

Towards a more objective inference of local and regional vegetation types using pollen assemblages from the East Africa Rift (Ethiopia)

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Abstract: Pollen grains are particularly important for the study of Hominini sites in the Plio-Pleistocene of East Africa Rift, which have long been known to be associated with riparian environments. These riparian environments allow the development of a wide variety of local vegetation that differs greatly from the surrounding regional vegetation. However, it is difficult to differentiate the local pollen signal from the regional pollen signal. In addition, in East Africa, an important part paleo-environmental reconstructions from pollen are based at least in part on empirical observations. These two elements make it difficult to appreciate the relevance of paleo-environmental interpretations, mainly their relevance regarding local vegetation. To help solve these problems, this study presents a proof of concept based on the calculation of pollen scores to objectively discriminate local vegetation (in particular riparian vegetation) from a modern data set of 283 surface pollen samples. The aim is to see and discuss the effectiveness and limitations of our concept in the modern data before potentially applying it to fossil data. Our approach allows the identification of 24 types and sub-types of vegetation, nine of which are local riparian and azonal vegetation. It differs from previous pollen scoring approaches in the spatial precision of the reconstruction of vegetation (lower than the regional

scale), the integration of riparian vegetation, and further treatment of aquatic plant pollen. We highlight that relative abundance is not the best way to deal with pollen data and the importance of treating pollen data differently depending on the conditions of deposit: relative abundance scale for non-dynamic deposition environments (e.g., soils or lake edges) and presence/absence for dynamic deposition environments (e.g., river beds). The interest of this approach is not limited to the determination of vegetation but also to the inference of climatic parameters, the percentage of tree cover, and river morphology.

1. Introduction

Many paleontological areas in Ethiopia have preserved abundant fauna, including hominin remains of Late Miocene, Pliocene, and Pleistocene age (e.g., Campisano et al., 2017; Haile-Selassie et al., 2015; Johanson, 2017; Saylor et al., 2019; Villmoare et al., 2015). Ethiopia is an important place to study the history of human evolution and to understand how the environment and climate may have influenced it (Levin, 2015). For some paleontological areas in Afar (Ethiopia), such as the *Australopithecus afarensis*-bearing Hadar Formation (Bonnefille et al., 1987), the *Ardipithecus ramidus*-bearing Aramis member (WoldeGabriel et al., 2009), more recently the *Australopithecus anamensis*-bearing deposits at Woranso-Mille (MRD locality) (Saylor et al., 2019), and other *Australopithecus* sites such as Meshellu/Dikika (Bonnefille, 2010), pollen were extensively studied and provided direct botanical evidences of the plant taxa associated with early hominids. By comparison with modern pollen data obtained from surface soil samples, fossil pollen relative abundances allowed to appraise the composition of past plant communities (Bonnefille et al., 1987). This comparative approach is possible because the floristic composition of plant ecosystems in East Africa has been relatively stable over the last 11 Ma. (Bonnefille, 2010).

In the 1990s, based on the work of Prentice et al. (1992) on the characterization of biomes in Europe by a combination of PFTs, Jolly et al. (1998) proposed a method of reconstructing vegetation and climate from pollen for Africa and the Arabian Peninsula: the Plant Functional Type (PFT) method. This method is based on the allocation of one or more PFTs to the different pollen types and was calibrated with 966 modern pollen samples from Africa, sub-Saharan Africa and the Arabian Peninsula. It excluded pollen from aquatic plants and allowed the reconstruction of biomes *in equilibrium* with climate (e.g., desert, steppe, savannah, warm mixed forest, tropical rain forest). The PFT method produced consistent reconstructions of past (mid-Holocene and Pleistocene) and present climate parameters (Bonnefille et al., 2004; Jolly et al., 1998; Peyron et al., 2000). However, the interpretation by the PFT method of past and present biome is unfortunately not efficient in East African (Bonnefille

et al., 2004; Peyron et al., 2000). Vincens et al. (2006) highlight the difficulties of reconstructing, with the PFT method, the mosaic of open and closed vegetation, which is the most frequently defined habitat type for early hominids (Reynolds et al., 2015; Stewart, 2014).). This method has shown its limitations. It is necessary to find another way to extract a more precise signal from the vegetation than the biome if we want to apply it to the reconstruction of hominid paleohabitats

