# Revision of the AF4 calibration experiment

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Asymmetrical flow field-flow fractionation, channel width void peak-determination, size determination,

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calibration

### 1. Introduction

- 40 AF4 (asymmetrical flow field-flow fractionation) is a chromatographic technique that can be used to
- 41 separate samples due to their diffusion coefficient[1]. It is a member of the FFF (field-flow fractionation)
- 42 family invented by J. Calvin Giddings [2]. Compared to more commonly applied separation methods like
- 43 SEC and HPLC[3], FFF techniques are based on the interaction of the analyte with a physical field which
- 44 separates the sample to a corresponding physical size[4]. In principle, the method is applicable to a huge
- 45 variety of samples, including small biomolecules [5,6], nanoparticles and polymers [3,4] up to big
- agglomerates like protein aggregates[7], virus-like particles [8] and drug carrier systems [9]. Nowadays,
- 47 AF4 is the most commonly used flow FFF method, where the separation channel is formed of a solid wall
- and a frit covered by a membrane.
- 49 The development of dedicated measurement protocols can be complicated due to the high number of
- 50 adjustable parameters[10]. This includes instrumental specifications like the channel shape, its related
- parameters length L, the channel width  $b_0$  and  $b_L$  and the choice of the membrane material. The three
- 52 typical variable experimental conditions are elution flow  $V_e$ , applied cross-flow  $V_c$  and the focusing flow
- 53 and period. AF4 has to be combined with at least one detection technique, typically MALLS (multi-angle
- 54 laser light scattering), UV/Vis and/or RI (refractive index) detection. Also, on-line NMR [11], mass
- 55 spectrometry [12] and SAXS (Small-angle X-ray scattering) [13] have been applied as detection
- 56 techniques.
- 57 In addition to the parameters listed above, the channel thickness w and the separation volume V have to be
- 58 known precisely in order to allow a correct determination of the diffusion coefficient, and, thereby, the
- size of a measured sample [1,10]. However, due to swelling of the applied membrane in the channel,
- 60 these magnitudes are not directly accessible with current devices, which reasons the necessity of suitable
- 61 calibration methods.
- 62 Although AF4 theory has been elaborated and well documented in literature, the transfer to its application
- 63 quantitative evaluation software lacks still lags behind compared to methods like AUC (analytical
- 64 ultracentrifugation). Here, several software solutions and a couple of evaluation methods are already
- available and can be used even without in-depth knowledge of the underlying algorithmic considerations
- 66 [14,15]. This may be also a reason, why the practical handling of AF4 data evaluation is handled very
- 67 inconsistently by different authors. While some authors suggest a direct conversion from measurement
- data is directly possible from a measurement [16], others suggest an external measurement using a
- 69 calibrant with a known diffusion coefficient [10,17] or an external size measurement via coupled MALLS
- 70 [18]
- 71 Therefore, we fill this gap with an implementation of the known procedures and compare them to each
- 72 other.
- In this work, we want to compare some of the different approaches reported up to now in literature. In the
- past, the validity of the no-field method in AF4 has already been disproved successfully [19]. A reported
- 75 approach, which aims to measure the thickness of the membrane directly by a micrometer [5,10,17] has to
- be questioned as the measurement cannot be not conducted *in situ*.

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### 2. Theory

The sample is injected into a flat channel with a solid upper wall and a lower wall that allows the streaming solvent to pass partially (Fig. 1). In current devices this wall is made of a frit covered by an ultrafiltration membrane. The inlet flow  $V_{\rm in}$  is, thereby, split to a crossflow  $V_{\rm c}$  (which is distributed uniformly over the horizontal section of the channel) and an elution flow  $V_{\rm e}$  forming parabolic flow profile typical for all FFF variants:

$$V_{in} = V_e + V_c \tag{1}$$

The "broadness" of the parabole representing the velocity gradient depends on the plate distance w, also designated as channel width.  $L_1$ ,  $L_2$ ,  $L_3$ ,  $b_0$  and  $b_L$  describe the channel dimensions as shown in Fig. 1.

 $V_{\rm c}$  transports the particles to the membrane. As a consequence, the opposed translational diffusion  $J_{\rm z}$  determines the average velocity zone and hence the time of elution. Mathematical descriptions of AF4 experiments and derivations have been described in literature [1,10,20,21]. They describe phenomena by physical and geometric approaches. As shown in the supporting information, rigorously applying a coordinate system to the channel shape allows to express these formulas for random channel shapes as

Thereby, we only state a short description of those formulas that are used in our evaluation approach which is essentially built up on existing theory. While the physical relationships are widely known and well documented, this is not always the case for their translations into an evaluation procedure. This might seem to be a trivial step as the physical content is well elaborated. However, having a closer look to the physical formalism, it turns out that the overdetermined nature of the system allows several possible evaluation procedures with different input parameters.

Considering the number of different approaches which exist for calibration [10,18] and their variations in detail, the implementation affects not only the evaluation but also the required measurement setup and, of course, the final measurement result. The lack of such standardized evaluation procedures impairs the reproducibility of measurements and may be one of the reasons why the analytical characterization potential is not exhausted to its potential up to now [3].

In total, 5 different calibration ways are described here briefly. The explicit derivation of the underlying formalisms and algorithmic considerations are stated in the supporting information.

## The retention ratio R, defined as

$$R = \frac{t_{\text{avoid}}}{t_{e}} \tag{2}$$

with the time of the void peak t<sub>θvoid</sub> (the time which is required for a particle to travel if no retention occurs) and any possible point of time during the evaluation) and the time of sample elution t<sub>e</sub>.
connected to the relative mean layer distance λ by the classical FFF retention equation:

$$R = 6\lambda \cdot \left(coth\left(\frac{1}{2\lambda}\right) - 2\lambda\right) \qquad , \qquad (3)$$

which is often simplified to

 $R_{approx} = 6\lambda$  . 116 (4)

118 The primary separation size of AF4 is the translational diffusion coefficient D. For AF4, the relevant

119 correlation of  $\lambda$  and D has been elaborated [1] as

$$\lambda = \frac{D \cdot V^0}{V_C w^2} \qquad . \tag{5}$$

For a typical AF4 measurement the channel volume  $V^0$  and the channel width w are critical sizes for the 121

122 evaluation.

