



Contents lists available at ScienceDirect

Advanced Drug Delivery Reviews

journal homepage: www.elsevier.com/locate/addr

Burn injury: Challenges and advances in burn wound healing, infection, pain and scarring

Yiwei Wang^a, Joanneke Beekman^a, Jonathan Hew^a, Stuart Jackson^a, Andrea C. Issler-Fisher^b, Roxanne Parungao^a, Sepher S. Lajevardi^b, Zhe Li^{a,b}, Peter K.M. Maitz^{a,b,*}

^a Burns Research Group, ANZAC Research Institute, Concord Hospital, University of Sydney, Concord West, NSW 2139, Australia

^b Burns and Reconstructive Surgery Unit, Concord Hospital, Concord West, NSW 2139, Australia

ARTICLE INFO

Article history:

Received 5 June 2017

Received in revised form 15 September 2017

Accepted 18 September 2017

Available online xxx

Keywords:

Severe burn injuries

Burn wound infections

Burn pain management

Scarring

Skin tissue engineering

Stem cells

Negative pressure wound therapy

Laser therapy for scarring

ABSTRACT

Severe burn injuries are the most traumatic and physically debilitating injuries affecting nearly every organ system and leading to significant morbidity and mortality. Early burn wound excision and skin grafting are common clinical practices that have significantly improved the outcomes for severe burn injured patients by reducing mortality rate and days of hospital stay. However, slow wound healing, infection, pain, and hypertrophic scarring continue to remain a major challenge in burn research and management. In the present article, we review and discuss issues in the current treatment of burn injuries; the advances and novel strategies developed in the past decade that have improved burn management; and also, pioneer ideas and studies in burn research which aims to enhance burn wound care with a focus on burn wound infection, pain management, treatments for scarring and skin tissue engineering.

© 2017 Elsevier B.V. All rights reserved.

1. Introduction

Severe burn injuries are the most traumatic and physically debilitating injuries affecting nearly every organ system and leading to significant morbidity and mortality. In Australia, over 10,000 people are hospitalised each year because of severe burn or scald injuries. According to the World Health Organisation (WHO), 180,000 deaths annually are related to burn injury and in 2004, nearly 11 million people worldwide were

severely burned and required medical treatment [1]. Both small burn and large severe burn injuries initiate the wound healing process which consists of several highly integrated and overlapping phases: inflammation, cell recruitment, matrix deposition, epithelialization and tissue remodeling. In addition to local wound repair, severe large burns also stimulate a persistent pathophysiological stress response and a systemic hypermetabolic-catabolic condition. These pathophysiological changes have great effects on the pharmacokinetics and pharmacodynamics of drug use in the treatment of severe burn injuries. Early burn wound excision and skin grafting are common clinical practices that have significantly improved the outcomes for severe burn patients by reducing mortality rate and days of hospital stay [2]. However, slow wound healing, infection, pain, and hypertrophic scarring continue to remain as major challenges in burn management and research. In the present article, we discuss the challenges, advances and novel strategies in burn management and research with a focus on burn wound infection, pain management, treatments of scarring and skin tissue engineering.

2. Clinical practice in the treatment of burn injury

2.1. Skin grafting

When burns or scald injuries are deep partial-thickness in the dermis or completely destroy all skin layers, wounds cannot be closed by

Abbreviations: WHO, World Health Organisation; TBSA, Total body surface area; NIKS, Near-diploid Immortalised Keratinocyte S; NPWT, Negative pressure wound therapy; NPWTi, Negative pressure wound therapy instillation; NPWTci, Negative pressure wound therapy continuous instillation; MDR, Multi-drug resistant; MRSA, Methicillin resistant staphylococcus aureus; VRE, Vancomycin resistant enterococci; ESBL, Extended-spectrum beta-lactamases; VISA, Vancomycin intermediate susceptible staphylococcus aureus; EPE, Enhanced permeability and retention effect; PK, Pharmacokinetics; PD, Pharmacodynamics; IV, Intravenous; IV-PCA, Intravenous patient-controlled analgesia; TRP, Transient receptor potential; TRPV1, Transient receptor potential vanilloid-1; KO, knockout; Na_v, Voltage gated sodium channels; MSCs, Mesenchymal stem cells; ADSCs, Adipose derived stem cells; TLRs, Toll-like receptors; TGFβ1, Transforming growth factor-beta1; MMPs, Matrix metalloproteinases; VEGF, Vascular endothelial growth factor; IL-6, Interleukin-6; ECM, Extracellular matrix; PDL, Pulsed dye lasers; Nd:YAG, Neodymium-doped yttrium-aluminium-garnet; NAFL, Non-ablative fractional lasers; AFL, Ablative fractional lasers; PCL, Poly (ε-caprolactone); PU, Polyurethane; PGA, Poly (glycolic acid); PLLA, Poly (L-lactide); 3D, 3-Dimensional; CAD, Computer-aided design; PLA, Poly (lactic acid); PLGA, Poly (lactide-co-glycolide).

* Corresponding author at: Concord Hospital, University of Sydney, Australia.

E-mail address: peter.maitz@sydney.edu.au (P.K.M. Maitz).

<https://doi.org/10.1016/j.addr.2017.09.018>

0169-409X/© 2017 Elsevier B.V. All rights reserved.

Please cite this article as: Y. Wang, et al., Burn injury: Challenges and advances in burn wound healing, infection, pain and scarring, Adv. Drug Deliv. Rev. (2017), <https://doi.org/10.1016/j.addr.2017.09.018>

the primary healing process or by suturing [3], rather additional surgical procedures are required. The gold-standard treatment for partial- and full-thickness burn injuries is early excision of necrotic tissue followed by autologous skin grafting (Fig. 1). Skin grafting involves the transplantation of healthy skin from an undamaged donor site of the patient to the wound site. Autologous skin grafts can be full thickness, consisting of epidermis and dermis, or split-thickness, consisting of the epidermis and upper part of the dermis. Unfortunately, in practice, donor skin is extremely limited for patients with severe burn injury >50% total body surface area (TBSA) [4,5]. This problem can be overcome by repeated harvesting of the donor sites over a period of time. However, healing of donor sites can be slow with additional scarring and possible pigmentation disorder [6,7]. While autografts can be meshed to increase the available surface area of the graft up to four times, the meshing process compromises the quality of the original graft and the recipient area heals with an irregular meshed pattern [8] and can result in severe scarring [9].

2.2. Skin substitutes

Skin substitutes could protect large burn wounds when donor skin is limited, to enhance wound healing, increase the dermal component of healed wound, reduce inflammatory responses and subsequent scarring [10–13]. Since 2000, over 30 new skin substitutes have been tested or used in the treatment of burn injuries (Table 1). Skin substitutes can be categorized into biological substitutes, synthetic substitutes or a combination of both. Biological substitutes can be further categorized into 1) natural scaffolds such as Alloderm, an intact, de-cellurised and dermal human matrix, 2) constructed scaffolds such as Integra, composed of lyophilized collagen, supplemented and cross-linked, or 3) cultured scaffolds, such as cultured epithelial graft, autologous cultured fibroblasts and keratinocytes. Biological skin substitutes have components that resemble natural skin, yet these skin substitutes are relatively simple compared to the complexities of human skin. The majority of skin substitutes available for clinical practice contain allogenic biological products, and the risk of disease transmission poses as a limitation particularly for natural biological skin substitutes. Despite extensive and strict sterilization procedures, current methods are insufficient to certify biological skin substitutes to be free of any unknown diseases or prion

diseases from animal material, such as Creutzfeldt-Jakob disease [14]. Furthermore, there is evidence to suggest that allogeneic skin is highly immunogenic and that cellular remnants in the extracellular matrices (ECM) may be responsible for reduced skin graft take or even rejection [9]. Human derived allogenic skin is further limited by its supply. Autologous biomaterials have the advantage of culturing cells for a large surface area from a small skin biopsy, however culturing cells is time consuming and may compromise wound healing [15,16].

In contrast to biological substitutes, synthetic substitutes are free from any risk of disease transmission. However, only a few synthetic skin substitutes are on the market today. Although synthetic materials have greater structural integrity compared to natural products, its poor bioactivity remains a major concern [17]. In addition, poor host response may lead to negative effects on scar quality. Despite limitations in materials and cost, the development of bio-engineered skin is increasing due to innovative possibilities with new techniques and biomaterials, providing a glimpse into a promising future. These steps have promoted a shift in focus from traditional surgical interventions to skin tissue-engineered regeneration.

2.3. Wound dressings

Wound dressings are developed for wound coverage to aid re-epithelialization, prevent wound infection, skin desiccation, and further skin damage. Wound dressings can be categorized into four groups: biological dressings; conventional dressings; biosynthetic dressings and antimicrobial dressings. Biological dressings include cadaver allograft skin (transplantation between individuals of the same species), xenograft (transplantation between individuals of different species) and human amnion, which have been adopted to temporarily cover wounds for reepithelialization. Although biological dressings are effective in terms of improving the quality of wounds for further skin grafting, they cannot be used as a permanent skin replacement due to immunological disparities [8]. Furthermore, many issues are associated with biological dressings, such as inconsistent quality, limited supply, and the risk of pathogen transfer [44]. Conventional dressings which do not contain antibiotics or medications, e.g. Vaseline gauze or silicone sheets are also widely used to temporarily cover wounds during reepithelialisation. However, these dressing tend to adhere to the



Fig. 1. Autologous skin grafting: (A) partial-thickness skin graft taken from the healthy donor site. (B) Skin graft is meshed to increase the surface area (C) Application of skin graft on the wound bed.

Table 1
Skin substitutes used in burn care.

Skin substitute	Composition	Cells incorporated
KaroSkin [18]	Human cadaver skin with dermal and epidermal cells	
GraftJacket® [18,19]	Human acellular pre-meshed dermis	
StrataGraft™ [20]	Human dermal fibroblasts and stratified epidermis derived from Near-diploid Immortalised Keratinocyte S (NIKS)	
GlyaDerm® [21,22]	Glycerol preserved acellular dermal collagen-elastin matrix	
OASIS® Wound Matrix [23]	Porcine acellular lyophilized small intestinal collagen matrix	
XenoDerm [24,25]	Lyophilised acellular porcine dermis	
Permacol™ [26]	Porcine acellular diisocyanite cross-linked dermis	
PermaDerm™ [27,28]	Autologous keratinocytes and fibroblasts cultured with bovine collagen	
OrCel® [29]	Bilayered type I collagen matrix	Allogenic neonatal foreskin keratinocytes and fibroblasts
RenoSkin® [30]	Bilayer dermal matrix – silicone film and porous crosslinked bovine collagen	
TransCyte® [31,32]	Porcine collagen-coated nylon mesh	Allogenic neonatal human foreskin fibroblasts
Integra® [33–35]	Cross-linked bovine tendon collagen and glycosaminoglycan, and polysiloxane (silicone)	
Pelnac® [36,37]	Porcine tendon derived atelocolla gen type I, sponge layer with silicone film	
Hyalograft 3D [23]	Hyaluronic acid membrane	Autologous fibroblasts
Dermagraft® [38]	Bioabsorbable polygalactin mesh matrix	Human neonatal fibroblasts
TissueTech Autograft System [18]	Microperforated hyaluronic acid membrane	Autologous keratinocytes and fibroblasts
Suprathel® [39–42]	Synthetic copolymer – dl-lactide (>70%), trimethylenecarbonate and ε-caprolactone	
Laserskin® [43]	Hyaluronic acid membrane	Autologous keratinocytes and fibroblasts
MySkin™ [18,28]	Silicone support with a specially formulated surface coating	Autologous keratinocytes

wound surface and their need for frequent changes traumatizes the newly epithelialized surface and delays healing [45,46]. Biosynthetic dressings are designed to use materials that mimic the function of skin by replacing the epidermis or dermis, or both. Examples include Biobrane® and TransCyte®. Antimicrobial dressings are widely used in burn management to prevent wound infection, by minimising bacterial colonisation. A number of antimicrobial dressings have been introduced to burn care. These products can contain either silver (e.g. Aquacel AG), nano-crystalline silver (e.g. ACTICOAT), cadexomer iodine (e.g. Iodosorb), or honey as antimicrobials. Application of silver compounds on burn wounds was a major milestone in topical burn therapy, which remarkably reduced the incidence of burn wound induced sepsis and death.

2.4. Negative pressure wound therapy (NPWT)

Negative pressure wound therapy (NPWT; also known as vacuum assisted closure, topical negative pressure therapy or microdeformational wound therapy) is currently utilised in wound care for both large and small burns. It was first demonstrated to be effective at halting partial thickness wound progression compared to most other pharmacological means available [47,48]. NPWT was further utilised as part of temporary abdominal closure in acute burn patients or in a high risk burns patient who underwent pre-operative optimisation prior to skin grafting [49, 50]. Considering burn injury as the most traumatic and composite wounds, the use of NPWT has been acknowledged to be helpful by allowing surgical teams to effectively manage acute burns, as well as their chronic sequelae. Numerous case reports have been published supporting the ability of NPWT to improve both split thickness or full

thickness skin graft take, allowing early mobilisation of patients [51–55]. Additionally, this effect has been demonstrated both in the acute setting and when excising burn scar contractures [54,56–58].

The use of NPWT with skin substitutes or templates, such as Integra® has been utilised in clinical practice to improve poor skin graft take and infection [59,60], as NPWT has been hypothesized to prevent shear. Additionally, the optimised NPWT environment is thought to contribute to accelerated wound healing [61–63]. More studies have also shown that NPWT can also be effective to promote wound healing using Matrigel® or Pelnac® skin substitute with split skin graft [64,65]. The positive outcomes of NPWT associated with improved graft and dermal substitute take in burn injury has been attributed to reduced wound infection [54,66–68]. A case control study demonstrated a significantly reduced number of infections in the NPWT treatment compared to normal therapy [68]. Additionally, a study using a porcine burn model with locally applied *Pseudomonas aeruginosa* infection [69] found that NPWT effectively reduced bacterial proliferation and alleviated sepsis progression compared to usual wound treatment. Cytokine analysis and histopathological examinations confirmed that NPWT reduces wound inflammation and infection.

Recent evolutions in NPWT involve combining use with instillation or continuous irrigation. NPWT instillation (NPWTi) is a cyclical process whereby a solution of choice is instilled into a wound covered by NPWT dressings. Comparatively, NPWT continuous irrigation (NPWTci) is non-cyclical. This method involves the continuous irrigation of solution into the foam and wound bed, while the suction action of NPWT is continuously applied at the same time. Although, there are no clinical case reports on NPWTi and NPWTci systems in burn injury, these advanced systems could play a beneficial role in future burn wound management

via reducing number of treatment days, accelerated clearance of infection and wound closure [70]. A study assessing NPWTi suggested that NPWTi may promote wound granulation rates compared to normal NPWT [71]. When antimicrobial solutions were used in conjunction with NPWTi, a lower bioburden load was found when compared to standard dressings soaked with the same antimicrobial solution. This finding suggests that NPWTi has the potential to increase the efficacy of the antimicrobial solutions in burn care.