Today, for the Afar and more generally for the East Africa Rift, the PFT method has been abandoned for paleo-palynological studies and we returned to a more empirical approach (R. Bonnefille, personal communication). This makes it common to see oppositions between different authors on how to interpret the presence of a pollen type. This is the case of *Hyphaene* pollen which are considered as a signal of edaphic forest by Barboni et al. (2019) and a signal of sub-desert steppe by Bonnefille et al. (1987). In addition, the same author may, without clear justification, interpret the same pollen spectra differently. For example, Pliocene pollen samples from sub-member SH-2 at Hadar (sample KMa to KMi) have been interpreted as swamp "close to evergreen bushland and forest" by Bonnefille et al. (1987) and swamp "close to wet/dry grassland" by Bonnefille et al. (2004). Moreover, the difficulty in separating the local pollen signal from the regional pollen signal in the East Africa Rift (Bonnefille et al., 1987) makes the environmental reconstructions based on pollen often unclear and subject to interpretation. This is reflected in the use of terms such as "dry shrubland with varying proportions of grassland, wetland and riparian forest" (Saylor et al., 2019) or "woodland-grassland landscape" (Bonnefille, 2010). In addition to this problem, the comparisons between modern and fossil pollen data are more often made in terms of relative abundance of taxa than in terms of presence/absence as it is generally the case with paleontological (faunal) data. Working with relative abundance pollen data can be a serious issue when dealing with fossil assemblages that definitely underwent significant taphonomical processes. Unfortunately, this is the case for the sites presented above, and more globally for the East Africa Rift sites of Late Miocene, Pliocene, and Pleistocene age (Bonnefille, 1984; El-Moslimany, 1983). On the other hand, ignoring the difference of pollen relative abundance resulting from a real difference in the vegetation between two sites is also not satisfactory. The use of a relative abundance scale is a compromise that still needs to be tested.

Finally, reconstructing vegetation (biomes) *in equilibrium* with climate may provide plausible paleo-climate estimates, but is a too coarse approach to capture the habitat diversity that likely made the attractive places for a wide variety of animals including several hominin species as recorded by the abundance and diversity of faunal taxa in this area (Faith et al., 2019; Reed et al., 2013). Although the presence of regional vegetation types and sub-types with different climatic conditions is clearly established by botanists in the East African Rift (Friis et al., 2010), the potential of pollen to characterize

96 them objectively has not been developed. In arid and semi-arid eastern Africa, surface water such as
97 rivers, lakes and groundwater discharge areas (springs) promotes the development of micro-habitats,
98 and strongly influences vegetation and wildlife distribution (e.g., Friis et al., 2010; Geheb and Abebe,
99 2003). These azonal wooded riparian vegetation associated with permanent water were particularly
100 attractive for Hominini in the past (Barboni et al., 2019). Yet, few vegetation proxies allow
101 reconstructing paleo-vegetation at the spatial scale of hominid sites. Phytoliths and organic
102 compounds were used for this purpose at a few sites in Ethiopia (WoldeGabriel et al., 2009) and
103 Tanzania (e.g., Arráiz et al., 2017; Magill et al., 2016), but their taxonomic resolution is too low to
104 characterize paleo-vegetation beyond its physiognomy (i.e., the paleo-tree cover), or beyond
105 indicating the presence of palms, sedges, ferns, or details within the grass paleo-communities. To our
106 knowledge, there is no more accurate proxy than pollen to describe the composition of past
107 vegetation. Yet, little interest has been given so far to the local vegetation (riverine and fed by springs)
108 signals contained in pollen assemblages despite the fact that “riverine forests are represented by a
109 long range of floristic sub-types from nearly all parts of Ethiopia, and differ in composition according
110 to topographic and climatic factors, and the surrounding vegetation with which they more or less share
111 similar species” (Friis et al., 2010). The potential of pollen proxy to characterize different riparian
112 vegetation types, which typically occur as narrow and strip-like habitats along perennial and non-
113 perennial rivers below 1800 m above mean sea level (a.s.l.) and associated with lakes and springs, has
114 yet to be formally tested.

115 To solve these problems and improve our understanding of Hominini habitats, it is necessary
116 to develop a new approach to reconstructing vegetation from pollen in East Africa. This new method
117 must respect several elements. (1) It must be based on pollen scores without excluding aquatic plants
118 in order to be objective and reproducible. (2) It must be able to separating the local pollen signal from
119 the regional pollen signal in the East Africa Rift, and thus to distinguish as clearly as possible between
120 regional (climatically driven) and local (edaphically-driven) vegetation. (3) It must minimize the use of
121 relative abundance data and maximize the use of relative abundance scales or presence/absence data,
122 in order to minimize taphonomic biases in future comparisons between modern and fossil pollen data
123 to better assess past vegetation.

124 With this in mind, we developed a new method based on a pollen score: the vegetation score
125 of sample β for the vegetation group α ($V_{\alpha,\beta}$ score). To see if the $V_{\alpha,\beta}$ effectively separates different
126 regional and local vegetations, we used modern pollen assemblages. This modern pollen assemblages
127 includes new lowlands samples from the Awash Valley and the southern extremity of the East
128 Highlands that complement existing data. Here, we will see (1) which types of data (relative

abundance, relative abundance scale, and presence/absence) for the $V_{\alpha,\beta}$ score are most effective in discriminating these 24 regional and local vegetation types and subtypes. (2) Whether the vegetation discriminations performed with the $V_{\alpha,\beta}$ score are good enough to consider its use to assist in the reconstruction of paleo-vegetation from pollen data. (3) The environmental information that can be obtained with the $V_{\alpha,\beta}$ score in addition to vegetation.