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123 Recently, we used a calibration method [22] that makes use of the volume calculation as reported by

124 Wahlund and Giddings [1] and then adjusts w by a simple bisection accordingly to Eq. (3) and (4). A

125 similar method was reported independently before [20,2,20]. This formalism does not include the steric

effect which has to be considered for larger particles. This can be quantified by a more refined version of

127 Eq. (3). [Giddings1978] An overview over possible "levels of sophistication" of variants of Eq. (3) has

128 been given by Håkansson et al. [20]

129 Fig. 2 shows that bisection is easily applicable due to the strict monotonicity monotony of the retention 130

equation within the relevant scope and sufficient for being used on modern CPUs. It can be replaced by

131 an even more efficient conversion if required [23]. Here, tT he separation volume  $V^0$  is estimated

132 according to [1]:

$$\lambda = \frac{V^{\oplus}}{V_{\text{cov}}^2} \tag{6}$$

In the following, we refer to this method as "classical" calibration method with the calculated separation

135 volume  $V^{\text{cla}}$  and channel height  $w^{\text{cla}}$ .  $z_{\%}$  designates the relative focus position of the sample at the

136 beginning of a measurement. It is given as ratio of the start position (i.e. the skipped channel length) and

137 the total channel length L. This description neglects the tapered ends and assumes a simple rectangular

138 shape of the channel.

139 The formalism for the relevant method has been adjusted for narrowing trapezoidal channel shapes as

140 follows [24] and It is used as such afor the second method similar to the first one. (Y is a correction term

for approximating the correct channel surface. In order to distinguish the calculated channel widths and

142 volumes, we refer to the results here as  $V^{\text{appGeo}}$  and  $w^{\text{appGeo}}$ 

$$t_0 = \frac{V^{app}}{V_c} ln \left( 1 + \frac{V_c}{V_e} \left( 1 - \frac{b_0 z - \frac{(z^2)(b_0 - b_L)}{2L} - Y}{A_L} \right) \right)$$
 (82)

 $V^{geo} = A_{zL} \cdot w^{geo}$ 

146 Thereby, it leads to the intuitive observation that variations of any parameters don't affect the calculated 147 volume and the channel width linearly equally.

$$\frac{\lambda V_c}{D} = \frac{V_g e V_{\theta} g^{eo}}{w^2} := S$$

$$\frac{109}{\text{in Eq. (4)}} \text{ and determine} S \text{ analogously to } w \text{ as in the classical approach } via bisection.} -In a second step,$$

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150 inserting inserting the geometrical definition of  $V^{\text{geo}}$  as indicated in Eq. (8)5

 $w = \frac{A_{Lz}}{S}$ 151 (1110)

152  $A_L$  can be obtained by simple geometric considerations as described in the supporting information. Using 153

- Economic Transport of the control of
- 154 already used to calculate S in this approach. By this approach it is ensured, that the calculated volume, width and the
- 155 channel area fit together geometrically by including the term into the calculation.-
- 156 A fourth way of calculating the width and the volume is based on considerations concerning the flow
- 157 velocities and hydrodynamic processes in the channel. The same rigorous equations for the description of
- 158 the channel shape were used as for the calculation of  $V^{\text{geo}}$ . This leads to a direct linear relationship of  $t_{\text{void}}$
- 159  $t_0$  and w:

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- The "conversion factor"  $C_F$  is determined via the hydrodynamic and geometric properties of the 161
- measurement. It can be obtained by solving the integral: 162

$$C_F = \int_{z_0}^L \frac{E(\xi)}{v_{in} - V_c \cdot \frac{2 \cdot \int_0^{\xi} E(x) dx}{A}} d\xi \qquad (132)$$

- 164 This expression is derived based on a known approach [24], but independent from the shape and more
- 165 suited to relate w and  $\frac{1}{40}$  directly. The function E(x) describes the shape of the channel in dependence
- 166 of its longitudinal position x. In eq. 13, x marks the positions in the channel to the current position  $\xi$  of a
- 167 hypothetical non-retained species, which moves with the mean velocity.  $A_L$  is the complete surface of the
- 168 channel including the non-separating part, i.e.

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$$A_L = 2 \int_0^L E(x) dx$$
 (143)

- 170 A detailed derivation and an attempt for an analytical solution of Eq. -1142-143 is given the supportings
- 171 information. Using Eq. 5,  $V^{hyd}$  can be calculated. The values obtained by this method are denoted as  $V^{hyd}$
- in the following. A detailed derivation for the factor  $C_F$  and  $V^{hyd}$  is given in the supporting information. 172
- 173 The obvious advantage of this method is the independence from an external diffusion coefficient i.e. no
- 174 calibration measurement is involved in this procedure. This is a formalized method which is equivalent to
- 175 calibration free conversion approaches [16,25,26].
- 176 The fifth algorithm also makes use of this conversion factor. Here, it is used to substitute the void time #0
- 177  $t_{\text{void}}$ . As no experimental void time is used in this approach, we designated the calculated results with  $w^{\text{noT}}$
- 178 and  $V^{\text{noT}}$ . This way, Eq. 2 can be written as

$$R = \frac{2C_F w^{noT}}{t_e} \tag{154}$$

180 By reformulating Eq. 5 with the substituted volume according to Eq. 108, λ can be calculated written as:

$$\lambda(w) = \frac{DA_{\pi L}}{V_{c}w} \tag{165}$$

Now Eq.  $\underline{35}$  can be merged with  $\underline{\underline{Eq. (144)}}$ : 182

$$\frac{2C_{FW}}{t_{e}} = 6\lambda(w) \cdot \left(\coth\left(\frac{1}{2\lambda(w)}\right) - 2\lambda(w)\right) \tag{16.7}$$

- 184  $w^{\text{noT}}$  can now be easily determined numerically from Eq. 15.6 and Eq. 17.6 This calibration calculation
- 185 procedure is advantageous as tvoid to does not have to be determined experimentally and be read from the
- 186 fractogram. However, it requires a calibration measurement for the determination of  $t_e$  and a known D.

#### Materials and methods

AF4Eval is our current version of hydrodynamic evaluation software for AF4 data. The user can create profiles for channel shapes and corresponding calibrations for a measurement set. Data are provided in a standardized csv-file format. Further input formats such as AniML[27,28,29] are planned to be added. As  $t_0 \, t_{\rm cold}$  and  $t_{\rm c}$  have to be determined manually for some calibration methods, we integrated a simple graphical element (Fig. 4) with movable bars to the user interface to enable the user to pick the peak position manually without transferring the chosen value as it would be required in a spreadsheet application.

AF4Eval is written in C++14. Its source code obtainable via github and can be used without charge for academic purposes: https://github.com/biocrystal777/AF4Eval. Data shown in this report where obtained with a version compiled with g++ 6.3 under Debian Gnu/Linux 9.5, using the framework Qt 5.7 and the plotting library Qwt 6.1.2[30].