3. Burn wound infection: challenges and innovations

Wound infection is a major challenge in burn care and is the most common cause of mortality after burn injury [72–74]. Pathogens have evolved overtime in line with innovations and antibiotic use [75]. Currently, multi-drug resistant (MDR) organisms and fungi present a major challenge [76]. With limited new antibiotics [77] innovations enhancing the effectiveness of currently available topical and systemic antimicrobials is paramount to improve morbidity and mortality in burn patients.

Bacterial organisms causing burn wound infection can be classified into two groups, gram negative and gram positive (Table 2). Gram negative bacteria cause most burn wound infections, with similar incidence, prevalence and pathogens regardless of geography or institution [78, 79]. Burn wound infection by *Pseudomonas aeruginosa*, *Klebsiella pneumoniae* and *Escherichia coli* and the gram positive organism *Staphylococcus aureus* are independent predictors of mortality [72]. *Staphylococcus aureus* is the major cause of gram-positive burn wound infections globally [74] and a common cause of septicemia [76,80]. Methicillin resistant *Staphylococcus aureus* (MRSA) is now the major pathogen in some burn centres [74] and vancomycin resistant enterococci (VRE) although not as common, appears to be highly virulent [74].

3.1. Topical antimicrobial agents

Topical application of antimicrobial agents began with the application of sulfathiazole dressing to burns in 1940's [81]. With the discovery of the antibacterial properties of silver [82], silver sulfadiazine dressings have been the standard of care since 1968 [83]. Recent systematic reviews and meta-analysis report silver sulfadiazine to have poorer healing outcomes and little evidence of effectiveness in preventing wound infection than alternate dressings [84,85]. Mechanisms believed to be responsible for these outcomes are the requirement for regular wound dressing changes, poor eschar penetration and cytotoxicity of silver to keratinocytes and fibroblasts delaying wound healing [85–87]. A further issue is the recent report of resistance to silver in clinical isolates [88].

Table 2
Common pathogens causing burn wound infection.

Group	Species	Antibiotic treatment
Gram negative	<i>Pseudomonas aeruginosa</i>	Piperacillin-tazobactam
	<i>Acinetobacter baumannii</i>	Carbapenems
	Enterobacteriaceae	Cephalosporins
Gram positive	<i>Staphylococcus aureus</i>	Penicillins
	<i>Streptococcus</i>	Penicillins
	<i>Enterococcus</i>	Penicillins
Fungi	<i>Candida</i> spp.	Amphotericin B, Micafungin
	Non-albicans <i>Candida</i>	Amphotericin B
	<i>Aspergillus</i>	Caspofungin
Viruses	Blastomycosis	Fluconazole
	Herpes simplex	Acyclovir
	Varicella-zoster	Acyclovir
Drug resistant strains	MRSA	Vancomycin
	VRE	Linezolid
	(Extended-spectrum beta-lactamases) ESBL	Carbapenems
	MDR <i>P. aeruginosa</i>	Colistin

Numerous innovative approaches have then been investigated in the past decade to achieve alternate topical antimicrobial treatment for burn wounds which do not compromise wound healing, require less dressing changes and induce minimal antibiotic resistance (Table 3). Innovations generally involve new methods of delivering well-established antibiotics, alternative antimicrobials such as curcumin [89,90] or synthetic antimicrobial agents such as LLKKK18 [91], an antimicrobial peptide. Clinical trials comparing novel dressings containing LLKKK18 to the silver dressings in human burn cohorts will likely challenge conventional practice in the near future. Further investigation of the effectiveness of novel dressings against MDR bacteria is also required.

3.2. Systemic antibiotics

Overtime treatment with various systemic antibiotics have propagated the evolution of micro-organisms causing morbidity and mortality in burns [76]. Currently, drug resistant organisms pose a difficult challenge. MRSA is the most common resistant organism encountered in burns and is normally treated with an ancient antibiotic, vancomycin [76,92]. Despite various measures to control and treat MRSA, the organism has continued to evolve with vancomycin-intermediate susceptible *Staphylococcus aureus* (VISA), a developing strain. New antimicrobials including oxazolidinones, streptogramins, tigecycline, daptomycin, dalbavancin [74,76] remain an important addition to the antibiotic armamentarium to combat such infections [93].

In many burn centers, the predominant organisms are the gram negative pathogens *Pseudomonas aeruginosa* and *Acinetobacter baumannii* [76,78,79]. Resistance of these organisms to antibiotics, including reserve antibiotics such as the carbapenams [76], has seen increasing reliance on the use of old antibiotics [74]. Colistin, a polymyxin antibiotic, was once abandoned due to extensive side effects (neurotoxicity and nephrotoxicity) but has recently been reinvigorated to combat MDR gram-negative organisms [94]. This new reliance on colistin has triggered various efforts to redevelop and re-engineer the antibiotic [94]. One such effort is the utilisation of nano-antibiotic polymer therapeutics in which colistin is conjugated to dextrin. The dextrin-colistin conjugate exploits the enhanced permeability and retention effect (EPR) [95] enabling the molecule to be delivered preferentially to burn wounds and in a greater concentration for a prolonged duration with less systemic side effects. Pre-clinical studies establishing antimicrobial spectrum, pharmacodynamics and pharmacokinetic properties in an animal model have shown a reduced toxicity profile with favourable pharmacological properties [96,97].

Efforts to improve drug delivery of current antibiotics also offers promise. Critically ill patients post severe burn injuries have substantially altered physiology, which may profoundly lower antibiotic tissue concentrations culminating in therapeutic failure and the emergence of resistance. Physiological changes include altered fluid balance, a hyperdynamic circulation, renal and hepatic dysfunction and organ support [98]. Methods to improve drug delivery involve regular monitoring of antibiotic concentrations and the delivery of antibiotics via continuous infusion rather than bolus regimes [99]. For example a recent study demonstrated improved outcomes when *P. aeruginosa* infection was treated via extended infusions of piperacillin-tazobactam [100].

3.3. Anti-fungal infection

The incidence of fungal infections is increasing [101–103] with candida species being the most common [104]. In the USA, fungi are responsible for approximately two thirds of invasive burn wound infections [75] and is the fourth most common organism isolated in blood cultures [104]. Fungal infections, particularly non-candida infections such as *Aspergillus* spp. and moulds are associated with high mortality due to sepsis with multi-organ failure [76]. Clinically, fungal infection has an added comorbid effect equivalent to an additional burn involving 33% of the total body surface [105]. The most effective

Table 3

A summary of studies in topical antimicrobials.

Studies	Antimicrobial	Delivery system	Organisms tested	Year
1	Ciprofloxacin	Keratin hydrogel	<i>S. aureus</i> , <i>Psuedomonas</i>	2016
2	Gentamicin	Soy protein films	<i>Psuedomonas</i>	2015
3	Curcumin	Gel-core hyalurosomes	Nil	2015
4	Curcumin	Nanoparticle encapsulation	MRSA, <i>Psuedomonas</i>	2015
5	LLKKK18	Carbopol® hydrogel	Nil	2015
6	Gentamicin	Hybrid bi-layer wound dressing	<i>Psuedomonas</i>	2015
7	Nitric oxide	Polymeric film	<i>Acinetobacter baumannii</i>	2014
8	Mupirocin	Liposomes in hydrogel	<i>S. aureus</i>	2013
9	Gentamicin	Polyglyconate mesh and PDLGA matrix	<i>Psuedomonas</i>	2011
10	Lysostaphin	Chitosan-collagen hydrogel	MRSA	2011
11	Fucoidan-chitosan	Hydrogel	Nil	2008
12	Minocyclin	Chitosan film and tegaderm	Nil	2007
13	Chlorhexidine	Bilayered hydrogel and chitosan foam	<i>S. aureus</i> , <i>Psuedomonas</i>	2000
14	Amphotericin B	Nanoparticle hydrogel	<i>Candida</i> spp.	2014
15	Boron/pluronic copolymers	Carbopol-based hydrogel	Bacteria, yeast, fungi	2015

treatment for fungal infection is prevention via removal of burned tissue and closure of wounds [74]. Positive fungal wound cultures and fungaemia particularly if *Aspergillus* spp. or other moulds are cultured, are generally indications for systemic therapy although specific indications for treatment remain unclear [103]. Currently three classes of systemic antifungal drugs are available for treating fungal burn wound infection, including polyenes, azoles and echinocandins [104].

Polyenes such as amphotericin B are first line agents for the treatment of *Candida*, but its use is often limited by significant nephrotoxicity [74,76,104]. Lipid soluble formulations of amphotericin B although causing less side effects, are substantially expensive [104]. Of the azoles, fluconazole is the empiric choice for invasive candidiasis, although developing resistance is an issue [104]. Echinocandins the newest antifungal agent introduced in 2001, are effective with a unique mechanism of action inhibiting the synthesis of β -1,3-D-glucan polymers and has a more favourable toxicity profile and pharmacokinetics [76,104,106]. As with antibiotics, future research addressing the impact of burn injury induced physiological alterations on the pharmacokinetic profile of systemic antifungal therapies has yet to be elucidated.

Pathogens causing morbidity and mortality in burn wounds continue to evolve in response to dynamic pressure from antibiotic use. Future clinical application of novel antimicrobial wound dressings in place of the established silver dressings is promising. Systemically, challenges include enhancing the effect of current antibiotics through the pursuit of individualized dosing regimens which adjust for in altered physiology after burn wound, and the utilisation of the EPE to enhance systemic delivery are encouraging innovations. Fungal pathogens are emerging as more common and troublesome infections, continued development and enhanced understanding of the clinical use of antifungal drugs will benefit burns patients.

4. Burn pain management: challenges and innovations

Control of burn pain is central to the recovery and reintegration of burn injury patients. Poor control of pain can hamper the healing process due to fear and anxiety induced elevation of stress hormones (such as glucocorticoids). This can lead to long lasting physical and psychological burdens and hospital stay. Three mechanisms are counted to burn pain– nociceptive, neuropathic and inflammatory. Intravenous morphine forms the basis of pain management through early stage post-burn injury, while shorter half-life opioids and nerve blockers are alternative options. During the wound healing process, surgical wound debridement, physiotherapy and dressing changes increase pain levels substantially. Nociceptive pain levels decline as scars mature and physical therapy continues. Additionally, neuropathic type pain

may increase at this stage along with pregabalin, gabapentin used for treatment of painful neuropathy.

4.1. Issues of pain management

Post severe burn injury at 48 h, a hypermetabolic response begins which is known to effect on drug clearance rates, pharmacokinetics (PK) and pharmacodynamics (PD). However, lack of literature has been reported on the effect of burn-induced hypermetabolism on drug PK and PD [107,108]. As a result, appropriate dosing for a majority of drugs used in burn pain remains an area requiring further investigation.

Opioid reliance continues to define burn pain care [109]. The side effects of opioid use include: nausea, vomiting, constipation, gastrointestinal dysmotility, dependence and tolerance. Severity of burn pain and duration of treatment also predisposes to tolerance, escalation of doses and high risk of addiction. Moreover, opioid can induce hyperalgesia, an additional overarching state of enhanced pain sensitivity along with increasing doses of opioid [110]. Issues of opioid induced hyperalgesia presents a clinical challenge in the burn injury population [110]. In consideration of these effects, further investigation of opioid treatment options, alternatives, and adjuncts is paramount for burn patient care.

4.2. Treatments for burn pain

4.2.1. Ketamine

Ketamine forms a part of burn pain management for a number of decades, playing a major role in burns procedures since the 1960's due to potent analgesic, sedative and amnesic properties [111–113]. Recent literature exhibited the capability of ketamine to reduce primary and secondary hyperalgesia; while also increasing thermal injury induced mechanical pain thresholds [114,115]. Delivery of ketamine in burns patients can be conducted by intravenous (IV) infusion, intravenous patient-controlled analgesia (IV-PCA), intramuscular, oral and intranasal administration [116–120]. Studies examining the combination of ketamine with other classes of burn-pain pharmacology have been prominent, particularly in an effort to reduce sympathomimetic, sedative and psychomimetic side effects. Ketamine was first combined with benzodiazepines in attempt to reduce these adverse effects [121]. A study reported the positive effects of this combination in burns patients, indicating that ketamine in combination with midazolam successfully reduces the side effects of ketamine [120]. Further study provided evidence that combination of ketamine with propofol can be used to attenuate ketamine's unexpected effects [122]. The range of pharmaceuticals utilised in burn pain management provides opportunity to further

investigate ketamine synergism, particularly in the areas of background and breakthrough pain which have not been studied in previous research.

4.2.2. Sedatives and anxiolytics

Benzodiazepines have no analgesic properties but are extensively utilised in burns patients as adjuncts for pain management. It has been shown in recent burns pain studies that anxiety decreases pain tolerance and increases use of pain medications [123,124]. Recent studies exploring benzodiazepines in burns patients have largely centered on combination with other sedatives (such as ketamine and dexamethasone). $\alpha 2$ -agonists, similar to benzodiazepines, are used as sedatives and anxiolytics in burns. A clinical trial study of $\alpha 2$ -agonists indicated the potential for an additional role in pain reduction. However, this must be in consideration of $\alpha 2$ -agonist side effects; reduced sympathetic outflow and hypotension, which have complicated several burn studies to date [125,126].

4.2.3. Neuropathic drugs and pruritus

Pregabalin and gabapentin are anticonvulsants used in the treatment of neuropathic pain of burn injury patients. A case series in 2010 established the efficacy of pregabalin in reducing post-burn neuropathic pain scores of outpatients [127]. Subsequent randomised controlled trial demonstrated the role of pregabalin in both the acute and healing phases of burn injury [128,129], demonstrating reductions of neuropathic pain during the first four weeks of treatment, while additionally reducing pain levels during procedural pain events. Another study found that pregabalin had positive effects in reducing post-burn pruritus, suggesting that pregabalin can be used in any patient with moderate to severe pruritus [128]. Secondary to this, recommendation was made to utilise pregabalin in mild itch to achieve the benefit of rapid and complete response.