2. Study area

This study focuses on the present-day vegetation and climate of the Horn of Africa, specifically in Ethiopia, for which pollen assemblages from surface soil/mud samples are available (**Figure 1**). In Ethiopia, topography, which is with the seasonal movement of the Intertropical Convergence Zone (ITCZ) one of the main factors influencing precipitation and precipitation patterns, is characterized by highlands generally between 1600 and 2600 m a.s.l. (up to 4500 m a.s.l.) of elevation on the western and eastern sides of the Rift valley. In Central Ethiopia, the floor of the Rift valley, where large lakes are located, lies at ca. 1650 m a.s.l., but falls down to -125 m below the sea level in the northeast, in the Awash Valley (Danakil depression). The Western Ethiopian highlands receive maximum rainfall (heavy rains coming from the west Atlantic front mainly from June to September, or almost all year round in the far west of the country), while the Eastern Ethiopian highlands receive monsoon precipitations coming from the Indian Ocean in two rainy seasons due to seasonal variation of the ITCZ (Diro et al., 2011). Rainfall is unimodal in the west of the Rift valley and bimodal everywhere else (Diro et al., 2009, 2008). Dry season, expressed here as the number of months with <30 mm rainfall lasts just three months in the western regions of Amhara and Benishangul-Gumuz Regional States, but is as long as 6 months in the Afar Region and Somali Regional State (Fick and Hijmans, 2017; Friis et al., 2010). In Ethiopia, temperature decreases with increasing elevation according to a thermal gradient of about -0.4°C/100 m. In the rift valley, mean annual temperature is about 30°-34°C in the northernmost area of the rift valley (Danakil Depression), and about 20°-23°C in the Great Rift Lakes Region. Mean annual temperature in Ethiopia is as low as 6-7°C above 4100 m a.s.l. where freezing is common.

Vegetation composition and physiognomy are largely influenced by precipitation and temperature. Friis et al. (2010) describe regional vegetation types and vegetation sub-types for Ethiopia based on dominant species, vegetation physiognomy, and environmental parameters (e.g., soil, climate, elevation). A georeferenced map of vegetation types and sub-types over Ethiopia and East Africa (with homogenized names and abbreviations) was produced under the Vegetation and Climate change in East Africa (VECEA) project (van Breugel et al., 2015), and has been used here. Vegetation distribution can be summarized as follows. Afroalpine vegetation (A) including Ericaceous zones (E) occur at high elevation (>3000 m a.s.l.) where mean annual temperature ranges between ca.

15° and 6°C (**Figure 2**). A complex of Afromontane forest with wooded grasslands and evergreen or semi-evergreen bushland and thicket (Fb/Wd) occurs below the Afroalpine A and E zones and covers most of the Western and Eastern Ethiopian highlands where mean annual temperature ranges is ca 22.5-12.5°C between ca 1800-3000 m a.s.l. (but upper and lower limits can vary). Afromontane rainforest (Fa) are restricted to the south-western Highlands (Illubator and Kefa regions), where mean annual temperature ranges between ca. 12.5° to 20°C, and mean annual precipitations are of ca 1800-2000 mm/yr. At low and mid-elevation (ca. 400-1800 m a.s.l.), *Combretum-Terminalia* woodland and wooded grassland (Wcd) (with small to moderate sized trees with large deciduous leaves) occur in the west where mean annual precipitation is ca. 600-2000 mm/yr, rainfall is unimodal with a dry season of 3 to 6 months, and mean annual temperature ranges from ca. 18°C to 28°C. At similar altitude (ca. 400-1800 m a.s.l.) and comparable mean annual temperature range (ca. 18°-30°C) but with lower annual precipitations (<1000 mm/yr) and a bimodal rainfall regime, the eastern and southern regions are covered with *Acacia-Commiphora* woodlands and bushlands (Bd). In the central part of the Rift Valley below 1800 m a.s.l., where mean annual temperature is ca. 15°-24°C, rainfall is also bimodal but higher (600-1000 mm/yr), the vegetation is less xerophytic and is described as Upland Acacia wooded grassland (We). Desert (D) and semi-desert grassland and shrubland (S) occur in northernmost area of Ethiopia rift valley and eastern Ethiopia below ca 400 m a.s.l., where mean annual temperature is ca. 25°-34°C. The mean annual precipitations is < 200 mm/yr for D and generally is 200-400 mm/yr for S but can but can be on a few S sites up to 1150 mm/yr. In this most arid land, small trees, shrubs and herbs may be succulent, geophytic or annual. We note that boundaries of main vegetation types match the bioclimatic boundaries except in south-western Ethiopia, where the *Acacia-Commiphora* woodland and bushland (Bd) (in the east) meets with *Combretum-Terminalia* woodland and wooded grassland (Wcd) (in the west) (Friis et al. 2010) (**Figure 2**). Edaphic vegetation associated with rivers, freshwater lakes and salty lakes was also studied in detail and mapped by Friis et al. (2010), who distinguished up to seven types and sub-types. They noted that riverine vegetation is quite similar to vegetation in the major type above 1800 m a.s.l., but shows very different floristic composition from the surrounding vegetation below 1800 m a.s.l. (Friis et al., 2010, p. 42). Riverine habitats below 1800 m a.s.l. may contain species found in forests under higher rainfall regimes at the same elevations. Floristic composition of riverine vegetation was estimated at 242 species. 26% of these (including 60 woody species) are exclusively or predominantly recorded in this habitat (Friis et al., 2010, p. 127).