Table 1: Required input parameters for the described calibration algorithms

|                | Classical        | Approximated Volume         | Geometrical                 | Hydrodyamic             | Without <u>t<sub>void</sub>to</u> |
|----------------|------------------|-----------------------------|-----------------------------|-------------------------|-----------------------------------|
|                | $D_{ m calib}$   | $D_{ m calib}$              | $D_{ m calib}$              |                         | $D_{ m calib}$                    |
|                | <u>tvoid_</u> ₹0 | <u>tvoid_</u> ₹0            | <u>t<sub>void_</sub>t</u> 0 | <u>tvoid_</u> ₹0        |                                   |
|                | $t_{\mathrm{e}}$ | $t_{ m e}$                  | $t_{ m e}$                  |                         | $t_{ m e}$                        |
|                | $V_{\mathrm{e}}$ | $V_{ m e}$                  |                             | $V_{ m e}$              | $V_{\mathrm{e}}$                  |
| Inputs         | $V_{\rm c}$      | $V_{\rm c}$                 | $V_{\rm c}$                 | $V_{\rm c}$             | $V_{\rm c}$                       |
|                | Z%               | Z%                          | <del>2</del> %              | Z%                      | Z%                                |
|                |                  | $L_1, L_2, \underline{L_3}$ | $L_1, L_2, L_3$             | $L_1, L_2, L_3$         | $L_1, L_2, L_3$                   |
|                |                  | $b_0,b_{ m L}$              | $b_0,b_{ m L}$              | $b_0,b_{ m L}$          | $b_0,b_{ m L}$                    |
| Side condition | -                | -                           | Eq. <u>8</u> 9              | Eq. <del>9</del> 8      | Eq. <del>9</del> 8                |
|                | w                | w (w <sup>appgeo</sup> )    | w (w <sup>geo</sup> )       | w (w <sup>hyd</sup> )   | w (w <sup>noT</sup> )             |
| Outputs        | $V^0$            | $V^{0}(V^{\text{appgeo}})$  | $V^{0}(V^{\text{geo}})$     | $V^{0}(V^{\text{hyd}})$ | $V^{0}(V^{\text{nof}})$           |

Based on the theory above we implemented 5 calibration algorithms. The calibration-dependent three methods enable an estimation of the void peak time from the geometrical properties of the calibrated channel. Thus, manual readout of the void peak is avoided entirely and the methods can be integrated in a fully automated procedure. The direct conversion turns out to be useful if no appropriate standard is available. These algorithms vary in their specific required input magnitudes (Tab. 1). A pseudocode of the applied calculations representation is given in the supporting information.

In addition, an error analysis function was implemented that allows to estimate the uncertainties of the methods. The error analysis allows to define a range R of the estimated uncertainty  $\delta X$  from -100% to 100% for the input quantity  $X_i$  and a grid resolution parameter. The method then iterates over  $X+\delta X$  of R while conducting the assigned algorithm and gives the deviation of the output quantity  $Y_i$ . The method seems trivial from a mathematical perspective, however, it gives a good overview, how the deviation of one quantity affects the result while the other quantities are kept constant. This is especially useful if the impact cannot be foreseen due to the involvement in several numerical procedures. This way, the

individual impact of the uncertainty of each variable can be easily quantified for each experimental condition. Other functionalities, such as the evaluation of size distributions based on the calibration, are not discussed in this publication.

The experiments were conducted with a Wyatt Eclipse DualTec Separation system. The setup was coupled with a degassing unit (G1322A), an isocratic pump (G1310B) and an autosampler (G1328C), all from the Agilent 1260 series. Signals were recorded using the software Astra 6.1.7.17 with a sample rate of 0.5 Hz. A detector chain, consisting of a Dawn Heleos 8<sup>+</sup> MALLS detector (wavelength = 663 nm) a UV/VisDAD detector (G1315C, Agilent series 1100) and an RI detector (G7162A, Agilent series 1260). Before usage, an interdetector distance correction was conducted according to the elution peak of a BSA measurement using the respective function in Wyatt. Only signals with a decent signal-to-noise ratio were considered.

In the channel, an SC W350 spacer from Wyatt was used. The detailed dimensions are given in Table 3, together with the measurement conditions and related input parameters for the algorithms. Two samples were used: BSA and PS (Polystyrene) Nnanospheres. 3 replicates for each condition were measured. The eluent was 50 mM NaNO<sub>3</sub> for measurements with BSA. The injected sample amount was 20 μl. For the measurement with PS (Polystyrene) nanospheres, a 0.1 % w/v of SDS (sodium dodecyl sulfate) was used. The nanosphere standard particles had a nominal diameter of 60 nm. For our calculations, we assumed a hydrodynamic radius of 32 nm, which included the estimated influence of the surface stabilizer.

<u>m</u>Weasurements were conducted at 293 K. For each spacer a new 5 kDa Millipore regenerated Cellulose membrane was used. Detailed measurement program and sequence setup is given in the supplementary information.

## 4. Results

### 4.1 Comparison of algorithms with test measurements

First, we tested the algorithms with data acquired from our own field-flow fractionation. Standard protocols with a constant crossflow were used. Before elution, the sample was focused at  $Z_{26}=12$ % of the total channel length. The details of the measurement protocols are given in the supporting information (Table S.1-S.3). BSA was measured at  $V_c$ =2.5 mlmin<sup>-1</sup> and  $V_c$ =3.5 mlmin<sup>-1</sup>. To provide an example for a lower crossflow, additional measurements with PS nanospheres were conducted at  $V_c$ =0.5 mlmin<sup>-1</sup>. For BSA, the signals of the UV-Vis detector were chosen, for the PS measurements, UV and RI were considered.