Gabapentin has also received attention from burn pain investigators. Regarding to the acute setting, conflicting evidence has arisen on the efficacy of gabapentin in reducing acute burn pain and opioid consumption. Studies have shown positive results with gabapentin via reduction of mechanical allodynia in experimental partial thickness wounds [130,131]. However, a randomised controlled trial study demonstrated that burn patients receiving gabapentin on day 1 of injury did not display any significant effects on either opioid consumption or acute burn pain [132]. This work is supported by a double-blinded, placebo-controlled, crossover study, showing no anti-hyperalgesic or opioid enhancing effects when gabapentin was used in an experimental superficial burn model [133]. Similar to pregabalin, gabapentin shows favourable results in the management of post-burn pruritus. Successful use of gabapentin in paediatric burns patients was reported as an alternative to antihistamines [134]. Subsequent studies further established gabapentin as a more effective alternative to antihistamine as monotherapy. Additionally, gabapentin was demonstrated as a first line therapy that leads to greater efficacy at each step up in therapy as part of a burns pruritus protocol. Other notable pharmacological forms of anti-pruritic therapy include naltrexone, ondansetron and botulinum toxin [135–138]. However, these are lacking substantial evidence and require further investigation in burns populations.

4.3. Future pain targets and considerations

Advances in pain and thermal sensation is essential to the development of future burn pain pharmacology. Identification of polymodal transient receptor potential (TRP) channels has been known to be the most important finding in pain research in the last few decades [139, 140]. As part of this family, TRP vanilloid-1 (TRPV1) is a key channel activated by heat, acidosis, chemical mediators and molecules such as capsaicin [141–145]. TRPV2 and TRP melastatin-3 have also been demonstrated to play a role in thermal nociceptive sensation [146,147].

Research using TRPV1 knockout (KO) mice showed that TRPV1 is a principle receptor responsible for thermal hyperalgesia during the

acute phase post burn injury (up to 24 h); while a recent study suggests a role for TRPV1 in thermally induced chronic pain [144,148–151]. TRPV1 channels were also found to be involved in thermally induced cell death [152], suggesting TRPV1 to be the target for both agonist and antagonist pain control in burn injury. However, systemic delivery may not be feasible as it has emerged that TRPV1 are implicit in homeostatic roles [153,154]. Overcoming the barriers of systemic TRPV1 modulation remains unclear in burn pain management, therefore, development and local application of TRPV1 modulators is expected to avoid undesirable systemic and central effects.

Voltage gated sodium channels (Na_v) have also garnered attention as pharmacological targets for pain control. While a number of channels have been identified in the literature, the three channel types which show most promise in burn injury are $\text{Na}_v 1.7$, $\text{Na}_v 1.8$ and $\text{Na}_v 1.9$. Both human studies and animal experiments have demonstrated the potential of $\text{Na}_v 1.7$ in burn patients demonstrating either complete absence of pain response or conversely pathological states of pain [155–158]. Animal models demonstrated the potential utility of $\text{Na}_v 1.7$ to diminish thermal and mechano-sensation in $\text{Na}_v 1.7$ knockout mice – particularly, in states of inflammation induced nociception [159,160]. A recent study further confirmed that tetrodotoxin, a $\text{Na}_v 1.7$ blocker, significantly reduced thermal hyperalgesia and mechanical allodynia in a thermal injury model [161].

Innovation in pain management continues to improve acute and long term outcomes for patients experiencing the trauma of burn injury. However, research surrounding burn pain analgesia can often suffer from a lack of specificity. Future work to improve the current state of management must not fall victim to a one-size-fits-all approach. The patient and burn characteristics, phase of injury, and category of burn pain must all be considered to adequately address the needs of burn teams in their clinical efforts. In consideration of this framework, the continued development and understanding of mechanisms which contribute to burn pain is an essential component for the development and improvement of novel therapies.

5. Scarring and management

With reduced mortality rates associated with severe burn injuries, the aim of burn wound care is shifting towards the management of burn scars [162]. Hypertrophic and keloid scarring in burn patients causes significant morbidity often with poor functional and cosmetic outcomes (Fig. 2). Burn scars can lead to causes many debilitating factors including pain, pruritus, dyspigmentation, heat intolerance, and limited range of motion due to scar contraction. The challenge in the treatment of post burn injury scarring lays on finding new therapeutic targets by an enhanced understanding of scar formation.



Fig. 2. 16-year-old patient with keloid scarring 6 months following a 25% TBSA flame burn injury. The wounds on his left upper arm and left chest were excised and grafted.

5.1. Stem cells

Stem cell based wound healing therapies represents as a new promising modality for the treatment of fibrosis, scarring and treatment for wound contraction. Post injury, stem cells are heavily involved in all overlapped phases of wound healing [163–165]. Endogenous stem cells migrate to the site of injury during the initial inflammatory phase, where they elicit immunomodulation effects, followed by accelerated wound closure, angiogenesis and re-epithelialization [166]. Mesenchymal stem cells (MSCs) and adipose derived stem cells (ADSCs) have been widely investigated in scar treatment, wound contraction and in the pathophysiology of scar formation [167]. MSCs are defined as self-renewing multipotent stem cells that can be differentiated into various lineages of mesenchymal origin [163]. A number of studies have shown the positive effects of bone marrow-derived MSCs in reducing hypertrophic scarring [168–170] via reduced expression of myofibroblast marker and the down-regulation of collagen I synthesis. Recent studies have further characterized two distinct phenotypes of MSCs involved in the wound healing process: the pro-inflammatory M1 and the anti-inflammatory M2 [166]. This polarization is mediated by different toll-like receptors (TLRs), which respond to various substances in the extracellular milieu and promote MSC phenotype switching accordingly to the current needs of the organism. Specifically, stimulation of TLR3 and TLR4 switches the cell between an anti-inflammatory and pro-inflammatory phenotype, respectively, resulting in control of cell migration, cytokine secretion and ECM deposition. This research leads to a new direction on the precise role of MSCs on wound healing and scar formation.

ADSCs have received considerable attention in skin regeneration as it has potential to regenerate hypodermis, dermis and epidermis [171]. An abundance of studies have supported the efficacy of fat grafting in both aesthetic and reconstructive cases [172–174], which has led to research into the utility of ADSCs in wound healing, regeneration of soft tissue, and reducing scarring [170,174,175] by its remodeling capacity provided by the unique cytokine and growth factor profiles. ADSCs treatment can diminish established hypertrophic scars and keloids by the inhibition of transforming growth factor-beta 1 (TGF- β 1) mediated fibroblast differentiation into myofibroblasts [164], and reducing collagen deposition by up-regulating matrix metalloproteinases (MMPs), which are capable of remodeling collagen in the wound site. In a recent study, burn wound exudate was shown to significantly increase ADSC secretion of vascular endothelial growth factor (VEGF) and interleukin-6 (IL-6), a pro-inflammatory cytokine [176]. ADSCs have also been found to promote vascularization and wound healing through VEGF secretion and a decrease in TGF- β 1 secretion in a mouse model [168]. In line with these conflicting results, there is evidence that ADSC stimulated cytokines promote granulation and angiogenesis in the wound bed, which has been linked to the production of hypertrophic scars in deep burns. However, few latest conflicting findings demonstrated in addition to increased inflammatory responses, ADSC stimulated cytokines promote granulation and angiogenesis in the wound bed, which linked to the production of hypertrophic scars in deeper burns [177–179]. Therefore, more research into the potential role of ADSCs in burn scar treatment is required.

Innovations for the utility of stem cells in treating scars involve surgical fat grafting with stem cells or introducing novel delivery methods of stem cells into the wound site. A systematic review article showed that fat grafting together with ADSCs enhanced wound angiogenesis, decreased inflammation, improved burn scar size and quality in both murine models and human studies [172]. Sundew-inspired adhesive hydrogels combined with ADSCs was recently reported as an innovative method to deliver ADSCs and was found to promote a “suturing” effect to accelerate wound closure, although the effects on long term tissue remodeling or scar formation is not clear yet [180]. The effective delivery of adipose-derived stromal cells using ECM patch also holds promises to reduce scarring. Stem cell delivery along with an ECM patch can improve cell survival and proliferation, and significantly reduce fibrosis

[175]. Whether stem-cell-based therapies can be translated in clinical practice is still in debate. Furthermore, practical barriers associated with stem-cell-based therapies might restrict their utility in scarless wound healing post burn injury [181]. To address this limitation, conditioned media from umbilical cord-MSC cultures may be used to treat wounds. Dermal fibroblasts were found to exhibit characteristics comparable to fetal fibroblasts, showing low capacity to form myofibroblasts [182]. Wounds treated with such umbilical cord-MSC-conditioned media healed faster with decreased collagen accumulation. Additionally, human amniotic-fluid-derived MSC-conditioned media also have the potential to inhibit the pro-fibrotic actions of TGF- β 1 and even reverse the myofibroblasts phenotype to a fibroblast-like state in vitro. Taken together, innovations in stem cell studies provide more therapeutic options for treating burn scars.

5.2. Pharmacological approaches

5.2.1. Silicone

Silicone has been one of the gold standard for non-invasive burn scar management of hypertrophic scars for years [183–185]. A recent randomized clinical trial showed that the use of silicone caused a significant reduction in pain and pruritus at 6 months post-burn injury. The use of pressure garments to reduce scar thickness, with silicone provided the most optimal results [186]. Silicone gel sheets, however, are difficult to use in anatomical areas subject to a high range of motion such as joints and can be undesired in exposed areas such as the face. To overcome these challenges, fluid silicone gel has been developed to allow for a thin layer of silicone gel to dry into a transparent silicone sheet [183–185]. Silicone is also being used to develop new devices such as the Embrace, a dynamic stress shielding device showing good results, however, evidence is still limited [187].

5.2.2. Corticosteroids

Injection of corticosteroids into hypertrophic scars has been considered a first-line of treatment for small hypertrophic scars scarring since 1960s. Corticosteroids are reported to reduce the height and volume of scars, decrease pain and pruritus and make scars more pliable [185,188]. Triamcinolone acetonide, a long acting corticosteroid, is commonly used to treat post burn scarring. It is thought that corticosteroids decrease collagen synthesis, attenuate cell proliferation of fibroblasts and keratinocytes, and accelerate collagen degradation by activating collagenase and suppressing TGF β [189–191]. However, the optimal dose of corticosteroids for treatment have yet to be determined [185,188], and the side effects of its use including hypopigmentation and subcutaneous atrophy [192] only represent a few limitations. A new approach combining corticosteroids (Triamcinolone) and 5-fluorouracil (a pyrimidine analog antimetabolite chemotherapy agent) was shown to reduce pain and have less side effects compared to the use of Triamcinolone alone [193,194]. Furthermore, this combination in use with a pulsed-dye laser has the potential to decrease scar volume and improve softening of hypertrophic scars and keloids [195].

5.2.3. Transforming growth factor- β modulators

TGF- β is a cytokine implicated in the pathogenesis of keloids and hypertrophic scarring. It is produced and released by platelets, fibroblasts, endothelial, epithelial and inflammatory cells and acts as a potent inducer of myofibroblasts differentiation [196]. Previous studies have confirmed that TGF- β 1 and TGF- β 2 promotes collagen synthesis and scar formation with significantly increased mRNA expression in keloids. In contrast, TGF- β 3 is involved in scar prevention. Numerous research and multiple therapies targeting the TGF- β pathway have been proposed as advanced treatments for scars, aiming to reduce TGF- β activities, increase TGF- β 3 expression, and utilize TGF- β inhibitors such as decorin [197], proteolytic inhibitors such as mannose-6-phosphate [198] and neutralized antibodies [199]. A recent study used antisense oligonucleotides to block the effect of TGF- β 1 over wound healing and

tissue remodeling [196], and showed significantly reduced expression of TGF- β 1 and β 2 with upregulation of TGF- β 3 [200]. However, to date, only the injection of TGF- β 3 into surgical wounds have been used to modulate the TGF- β pathway, demonstrating significant improvements in scar appearance in a randomized, controlled clinical trial [201].

5.2.4. Botulinum toxin A

Botulinum toxin A is a neurotoxin produced by bacteria that inhibits acetylcholine release from a neuron, and suppresses muscle contraction for approximate 2–6 months. Botulinum toxin has shown great potential as an additional tool to improve scar formation. Gassner and colleagues first reported that injection of botulinum toxin into the underlying musculature of wound sites significantly enhanced wound healing and minimized scar formation [202]. When botulinum toxin A is injected into a wound, it causes flaccid paralysis of the surrounding muscle, decreasing mechanical forces and thereby reducing wound tension [196,203]. Increased wound tension has been theorized to stimulate fibroblasts to differentiate into myofibroblasts and increase alpha-smooth muscle actin production, causing the wound to contract. Thereafter, many studies were conducted by intralesional injection of botulinum toxin A to treat keloids or mature scar [204]. Clinical studies followed up to 1 year showed that all patients that received botulinum toxin A had excellent scar regression and flattening of the lesions. Although the underlying mechanisms are still under investigation, researchers suggest that botulinum toxin suppresses fibroblast proliferation and differentiation by inhibiting the expression of TGF- β 1 [205]. In a recent study, reduction of the tensile force by botulinum toxin A injection into the musculature adjacent to the wounds may represent a comprehensive mechanism of action for aesthetic improvement of post-surgical scar [206]. All reported studies of botulinum toxin A have focused on the improvement of scars however, whether botulinum toxin A can prevent scar formation has yet to be established, and more extensive research is required.

5.3. Surgical approaches

5.3.1. Fat grafting

Fat grafting as a surgical approach in treatment of burns has several documented benefits and is a rapidly evolving field for scar modulation [207–209]. Functional and aesthetic improvements were shown in split-scar patients with reduced scar thickness and significantly better results in pain, colour, thickness, pliability/movement and shape/relief after 3 months [208]. A study on adherent burn scars showed a significant improvement in elasticity and maximal extension, and of the patient observer scales score at the 3-month follow up [210]. In addition, colour difference remained unaffected [210]. Fat grafting was also reported to enhance wound closure and improve neuropathic pain [211–215], and is further supported by a study that evaluated the effects of fat-grafting on burn-induced neuropathic pain in rat-models [216, 217]. Moreover, several case series have demonstrated great improvements in subjective patient ratings of burn scars, with improvements in texture, softness, thickness, colour, and wound healing which were also correlated histological findings [208,218–221]. ADSCs are believed to play a key role in fat grafting as they reduced collagen deposition and promoted vascularization and wound healing.