3. Material and method

3.1. Modern pollen samples

Pollen names were homogenized according to Vincens et al. (2007) (**Table S1**). Pollen types difficult to differentiate in the modern samples and rarely differentiated in the fossil register were grouped together (e.g., *Nuxia/Dobera*). Taxonomic precision was limited at the subfamily level for Asteraceae. Spores were excluded from our analyses because it is not clear if they were consistently identified and counted by all analysts. *Prosopis*-type and *Prosopis*-type *africana*, which exhibit little morphological differences were also removed from the dataset after the calculation of relative abundance, because *Prosopis*-type is produced by the recently introduced and very invasive species *Prosopis chilensis*.

This study includes 283 modern pollen assemblages (**Table S2**) obtained from 207 soil samples and 76 mud samples from Ethiopia (251 assemblages), the Arabian Peninsula (17 assemblages from Yemen, Saudi Arabia, and Qatar) and Tanzania (15 assemblages) (**Figure 1** and **Table S3**). Out of these 283 assemblages, 182 assemblages were extracted from the APD (African Pollen Database, 2019), 93 assemblages were extracted from the literature, and 62 assemblages (22% of the dataset) are new. This dataset samples most of the climate domain of Ethiopia, except where annual temperature is above 30°C (cf., the Danakil Depression) (**Figure 2** and **Table S4**). The 15 samples from Tanzania were added to improve representation of spring-associated wetlands and forests (Barboni et al., 2019). The 17 samples from the Arabian Peninsula (Bonnefille and Riollet, 1988; El-Moslimany, 1983; Lézine et al., 1998) were included to represent very arid conditions (annual precipitation < 125 mm/yr) that are not currently found in the modern Ethiopian Rift Valley.

The 62 new assemblages were collected in 2009 by D. Barboni and R. Bonnefille (Ethiopian samples DB09) and in 2017 by D. Barboni, B. Bourel and J.-C. Mazur (Ethiopian samples BB17). Our new samples fill several gaps. First, they document the driest range of the *Acacia-Commiphora* deciduous bushland and thicket (Bd), which occurs under 400-600 mm/yr precipitation within the 20°-30°C mean annual temperature zone (**Figure 2**). Second, they document the hottest and driest range of occurrence of the Complex of Afromontane forest with wooded grasslands and evergreen or semi-evergreen bushland and thicket (Fb/Wd) where mean annual temperature is 18-21°C and mean annual precipitation 700-800 mm/yr. And third, they document the lowland azonal vegetation fed by springs or groundwater. Despite these additions, sampling of the climate spaces could still be improved for Afroalpine vegetation and Ericaceous belt (A.E), Dry *Combretum* wooded grassland (Wcd), Complex of Afromontane forest with wooded grasslands and evergreen or semi-evergreen bushland and thicket (Fb/Wd), and Semi-desert grassland and shrubland (S) and Desert (D).

3.2. Geopositioning and assignment of environmental data

To make local vegetation reconstructions possible, geolocation of some pollen samples required to be reworked. This was especially true for samples that were collected at a time when GPS was not available or not very accurate (Bonnefille, 2018). To do so, we used all material available: authors' notes, descriptions of vegetation at sampling sites, mission preparation documents, road maps, elevation data obtained with altimeters, and mission reports, in addition to information provided in the original publications. The data we used are from (Barboni et al., 2019; Bonnefille et al., 1987, 2004, 1993; Bonnefille, 1972; Bonnefille and Buchet, 1986; Bonnefille and Riollet, 1988; Buchet, 1982; El-Moslimany, 1983; Garnier et al., 2013; Lézine, 1981; Lézine et al., 1998; Mohammed, 1992; Mohammed et al., 2007; Mohammed and Bonnefille, 2002) (column 6 and 9 in **Table S5**). For example, in the Hadar area (north of the Awash valley), samples E010 to E026 are archived under the same geographical coordinates in the APD, although author's notes and a map in the original (Bonnefille et al., 1987) show that they were collected at different locations along the Awash River, and not some 10 to 16 km away in the xerophytic bushland. For the purpose of our study, their geographical coordinates were therefore corrected. In total, we corrected the geographical coordinates for 68% of the "old" pollen samples (150/221 pollen samples) with an average correction of 9.7 km (column 9 in **Table S3**).

