* 25, 597–603.
- Marinova, D., Ribarova, F., Atanasova, M., 2005. Total phenolics and total flavonoids in Bulgarian fruits and vegetables. *Journal of the University of Chemical Technology and Metallurgy* 40, 255–260.
- Marxen, K., Vanselow, K.H., Lippemeier, S., Hintze, R., Ruser, A., Hansen, U., 2007. Determination of DPPH radical oxidation caused by methanolic extracts of some microalgal species by linear regression analysis of spectrophotometric measurements. *Sensors* 7, 2080–2095.
- Mbaveng, A.T., Kuete, V., Mapunya, B.M., Beng, V.P., Nkengfack, A.E., Meyer, J.J.M., Lall, N., 2011. Evaluation of four Cameroonian medicinal plants for anticancer, anticonorrheal and antireverse transcriptase activities. *Environmental Toxicology and Pharmacology* 32, 162–167.
- Mir, M.A., Sawhney, S.S., Jassal, M.M.S., 2013. Qualitative and quantitative analysis of phytochemicals of *Taraxacum officinale*. *The Journal of Pharmacy and Pharmacology* 2, 1–5.
- Moylan, S., Berk, M., Dean, O.M., Samuni, Y., Williams, L.J., O'Neil, A., Hayley, A.C., Pasco, J.A., Anderson, G., Jacka, F.N., Maes, M., 2014. Oxidative & nitrosative stress in depression: why so much stress? *Neuroscience and Biobehavioral Reviews* 45, 46–62.
- Omoruyi, B.E., Bradley, G., Afolayan, A.J., 2012a. Ethnomedicinal survey of medicinal plants used for the management of HIV/AIDS infection among local communities of Nkonkobe Municipality, Eastern Cape. *South Africa Journal Medicinal Plants Research* 6, 3603–3608.
- Omoruyi, B.E., Bradley, G., Afolayan, A.J., 2012b. Antioxidant and phytochemical properties of *Carpobrotus edulis* (L.) bolus leaf used for the management of common infections in HIV/AIDS patients in Eastern Cape Province. *BMC Complementary and Alternative Medicine* 12. <http://dx.doi.org/10.1186/1472-6882-12-215>.
- Panat, N.A., Maurya, D.K., Ghaskadbi, S.S., Sandur, S.K., 2016. Troxerutin, a plant flavonoid, protects cells against oxidative stress-induced cell death through radical scavenging mechanism. *Food Chemistry* 194, 32–45.
- Shikanga, E.A., Kamatou, G.P.P., Chen, W., Combrinck, S., Viljoen, A.M., 2012. Validated RP-UHPLC PDA and GC-MS methods for the analysis of psychoactive alkaloids in *Scelletium tortuosum*. *South African Journal Botany* 82, 99–107.
- Smaga, I., Niedzielska, E., Gawlik, M., Moniczewski, A., Krzek, J., Przegalski, E., Pera, J., Filip, M., 2015. Oxidative stress as an etiological factor and a potential treatment target of psychiatric disorders. Part 2. Depression, anxiety, schizophrenia and autism. *Pharmacological Reports* 67, 569–580.
- Smith, M.T., Crouch, N.R., Gericke, N., Hirst, M., 1996. Psychoactive constituents of the genus *Scelletium* N.E.Br. and other Mesembryanthemaceae: a review. *Journal of Ethnopharmacology* 50, 119–130.
- Ugochukwu, S.C., Arukwe Uche, I., Onuoha, I., 2013. Preliminary phytochemical screening of different solvent extracts of stem bark and roots of *Dennetia tripetala* G. Baker. *Asian Journal Plant Science Research* 3, 10–13.
- Van Wyk, B.-E., 2011. The potential of South African plants in the development of new medicinal products. *South African Journal Botany* 77, 812–829. <http://dx.doi.org/10.1016/j.sajb.2011.08.011>.
- Van Wyk, B.-E., Gericke, N., 2000. *People's Plants: a Guide to Useful Plants of Southern Africa*. Briza Publications, Pretoria.
- Vuuren, S.F.V., Naidoo, D., 2010. An antimicrobial investigation of plants used traditionally in southern Africa to treat sexually transmitted infections. *Journal of Ethnopharmacology* 130, 552–558.
- WHO, 2005. National policy on traditional medicine and regulation of herbal medicines. Report of a WHO Global Survey World Health Organization.
- Zielińska-Przyjemská, M., Ignatowicz, E., Krajka-Kuźniak, V., Baer-Dubowska, W., 2015. Effect of tannic acid, resveratrol and its derivatives, on oxidative damage and apoptosis in human neutrophils. *Food and Chemical Toxicology* 84, 37–46.