5.3.2. Laser therapy for scar modulation

Keloid and hypertrophic scarring occurs by the loss of control mechanisms involved in biosynthesis and tissue degradation during the wound healing process, resulting in excessive collagen production and fibroblast proliferation [222]. Although contraction is part of the regular wound healing process [222], the contraction of healing burn wounds, grafts and scar tissue results in abnormal scar formation [223,224]. In addition, prolonged wound-healing and the inflammatory nature of burn wounds lead to hypertrophic scarring [225], followed by pigment

alteration due to a dysbalanced amount of melanin or erythema [226, 227]. Laser treatment may serve as a useful modality for the treatment of hypertrophic and keloid scars [228] that have shown very encouraging results [207,229]. Over the last decade, laser therapy has become increasingly popular and is increasingly a crucial part in reconstructive burn surgery [224,230].

5.3.3. Pulsed dye lasers (PDL) & neodymium-doped yttrium-aluminium-garnet (Nd:YAG) lasers

Wavelength-specific lasers, such as the PDL and Nd:YAG laser, that selectively absorb oxyhaemoglobin and thus ablate blood vessels, have been widely analysed for the treatment of hypertrophic burn scars [151,207,231]. The mechanism of these vascular lasers still remains unclear, however, it has been postulated that selective photothermolysis by targeted vascular destruction leads to tissue hypoxia, collagen fibre heating, dissociation of disulfide bonds, catabolism and decreased cellular function, which consequently prevents excessive collagen deposition, and results in collagen fibre realignment and remodelling [223, 232]. The relative hypoxia of fetal skin leads to an upregulation of hypoxia-inducible factor I, which subsequently upregulates oxygen-dependent genes such as VEGF and TGF- β 3, that results in a decrease in fibroblast activity [233]. Histological analysis of scars treated with a 585-nm PDL revealed that thick hyalinised dermal collagen with haphazardly arranged fibroblasts became looser, with less coarse collagen fibres and a normal number of fibroblasts [234]. A meta-analysis including 28 well-designed clinical trials [232] showed that the 585/595-nm PDL is effective in improving overall scar appearance and reducing both scar height and degree of erythema of hypertrophic scars. It was also found that to 585/595-nm PDL decrease scar pruritus [224,235], and may result in a synergistic action when combined with intralesional steroid or 5-fluorouracil injections [236].

5.3.4. Non-ablative and ablative fractional lasers

Novel techniques such as non-ablative and ablative fractional resurfacing techniques are also developed to rehabilitate burn scars [224,230,237]. Fractional lasers affect the scars only in small areas leaving the surrounding skin uninjured, which serve as a reservoir of viable tissue to stimulate neocollagenesis and tissue remodelling [229,237]. Non-ablative fractional lasers (NAFL) are particularly effective for the treatment of atrophic, flat or mature scars [237]. A clinical trial demonstrated that treatment of burn scars with NAFLs significantly improved scar quality over a 6-month follow-up, and through histological analysis collagen resembled normal skin [238]. However, it seems that treatment of burn scars with an ablative fractional laser (AFL) triggers a more vigorous reaction of neocollagenesis and tissue-rearrangement when compared to NAFL [237,239].

Histological analysis demonstrates that the distinct AFL injury promotes a cascade of heat shock proteins, matrix metalloproteinases, and inflammatory processes, which result in a rapid wound healing response with prolonged neocollagenesis and ensuing collagen remodelling [240–243]. Tension of the scar is released and the dermal architecture is rearranged by this remodelling process, leading to improved pliability, decreased thickness, and better function [243–249]. Several groups reported a significant decrease in scar assessment scores and a significant decrease in scar thickness when measured by ultrasound [230,240,242–244,250,251]. Recent reports demonstrate that even one treatment with the AFL can lead to significant improvements in scar quality (Fig. 3) and symptoms (such as pruritus and neuropathic pain), heat-sensitivity, and a measured decrease in scar thickness, irrespective of the scar maturation status [230,244].

Laser-facilitated drug delivery combined with fractional resurfacing is another advanced technique to further enhance pliability and reduce thickness [252,253]. This combination can help to reduce adverse effects from the local accumulation of intralesional injected steroids or 5-fluorouracil by assisting the even distribution of the drug in the dermis [252, 254–256]. Laser treatment represents a therapy that has the potential to

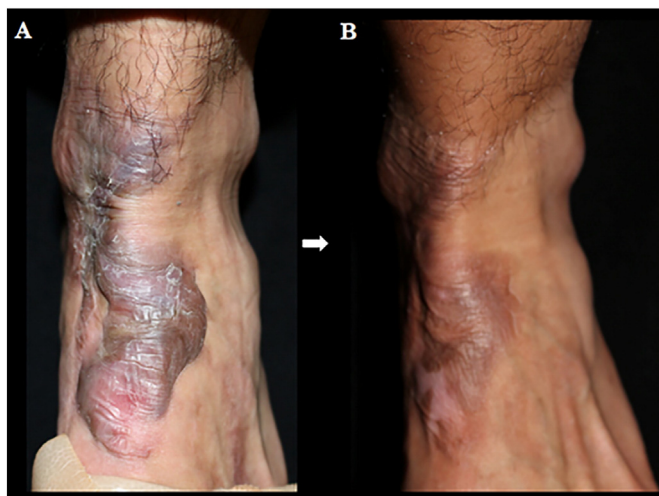


Fig. 3. (A) A 19 year old patient with hypertrophic scarring on his left ankle 7 months following a burn injury with hot oil. The scar was unstable and led to frequent wound breakdown depending on the footwear. (B) 6 weeks following one treatment with the ablative fractional Ultrapulse® CO₂ laser and laser facilitated steroid infiltration, the scar has flattened, became more pliable, pain and itchiness improved, and wound breakdown subsided.

fill the gap between non-surgical treatment options and surgical interventions for scar management [237]. It can also be used before surgery to make scars more pliable, to potentially improve surgical outcomes [230] or in scar rehabilitation to limit the development of contractures, promoting early mobility and thus accelerating the entire rehabilitative process [237,249]. However, the exact mechanisms by which lasers can achieve beneficial scar modulation remain unclear. Most data supporting its clinical application stem from retrospective series, thus being prone to selection bias and similar. Therefore, more prospective studies using in depth molecular analyses to explain treatment effects and patient outcomes are required to determine the exact role for this novel, promising therapeutic strategy for burn scars.

6. Skin tissue engineering in treatment of burn injury

Skin tissue engineering provides a new tool in treatment of burn injury, aiming to enhance wound healing, trigger skin regeneration and reduce the long term consequences of scarring [257]. The ultimate goal of skin tissue engineering is to produce a 'living skin' which offers completely functional skin, including all skin appendages to replace human skin in its entirety. Challenges and problems in skin tissue engineering include poor vascularization; delayed cell infiltration and migration due to undesirable surface structure or lack of cell-recognition signals or poor mechanical strengths. Researchers therefore pay their attentions into developing innovative biomaterials, advanced formulation technologies or new methods to incorporate autologous cells (e.g. keratinocytes, dermal fibroblasts, melanocytes) in order to increase the complexity of the skin substitutes. The applications of skin tissue engineering are broad, ranging from basic scaffolds to aid the growth of the neodermis, to the delivery of biomolecules such as growth factors, and stem cells to enhance wound healing and skin regeneration with full functional skin.

6.1. Biomaterials in skin tissue engineering

Both natural polymers and synthetic polymers have been widely investigated in skin tissue engineering. Natural polymers are isolated from a variety of sources including animal (collagen, hyaluronic acid), seaweed (agarose, alginate) or bacteria. They are soluble in aqueous or buffer solutions and are biodegradable *in vivo*. Collagen and hyaluronic acid (HA) are two major components for skin regeneration

and have been fabricated as sponges, films, matrices and gels to facilitate wound healing utilising the biodegradability and biocompatibility properties that make these polymers favourable [258–262]. Collagen scaffolds promote the strong attachment and proliferation of keratinocytes and dermal fibroblasts with various collagen based skin templates such as Biobrane™, MatriDerm®, Apligraf® and Integra® currently used in burn injury wound care. Hyaluronic acid (HA) is a natural polysaccharide composed of *N*-acetylglucosamine and glucuronic acid sugar units. HA is a structural component of skin ECM and due to its low immunogenicity, it has been considered as an ideal biomaterial for wound repair [260–262]. However, poor physical properties of HA represent serious limitations for its medical application. Chemical modification with enhanced resistance to degradation has significantly improved physical properties of HA, resulting in two commercial products: Hyalomatrix and Hyalofase that are currently used in burn treatment as dermal substitutes.

6.1.1. Elastin

Elastin is an essential component of native human skin, providing structural and cell mediating functions [263]. Recent studies demonstrated that incorporation of elastin in skin substitutes can improve scar quality and enhances angiogenesis [264,265]. In a clinical study, collagen-elastin scaffolds were found to reduce wound contraction and improve scar tissue architecture [265]. Reduced wound contraction was attributed to a decrease in the number of fibroblast differentiating into myofibroblasts [263,266]. The ability of elastin to enhance angiogenesis has led to the introduction of one-step procedures in burn surgery. Currently, many dermal skin substitutes involve two-step procedures as skin substitutes need time to be fully integrated with the wound bed and vascularized, prior to a secondary surgical step of split-thickness skin grafting. Otherwise, skin grafting will fail and result in skin graft loss and poor take. MatriDerm® is the first commercialized skin substitute containing elastin. MatriDerm® has been proven to be applicable in one-step procedures without a reduction in skin graft taken rate [267–271] and it was also found effective in a range of acute and reconstructive burn wounds.

6.1.2. Silk

Silk is composed of two proteins, fibroin and sericin. The favourable characteristics of silk make it as an optimal biomaterial in skin tissue engineering to enhance biocompatibility, mechanical structure with controllable biodegradability [272,273]. Silk are widely investigated in tissue engineering studies, including hard tissue, e.g. bone/cartilage and soft tissue, e.g. eye and neuron regeneration. In skin tissue engineering, silk was found to promote collagen synthesis from dermal fibroblasts [274] and 3D silk scaffolds are being optimized as a new generation of skin substitutes. A recent study showed that scaffold incorporation of silk and hair-derived keratin as a dermal substitute, resulted in better cell adhesion and proliferation of fibroblasts, and increased collagen protein expression [275]. The advantages of using silk in skin tissue engineering are further expanded in combination with keratin, collagen, elastin, chitosan, fibronectin and synthetic materials [272–274].

6.2. Advanced technologies in skin tissue engineering

6.2.1. Recombinant proteins

In addition to advancements in biomaterials, artificial ECM proteins are being produced to diminish the risk of disease transmission and to reduce host immunological responses, yet maintain a high bioactivity for wound healing [17]. Typically using *E. coli* or yeast systems, recombinant proteins with designed structures and functions can be produced by genetic manipulation. A variety of recombinant proteins are currently being investigated, such as tropoelastin, a precursor to elastin. Our research team recently investigated the effects of tropoelastin incorporated into Integra®, and found that tropoelastin has a comparable

function to bovine derived elastin [264]. Tropoelastin incorporated Integra® significantly increased angiogenesis in both mouse and porcine wound repair models compared to skin grafting only [264]. In contrast, injection of tropoelastin into a porcine hypertrophic burn scar showed no changes in scar flexibility [276]. Research into recombinant collagen is also under taking, but the complex structure of collagen makes it more challenging in tissue engineering. The production of recombinant collagen is limited by the lack of an animal-cell-specific, post-translational enzyme system. Although the gram-positive bacterium, *Streptococcus pyogenes*, has been found to produce a stable and thermal resistant collagen, it lacks specific binding sites native to human collagen [277,278]. This represents the framework for recombinant collagen engineering and further studies aim to improve its complexity in comparing to native human collagen. Recombinant proteins as building blocks for skin substitution is progressing, limitations including the lack of production in commercially required quantities and insufficient knowledge regarding protein structure-function relationships needs to be overcome [278].

6.2.2. Electrospinning

Electrospinning was developed over 30 years ago but has attracted new interest in the last decade [279]. Using this technique, nano-scale fibers can be produced to mimic ECM in native human skin. A major advantage of electrospinning technology is the ability to spin both natural polymer such as silk, chitosan, gelatin, fibrinogen and synthetic materials such as Poly (ϵ -caprolactone (PCL), polyurethane (PU), poly (glycolic acid) (PGA) and poly (L-lactide) PLLA [280–282] or copolymer together. Research attempts to combine various natural and synthetic materials to enhance the biocompatibility as well as the mechanical strength of scaffolds [282]. In order to overcome poor infiltration of cells into electrospun scaffolds due to low porosity and small pore size, many advanced methods have been developed, including leaching of selective water-soluble fibers [283] and cell-nanofiber fabrication. A successful approach involved repeatedly seeding fibroblasts layer-by-layer into electrospun scaffolds until the desired number of cell-nanofiber layers were achieved [284]. Adoption of coaxial electrospinning system has also shown excellent results in cell infiltration and proliferation [285]. Additionally, the unique properties of coaxial electrospinning system has been utilised in drug and bioactive molecule delivery as it can protect the bioactivity of bioactive molecules by spinning two separate layers of nanoscale fibers that contain both polymer solution and bioactive molecules [281].

6.2.3. Three-dimensional (3D) bioprinting

3D bioprinting is an advanced manufacturing platform that enables the predefined deposition of living cells, biomaterials and growth factors using computer-aided design (CAD) technology. Bioprinting technology has the potential to directly create graded macroscale architectures to better mimic the natural ECM, thereby augmenting cell attachment and proliferation [286]. Bioprinting is also to produce microfeatures, such as ridges and modulated surfaces by using multiple material platforms with various dispensing mechanisms [287]. There are three important steps in 3D bioprinting, including pre-processing (3D model generation and bio-ink preparation); 3D bioprinting and post-processing (using a bioreactor for tissue regeneration). 3D bioprinting of skin is receiving great attention as it can rapidly produce an even landscape by allowing an increasingly controlled and precise deposition of cells into a predefined tissue structure. Various biomaterials have been evaluated for bioprinting skin such as collagen [288], bovine gelatin [289], chitosan and silk fibrin [290], also synthetic biopolymers being PCL, poly (lactic acid) PLA, poly (lactide-co-glycolide) PLGA and PLLA [291] or human plasma [292]. Additionally, keratinocytes and fibroblasts are two widely studied cell types in 3D bioprinting skin [289]. A recent study successfully demonstrated the deposition of 20 layers of fibroblasts and 20 layers of keratinocytes, respectively, in collagen using laser-based bioprinting. The results showed the

presence of cadherin and connexin 43 in the epidermis, which are fundamental for tissue morphogenesis and cohesion [293]. Furthermore, a 3D bioprinted skin using keratinocytes and fibroblasts in collagen showed good graft take, and blood vessel formation/growth in the surrounding wound area in nude mice at 11 days post transplantation [294]. In order to regenerate fully functional skin, co-bioprinting melanocytes with keratinocyte; or incorporating endothelial cells to facilitate angiogenesis should be evaluated in future studies [291]. The integration of pilosebaceous units, with hair follicles and sebaceous glands in skin substitutes remains an unsolved challenge.