Using a geographic information system software (GIS QGIS, v3.2.2), we assigned to all 283 sites (with corrected coordinates) 12 soil parameters (SoilGrids, 2000-2015, 0.25×0.25 km, Hengl et al., 2017), 32 climatic parameters (WorldClim2, 1970-2000, 0.93×0.93 km, Fick and Hijmans, 2017), five aridity parameters (Global High-Resolution Soil-Water Balance, 1950-2000, 0.93×0.93 km, Trabucco and Zomer, 2019), Net Primary Productivity (NPP) (MODIS MOD17A3, 2000-2014, 0.93×0.93 km, Zhao et al., 2005), and percent of tree cover (Global Maps - Vegetation, year 2008, 0.46×0.46 km, Kobayashi et al., 2017) (**Table S4** *Erreur ! Source du renvoi introuvable.*). Satellite percent tree cover values for the 2000-2014 period were sometimes very different from the authors' notes at the time of sampling (some 40 years ago in some cases). Coherence between modern satellite tree cover values and authors' notes was therefore checked for every site, and when deforestation was observable on Google Earth images ("time travel" function) (column 10 in **Table S5**), then tree cover values of neighboring, unaffected or less affected areas within 1-kilometer radius of the sampling point were considered. Similarly, we corrected tree cover values when a slight discrepancy (<250 m) was observed between Google Earth images and the tree cover grid. In total, tree cover values were corrected for 116 sites (over 283). GIS was also used to attribute to each surface sample (with corrected geographical coordinates) a potential vegetation type according to the VECEA project (van Breugel et al., 2015) for African samples and according to vegetation map of Frey and Kürschner (1989) for Arabian samples

(column 7 in **Table S5**). We gave priority to the collector's notes and vegetation observed with Google Earth to correct the assigned potential vegetation accordingly (column 6 and 8 in **Table S5**).

3.3. Pre-processing of data

To define a threshold pollen percentage optimized to our study area, data transformations and analyses were carried out using R (R Development Core Team, 2018). Pollen type relative abundance (without spores and undetermined pollen) were calculated for each of the 283 sites and compiled in double entry matrix of n pollen taxa \times m sites (data in **Table S2**). The double entry matrix was then transformed into a column vector \vec{V} . Classified in ascending order, values of \vec{V} show a break at 0.2% in the evolution of pollen percentages. We interpret this break as the signal of allochthonous taxa, which occurrence in pollen assemblages is erratic. Before further processing of the data, all \vec{V} entries with values $< 0.2\%$ were therefore excluded from the column vector to reduce the noise associated with the presence of allochthonous pollen (**Figure 3**). In other words, we chosen a threshold pollen percentage of 0.2% for this study.

3.4. Determination of the relative abundance scale

To define the relative abundance scale optimized to our study area, after removal of pollen percentage $< 0.2\%$ in each pollen sample, we used the Vegan function `cascadeKM()` (Vegan v2.5-2: Oksanen et al., 2010) that applies the k -means partition comparison method and the simple structure index (ssi) criterion to identify the minimum number of classes of relative abundances that keep a maximum of information (Dolnicar et al., 1999). The larger the ssi, the more significant is the partitioning (Borcard et al., 2018). Based on this index, 2 to 6 classes were optimal (**Figure S1**). We retained six classes. To define the classes we used the Fisher-Jenks method of the Cartography function `getBreaks()` (Cartography v2.2.0: Giraud and Lambert, 2017): relative abundance class 1 $\in]0.2\% ; 3\%]$, relative abundance class 2 $\in]3\% ; 11\%]$, relative abundance class 3 $\in]11\% ; 23\%]$, relative abundance class 4 $\in]23\% ; 38\%]$, relative abundance class 5 $\in]38\% ; 60\%]$ and relative abundance class 6 $\in]60\% ; 100\%]$. A 7th class was added for the threshold pollen percentage of 0.2%: relative abundance classes $0 \in]0\% ; 0.2\%]$.

3.5. Inference of vegetation groups

The pollen assemblages were grouped into vegetation groups from 6 observations: (1) the partition of the pollen assemblages according to the pollen data using a k -means partitioning constrained by a hierarchical classification with the function `vegdist()`, function `cascadeKM()` and function `hclust()` of Vegan v2.5-2 and Stats v3.5.1 R package (Oksanen et al., 2010; R Core Team, 2010), (2) the sample type (mud or soil), (3) the potential natural vegetation (Frey and Kürschner, 1989; Friis

et al., 2010; van Breugel et al., 2015), (4) the vegetation described at the sampling sites in the original publications (Barboni et al., 2019; Bonnefille et al., 1987, 1993; Bonnefille, 1972; Bonnefille et al., 2004; Bonnefille and Buchet, 1986; Bonnefille and Riollet, 1988; El-Moslimany, 1983; Garnier et al., 2013; Lézine, 1981; Lézine et al., 1998; Mohammed, 1992; Mohammed et al., 2007; Mohammed and Bonnefille, 2002), (5) the vegetation described at the sampling sites in mission reports, (6) and some hydrological studies providing information on the edaphic context at the sampling sites (e.g., spring) (Ayenew et al., 2008; Cuthbert et al., 2017) (**Table S5**). Of the 283 pollen assemblages, 21 were removed: 7% of the dataset. Within these 7%, 12 samples are from anthropic sites (e.g., DB09-04, Karasoditi coffee forest) and 9 samples are from very poorly sampled vegetation (e.g., BB17-41, pioneer vegetation growing in small patches on 200 years-old lava flow) (**Table S5**). In the end, the pollen assemblages were distributed in 24 vegetations types or subtypes, including nine local riparian and azonal vegetation. (**Table 2**). These 24 groups are divided into two sets: 18 groups composed of pollen samples from non-dynamic depositional environments (soils, muds of lake edges and soils/muds of swamps) and 6 groups composed of pollen samples from dynamic depositional environments (muds of river beds or edges).

