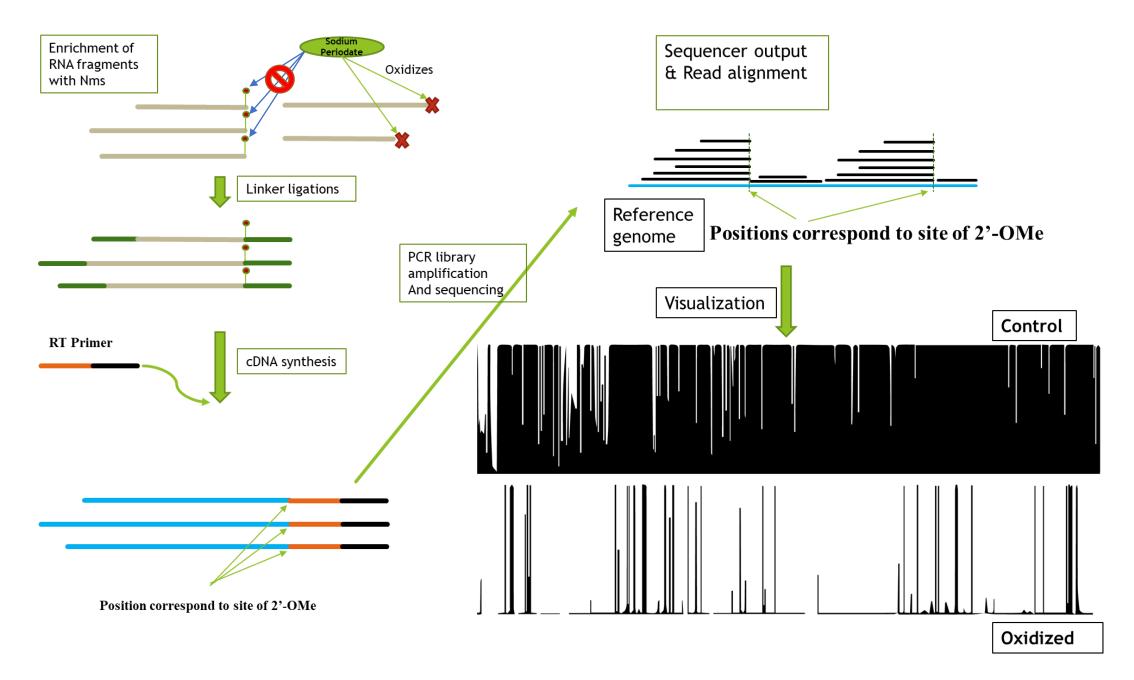
Demonstration of basic RibOxi-seq concept



RNA structure after 3' and 5' linker ligations

5'-/Biosg/ACACUCUUUCCCUACACGACGCUCUUCCGAUCUNNNN------insert------ATCACGCTGTAGGCACCATCAATGACAG/SpC3/- 3'

cDNA structure

Randomer for deduplication

3'-linker sequence

3'-TGTGAGAAAGGGATGTGCTGCGAGAAGGCTAGANNNN----insert----TAGTGCGACATCCGTGGTAGTTACTGTCTAGCCTTCTCGTGTGCAGACTTGAGGTCAGTG-5'

Complementary to partial Illumina i5 PCR primer

In-line barcode for filtering out RT mispriming

<u>Identical</u> to partial Illumina i7 PCR primer

Sequencing Output (example using 75x75 Paired end sequencing)

Read1: Read2:

@Sequence_identifier+i7_index+i5_index

GTCATTGATGGTGCCTACAGCGTGAT_SOME_INSERT_AND READ_THROUGH_ADAPTER

+

QUALITY OF THE READ

1. cutadapt paired-end mode remove read-through adapters:

Read1 output:

Read2 output:

@ Sequence_identifier+i7_index+i5_index
NNNN_SOME_INSERT
+
QUAL

@Sequence_identifier+i7_index+i5_index
GTCATTGATGGTGCCTACAGCGTGAT_SOME_INSERT
+
QUALITY OF THE REA

2. *pear* to merge read1 and read2 into a single read:

@ Sequence_identifier+i7_index+i5_index
NNNN_SOME_INSERT_ATCACGCTGTAGGCACCATCAATGACAG
+
____QUALAER_EHT_FO_YTILAUQ_____

3. move_umi.py moves randomer sequence from read to read identifier and discard reads lacking ATCACG:

@ Sequence_identifier_NNNN+i7_index+i5_index
SOME_INSERT_ATCACGCTGTAGGCACCATCAATGACAG
+
____QUALAER_EHT_FO_YTILAUQ______

4. *cutadapt* removes the rest of the linker sequence:

Not duplicates

```
@ Sequence_identifier_NNNN+i7_index+i5_index
SOME_INSERT
+
____QUAL
```

- 5. *STAR* alignment to genome of interest
- 6. samtools indexing the bam output
- 7. umi_tools deduplication:

Example aligned read #1:

UMI: AATC

Sequence: GGTTACG

Example aligned read #2:

UMI:CGTA

Sequence: GGTTACG

Example aligned read #1:

UMI: AATC

Sequence: GGTTACG

Example aligned read #2:

UMI: AATC

Sequence: GGTTACG

Duplicates, discard

- 8. bedtools converts deduplicated bam to bed file format
- 9. genomecov generates genome coverage track of 3'-end only to visualize Nm sites on genome browser
- 9. riboxi_bed_parsing.py counts all 3'-end alignments and generates a tab delimited file listing chromosome number, base position, gene name, (+15,base,-15) nucleotide sequence and counts for all samples:

1	chr	base gene	seq	ms_wt_186_0x	
2	chr13	97190585	Hexb	000000000000000000000000000000000000000	13263
3	chr9	123462160	Lars2	gttgttgccatggtaatcctgctcagtacga	11174
4	chr2	102829412	Cd44	CCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCC	10624
5	chr17	39848136	Rn45s	gaagacggtcgaacttgactatctagaggaa	9501
6	chr17	39847806	Rn45s	tcccccaacttcttagagggacaagtggcgt	4412
7	chr17	39846960	Rn45s	gaggatccattggagggcaagtctggtgcca	3148
8	chr7	19697484	Apoe	CCAAGTCACACAAGAACTGACGTGAGTGTCC	2871
9	chr5	136932984	493340	4012Rik gtctccaaggtgaacagcctctgg	cacattg 2850
10	chr1	100180179	Cntnap	5b CCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCC	CC 2801
11	chr17	39847687	Rn45s	attccgtgggtggtggtgcatggccgttctt	2405
12	chr11	17221313	Wdr92	gtctccaaggtgaacagcctctggcatgttg	2286

Downstream analysis can then be performed

RibOxi-seq Counts Visualization