Table 2: Parameters used for calibration experiments with test measurements

| Parameter        | unit                            | BSA_Vc2_5            |      |      | BSA_Vc3_5 |               |      |                | PS_Vc0_5 |      |  |
|------------------|---------------------------------|----------------------|------|------|-----------|---------------|------|----------------|----------|------|--|
|                  |                                 | r1                   | r2   | r3   | r1        | r2            | r3   | r1             | r2       | r3   |  |
| tvoid_t0         | min                             | 0.53                 | 0.55 | 0.54 | 0.6       | 0.5           | 0.51 | 0.73           | 0.72     | 0.72 |  |
| $t_{\mathrm{e}}$ | min                             | 3.65                 | 3.65 | 3.35 | 4.6       | 4.5           | 4.5  | 15.8           | 15       | 15.1 |  |
| $V_{\rm c}$      | mlmin <sup>-1</sup>             | 2.5                  |      |      | 3.5       |               |      | 0.5            |          |      |  |
| $V_{\mathrm{e}}$ | mlmin <sup>-1</sup>             | 1.0                  |      |      | 1.0       |               | 1.0  |                |          |      |  |
| $D_{ m calib}$   | cm <sup>2</sup> s <sup>-1</sup> | 6.1*10 <sup>-7</sup> |      |      |           | $6.1*10^{-7}$ |      | $6.74*10^{-8}$ |          |      |  |
| z%               | %                               | 12                   |      |      | 12        |               | 12   |                |          |      |  |
| $b_0$            | mm                              | 22                   |      |      | 22        |               | 22   |                |          |      |  |
| $b_{ m L}$       | mm                              | 3                    |      |      | 3         |               | 3    |                |          |      |  |

| $L_1$ | mm | 20  | 20  | 20  |
|-------|----|-----|-----|-----|
| $L_2$ | mm | 150 | 150 | 150 |
| $L_3$ | mm | 3   | 3   | 3   |

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The plotted signals of the fractograms were used to pick positions of two the BSA monomer peak, as shown exemplarily in Fig. 5. The fractograms have been corrected in time by its offset due to the focusing-related steps. I.e. "0" on the time axis displays the actual time of elution from the focusing position  $z_{\%}$ .

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The complete fractograms and extracted peak positions of all 9 measurements are given in the supporting information (Fig. S.<u>6.1</u>9-S.<u>6.5</u>13). All information was collected (Table 2) and processed by the 5 calibration algorithms of AF4Eval. The calculated channel widths and volumes are displayed in Fig. 6. The acquired retention times were typical for our AF4 setup and comparable to a lot of other measurements with similar conditions. In all three cases,  $V^{\text{app}Ggeo}$  is smaller than the calculated- $V^{\text{cla}}$ . This is simply reasoned by the smaller surface, when the trapezoidal tapering is considered in the calculation.

Consequently, the same observation is made for the relation of  $w^{app \text{Ggeo}}$  and  $w^{cla}$ . As each of our calibration calculations was deduced from essentially the same established AF4 theory, we would have expected each of the calculations to lead to a very similar result. Instead, introducing the geometrical side condition of Eq. 89 (w<sup>geo</sup> and V<sup>geo</sup>) made the calculated channel volume and related width shrink dramatically compared to the other calibration results.

Whereas removing the dependence on the known  $D_{\text{calib}}$  (for determination of  $w^{\text{hyd}}$ ) led to dramatically increased calculated channel widths, which are highly unrealistic as the spacer thickness of 350 µm represents the maximal possible distance between membrane and upper wall. Interestingly, the related separation volume V<sup>hyd</sup> was nearly the same as for the established first two variants as they use a very similar assumption for the derivation of the respective volume. This indicates that the formalism is indeed correct. Finally, removing twid to from the list input parameters by making use of Eq. 112 led to a result, which preserves the correct geometric relation, and resulted in a channel thickness within the expected

Also, it could be observed that the ratio the calculated  $V^{\text{cla}}$ ,  $V^{\text{appGeo}}$  their respective calculated w were increased when a higher crossflow was applied although the membrane area should be independent from  $V_c$  For  $w^{hyd}$  and  $V^{hyd}$  also an increase with constant ratio was observed. The difference was too big to be explained by the higher pressure on the membrane. The discrepancy was not observed for the other algorithms.

277 In order to exclude potential errors in our own experimental setups, we applied the algorithms on 278 published fractogram data.

4.2 Comparison of algorithms using published data in literature

For the reevaluation we screened the respective AF4-related literature for data sets which provided 281 fractograms and required information on the applied measurement conditions if possible. Especially the focus position z<sub>%</sub> was not always indicated. For our investigations, we decided, to assume a value which 282 283 is typically used for channels without frit-inlet. The implication of possible deviations of this real value 284 are discussed below separately. We chose to work with measurements made with spherical gold 285 nanoparticles [16,31], Silica particles [16] and silver nanospheres [32,33]. Measurements in the used

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literature had been conducted using trapezoidal channel geometries with different channel dimensions and AF4 devices from different vendors. The respective diffusion coefficient was calculated according to the specifications made in the respective publication and the expected shell thickness for polymer coatings[34,35]. All used parameters are displayed in Table 3 and Table 4. We applied the algorithms in the same way as we did for our own measured data.

Table 3: Parameters used for calibration calculation with literature data

| Parameter        | unit                            | Lit_Au1[30]   | Lit_Au2[16]   |        | LitAu3[16] |           | Lit_Sil[16] |
|------------------|---------------------------------|---------------|---------------|--------|------------|-----------|-------------|
|                  |                                 |               | Vc1_0         | Vc0_75 | Vc1_0      | Vc0_75    |             |
| <u>tvoid_</u> t0 | min                             | 0.593         | 0.89          | 0.89   | 0.992      | 0.992     | 0.96        |
| $t_{ m e}$       | min                             | 4.42          | 4.55          | 0.992  | 9.45       | 9.45      | 13.3        |
| $V_{\rm c}$      | mlmin <sup>-1</sup>             | 0.8           | 1.0           | 0.75   | 1.0        | 0.75      | 0.2         |
| $V_{\mathrm{e}}$ | mlmin <sup>-1</sup>             | 0.5           | 1.0           |        | 1.0        |           | 0.2         |
| $D_{ m calib}$   | cm <sup>2</sup> s <sup>-1</sup> | $6.1*10^{-7}$ | $6.1*10^{-7}$ |        | 6.74       | 6.74*10-8 |             |
| z%               | %                               | 12            | 12            |        | 12         |           | 12          |
| $b_0$            | mm                              | 22            | 22            |        | 22         |           | 22          |
| $b_{ m L}$       | mm                              | 3             | 3             |        | 3          |           | 5           |
| $L_1$            | mm                              | 20            | 20            |        | 20         |           | 22          |
| $L_2$            | mm                              | 150           | 150           |        | 150        |           | 213         |
| $L_3$            | mm                              | 3             | 3             |        | 3          |           | 5           |