3.6. Environmental parameters for inferred vegetation groups

To see which environmental, climatic, and/or edaphic variables would best relate to the 24 vegetation groups (**Table 2**), we ran a Random Forest model of 10000 iterations with the function Random Forest () of the Random Forest v4.6 R package. We used 10 variables measuring temperature, 10 for precipitation, 5 for aridity, 11 for soil, tree cover percentage, and net primary production (NPP) (columns 8-26 ; 39-44 ; 46 ; 48-59, **Table S4**). We then applied the function varImpPlot () of the same R package with our model to see which variables best explain the distribution of vegetation with the function.

3.7. Vegetation score ($V_{\alpha,\beta}$ score)

Pollen taxa affinities for a vegetation group, were estimated with the Value Test (VT) (Lebart, 2000). VT compares for one variable (pollen type) the values of descriptive statistic indicators (means, variances, and group size) computed on all groups (the full dataset: all vegetation groups tested) and on a subsample related to one group (one vegetation group). The VT of a pollen taxon for a vegetation group is calculated as follows:

$$VT = \frac{\mu_g - \mu}{\sqrt{\frac{n - n_g}{n - 1} \times \frac{\sigma^2}{n_g}}}$$

where μ_g is the mean relative abundance of the pollen taxon within the vegetation group, μ the mean of the pollen type (in relative abundance, relative abundance scale, or presence/absence) in the whole dataset, n_g the size of the group (number of samples in the vegetation group), n the size of the whole dataset (total number of samples), and σ^2 the variance associated with mean of the pollen type (in relative abundance, relative abundance scale, or presence/absence) in all vegetation groups tested (the whole dataset) (Lebart et al., 1995). The VT of a pollen type for a vegetation group is a simple measure of similarity between the pollen type and the vegetation group. Similarity is indicated by positive VT, and dissimilarity by negative VT (**Table S6**).

Using VT values of the pollen taxa for the vegetation groups, we then developed the calculation of a score to measure the affinity of the pollen samples for vegetation groups. The affinity of a pollen assemblage to a vegetation group is calculated as follows:

$$A_{\alpha,\beta} = \frac{\sum_{i=0}^9 \left(\sum_{VT\% \in \left[\frac{10i \times VT_{max}}{100}, \frac{10(i+1) \times VT_{max}}{100} \right]} P \times i \right)}{\sum_{i=0}^9 \left(\sum_{VT\% \in \left[\frac{10i \times VT_{max}}{100}, \frac{10(i+1) \times VT_{max}}{100} \right]} N \times i \right)} - \frac{\sum_{i=0}^9 \left(\sum_{VT\% \in \left[\frac{10(i+1) \times VT_{min}}{100}, \frac{10i \times VT_{min}}{100} \right]} P \times i \right)}{\sum_{i=0}^9 \left(\sum_{VT\% \in \left[\frac{10(i+1) \times VT_{min}}{100}, \frac{10i \times VT_{min}}{100} \right]} N \times i \right)}$$

where $A_{\alpha,\beta}$ is affinity of sample β for the vegetation group α , $P_{VT\% \in [x;y]}$ is the relative abundance scale of a pollen type in sample β with VT percentage between $[x; y]$ for the vegetation group α , $N_{VT\% \in [x;y]}$ is the number of pollen types in the sample β with VT percentage between $[x; y]$ for the km-group α . The VT percentage of a pollen type for the vegetation group α is $[VT_{\text{pollen type}} \times 100] / VT_{max}$, VT_{max} is the most positive VT of pollen type for the vegetation group α , and VT_{min} is more negative VT of pollen type for the vegetation group α . In this formula, the first member of the formula (left) is the weighted mean of the relative abundance scale of all pollen types in sample β with a positive VT for the vegetation group α . The weight of each relative abundance scale is based on the VT percentage of its pollen type such as: $VT\% \in [10n; 10(i+1)]$ for weight $= i+1$. The second member of the formula (right) does the same thing but for negative VT. $A_{\alpha,\beta}$ therefore represents the weighted mean of pollen of sample β with a similarity for the vegetation group α minus the weighted mean of pollen of sample β with a dissimilarity for the km-group α .