Skin tissue engineering has evolved from two dimensional supports to multifunctional complex systems, providing platforms for support, drug delivery, and the foundation for tissue regeneration. Limitations in current available products have led to developments in the use and applications of new materials and techniques. These advancements aim to optimize skin substitutes and ultimately improve patient quality of life.

7. Conclusion

Significant advancements have been made in burn injury management and research in the past decade, including developments in novel skin substitutes; application of new antimicrobial wound dressings and enhanced systemic drug delivery for wound infection; testing new pharmacological interventions and finding new targets for wound pain control; together with advanced surgical approaches such as laser therapy, fat grafting, skin grafting and coverage options. As a result, the survival rate of severe burn injury patients has been improved with significantly reduced hospital stay, resulting in decreased costs to patients and medical providers. However, several challenges still need to be solved to continue to improve current burn care. In particular, investigating how to accelerate wound healing, attenuate burn induced hypermetabolic-catabolic conditions, control systemic infection and reduce the overall time for functional recovery, should be prioritized. Further research will continue to optimize current treatment paradigms and identify novel targets in burn care to ultimately enhance the outcome for severe burn injury patients.

References

- [1] R.F. Pereira, et al., Advanced biofabrication strategies for skin regeneration and repair, *Nanomedicine* 8 (4) (2013) 603–621.
- [2] F.M. Wood, Skin regeneration: the complexities of translation into clinical practise, *Int. J. Biochem. Cell Biol.* 56 (2014) 133–140.
- [3] Z. Ruzczak, Effect of collagen matrices on dermal wound healing, *Adv. Drug Deliv. Rev.* 55 (12) (2003) 1595–1611.
- [4] S.L. Hansen, et al., Using skin replacement products to treat burns and wounds, *Adv. Skin Wound Care* 14 (1) (2001) 37–46.
- [5] B.A. Rubis, et al., The use of split-thickness dermal grafts to resurface full thickness skin defects, *Burns* 28 (8) (2002) 752–759.
- [6] J. Schulz Iii, R. Tompkins, J. Burke, Artificial skin, *Annu. Rev. Med.* 51 (1) (2000) 231–244.
- [7] S.T. Boyce, et al., Cultured skin substitutes reduce donor skin harvesting for closure of excised, full-thickness burns, *Ann. Surg.* 235 (2) (2002) 269–279.
- [8] E.S. Garfein, D.P. Orgill, J.J. Pribaz, Clinical applications of tissue engineered constructs, *Clin. Plast. Surg.* 30 (4) (2003) 485–498.
- [9] V.C. van der Veen, et al., Biological background of dermal substitutes, *Burns* 36 (3) (2010) 305–321.
- [10] A.S. Halim, T.L. Khoo, S.J. Mohd Yusoff, Biologic and synthetic skin substitutes: an overview, *Indian J. Plast. Surg.* 43 (S1) (2010) S23–S28.
- [11] J.T. Shores, A. Gabriel, S. Gupta, Skin substitutes and alternatives: a review, *Adv. Skin Wound Care* 20 (9) (2007) 509–511.
- [12] D.M. Supp, S.T. Boyce, Engineered skin substitutes: practices and potentials, *Clin. Dermatol.* 23 (4) (2005) 403–412.
- [13] S. Böttcher-Haberzeth, T. Biedermann, E. Reichmann, Tissue engineering of skin, *Burns* 36 (4) (2010) 450–460.
- [14] S. Enoch, J.E. Grey, K.G. Harding, Abc of wound healing: recent advances and emerging treatments, *Br. Med. J.* 332 (7547) (2006) 962–965.
- [15] B.S. Atiyeh, S.N. Hayek, S.W. Gunn, New technologies for burn wound closure and healing – review of the literature, *Burns* 31 (8) (2005) 944–956.
- [16] P. van Zuijlen, et al., Tissue engineering in burn scar reconstruction, *Burns Trauma* 3 (1) (2015) 18.

- [17] P.S. Low, M.S. Tjin, E. Fong, Design and construction of artificial extracellular matrix (aECM) proteins from *Escherichia coli* for skin tissue engineering, *J. Vis. Exp.* 100 (2015).
- [18] R.V. Shevchenko, S.L. James, S.E. James, A review of tissue-engineered skin bioconstructs available for skin reconstruction, *J. R. Soc. Interface* 7 (43) (2010) 229–258.
- [19] R. Nathoo, N. Howe, G. Cohen, Skin substitutes: an overview of the key players in wound management, *J. Clin. Aesthet. Dermatol.* 7 (10) (2014) 44–48.
- [20] M.J. Schurr, et al., Phase I/II clinical evaluation of StrataGraft: a consistent, pathogen-free human skin substitute, *J. Trauma Injury Infect. Crit. Care* 66 (3) (2009) 866–874.
- [21] A. Pirayesh, et al., Development of a novel dermal substitute based on glycerinated allo graft: clinical (phase i) and experimental evaluation, *J. Burn Care Res.* 27 (2 Supplement) (2006) S58.
- [22] A. Pirayesh, et al., Glyaderm® dermal substitute: clinical application and long-term results in 55 patients, *Burns* 41 (1) (2015) 132–144.
- [23] S. Shahrokhi, A. Arno, M.G. Jeschke, The use of dermal substitutes in burn surgery: acute phase, *Wound Repair Regen.* 22 (1) (2014) 14–22.
- [24] S.N. Hosseini, S.N. Mousavinasab, M. Fallahnezhad, Xenoderm dressing in the treatment of second degree burns, *Burns* 33 (6) (2007) 776–781.
- [25] S.N. Hosseini, et al., Xenoderm versus 1% silver sulfadiazine in partial-thickness burns, *Asian J. Surg.* 32 (4) (2009) 234–239.
- [26] T.M. MacLeod, et al., Evaluation of Permacol™ as a cultured skin equivalent, *Burns* 34 (8) (2008) 1169–1175.
- [27] M. Varkey, J. Ding, E.E. Tredget, Advances in skin substitutes—potential of tissue engineered skin for facilitating anti-fibrotic healing, *J. Funct. Biomater.* 6 (3) (2015) 547–563.
- [28] K. Vig, et al., Advances in skin regeneration using tissue engineering, *Int. J. Mol. Sci.* 18 (4) (2017) 789.
- [29] J. Still, et al., The use of a collagen sponge/living cell composite material to treat donor sites in burn patients, *Burns* 29 (8) (2003) 837–841.
- [30] B. Fabienne, et al., Development of a new bilayer dermal matrix, RENOSKIN®: pre-clinical data, *Burns* 33S (1) (2007) S105.
- [31] J. Noordenbos, C. Doré, J.F. Hansbrough, Safety and efficacy of Transcyte for the treatment of partial-thickness burns, *J. Burn Care Rehabil.* 20 (4) (1999) 275–281.
- [32] H. Amani, W.R. Dougherty, S. Blome-Eberwein, Use of Transcyte® and dermabration to treat burns reduces length of stay in burns of all size and etiology, *Burns* 32 (7) (2006) 828–832.
- [33] E. Dantzer, F.M. Braye, Reconstructive surgery using an artificial dermis (Integra): results with 39 grafts, *Br. J. Plast. Surg.* 54 (8) (2001) 659–664.
- [34] J.D. Frame, et al., Use of dermal regeneration template in contracture release procedures: a multicenter evaluation, *Plast. Reconstr. Surg.* 113 (5) (2004) 1330–1338.
- [35] D.Q.A. Nguyen, T.S. Potokar, P. Price, An objective long-term evaluation of Integra (a dermal skin substitute) and split thickness skin grafts, in acute burns and reconstructive surgery, *Burns* 36 (1) (2010) 23–28.
- [36] S. Suzuki, et al., Review of acellular and cellular artificial skins, *Tissue Eng.* 2 (4) (1996) 267–275.
- [37] W. Widjaja, P. Maitz, The use of dermal regeneration template (Pelnac®) in acute full-thickness wound closure: a case series, *Eur. J. Plast. Surg.* 39 (2) (2016) 125–132.
- [38] J.F. Hansbrough, C. Doré, W.B. Hansbrough, Clinical trials of a living dermal tissue replacement placed beneath meshed, split-thickness skin grafts on excised burn wounds, *J. Burn Care Rehabil.* 13 (5) (1992) 519–529.
- [39] C. Uhlig, et al., Suprathel®—an innovative, resorbable skin substitute for the treatment of burn victims, *Burns* 33 (2) (2007) 221–229.
- [40] M. Keck, et al., The use of Suprathel® in deep dermal burns: first results of a prospective study, *Burns* 38 (3) (2012) 388–395.
- [41] L. Highton, C. Wallace, M. Shah, Use of Suprathel® for partial thickness burns in children, *Burns* 39 (1) (2013) 136–141.
- [42] S. Fischer, et al., Suprathel® for severe burns in the elderly: case report and review of the literature, *Burns* 42 (5) (2016) e86–e92.
- [43] P.K. Lam, et al., Development and evaluation of a new composite laserskin graft, *J. Trauma Injury Infect. Crit. Care* 47 (5) (1999) 918–922.
- [44] B.S. Atiyeh, S.W. Gunn, S.N. Hayek, State of the art in burn treatment, *World J. Surg.* 29 (2) (2005) 131–148.
- [45] H.-F. Liu, F. Zhang, W.C. Lineaweaver, History and advancement of burn treatments, *Ann. Plast. Surg.* 78 (2) (2017) S2–S8.
- [46] J. Boateng, O. Catanzano, Advanced therapeutic dressings for effective wound healing—a review, *J. Pharm. Sci.* 104 (11) (2015) 3653–3680.
- [47] M.J. Morykwas, et al., Use of subatmospheric pressure to prevent progression of partial-thickness burns in a swine model, *J. Burn Care Rehabil.* 20 (1) (1999) 15–21.
- [48] L.P. Kamolz, et al., Use of subatmospheric pressure therapy to prevent burn wound progression in human: first experiences, *Burns* 30 (3) (2004) 253–258.
- [49] M.O. Hardin, et al., An experience in the management of the open abdomen in severely injured burn patients, *J. Burn Care Res.* 33 (4) (2012) 491–496.
- [50] B.M. Parrett, et al., Fourth-degree burns to the lower extremity with exposed tendon and bone: a ten-year experience, *J. Burn Care Res.* 27 (1) (2006) 34–39.
- [51] N. Gumus, Negative pressure dressing for the treatment of high-voltage electrical burn injury, *J. Burn Care Res.* 31 (1) (2010) 215.
- [52] J.R. Heugel, et al., Treatment of the exposed achilles tendon using negative pressure wound therapy: a case report, *J. Burn Care Rehabil.* 23 (3) (2002) 167–171.
- [53] C. Koehler, et al., Wound therapy using the vacuum-assisted closure device: clinical experience with novel indications, *J. Trauma Injury Infect. Crit. Care* 65 (3) (2008) 722–731.
- [54] A.G. Landau, et al., Full-thickness skin grafts: maximizing graft take using negative pressure dressings to prepare the graft bed, *Ann. Plast. Surg.* 60 (6) (2008) 661–666.
- [55] N. Nugent, D. Lannon, M. O'Donnell, Vacuum-assisted closure — a management option for the burns patient with exposed bone, *Burns* 31 (3) (2005) 390–393.
- [56] S.J. Chong, et al., Full thickness burns over bilateral patella tendons — adjunctive hyperbaric oxygen therapy and negative pressure wound therapy for wound bed preparation and improved graft take, *Ann. Acad. Med. Singap.* 40 (10) (2011) 471–472.
- [57] N. Gumus, Negative pressure dressing combined with a traditional approach for the treatment of skull burn, *Niger. J. Clin. Pract.* 15 (4) (2012) 494–497.
- [58] I. Tevanov, et al., Negative pressure wound therapy (NPWT) to treat complex defect of the leg after electrical burn, *Chirurg* 111 (2) (2016) 175–179.
- [59] J.B. Lynch, T.S. Ismael, J.L. Kelly, Release of anterior neck burn contracture using artificial dermis and vacuum-assisted closure, *Plast. Reconstr. Surg.* 121 (1) (2008) 352–353.
- [60] W. McEwan, et al., Suction dressings to secure a dermal substitute, *Burns* 30 (3) (2004) 259–261.
- [61] M. Leffler, et al., The use of the artificial dermis (Integra®) in combination with vacuum assisted closure for reconstruction of an extensive burn scar—a case report, *J. Plast. Reconstr. Aesthet. Surg.* 63 (1) (2010) e32–e35.
- [62] N.S. Moiemien, et al., Topical negative pressure therapy: does it accelerate neovascularisation within the dermal regeneration template, Integra? A prospective histological in vivo study, *Burns* 36 (6) (2010) 764–768.
- [63] C.A. Park, et al., Outpatient reconstruction using Integra® and subatmospheric pressure, *Ann. Plast. Surg.* 62 (2) (2009) 164–169.
- [64] M.C. Bloemen, et al., Clinical effectiveness of dermal substitution in burns by topical negative pressure: a multicenter randomized controlled trial, *Wound Repair Regen.* 20 (6) (2012) 797–805.
- [65] V. Harish, P.K. Maitz, Uninterrupted continuous negative pressure wound therapy is safe and can facilitate engraftment of dermal regeneration templates, *J. Plast. Reconstr. Aesthet. Surg.* 67 (7) (2014) 1011–1013.
- [66] S. Poulakidas, A. Kowal-Vern, Facilitating residual wound closure after partial graft loss with vacuum assisted closure therapy, *J. Burn Care Res.* 29 (4) (2008) 663–665.
- [67] C.M. Psionos, et al., Use of gauze-based negative pressure wound therapy in a pediatric burn patient, *J. Pediatr. Surg.* 44 (12) (2009) e23–e26.
- [68] F. Zhang, et al., Using negative pressure wound therapy on microskin autograft wounds, *J. Surg. Res.* 195 (1) (2015) 344–350.
- [69] Y. Liu, et al., Negative pressure wound therapy decreases mortality in a murine model of burn-wound sepsis involving *Pseudomonas aeruginosa* infection, *PLoS One* 9 (2) (2014), e90494.
- [70] A. Gabriel, et al., Negative pressure wound therapy with instillation: a pilot study describing a new method for treating infected wounds, *Int. Wound J.* 5 (3) (2008) 399–413.
- [71] C. Lessing, R. James, S. Ingram, Comparison of the effects of different negative pressure wound therapy modes—continuous, noncontinuous, and with instillation—on porcine excisional wounds, *Eplasty* 13 (2013) 443–454.
- [72] L.C. D'Avignon, et al., Contribution of bacterial and viral infections to attributable mortality in patients with severe burns: an autopsy series, *Burns* 36 (6) (2010) 773–779.
- [73] N. Merchant, K. Smith, M.G. Jeschke, An ounce of prevention saves tons of lives: infection in burns, *Surg. Infect.* 16 (4) (2015) 380–387.
- [74] W. Norbury, et al., Infection in burns, *Surg. Infect.* 17 (2) (2016) 250–255.
- [75] B.A. Pruitt Jr., Reflection: evolution of the field over seven decades, *Surg. Clin. N. Am.* 94 (4) (2014) 721–740.
- [76] L.K. Branski, et al., Emerging infections in burns, *Surg. Infect.* 10 (5) (2009) 389–397.
- [77] M.S. Kinch, et al., An analysis of FDA-approved drugs for infectious disease: antibacterial agents, *Drug Discov. Today* 19 (9) (2014) 1283–1287.
- [78] E.A. Azzopardi, et al., Gram negative wound infection in hospitalised adult burn patients—systematic review and meta-analysis, *PLoS One* 9 (4) (2014), e95042.
- [79] A.C. Issler-Fisher, et al., Microbiological findings in burn patients treated in a general versus a designated intensive care unit: effect on length of stay, *Burns* 42 (8) (2016) 1805–1818.
- [80] R.L. Bang, et al., Burn septicemia in Kuwait: associated demographic and clinical factors, *Med. Princ. Pract.* 13 (3) (2004) 136–141.
- [81] H.J. Klasen, A historical review of the use of silver in the treatment of burns. II. Renewed interest for silver, *Burns* 26 (2) (2000) 131–138.
- [82] D.E. Marx, D.J. Barillo, Silver in medicine: the basic science, *Burns* 40S (1) (2014) S9–S18.
- [83] C.L. Fox, Silver sulfadiazine — a new topical therapy of *Pseudomonas* infection, *Arch. Surg.* 96 (2) (1968) 184–188.
- [84] J. Wasiak, et al., Dressings for superficial and partial thickness burns, *Cochrane Database Syst. Rev.* 3 (3) (2013), CD002106.
- [85] Z. Aziz, S.F. Abu, N.J. Chong, A systematic review of silver-containing dressings and topical silver agents (used with dressings) for burn wounds, *Burns* 38 (3) (2012) 307–318.
- [86] B.S. Atiyeh, et al., Effect of silver on burn wound infection control and healing: review of the literature, *Burns* 33 (2) (2007) 139–148.
- [87] Y. Yoshino, et al., The wound/burn guidelines - 6: guidelines for the management of burns, *J. Dermatol.* 43 (9) (2016) 989–1010.
- [88] P.J. Finley, et al., Unprecedented silver resistance in clinically isolated enterobacteriaceae: major implications for burn and wound management, *Antimicrob. Agents Chemother.* 59 (8) (2015) 4734–4741.