Table 4: Parameters used for calibration calculation with literature data

| Parameter        | unit                            | LitAG[31] |                    |                       |         |                         | Lit_Ag                         | _42nm[32]             |
|------------------|---------------------------------|-----------|--------------------|-----------------------|---------|-------------------------|--------------------------------|-----------------------|
|                  |                                 | 10        | 20nm               | 30nm                  | 50nm    | 70nm                    | 90nm                           |                       |
|                  |                                 | nm        |                    |                       |         |                         |                                |                       |
| <u>tvoid</u> ŧ0  | min                             | 0.75      | 0.75               | 0.75                  | 0.75    | 0.75                    | 0.75                           | 0.75                  |
| $t_{ m e}$       | min                             | 1.81      | 4.32               | 7.85                  | 10.11   | 12.02                   | 14.32                          | 10.39                 |
| $V_{\rm c}$      | mlmin <sup>-1</sup>             |           |                    |                       | 0.4     |                         |                                | 1                     |
| $V_{\mathrm{e}}$ | mlmin <sup>-1</sup>             |           |                    |                       | 1.0     |                         |                                | 0.4                   |
| $D_{ m calib}$   | cm <sup>2</sup> s <sup>-1</sup> | 4.29*10   | <sup>7</sup> 2.14* | 1.43*10 <sup>-7</sup> | 8.58*10 | 0 <sup>-8</sup> 6.13*10 | $0^{-8}$ 4.76*10 <sup>-8</sup> | 1.02*10 <sup>-7</sup> |
|                  |                                 |           | $10^{-7}$          |                       |         |                         |                                |                       |
| z%               | %                               |           |                    |                       | 12      |                         |                                | 12                    |
| $b_0$            | mm                              |           |                    |                       | 20      |                         |                                | 24                    |
| $b_{ m L}$       | mm                              |           |                    |                       | 5       |                         |                                | 3                     |
| $L_1$            | mm                              |           |                    |                       | 20      |                         |                                | 22                    |
| $L_2$            | mm                              | 250 152   |                    |                       |         |                         |                                | 152                   |
| $L_3$            | mm                              |           |                    |                       | 5       |                         |                                | 3                     |

 The results of our evaluations are displayed in Fig.7. For the data set Lit\_Au1, we could confirm all effects of previous findings:  $w^{\text{geo}}$  was usually significantly smaller than the other calculated channel distances, while the calculated value for  $w^{\text{hyd}}$  increased. Only in two cases we found the opposite result, but also here, the presence of differences as such was remarkable.

Due to our lack of knowledge of  $\epsilon_{k}$ , we repeated the analysis for our own data as well for the literature data also for the alternative values 8% and 16%. The respective results do not vary significantly from the observations made for 12% and are displayed in the supporting information (S.6.19).

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- 302 It can be summarized that applying AF4 theory blindly leads to conspicuously inconsistent results. As we
- 303 counterchecked the derivation of our formulas and implementation into the software multiple times, we
- 304 speculated this behavior to be reasoned by a systematic measurement error. For this reason, we continued
- and investigated the influence of measurement uncertainties on the resulting sizes.
- 306 Influence of measurement uncertainties on the calibration result
- 307 In the following, we implemented an additional feature which displays the effect of deviations to the final
- 308 results. The task was implemented by a generic preprocessor macro, which keeps all but one parameter
- 309 constant and iterates over a grid of the remaining parameters to be investigated. Thereby, we plot the
- impact of a deviation  $\delta X$  of a parameter X to the relative deviations of the resulting volume and channel
- 311 widths ( $\delta Y$ ). This analysis is useful as due to the included numeric procedures and the relationship
- between the two output variables and its inputs is not obvious, especially when an input variable is used
- 313 multiple times. In addition, this analysis can be used as a base for an estimation of the total propagation of
- 314 uncertainty. Here, we show the result of this analysis, where the central parameters  $\delta X=0$  are taken from
- 315 the parameter set BSA\_Vc2\_5\_r1.
- 316 Fig. 8 shows the influence of deviations to 4 of the parameters, we expected to exhibit the highest
- 317 possible uncertainty.  $t_0 t_{\text{void}}$  and  $t_0$  have to be considered as they are the primary raw measured results. The
- 318 physical dimensions of the channel area and the applied flow volumes are assumed to be measured with
- 319 high accuracy, thereby, we don't discuss them further here in detail. Also, we skip the discussion on the
- 320 first algorithm due to its similarity to the second one. However, the resulting data of full analysis of all 5
- 321 algorithms and 10 parameter sets is shown in the supporting information. The diffusion coefficient
- 322 externally acquired has to be considered here, as for many samples, different numerical values have been
- published [36,37,38,39,40]. Also, side conditions like temperature and the temperature-dependent
- viscosity can influence the effective diffusion coefficient[10], where the latter one is often not known
- 325 exactly in practice. The analysis shows that all procedures but the fourth (where D is not used) scale with
- 326 D. However, the dependence was significantly stronger for the third procedure, while the volume is
- completely neglected for the second. The same relationships are shown for the input variable  $t_e$ .
- 328 Interestingly, a deviation of the focus position seems only to contribute significantly if the actual position
- is strongly displaced from its designated position. The relation of the deviations of to deviations of the deviations of
- 330 remarkable. While the linear relationship of the calculated volume was obvious, there was almost no
- impact on  $w^{appGeo}$ . For the third and fourth procedures, we found a proportional and a nearly inverse
- proportional relationship. As logical consequence a deviation of <u>to\_tvoid</u> leads to a divergence of the results
- 333 in these procedures as it could be seen in the experimental results. This consideration lead to a closer
- 334 consideration of an impact of the void peak time.
- 335 4.3 Convergence of the calibration calculations via the adjustment of to two
- 336 Similarly, to the analysis above, # tvoid was varied for all 5 algorithms and the resulting channel widths.
- 337 The analysis was conducted for one measurement per triplicate. The results, shown in Fig. 9, indicate a
- convergence of all algorithms for w and V for a value of  $\frac{1}{2}$  a range of  $\frac{1}{2}$  which is significantly
- dislocated from the respective measured value. We concluded that the position of the intersection point
- rather designates the position which should represent the actual position of  $t_{\text{void}}$  by its definition (the time
- used of passing the channel with the average migration velocity, starting from the focus position). This
- confirms prior observations that the measured void peak might yield wrong results [1,10].
- The results of an analysis for this intersectional  $t_{\text{void}}$  are shown in Fig. 10. Besides the balanced results for
- 344 <u>the single measurements we can also show that the discrepancies of the results between the different</u>
- 345 crossflows now completely vanish. Similar results could be obtained for the evaluation of the given

- 346 literature data (Supporting information). Consequently, only a much smaller value can be considered as
- 347 the "true" void peak position by its definition (the time used of passing the channel with the average
- 348 migration velocity, starting from the focus position).