In order to have a Vegetation score ($V_{\alpha,\beta}$ score), the standard score of $A_{\alpha,\beta}$ for $A_{all,\beta}$ (all A scores of sample β for all vegetation groups tested) is expressed as follows:

$$V_{\alpha,\beta} = \frac{A_{\alpha,\beta} - \mu_{all,\beta}}{\sigma_{all,\beta}}$$

where $V_{\alpha,\beta}$ is the vegetation score of sample β for the vegetation group α , $A_{\alpha,\beta}$ is affinity of pollen sample β for the vegetation group α , $\mu_{all,\beta}$ is the mean of all A scores of sample β for all vegetation groups tested, and $\sigma_{all,\beta}$ is the standard deviation of $A_{all,\beta}$ (all A scores of sample β for all vegetation groups tested). This last step was added in anticipation of using this method on fossil data. Indeed, for similar vegetation, fossil pollen samples may have a lower taxonomic diversity than their modern analogues due to taphonomic bias. In this particular case, the A-scores of fossils will be lower than the A-scores of their modern analogues. The numeric centering and scaling used in V-scores minimize this effect.

3.8. Significant and non-significant score ranges for $V_{\alpha,\beta}$ score

For a selected vegetation group in all vegetation groups tested, we calculate the $V_{\alpha,\beta}$ score values of this selected vegetation group (α) for all pollen assemblages of all vegetation groups tested: $V_{\alpha,\beta \in all \text{ tested groups}}$. The $V_{\alpha,\beta}$ score values of the pollen assemblages classified in this vegetation group ($V_{\alpha,\beta \in \alpha}$) define the range of significant values for $V_{\alpha,\beta}$ score of this selected vegetation group. The $V_{\alpha,\beta}$ score values of the pollen assemblages that are not classified in this selected vegetation group ($V_{\alpha,\beta \notin \alpha}$) define the range of non-significant values for $V_{\alpha,\beta}$ score of this selected vegetation group.

4. Results

4.1. $V_{\alpha,\beta}$ score applied to the vegetation groups

The $V_{\alpha,\beta}$ score was applied to pollen assemblages from the 24 vegetation groups but by grouping the six groups linked to dynamic depositional environments into two sets: the S-, Bd- and, We-river grouped as We/Bd/S-river and the Fb/Wd-river, Wcd-river 1, and Wcd-river 2 grouped as Wcd/Fb/Wd-river (**Figure 4**). We have chosen this grouping because the pollen assemblages of these 6 groups linked to dynamic depositional environments include a high proportion of exogenous pollen (pollen from vegetation groups not linked to dynamic depositional environments) which biases the determination of pollen taxa affinities with the VT score.

For pollen data in relative abundance, the $V_{\alpha,\beta}$ score of 18 vegetation groups (all groups except the Fb/Wd-low/middle and Fa-w) have overlapping significant and non-significant values (**Figure 4**). $V_{\alpha,\beta}$ score of 10 vegetation show large overlap of significant values by non-significant values ($\geq 50\%$). $V_{\alpha,\beta}$ score of S-xs ($V_{S-xs,\beta}$) has non-significant values that completely overlap with significant values. Depending on the vegetation (α) of the $V_{\alpha,\beta}$ score, the pollen assemblages that cause overlaps with significant values come from one to 10 different vegetation groups (on average 4). The taxonomic and geographical proximity of the vegetation groups explain some overlaps. This is the case for the close

$V_{Bd-wb\ 2,\beta}$ score values between pollen assemblages from vegetation subtypes Bd-wb-1, -2, and -3 (e.g., overlap of $V_{Bd-wb\ 2,\beta \in Bd-wb\ 2}$ linked to $V_{Bd-wb\ 2,\beta \in (Bd-wb\ 1 \cup Bd-wb\ 3)}$). But 25% of these overlaps, such as the overlap of V score values of the pollen assemblages from S-xs and Fa-upper (e.g., overlap of $V_{S-xs,\beta \in S-xs}$ related $V_{S-xs,\beta \in Fa-upper}$), cannot be explained. The use of the $V_{\alpha,\beta}$ score with pollen data in relative abundance seems to discriminate 10 vegetation in an acceptable way but for the other 10 vegetation the discrimination is not good (overlap of significant values by non-significant values $\geq 50\%$).

For pollen data in relative abundance scale, the $V_{\alpha,\beta}$ score of only four vegetation groups (We/Bd/S-river, Wcd/Fd/Wd-river, Bd-wb 2, and Bd-wb 3) have overlapping significant and non-significant values (**Figure 4**). These are slight overlaps of significant scores by non-significant scores ($< 50\%$). Depending on the vegetation (α) of the $V_{\alpha,\beta}$ score, pollen assemblages that cause overlaps with significant values come from one to two different vegetation groups (on average 1). The taxonomic and geographical proximity of the vegetation groups explain all overlaps. This is the case for the close $V_{Bd-wb\ 2,\beta}$ score values between pollen assemblages from vegetation subtypes Bd-wb-1 and-3 (e.g., overlap of $V_{Bd-wb\ 3,\beta \in Bd-wb\ 3}$ linked to $V_{Bd-wb\ 3,\beta \in Bd-wb\ 1}$). In contrast to the relative abundance pollen data,, significant values of $V_{Wcd/Fb/Wd-river,\beta}$ score overlap only with pollen assemblages from We/Bd/S-river group and the significant values of $V_{We/Bd/S-river,\ all}$ score overlap only with the pollen assemblages from Wcd/Fd/Wd-river group. Despite these overlaps, all pollen assemblages with a significant value for $V_{We/Bd/S-river,\beta}$ or/and $V_{Wcd/Fb/Wd-river,\beta}$ scores can only be from We/Bd/S-river or Wcd/Fd/Wd-river groups, and by extension they are necessarily from the set of dynamic depositional environments (the S-, Bd-, We-, Fb/Wd-, and the two Wcd-river vegetation group). The use of the $V_{\alpha,\beta}$ score with pollen data in relative abundance scale is therefore appropriate to discriminated vegetation linked with non-dynamic depositional environments and for isolating the pollen assemblages from dynamic depositional environments.