- [89] A.E. Krausz, et al., Curcumin-encapsulated nanoparticles as innovative antimicrobial and wound healing agent, *Nanomedicine* 11 (1) (2015) 195–206.
- [90] W.M. El-Rafea, et al., Novel curcumin-loaded gel-core hyalurosomes with promising burn-wound healing potential: development, in-vitro appraisal and in-vivo studies, *Int. J. Pharm.* 486 (1–2) (2015) 88–98.
- [91] J.P. Silva, et al., Improved burn wound healing by the antimicrobial peptide LLKKK18 released from conjugates with dextrin embedded in a carbopol gel, *Acta Biomater.* 26 (2015) 249–262.
- [92] B.M. Diederien, et al., Epidemiology of clinically relevant bacterial pathogens in a burn center in the Netherlands between 2005 and 2011, *J. Burn Care Res.* 36 (3) (2015) 446–453.
- [93] S.E. Trevino, M.H. Kollef, Management of infections with drug-resistant organisms in critical care: an ongoing battle, *Clin. Chest Med.* 36 (3) (2015) 531–541.
- [94] E.A. Azzopardi, et al., Colistin in burn intensive care: back to the future? *Burns* 39 (1) (2013) 7–15.
- [95] E.A. Azzopardi, E.L. Ferguson, D.W. Thomas, The enhanced permeability retention effect: a new paradigm for drug targeting in infection, *J. Antimicrob. Chemother.* 68 (2) (2013) 257–274.
- [96] E.A. Azzopardi, E.L. Ferguson, D.W. Thomas, Development and validation of an in vitro pharmacokinetic/pharmacodynamic model to test the antibacterial efficacy of antibiotic polymer conjugates, *Antimicrob. Agents Chemother.* 59 (4) (2015) 1837–1843.
- [97] E.L. Ferguson, et al., Dextrin-colistin conjugates as a model bioresponsive treatment for multidrug resistant bacterial infections, *Mol. Pharm.* 11 (12) (2014) 4437–4447.
- [98] J.A. Roberts, et al., Individualised antibiotic dosing for patients who are critically ill- challenges and potential solutions, *Lancet Infect. Dis.* 14 (6) (2014) 498–509.
- [99] F. Ravat, et al., Antibiotics and the burn patient, *Burns* 37 (1) (2011) 16–26.
- [100] T.P.J. Lodise, B. Lomaestro, G.L. Drusano, Piperacillin-Tazobactam for *Pseudomonas aeruginosa* infection: clinical implications of an extended-infusion dosing strategy, *Clin. Infect. Dis.* 44 (3) (2007) 357–363.
- [101] M.R. Capoor, et al., Epidemiological and clinico-mycological profile of fungal wound infection from largest burn centre in Asia, *Mycoses* 55 (2) (2012) 181–188.
- [102] J.V. Schaal, et al., Epidemiology of filamentous fungal infections in burned patients: A French retrospective study, *Burns* 41 (4) (2015) 853–863.
- [103] J. Ballard, et al., Positive fungal cultures in burn patients: a multicenter review, *J. Burn Care Res.* 29 (1) (2008) 213–221.
- [104] A.F. Pedrosa, A.G. Rodrigues, Candidemia in burn patients: figures and facts, *J. Trauma Injury Infect. Crit. Care* 70 (2) (2011) 498–506.
- [105] E.E. Horvath, et al., Fungal wound infection (not colonization) is independently associated with mortality in burn patients, *Ann. Surg.* 245 (6) (2007) 978–985.
- [106] J.C. Song, D.A. Stevens, Caspofungin: pharmacodynamics, pharmacokinetics, clinical uses and treatment outcomes, *Crit. Rev. Microbiol.* 42 (5) (2016) 813–846.
- [107] B. Blanchet, et al., Influence of burns on pharmacokinetics and pharmacodynamics of drugs used in the care of burn patients, *Clin. Pharmacokinet.* 47 (10) (2008) 635–654.
- [108] A.N. Steele, et al., Gap analysis of pharmacokinetics and pharmacodynamics in burn patients: a review, *J. Burn Care Res.* 36 (3) (2015) e194–e211.
- [109] J. Martyn, Clinical pharmacology and drug therapy in the burned patient, *Anesthesiology* 65 (1) (1986) 67–75.
- [110] J.R. Holtman Jr., W.S. Jellish, Opioid-induced hyperalgesia and burn pain, *J. Burn Care Res.* 33 (6) (2012) 692–701.
- [111] M. Sage, S.M. Laird, Ketamine anaesthesia for burns surgery, *Postgrad. Med. J.* 48 (557) (1972) 156–161.
- [112] C.M. Ward, A.W. Diamond, An appraisal of ketamine in the dressing of burns, *Postgrad. Med. J.* 52 (606) (1976) 222–223.
- [113] R.H. Demling, S. Ellerbe, F. Jarrett, Ketamine anesthesia for tangential excision of burn eschar: a burn unit procedure, *J. Trauma Injury Infect. Crit. Care* 18 (4) (1978) 269–270.
- [114] T. Warncke, A. Stubhaug, E. Jørum, Preinjury treatment with morphine or ketamine inhibits the development of experimentally induced secondary hyperalgesia in man, *Pain* 86 (3) (2000) 293–303.
- [115] S.K. McGuinness, et al., A systematic review of ketamine as an analgesic agent in adult burn injuries, *Pain Med.* 12 (10) (2011) 1551–1558.
- [116] T. Edrich, et al., Ketamine for long-term sedation and analgesia of a burn patient, *Anesth. Analg.* 99 (3) (2004) 893–895.
- [117] R.D. MacPherson, D. Woods, J. Penfold, Ketamine and midazolam delivered by patient-controlled analgesia in relieving pain associated with burns dressings, *Clin. J. Pain* 24 (7) (2008) 568–571.
- [118] C. Norambuena, et al., Oral ketamine and midazolam for pediatric burn patients: a prospective, randomized, double-blind study, *J. Pediatr. Surg.* 48 (3) (2013) 629–634.
- [119] C. Reid, R. Hutton, P. Middleton, Case report: prehospital use of intranasal ketamine for paediatric burn injury, *Emerg. Med. J.* 28 (4) (2011) 328–329.
- [120] F. Zor, et al., Pain relief during dressing changes of major adult burns: ideal analgesic combination with ketamine, *Burns* 36 (4) (2010) 501–505.
- [121] J.K. Lilburn, et al., Ketamine sequelae. Evaluation of the ability of various premedications to attenuate its psychic actions, *Anaesthesia* 33 (4) (1978) 307–311.
- [122] Z. Tosun, A. Esmaoglu, A. Coruh, Propofol-ketamine vs propofol-fentanyl combinations for deep sedation and analgesia in pediatric patients undergoing burn dressing changes, *Paediatr. Anaesth.* 18 (1) (2008) 43–47.
- [123] R.R. Edwards, et al., Symptoms of depression and anxiety as unique predictors of pain-related outcomes following burn injury, *Ann. Behav. Med.* 34 (3) (2007) 313–322.
- [124] L.A. Aaron, et al., The utility of a burn specific measure of pain anxiety to prospectively predict pain and function: a comparative analysis, *Burns* 27 (4) (2001) 329–334.
- [125] G. Pretto, G.A. Westphal, E. Silva, Clonidine for reduction of hemodynamic and psychological effects of S + ketamine anesthesia for dressing changes in patients with major burns: an RCT, *Burns* 40 (7) (2014) 1300–1307.
- [126] A. Fagin, et al., A comparison of dexmedetomidine and midazolam for sedation in severe pediatric burn injury, *J. Burn Care Res.* 33 (6) (2012) 759–763.
- [127] L. Wong, L. Turner, Treatment of post-burn neuropathic pain: evaluation of pregabalin, *Burns* 36 (6) (2010) 769–772.
- [128] R.B. Ahuja, G.K. Gupta, A four arm, double blind, randomized and placebo controlled study of pregabalin in the management of post-burn pruritus, *Burns* 39 (1) (2013) 24–29.
- [129] P. Gray, et al., Pregabalin in severe burn injury pain: a double-blind, randomised placebo-controlled trial, *Pain* 152 (6) (2011) 1279–1288.
- [130] M.U. Werner, et al., Effects of gabapentin in acute inflammatory pain in humans, *Reg. Anesth. Pain Med.* 26 (4) (2001) 322–328.
- [131] O. Cuiquet, et al., Effects of gabapentin on morphine consumption and pain in severely burned patients, *Burns* 33 (1) (2007) 81–86.
- [132] L. Wibbenmeyer, et al., Gabapentin is ineffective as an analgesic adjunct in the immediate postburn period, *J. Burn Care Res.* 35 (2) (2014) 136–142.
- [133] B. Gustorff, et al., The effects of remifentanyl and gabapentin on hyperalgesia in a new extended inflammatory skin pain model in healthy volunteers, *Anesth. Analg.* 98 (2) (2004) 401–407.
- [134] J.E. Mendham, Gabapentin for the treatment of itching produced by burns and wound healing in children: a pilot study, *Burns* 30 (8) (2004) 851–853.
- [135] N. Akhtar, P. Brooks, The use of botulinum toxin in the management of burn itching: preliminary results, *Burns* 38 (8) (2012) 1119–1123.
- [136] S. Gross, R. Overbaugh, R. Jansen, Ondansetron for treating itch in healing burns, *Internet J. Pain Symptom Control Palliat. Care* 5 (1) (2006) 1–4.
- [137] S.I. Jung, et al., Efficacy of naltrexone in the treatment of chronic refractory itching in burn patients: preliminary report of an open trial, *J. Burn Care Res.* 30 (2) (2009) 257–260.
- [138] L. LaSalle, G. Rachelska, B. Nedelec, Naltrexone for the management of post-burn pruritus: a preliminary report, *Burns* 34 (6) (2008) 797–802.
- [139] P. Holzer, The pharmacological challenge to tame the transient receptor potential vanilloid-1 (TRPV1) nociceptor, *Br. J. Pharmacol.* 155 (8) (2008) 1145–1162.
- [140] I. Nagy, et al., Pharmacology of the capsaicin receptor, transient receptor potential vanilloid type-1 ion channel, *Prog. Drug Res.* 68 (2014) 39–76.
- [141] I. Nagy, H.P. Rang, Similarities and differences between the responses of rat sensory neurons to noxious heat and capsaicin, *J. Neurosci.* 19 (24) (1999) 10647–10655.
- [142] M.J. Caterina, et al., A capsaicin-receptor homologue with a high threshold for noxious heat, *Nature* 398 (6726) (1999) 436–441.
- [143] M.J. Caterina, et al., The capsaicin receptor: a heat-activated ion channel in the pain pathway, *Nature* 389 (6653) (1997) 816–824.
- [144] J.B. Davis, et al., Vanilloid receptor-1 is essential for inflammatory thermal hyperalgesia, *Nature* 405 (6783) (2000) 183–187.
- [145] N. Hellwig, et al., TRPV1 acts as proton channel to induce acidification in nociceptive neurons, *J. Biol. Chem.* 279 (33) (2004) 34553–34561.
- [146] J. Vriens, et al., TRPM3 is a nociceptor channel involved in the detection of noxious heat, *Neuron* 70 (3) (2011) 482–494.
- [147] N. Qin, et al., TRPV2 is activated by cannabidiol and mediates cgrp release in cultured rat dorsal root ganglion neurons, *J. Neurosci.* 28 (24) (2008) 6231–6238.
- [148] M.J. Caterina, et al., Impaired nociception and pain sensation in mice lacking the capsaicin receptor, *Science* 288 (5464) (2000) 306–313.
- [149] K. Bölskei, et al., Investigation of the role of TRPV1 receptors in acute and chronic nociceptive processes using gene-deficient mice, *Pain* 117 (3) (2005) 368–376.
- [150] D.P. Green, et al., Role of endogenous TRPV1 agonists in a postburn pain model of partial-thickness injury, *Pain* 154 (11) (2013) 2512–2520.
- [151] Y.S. Kim, et al., Central terminal sensitization of trpv1 by descending serotonergic facilitation modulates chronic pain, *Neuron* 81 (4) (2014) 873–887.
- [152] C. Radtke, et al., TRPV channel expression in human skin and possible role in thermally induced cell death, *J. Burn Care Res.* 32 (1) (2011) 150–159.
- [153] A.A. Steiner, et al., Nonthermal activation of transient receptor potential vanilloid-1 channels in abdominal viscera tonically inhibits autonomic cold-defense effectors, *J. Neurosci.* 27 (28) (2007) 7459–7468.
- [154] H.E. Gibson, et al., TRPV1 channels mediate long-term depression at synapses on hippocampal interneurons, *Neuron* 57 (5) (2008) 746–759.
- [155] Y.P. Goldberg, et al., Loss-of-function mutations in the $Na_v1.7$ gene underlie congenital indifference to pain in multiple human populations, *Clin. Genet.* 71 (4) (2007) 311–319.
- [156] S. Ahmad, et al., A stop codon mutation in SCN9A causes lack of pain sensation, *Hum. Mol. Genet.* 16 (17) (2007) 2114–2121.
- [157] C. Han, et al., Sporadic onset of erythralgia: a gain-of-function mutation in $Na_v1.7$, *Ann. Neurol.* 59 (3) (2006) 553–558.
- [158] J.J. Cox, et al., An SCN9A channelopathy causes congenital inability to experience pain, *Nature* 444 (7121) (2006) 894–898.
- [159] M.A. Nassar, et al., Nociceptor-specific gene deletion reveals a major role for $Na_v1.7$ (PN1) in acute and inflammatory pain, *Proc. Natl. Acad. Sci. U. S. A.* 101 (34) (2004) 12706–12711.
- [160] S.D. Shields, et al., Sodium channel $Na_v1.7$ is essential for lowering heat pain threshold after burn injury, *J. Neurosci.* 32 (32) (2012) 10819–10832.
- [161] M.M. Salas, et al., Tetrodotoxin suppresses thermal hyperalgesia and mechanical allodynia in a rat full thickness thermal injury pain model, *Neurosci. Lett.* 607 (2015) 108–113.