#### 5. Discussion

- 350 5.1 Invalidity of the measured void peak
- 351 Our results show several implications towards working out best practices in AF4 calibration. As it could
- 352 be demonstrated, the measured "void peaks" are not suited to reproduce calibration results which are
- 353 consistent with the elaborated AF4 theory. An obvious potential error might be a bad correction of the
- 354 offset volume in our measurement setup. However, the error has been shown to occur abundantly in
- 355 literature data as well. As a consequence, 40 tvoid would be a very unreliable source of information just
- 356 because of this pitfall. Moreover, we even believe the observed small peak in AF4 should not be related to
- 357 this magnitude at all.
- 358 5.2 Alternate hypothesis for the occurrence of the "void peak"
- 359 Finally, we want to give another possible explanation for the reason behind the occurrence of the little
- observable peak, which has been considered as the void peak. To the best of our knowledge this
- 361 explanation has not been reported elsewhere up to now. FFF is fundamentally based on the friction
- between the solvent and the channel wall. This friction is responsible for forming the parabolic flow
- 363 profile. However, the friction to the side walls is usually not considered at all. Still, it should be assumed
- that a similar velocity gradient is present immediately at these side channel walls. While we assume the
- cross-flow to be uniformly distributed over the channel area and the migration velocity to be dependent
- 366 solely on x- and z position referring to Fig. 3, this is probably not true at these border regions. A more
- sophisticated hypothetical flow distribution is shown in Fig.  $1\underline{10}$ .
- 368 Because of the friction of solvent and channel side wall, 4 regions with different flow are present in this
- model. Due to diffusion, a sample particle can easily migrate from one of these zones into another. By
- this effect, a small amount of the sample partially eludes the retention effect of the crossflow. This portion
- 371 is then detected much earlier than the retained sample. However, it does not accompany a "solvent front",
- a term used for the description other chromatography techniques, which has no actual correspondent in
- field-flow-fractionation: Due to the parabolic flow distribution, the single laminar layers of the solvent
- flow are permanently displaced against each other. As a result, no common elution speed for all layers
- 375 should exist. It has been reported that the observed peak intensity increases with decreasing  $V_c$  [24,41].
- This might be reasoned by an increase of the permeable (i.e. unaffected from crossflow) area in the
- 377 presented model.
- 378 To give experimental evidence, dedicated mathematical models will have to be developed in the future.
- Therefore, the physical nature of hadvoid has to be considered as a theoretical auxiliary construct (i.e. "the
- time needed to flush the separation volume if there was no flow gradient along the diameter" as it was the
- case for very big wall distance). One should not rely on an experimental value for this parameter
- 382 consequently. Approaches already exist, which substitute its value by the information on the channel
- shape and the external calibration measurement using a calibrant with known diffusion coefficient. Our
- fifth algorithm belongs to this group as well as some simplified versions in literature [10,17,20,42,43] or
- the replacement of the known D with a related property of the calibrant [18]. According to our study, this
- groups remains the only valid approach to conduct the calibration properly.
- 387 **6. Conclusions**

- 388 We have shown that the measured void peak is not a reliable value for the evaluation of field-flow-
- 389 fractionation data. Based on this observation, we could confirm this hypothesis by Wahlund [10] and give
- 390 strong evidence how a gold standard calibration for the evaluation of AF4 data with current instruments
- 391 can be performed. Our dedicated software for the evaluation has the potential to greatly improve the
- 392 practical handling of AF4 data. As already known, the information about channel volume and channel
- width are the critical quantities for contemporary machines. Avoiding 40 Lvoid as an error-prone calibration 393
- 394 input is likely to improve accuracy of AF4 measurements. The development of our software will be
- 395 continued, considering a list of features for numerous possible extensions:
  - Alternative calibration methods as investigated recently [18]. Also, distance measurement has improved continuously [44], therefore we think that further calibration methods based on combination of the channel with such a device might be an additional orthogonal tool for the calibration.
  - Crossflow gradients [5,45,46], steric effects and decays [10,20,21]. Our final goal will be to provide an open and extensible reference implementation, which gathers all these state-of-the-art evaluation methods.
  - Advanced handling of light scattering data.
  - AF4 related deconvolution techniques [22,47].
  - Improved focus point determination as recently presented [43].
  - An API for scripting languages.
- 407 As distance measurement by optical methods has improved continuously over the last decades up to
- 408 submicrometer precision a direct measurement of w will be an additional improvement. If the width could
- be provided directly, the presented formalism could be applied easily to determine D without prior 409
- calibration. It was stated by Wahlund[10] that, unfortunately, hydrodynamic calculations were not 410
- 411 available in commercial software. Our platform-independent and vendor-agnostic implementation is
- 412 designed to fill this gap, relying on a state-of-the-art technology stack. We encourage users of AF4 get in
- 413 contact to discuss possible extensions for their specific needs.

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#### 418 8. References

- 419 [1] K.-G. Wahlund, J. C. Giddings, Properties of an asymmetrical flow field-flow fractionation channel having one
- 420 permeable wall, Anal. Chem. 1987, 59(9), 1332-1339; https://doi.org/10.1021/ac00136a016
- 422 [2] J. C. Giddings, F. J. Yang, M. N. Myers, Flow field-flow fractionation: new method for separating, purifying,
- 423 and characterizing the diffusivity of viruses, J. Virol .1977, 21(1), 131-138;
- 425 [3] H. Cölfen, M. Antonietti, Field-flow fractionation techniques for polymer and colloid analysis, in: New
- 426 Developments in Polymer Analytics I, Editor: Manfred Schmidt, Springer, 2000, Adv. Pol. Sci., 150, 67-187
- 428 [4] J. C. Giddings, Field-Flow Fractionation: Analysis of Macromolecular, Colloidal, and Particulate Materials,
- 429 Science 1993, 260, 1456-1465; https://doi.org/10.1126/science.8502990