- [162] M.E. van Baar, et al., Functional outcome after burns: a review, *Burns* 32 (1) (2006) 1–9.
- [163] A.C. Drost, et al., Plasma cytokines following thermal injury and their relationship with patient mortality, burn size, and time postburn, *J. Trauma Injury Infect. Crit. Care* 35 (3) (1993) 335–339.
- [164] M. Spiekman, et al., Adipose tissue-derived stromal cells inhibit tgf- β 1-induced differentiation of human dermal fibroblasts and keloid scar-derived fibroblasts in a paracrine fashion, *Plast. Reconstr. Surg.* 134 (4) (2014) 699–712.
- [165] F. Ghieh, et al., The use of stem cells in burn wound healing: a review, *Biomed. Res. Int.* 2015 (2015) 684084.
- [166] R.S. Waterman, et al., A new mesenchymal stem cell (MSC) paradigm: polarization into a pro-inflammatory MSC1 or an immunosuppressive MSC2 phenotype, *PLoS One* 5 (4) (2010), e10088.
- [167] R.S. Chiang, et al., Current concepts related to hypertrophic scarring in burn injuries, *Wound Repair Regen.* 24 (3) (2016) 466–477.
- [168] S.M. Sultan, et al., Fat grafting accelerates revascularisation and decreases fibrosis following thermal injury, *J. Plast. Reconstr. Aesthet. Surg.* 65 (2) (2012) 219–227.
- [169] Y. Wu, et al., Bone marrow-derived mesenchymal stem cell attenuates skin fibrosis development in mice, *Int. Wound J.* 11 (6) (2014) 701–710.
- [170] Q. Zhang, et al., Intraleisional injection of adipose-derived stem cells reduces hypertrophic scarring in a rabbit ear model, *Stem Cell Res Ther* 6 (1) (2015) 145.
- [171] V. Trotter, et al., IFATS collection: using human adipose-derived stem/stromal cells for the production of new skin substitutes, *Stem Cells* 26 (10) (2008) 2713–2723.
- [172] A. Condé-Green, et al., Fat grafting and adipose-derived regenerative cells in burn wound healing and scarring: a systematic review of the literature, *Plast. Reconstr. Surg.* 137 (1) (2016) 302–312.
- [173] P. Gentile, et al., Adipose-derived stromal vascular fraction cells and platelet-rich plasma: basic and clinical evaluation for cell-based therapies in patients with scars on the face, *J. Craniofac. Surg.* 25 (1) (2014) 267–272.
- [174] I.S. Yun, et al., Effect of human adipose derived stem cells on scar formation and remodeling in a pig model: a pilot study, *Dermatol. Surg.* 38 (10) (2012) 1678–1688.
- [175] M.T. Lam, et al., Effective delivery of stem cells using an extracellular matrix patch results in increased cell survival and proliferation and reduced scarring in skin wound healing, *Tissue Eng. A* 19 (5–6) (2012) 738–747.
- [176] L.J. van den Broek, et al., Differential response of human adipose tissue-derived mesenchymal stem cells, dermal fibroblasts, and keratinocytes to burn wound exudates: potential role of skin-specific chemokine CCL27, *Tissue Eng. A* 20 (1–2) (2013) 197–209.
- [177] G.-Y. Zhang, et al., Contribution of epidermal stem cells to hypertrophic scars pathogenesis, *Med. Hypotheses* 73 (3) (2009) 332–333.
- [178] A.J. van Den Bogaerd, et al., Collagen cross-linking by adipose-derived mesenchymal stromal cells and scar-derived mesenchymal cells: are mesenchymal stromal cells involved in scar formation? *Wound Repair Regen.* 17 (4) (2009) 548–558.
- [179] J. Ding, et al., Deep dermal fibroblast profibrotic characteristics are enhanced by bone marrow-derived mesenchymal stem cells, *Wound Repair Regen.* 21 (3) (2013) 448–455.
- [180] L. Sun, et al., Sundew-inspired adhesive hydrogels combined with adipose-derived stem cells for wound healing, *ACS Appl. Mater. Interfaces* 8 (3) (2016) 2423–2434.
- [181] A. Nuschke, Activity of mesenchymal stem cells in therapies for chronic skin wound healing, *Organ* 10 (1) (2014) 29–37.
- [182] M. Li, et al., Mesenchymal stem cell-conditioned medium accelerates wound healing with fewer scars, *Int. Wound J.* 14 (1) (2017) 64–73.
- [183] T.A. Mustoe, Evolution of silicone therapy and mechanism of action in scar management, *Aesthet. Plast. Surg.* 32 (1) (2008) 82–92.
- [184] S. Kim, et al., Update on scar management: guidelines for treating Asian patients, *Plast. Reconstr. Surg.* 132 (6) (2013) 1580–1589.
- [185] S. Monstrey, et al., Updated scar management practical guidelines: non-invasive and invasive measures, *J. Plast. Reconstr. Aesthet. Surg.* 67 (8) (2014) 1017–1025.
- [186] C.W. Li-Tsang, Y.P. Zheng, J.C. Lau, A randomized clinical trial to study the effect of silicone gel dressing and pressure therapy on posttraumatic hypertrophic scars, *J. Burn Care Res.* 31 (3) (2010) 448–457.
- [187] A.F. Lim, et al., The embrace device significantly decreases scarring following scar revision surgery in a randomized controlled trial, *Plast. Reconstr. Surg.* 133 (2) (2014) 398–405.
- [188] R. Ogawa, The most current algorithms for the treatment and prevention of hypertrophic scars and keloids, *Plast. Reconstr. Surg.* 125 (2) (2010) 557–568.
- [189] D. Wolfram, et al., Hypertrophic scars and keloids—a review of their pathophysiology, risk factors, and therapeutic management, *Dermatol. Surg.* 35 (2) (2009) 171–181.
- [190] A. Al-Attar, et al., Keloid pathogenesis and treatment, *Plast. Reconstr. Surg.* 117 (1) (2006) 286–300.
- [191] C.C. Finnerty, et al., Hypertrophic scarring: the greatest unmet challenge after burn injury, *Lancet* 388 (10052) (2016) 1427–1436.
- [192] B. Berman, et al., Prevention and management of hypertrophic scars and keloids after burns in children, *J. Craniofac. Surg.* 19 (4) (2008) 989–1006.
- [193] R.E. Fitzpatrick, Treatment of inflamed hypertrophic scars using intralesional 5-FU, *Dermatol. Surg.* 25 (3) (1999) 224–232.
- [194] M. Apikian, G. Goodman, Intralesional 5-fluorouracil in the treatment of keloid scars, *Australas. J. Dermatol.* 45 (2) (2004) 140–143.
- [195] A. Asilian, A. Darougheh, F. Shariati, New combination of triamcinolone, 5-Fluorouracil, and pulsed-dye laser for treatment of keloid and hypertrophic scars, *Dermatol. Surg.* 32 (7) (2006) 907–915.
- [196] M.H. Viera, et al., Innovative therapies in the treatment of keloids and hypertrophic scars, *J. Clin. Aesthet. Dermatol.* 3 (5) (2010) 20–26.
- [197] Y. Yamaguchi, D.M. Mann, E. Ruoslahti, Negative regulation of transforming growth factor-(beta) by the proteoglycan decorin, *Nature* 346 (6281) (1990) 281–284.
- [198] J. Zhang, et al., A cationic-independent mannose 6-phosphate receptor inhibitor (PXS64) ameliorates kidney fibrosis by inhibiting activation of transforming growth factor- β 1, *PLoS One* 10 (2) (2015), e0116888.
- [199] F. Wehrhan, et al., Transforming growth factor beta 1 dependent regulation of Tenascin-C in radiation impaired wound healing, *Radiother. Oncol.* 72 (3) (2004) 297–303.
- [200] G.M. Bran, et al., Effect of the abrogation of TGF- β 1 by antisense oligonucleotides on the expression of TGF- β -isoforms and their receptors I and II in isolated fibroblasts from keloid scars, *Int. J. Mol. Med.* 25 (6) (2010) 915–921.
- [201] M.W. Ferguson, et al., Prophylactic administration of avotermine for improvement of skin scarring: three double-blind, placebo-controlled, phase I/II studies, *Lancet* 373 (9671) (2009) 1264–1274.
- [202] H.G. Gassner, D.A. Sherris, Chemoimmobilization: improving predictability in the treatment of facial scars, *Plast. Reconstr. Surg.* 112 (5) (2003) 1464–1466.
- [203] L. Block, A. Gosain, T.W. King, Emerging therapies for scar prevention, *Adv. Wound Care* 4 (10) (2015) 607–614.
- [204] X. Zhibo, Z. Miao, Intraleisional botulinum toxin type A injection as a new treatment measure for keloids, *Plast. Reconstr. Surg.* 124 (5) (2009) 275e–277e.
- [205] Z. Xiao, et al., Effect of botulinum toxin type A on transforming growth factor β 1 in fibroblasts derived from hypertrophic scar: a preliminary report, *Aesthet. Plast. Surg.* 34 (4) (2010) 424–427.
- [206] G.G. Gauglitz, Management of keloids and hypertrophic scars: current and emerging options, *Clin. Cosmet. Investig. Dermatol.* 6 (2013) 103–114.
- [207] I. Khansa, B. Harrison, J.E. Janis, Evidence-based scar management: how to improve results with technique and technology, *Plast. Reconstr. Surg.* 138 (3S) (2016) 165S–178S.
- [208] M. Klinger, et al., Autologous fat graft in scar treatment, *J. Craniofac. Surg.* 24 (5) (2013) 1610–1615.
- [209] A. Bruno, et al., Burn scar lipofilling: immunohistochemical and clinical outcomes, *J. Craniofac. Surg.* 24 (5) (2013) 1806–1814.
- [210] M.E. Jaspers, et al., Effectiveness of autologous fat grafting in adherent scars: results obtained by a comprehensive scar evaluation protocol, *Plast. Reconstr. Surg.* 139 (1) (2017) 212–219.
- [211] R. Fredman, R.E. Edkins, C.S. Hultman, Fat grafting for neuropathic pain after severe burns, *Ann. Plast. Surg.* 77 (Suppl. 4) (2016) S298–S303.
- [212] F. Caviggioli, et al., Autologous fat grafting reduces pain in irradiated breast: a review of our experience, *Stem Cells Int.* 2016 (2016) 2527349.
- [213] S.-H. Huang, et al., Alleviation of neuropathic scar pain using autologous fat grafting, *Ann. Plast. Surg.* 74 (S2) (2015) S99–S104.
- [214] A.A. Juhl, P. Karlsson, T.E. Damsgaard, Fat grafting for alleviating persistent pain after breast cancer treatment: a randomized controlled trial, *J. Plast. Reconstr. Aesthet. Surg.* 69 (9) (2016) 1192–1202.
- [215] F. Caviggioli, et al., Autologous fat graft in postmastectomy pain syndrome, *Plast. Reconstr. Surg.* 128 (2) (2011) 349–352.
- [216] S.H. Huang, et al., Autologous fat grafting alleviates burn-induced neuropathic pain in rats, *Plast. Reconstr. Surg.* 133 (6) (2014) 1396–1405.
- [217] S.H. Huang, et al., Fat grafting in burn scar alleviates neuropathic pain via anti-inflammation effect in scar and spinal cord, *PLoS One* 10 (9) (2015), e0137563.
- [218] F. Caviggioli, et al., Nipple resuscitation by lipofilling in burn sequelae and scar retraction, *Plast. Reconstr. Surg.* 125 (4) (2010) 174e–176e.
- [219] S. Brongo, et al., Use of lipofilling for the treatment of severe burn outcomes, *Plast. Reconstr. Surg.* 130 (2) (2012) 374e–376e.
- [220] R. Viard, et al., Fat grafting in facial burns sequelae, *Ann. Chir. Plast. Esthet.* 57 (3) (2012) 217–229.
- [221] N.S. Piccolo, M.S. Piccolo, M.T. Piccolo, Fat grafting for treatment of burns, burn scars, and other difficult wounds, *Clin. Plast. Surg.* 42 (2) (2015) 263–283.
- [222] A.J. Singer, R.A.F. Clark, Cutaneous wound healing, *N. Engl. J. Med.* 341 (10) (1999) 738–746.
- [223] B.M. Parrett, M.B. Donelan, Pulsed dye laser in burn scars: current concepts and future directions, *Burns* 36 (4) (2010) 443–449.
- [224] M.B. Donelan, Principles of burn reconstruction, in: T.C. H. (Ed.), *Grabb and Smith's Plastic Surgery*, Lippincott Williams & Wilkins, Philadelphia, PA 2014, pp. 142–154.
- [225] C.S. Dunkin, et al., Scarring occurs at a critical depth of skin injury: precise measurement in a graduated dermal scratch in human volunteers, *Plast. Reconstr. Surg.* 119 (6) (2007) 1722–1732.
- [226] L.H. Engrav, W.L. Garner, E.E. Tredget, Hypertrophic scar, wound contraction and hyper-hypopigmentation, *J. Burn Care Res.* 28 (4) (2007) 593–597.
- [227] Z.F. Tyack, S. Pegg, J. Ziviani, Postburn dyspigmentation: its assessment, management, and relationship to scarring — a review of the literature, *J. Burn Care Rehabil.* 18 (5) (1997) 435–440.
- [228] D.J. Castro, et al., Effects of the Nd:YAG laser on DNA synthesis and collagen production in human skin fibroblast cultures, *Ann. Plast. Surg.* 11 (3) (1983) 214–222.
- [229] D. Manstein, et al., Fractional photothermolysis: a new concept for cutaneous remodeling using microscopic patterns of thermal injury, *Lasers Surg. Med.* 34 (5) (2004) 426–438.
- [230] A.C. Issler-Fisher, et al., Ablative fractional CO₂ laser for burn scar reconstruction: an extensive subjective and objective short-term outcome analysis of a prospective treatment cohort, *Burns* 43 (3) (2016) 573–582.
- [231] T.A. Mustoe, et al., International clinical recommendations on scar management, *Plast. Reconstr. Surg.* 110 (2) (2002) 560–571.
- [232] R. Jin, et al., Laser therapy for prevention and treatment of pathologic excessive scars, *Plast. Reconstr. Surg.* 132 (6) (2013) 1747–1758.
- [233] A. Scheid, et al., Physiologically low oxygen concentrations in fetal skin regulate hypoxia-inducible factor 1 and transforming growth factor-beta3, *FASEB J.* 16 (3) (2002) 411–413.

- [234] T.S. Alster, C.M. Williams, Treatment of keloid sternotomy scars with 585 nm flashlamp-pumped pulsed-dye laser, *Lancet* 345 (8959) (1995) 1198–1200.
- [235] K.P. Allison, et al., Pulsed dye laser treatment of burn scars. Alleviation or irritation? *Burns* 29 (3) (2003) 207–213.
- [236] W. Manuskiaiti, R.E. Fitzpatrick, Treatment response of keloidal and hypertrophic sternotomy scars: comparison among intralesional corticosteroid, 5-fluorouracil, and 585-nm flashlamp-pumped pulsed-dye laser treatments, *Arch. Dermatol.* 138 (9) (2002) 1149–1155.
- [237] R.R. Anderson, et al., Laser treatment of traumatic scars with an emphasis on ablative fractional laser resurfacing: consensus report, *JAMA Dermatol.* 150 (2) (2014) 187–193.
- [238] E.H. Taudorf, et al., Non-ablative fractional laser provides long-term improvement of mature burn scars—a randomized controlled trial with histological assessment, *Lasers Surg. Med.* 47 (2) (2015) 141–147.
- [239] H.S. Kim, et al., Comparison of the effectiveness of nonablative fractional laser versus ablative fractional laser in thyroidectomy scar prevention: a pilot study, *J. Cosmet. Laser Ther.* 14 (2) (2012) 89–93.
- [240] B.M. El-Zawahry, et al., Ablative CO₂ fractional resurfacing in treatment of thermal burn scars: an open-label controlled clinical and histopathological study, *J. Cosmet. Dermatol.* 14 (4) (2015) 324–331.
- [241] O.A. Azzam, et al., Treatment of hypertrophic scars and keloids by fractional carbon dioxide laser: a clinical, histological, and immunohistochemical study, *Lasers Med. Sci.* 31 (1) (2016) 9–18.
- [242] L. Qu, et al., Clinical and molecular effects on mature burn scars after treatment with a fractional CO₂ laser, *Lasers Surg. Med.* 44 (7) (2012) 517–524.
- [243] D.M. Ozog, et al., Evaluation of clinical results, histological architecture, and collagen expression following treatment of mature burn scars with a fractional carbon dioxide laser, *JAMA Dermatol.* 149 (1) (2013) 50–57.
- [244] C.S. Hultman, et al., Laser resurfacing and remodeling of hypertrophic burn scars: the results of a large, prospective, before-after cohort study, with long-term follow-up, *Ann. Surg.* 260 (3) (2014) 519–532.
- [245] V. Cervelli, et al., Ultrapulsed fractional CO₂ laser for the treatment of post-traumatic and pathological scars, *J. Drugs Dermatol.* 9 (11) (2010) 1328–1331.
- [246] N.S. Uebelhoefer, E.V. Ross, P.R. Shumaker, Ablative fractional resurfacing for the treatment of traumatic scars and contractures, *Semin. Cutan. Med. Surg.* 31 (2) (2012) 110–120.
- [247] J. Waibel, K. Beer, Ablative fractional laser resurfacing for the treatment of a third-degree burn, *J. Drugs Dermatol.* 8 (3) (2009) 294–297.
- [248] P.R. Shumaker, et al., Rapid healing of scar-associated chronic wounds after ablative fractional resurfacing, *Arch. Dermatol.* 148 (11) (2012) 1289–1293.
- [249] P.R. Shumaker, et al., Functional improvements in traumatic scars and scar contractures using an ablative fractional laser protocol, *J. Trauma Acute Care Surg.* 73 (2 S1) (2012) S116–S121.
- [250] A. Khandelwal, et al., Ablative fractional photothermolysis for the treatment of hypertrophic burn scars in adult and pediatric patients: a single surgeon's experience, *J. Burn Care Res.* 35 (5) (2014) 455–463.
- [251] S. Blome-Eberwein, et al., Prospective evaluation of fractional CO₂ laser treatment of mature burn scars, *J. Burn Care Res.* 37 (6) (2016) 379–387.
- [252] J.S. Waibel, A.J. Wulkan, P.R. Shumaker, Treatment of hypertrophic scars using laser and laser assisted corticosteroid delivery, *Lasers Surg. Med.* 45 (3) (2013) 135–140.
- [253] L.R. Sklar, et al., Laser assisted drug delivery: a review of an evolving technology, *Lasers Surg. Med.* 46 (4) (2014) 249–262.
- [254] M. Cavalié, et al., Treatment of keloids with laser-assisted topical steroid delivery: a retrospective study of 23 cases, *Dermatol. Ther.* 28 (2) (2015) 74–78.
- [255] M.C. Issa, et al., Topical delivery of triamcinolone via skin pretreated with ablative radiofrequency: a new method in hypertrophic scar treatment, *Int. J. Dermatol.* 52 (3) (2013) 367–370.
- [256] M. Hædersdal, et al., Fractional CO₂ laser-assisted drug delivery, *Lasers Surg. Med.* 42 (2) (2010) 113–122.
- [257] A.W.C. Chua, et al., Skin tissue engineering advances in severe burns: review and therapeutic applications, *Burns Trauma* 4 (1) (2016) 3.
- [258] K. Kawai, et al., Accelerated tissue regeneration through incorporation of basic fibroblast growth factor-impregnated gelatin microspheres into artificial dermis, *Biomaterials* 21 (5) (2000) 489–499.
- [259] M. Yamamoto, Y. Ikada, Y. Tabata, Controlled release of growth factors based on biodegradation of gelatin hydrogel, *J. Biomater. Sci. Polym. Ed.* 12 (1) (2001) 77–88.
- [260] R. Slavkovsky, et al., Effects of hyaluronan and iodine on wound contraction and granulation tissue formation in rat skin wounds, *Clin. Exp. Dermatol.* 35 (4) (2010) 373–379.
- [261] F. Gao, et al., Hyaluronan oligosaccharides promote excisional wound healing through enhanced angiogenesis, *Matrix Biol.* 29 (2) (2010) 107–116.
- [262] T.V. Anilkumar, et al., Advantages of hyaluronic acid as a component of fibrin sheet for care of acute wound, *Biologicals* 39 (2) (2011) 81–88.
- [263] J. Rnjak, et al., Severe burn injuries and the role of elastin in the design of dermal substitutes, *Tissue Eng. B Rev.* 17 (2) (2011) 81–91.
- [264] Y. Wang, et al., Tropoelastin incorporation into a dermal regeneration template promotes wound angiogenesis, *Adv. Healthcare Mater.* 4 (4) (2015) 577–584.
- [265] H.J. De Vries, et al., Reduced wound contraction and scar formation in punch biopsy wounds. Native collagen dermal substitutes. A clinical study, *Br. J. Dermatol.* 132 (5) (1995) 690–697.
- [266] B. Hinz, et al., α -Smooth muscle actin is crucial for focal adhesion maturation in myofibroblasts, *Mol. Biol. Cell* 14 (6) (2003) 2508–2519.
- [267] W. Haslik, et al., First experiences with the collagen-elastin matrix Matriderm® as a dermal substitute in severe burn injuries of the hand, *Burns* 33 (3) (2007) 364–368.
- [268] U. Wollina, A. Meseg, A. Weber, Use of a collagen-elastin matrix for hard to treat soft tissue defects, *Int. Wound J.* 8 (3) (2011) 291–296.
- [269] W. Haslik, et al., Management of full-thickness skin defects in the hand and wrist region: first long-term experiences with the dermal matrix Matriderm®, *J. Plast. Reconstr. Aesthet. Surg.* 63 (2) (2010) 360–364.
- [270] H. Ryssel, et al., The use of Matriderm® in early excision and simultaneous autologous skin grafting in burns — a pilot study, *Burns* 34 (1) (2008) 93–97.
- [271] H. Ryssel, et al., Matriderm® in depth-adjusted reconstruction of necrotising fasciitis defects, *Burns* 36 (7) (2010) 1107–1111.
- [272] B. Kundu, et al., Isolation and processing of silk proteins for biomedical applications, *Int. J. Biol. Macromol.* 70 (2014) 70–77.
- [273] L.-D. Koh, et al., Structures, mechanical properties and applications of silk fibroin materials, *Prog. Polym. Sci.* 46 (2015) 86–110.
- [274] D. Jao, X. Mou, X. Hu, Tissue regeneration: a silk road, *J. Funct. Biomater.* 7 (3) (2016) 22.
- [275] N. Bhardwaj, et al., Silk fibroin-keratin based 3D scaffolds as a dermal substitute for skin tissue engineering, *Integr. Biol.* 7 (1) (2015) 53–63.
- [276] H. Xie, et al., Treatment of burn and surgical wounds with recombinant human tropoelastin produces new elastin fibers in scars, *J. Burn Care Res.* 38 (5) (2017) e859–e867.
- [277] Z. Yu, et al., Bacterial collagen-like proteins that form triple-helical structures, *J. Struct. Biol.* 186 (3) (2014) 451–461.
- [278] T.D. Sutherland, et al., Recombinant structural proteins and their use in future materials, in: D.A.D. Parry, J.M. Squire (Eds.), *Fibrous Proteins: Structures and Mechanisms*, Springer, Cham, Switzerland 2017, pp. 491–526.
- [279] T.J. Sill, H.A. von Recum, Electrospinning: applications in drug delivery and tissue engineering, *Biomaterials* 29 (13) (2008) 1989–2006.
- [280] Y. Pilehvar-Soltanahmadi, et al., An update on clinical applications of electrospun nanofibers for skin bioengineering, *Artif. Cells Nanomed. Biotechnol.* 44 (6) (2016) 1350–1364.
- [281] Y.-F. Goh, I. Shakir, R. Hussain, Electrospun fibers for tissue engineering, drug delivery, and wound dressing, *J. Mater. Sci.* 48 (8) (2013) 3027–3054.
- [282] S.P. Zhong, Y.Z. Zhang, C.T. Lim, Tissue scaffolds for skin wound healing and dermal reconstruction, *Wiley Interdiscip. Rev. Nanomed. Nanobiotechnol.* 2 (5) (2010) 510–525.
- [283] S.N. Jayasinghe, Cell electrospinning: a novel tool for functionalising fibres, scaffolds and membranes with living cells and other advanced materials for regenerative biology and medicine, *Analyst* 138 (8) (2013) 2215–2223.
- [284] X. Yang, J.D. Shah, H. Wang, Nanofiber enabled layer-by-layer approach toward three-dimensional tissue formation, *Tissue Eng. A* 15 (4) (2009) 945–956.
- [285] S.L. Sampson, et al., Cell electrospinning: an in vitro and in vivo study, *Small* 10 (1) (2014) 78–82.
- [286] W.L. Ng, et al., Skin bioprinting: impending reality or fantasy? *Trends Biotechnol.* 34 (9) (2016) 689–699.
- [287] W. Yeong, et al., State-of-the-art review on selective laser melting of ceramics, *High Value Manufacturing: Advanced Research in Virtual and Rapid Prototyping-Proceedings of the 6th International Conference on Advanced Research in Virtual and Rapid Prototyping, High Value Manufacturing: Advanced Research in Virtual and Rapid Prototyping, Leiria, Portugal, (2013) 65–70.*
- [288] L. Koch, et al., Skin tissue generation by laser cell printing, *Biotechnol. Bioeng.* 109 (7) (2012) 1855–1863.
- [289] L.J. Pourchet, et al., Human skin 3D bioprinting using scaffold-free approach, *Adv. Healthcare Mater.* 6 (4) (2017).
- [290] A. Skardal, et al., Bioprinted amniotic fluid-derived stem cells accelerate healing of large skin wounds, *Stem Cells Transl. Med.* 1 (11) (2012) 792–802.
- [291] S. Vijayavenkataraman, W. Lu, J. Fuh, 3D bioprinting of skin: a state-of-the-art review on modelling, materials, and processes, *Biofabrication* 8 (3) (2016), 032001.
- [292] N. Cubo, et al., 3D bioprinting of functional human skin: production and in vivo analysis, *Biofabrication* 9 (1) (2016), 015006.
- [293] M. Gruene, et al., Adipogenic differentiation of laser-printed 3D tissue grafts consisting of human adipose-derived stem cells, *Biofabrication* 3 (1) (2011), 015005.
- [294] S. Michael, et al., Tissue engineered skin substitutes created by laser-assisted bioprinting form skin-like structures in the dorsal skin fold chamber in mice, *PLoS One* 8 (3) (2013), e57741.