- 431 [5] A. Litzén, K. G. Wahlund, Improved separation speed and efficiency for proteins, nucleic acids and viruses in
- 432 asymmetrical flow field flow fractionation, J. Chromatogr. A 1989, 476, 413-42 https://doi.org/10.1016/S0021-
- 433 9673(01)93885-3
- 434 435 [6] A. Litzán, L.
- 435 [6] A. Litzén, J.K. Walter, H. Krischollek, K.-G. Wahlund, Separation and quantitation of monoclonal antibody
- 436 aggregates by asymmetrical flow field-flow fractionation and comparison to gel permeation chromatography, Anal.
- 437 Biochem., 1993, 212(2), 469-480; https://doi.org/10.1006/abio.1993.1356
- 438
- [7] G.Yohannes, S. K. Wiedmer, M. Elomaa, M. Jussilal, V. Aseyev, M.-L. Riekkola, Thermal aggregation of
- bovine serum albumin studied by asymmetrical flow field-flow fractionation, Anal. Chim. Acta 2010, 675, 191-198;
- 441 https://doi.org/10.1016/j.aca.2010.07.016
- 442
- 443 [8] L. F. Pease, D. I. Lipin, D.-H. Tsai, M. R. Zachariah, L. H. L. Lua, M. J. Tarlov, A. P. J. Middelberg,
- 444 Quantitative characterization of virus-like particles by asymmetrical flow field flow fractionation, electrospray
- 445 differential mobility analysis, and transmission electron microscopy, Biotechnol. Bioeng. 2008, 102(3), 845-855;
- 446 http://dx.doi.org/10.1002/bit.22085
- 447
- 448 [9] W. Fraunhofer, G. Winter, C. Coester, Asymmetrical flow field-flow fractionation and multiangle light
- scattering for analysis of gelatin nanoparticle drug carrier systems, Anal. Chem. 2004, 76(7), 1909-1920;
- 450 http://dx.doi.org/10.1021/ac0353031
- 451
- 452 [10] K.-G. Wahlund, Flow field-flow fractionation: critical overview, J. Chromatogr. A 2013, 1287, 97-112;
- 453 https://doi.org/10.1016/j.chroma.2013.02.028
- 454
- 455 [11] W. Hiller, W. van Aswegen, M. Hehn, H. Pasch, Online ThFFF-NMR: A Novel Tool for Molar Mass and
- 456 Chemical Composition Analysis of Complex Macromolecule, Macromolecules 2013, 46(7), 2544-2552,
- 457 https://doi.org/10.1021/ma400350y
- 458
- 459 [12] G.Yohannes, M. Jussila, K. Hartonen, M-L Riekkola, Asymmetrical flow field-flow fractionation technique for
- 460 separation and characterization of biopolymers and bioparticles, J Chromatogr A 2011, 1218, 4104-4116;
- 461 https://doi.org/10.1016/j.chroma.2010.12.110
- 462

- 463 [13] A. F. Thünemann, P. Knappe, R. Bienert and S. Weidner, Online coupling of field-flow fractionation with
- 464 SAXS and DLS for polymer analysis, Anal. Methods 2009, 1(3), 153-228, https://doi.org/10.1039/B9AY00107G
- 466 [14] P. Schuck, Size-distribution analysis of macromolecules by sedimentation velocity ultracentrifugation and
- 467 lamm equation modeling, Biophys. J. 2000, 78(3), 1606-1619, https://doi.org/10.1016/S0006-3495(00)76713-0
- 468
- 469 [15] B. Demeler, UltraScan A Comprehensive Data Analysis Software Package for Analytical Ultracentrifugation
- 470 Experiments, in: Analytical Ultracentrifugation: Techniques and Methods, 2005, Editor: D. J. Scott, S. E. Harding
- 471 and A. J. Rowe
- 472
- 473 [16] A. Zattoni, D. C. Rambaldi, P. Reschiglian, M. Melucci, S. Krol, A. M. Coto-Garcia, A. Sanz-Medel, D.
- 474 Roessner, C. Johann, Asymmetrical flow field-flow fractionation with multi-angle light scattering detection for the
- analysis of structured nanoparticles, J. Chromatogr. A 2009, 1216, 9106-9112;
- 476 https://doi.org/10.1016/j.chroma.2009.06.037

478 [17] A. Litzén, Separation Speed, Retention, and Dispersion in Asymmetrical Flow Field-Flow Fractionation as 479 Functions of Channel Dimensions and Flow Rates, Anal. Chem. 1993, 65(4), https://doi.org/10.1021/ac00052a025 480 481 [18] H. Bolinsson, Y. Lu, S. Hall, L. Nilsson, A. Håkannsson, An alternative method for calibration of flow-field-482 flow fractionation channels for hydrodrnamic radius determination: The nanoemulsion method, J. Chromatogr. A 483 2018, 1553, 155-163, https://doi.org/10.1016/j.chroma.2017.12.026 484 485 [19] M. Martin, M. Hoyos, On the no-fied method for void time determination in flow field-flow fractionation 486 J. Chromatogr. A, 1218, 4711-4125, https://doi.org/10.1016/j.chroma.2011.01.010 487 488 [20] A. Håkansson, E. Magnusson, B. Bergenståhl, L. Nilsson, Hydrodynamic radius determination with 489 asymmetrical flow field-flow fractionation using decaying cross-flows. Part I. A theoretical approach J. Chromatogr. 490 A 2012, 1253, 120-126, https://doi.org/10.1016/j.chroma.2012.07.029 491 492 [21] E. Magnusson, A. Håkansson, J. Janiak, B. Bergenståhl, L. Nilsson, Hydrodynamic radius determination with 493 asymmetrical flow field-flow fractionation using decaying cross-flows. Part II. Experimental evaluation, 494 J. Chromatogr. A 2012, 1253, 127-153, https://doi.org/10.1016/j.chroma.2012.07.005https://doi.org/10.1016/j.chroma.2012.07.005 495 496 [22] M. Schmid, B. Häusele, M. Junk, E. Brookes, J. Frank, H. Cölfen, High-Resolution Asymmetrical Flow Field-497 Flow Fractionation Data Evaluation via Richardson-Lucy-Based Fractogram Correction, Anal. Chem. 2018, 90, 498 43978-13986, https://doi.org/10.1021/acs.analchem.8b03483 499 500 [XX] J. C. Giddings, M. N. Myers, Steric Field-Flow-Fractionation: A new method for separating 1 to 501 100 µm particles, Sep. Sci. Technol. 1978, 13(8), 673-645, https://doi.org/10.1080/01496397808057119 502 503 [23] M. R. Schure, Fast Algorithm for the Conversion of R to Lambda Values in Field-Flow Fractionation, Sep. 504 Sci. Technol, Separation Science and Technology 1987, 22(12), 2403-2411, https://doi.org/10.1080/01496398708057194 505 506 [24] A. Litzén, K.-G. Wahlund, Zone Broadening and Dilution in Rectangular and Trapezoidal — Asymmetrical Flow 507 Field-Flow Fractionation Channels, J. Chromatogr. A 1991, Analytical Chemistry, 63, 1001-1007 508 509 [25] Katri Eskelin, Minna M. Poranen, Hanna M. Oksanen, Asymmetrical Flow Field-Flow Fractionation on Virus 510 and Virus-Like Particle Applications, Microorganisms 2019, 7(11), 1-20; 511 https://doi.org/10.3390/microorganisms7110555 512 513 514 [26] V. de Carsalade du pont, E. Alasonati, S. Vaslin-Reimann, M, Martin, M. Hoyos, P. Fisicaro, Asymmetric field flow fractionation applied to the nanoparticles characterization: Study of the parameters governing 515 the retention in the channel: 19th International Congress of Metrology 2019, 516 https://doi.org/10.1051/metrology/201923001 517 518 [27] B. A. Schäfer, D. Poetz, G. W. Kramer, Documenting Laboratory Workflows Using the Analytical Information 519 Markup Language, J. Lab. Autom. 2004, 9, 375-381; https://doi.org/10.1016/j.jala.2004.10.003

477

520 521

[28] A. Roth, R. Jopp, R. Schäfer, G. W. Kramer, Automated Generation of AnIML Documents by Analytical

522 Instruments, J. Lab. Autom. 2006, 11, 247-253, https://doi.org/10.1016/j.jala.2006.05.013 523 524 [29] T. Davies, Herding AnIMLs, Chemistry International, 29(6), 21-23, 525 http://publications.iupac.org/ci/2007/2906/pp1\_animls.html 526 527 [30] U. Rathmann U. Qwt - Qt Widgets for Technical Applications, 2014 528 [31] A.-R. Jochem, G. N. Ankah, L.-A. Meyer, S. Elsenberg, C. Johann and T. Kraus, Colloidal Mechanisms of 529 530 Gold Nanoparticle Loss in Asymmetric Flow Field-Flow Fractionation, Anal. Chem. 2016, 88, 10065-10073, 531 https://doi.org/10.1021/acs.analchem.6b02397 532 533 [32] G.F. Koopmans, T. Hiemstra, I.C. Regelink, B. Molleman, R.N.J. Comans, Asymmetric flow field-flow 534 fractionation of manufactured silver nanoparticles spiked into soil solution, J. Chromatogr. A 2015, 1392, 100-109; 535 https://doi.org/10.1016/j.chroma.2015.02.073 536 537 [33] K. Loeschner, J. Navratilova, C. Købler, K. Mølhave, S. Wagner, F. von der Kammer, E. H. Larsen, Detection 538 and characterization of silver nanoparticles in chicken meat by asymmetric flow field flow fractionation with 539 detection by conventional or single particle ICP-MS, Anal. Bioanal. Chem. 2013, 405, 8185-8195; 540 https://doi.org/10.1007/s00216-013-7228-z541 542 [34] G. Duplâtre, M. F. Ferreira Marques, M. da Graça Miguel, Size of Sodium Dodecyl Sulfate Micelles in 543 Aqueous Solutions as Studied by Positron Annihilation Lifetime Spectroscopy, J. Phys. Chem. 1996; 544 https://doi.org/10.1021/jp960644m 545 546 [35] F. Bockstahl, E. Pachoud, G. Duplâtre, I. Billard, Size of sodium dodecyl sulphate micelles in aqueous NaCl 547 solutions as studied by positron annihilation lifetime spectroscopy, Chem. Phys. 2000, 256, 307-313, 548 https://doi.org/10.1016/S0301-0104(00)00126-9 549 550 [36] T. Raj, W. H. Flygare, Diffusion Studies of Bovine Serum Albumin by Quasielastic Light Scattering, 551 Biochemistry 1974, 13(16), 3336-3340; https://doi.org/10.1021/bi00713a024 552 553 [37] L. A. Larew R. R. Walter, A Kinetic, Chromatographic Method for Studying Protein Hydrodynamic Behavior, 554 Anal. Biochem. 1987, 164, 537-546, https://doi.org/10.1016/0003-2697(87)90530-6 555 556 [38] K. J. Stelzer D. F. Hastings M. A. Gordon, Treatment of Mobile Phase Particulate Matter in Low-Angle Quasi-557 elastic Light Scattering, Anal. Biochem. 1984, 136, 251-257; https://doi.org/10.1016/0003-2697(84)90332-4 558 559 [39] C. B. Fuh, S. Levin, J. C. Giddings, Rapid Diffusion Coefficient Measurements Using Analytical SPLITT 560 Fractionaction: Application to Proteins, Anal. Biochem. 1993, 208, 80-87; https://doi.org/10.1006/abio.1993.1011 561 562 [40] M.-K. Liu, P. Li, J. C. Giddings, Rapid protein separation and diffusion coefficient measurement by frit inlet 563 flow field-flow fractionations, Protein Sci. 1993, 2, 1520-153; https://doi.org/10.1002/pro.5560020917 564

[41] Wim T. Kok, Rashid. N.Qureshi, Optimization of Asymmetrical Flow Field-Flow Fractionation

565

566

567

(AF4), LG GC Eur. 2010, 23(1), 18-25

571 572 [43] J.-L. Wang and E. Alasonati, P. Fisicaro, M. F. Benedetti, M. Martin, Theoretical and experimental 573 investigation of the focusing position in asymmetrical flow field-flow fractionation (AF4), J. Chromatogr. A 2018, 574 1561, 67-75; https://doi.org/10.1016/j.chroma.2018.04.056 575 576 [44] G. Berkovic, E. Shafir, Optical methods for distance and displacement measurements, Adv. Opt. Photonics, 577 2012, 441-471 https://doi.org/10.1364/AOP.4.000441 578 579 [45] J. J. Kirkland, E. I. DuPont de Nemours, C. H. Dilks Jr., S. W. Rementer, W. W. Yau, Asymmetric-channel flow field-flow fractionation with exponential force-field programming, J. Chromatogr. A 1992, 339-355; 580 581 https://doi.org/10.1016/0021-9673(92)80303-C 582 583 [46] P.S. Williams, M. C. Giddings, J.C. Giddings, A data analysis algorithm for programmed field-flow 584 fractionation, Anal. Chem. 2001, 73(17), 4202-4211; https://doi.org/10.1021/ac010305b 585 586 [47] M. R. Schure, B. N. Barman, J. C. Giddings, Deconvolution of Nonequilibrium Band Broadening Effects for 587 Accurate Particle Size Distributions by Sedimentation Field-Flow Fractionation, Anal. Chem. 1989, 61(24), 2735-

[42] B. Wittgren, K.-G. Wahlund, H. Dérand, B. Wesslén, Aggregation Behavior of an Amphiphilic Graft

29, 268-276; https://doi.org/10.1021/ma950837s

2743, https://doi.org/ 10.1021/ac00199a011

Copolymer in Aqueous Medium Studied by Asymmetrical Flow Field-Flow Fractionation, Macromolecules 1996,

568569

570