For pollen data in presence/absence, $V_{\alpha,\beta}$ scores of four vegetation (We/Bd/S-river, Bd-spring1, Bd-wb 1, and Fa-upper) show large overlap of significant values by non-significant values ($\geq 50\%$) (**Figure 4**). $V_{\alpha,\beta}$ score of 6 vegetation (Wcd/Fb/Wd-river, Bd-wb 2, Bd-wb 3, Bd-rtf, Fb/Wd-upper, and Fa-sc 1) are slight overlaps of significant scores by non-significant scores ($< 50\%$). Depending on the vegetation (α) of the $V_{\alpha,\beta}$ score, the pollen assemblages that cause overlaps with significant values come from one to 11 different vegetation groups (on average 3). Except for $V_{We/Bd/S-river,\beta}$, the taxonomic and geographical proximity of the vegetation groups explain the overlaps. As a result, to discriminate vegetation, $V_{\alpha,\beta}$ score with pollen data in presence/absence is more efficient than $V_{\alpha,\beta}$ score with pollen data in relative abundance scale but less efficient than the $V_{\alpha,\beta}$ score with pollen data in relative abundance. Moreover, as $V_{\alpha,\beta}$ score with pollen data in relative abundance and unlike $V_{\alpha,\beta}$ score with

pollen data in relative abundance, $V_{\alpha,\beta}$ score with pollen data in presence/absence does not efficiently isolate the pollen assemblages from dynamic depositional environments.

4.2. $V_{\alpha,\beta}$ score applied only to the vegetation groups linked to dynamic depositional environments

As seen in the previous section, in some cases, it is possible to isolate with $V_{\alpha,\beta}$ score the set of vegetation groups related to dynamic depositional environments (the S-, Bd-, We-, Fb/Wd-, and the two Wcd-river vegetation group). The question, then, is whether it is possible to further improve the discrimination of vegetation groups with the $V_{\alpha,\beta}$ score once this set is isolated. We present here the results of the $V_{\alpha,\beta}$ score applied only to the six vegetation groups linked to dynamic depositional environments: S-, Bd-, We-, Fb/Wd-, and both Wcd- vegetation groups.

For pollen data in relative abundance, the $V_{\alpha,\beta}$ score of all vegetation groups have overlapping significant and non-significant values (**Figure 5**). Depending on the vegetation (α) of the $V_{\alpha,\beta}$ score, the pollen assemblages that cause overlaps with significant values come from one to five different vegetation groups (on average 3). $V_{\alpha,\beta}$ score of four vegetation groups show large overlap of significant values by non-significant values ($\geq 50\%$). Using the $V_{\alpha,\beta}$ score with pollen data in relative abundance discriminates acceptably only two of the six vegetation groups.

For pollen data in relative abundance scale, the $V_{\alpha,\beta}$ score of only two vegetation groups have overlapping significant and non-significant values (**Figure 5**). Depending on the vegetation (α) of the $V_{\alpha,\beta}$ score, the pollen assemblages that cause overlaps with significant values come from one to three different vegetation groups (on average 2). $V_{\alpha,\beta}$ score of only one vegetation group show large overlap of significant values by non-significant values ($\geq 50\%$). Using the $V_{\alpha,\beta}$ score with pollen data in relative abundance scale discriminates acceptably five of the six vegetation groups. This is much better than with the data in relative abundance.

For pollen data in presence/absence, there is no overlap between the significant and non-significant values of $V_{\alpha,\beta}$ score (**Figure 5**). The presence/absence data allow us to better discriminate between vegetation groups than the relative abundance data.

4.3. Relation between vegetation groups and environmental parameters

The Random Forest model was able to recreate the 24 vegetation groups (**Table 2**) with a success rate of 80%. This indicates that 80% of our dataset has a pollen signal that provides information about one or several environmental parameters. Three parameters emerge as most correlated with vegetation groups inferred from their pollen signature: the tree cover percentage, mean temperature

of warmest quarter, and mean precipitation of driest quarter (**Figure S2**). Globally, the climatic parameters and the NPP are more correlated with the vegetation groups than the edaphic parameters.

For the two climatic parameters that best explain our vegetation groups (mean temperature of warmest quarter and mean precipitation of driest quarter) (**Figure 6**), it appears that the 24 groups have well-defined and very diverse climatic spaces. The climatic spaces of these vegetation groups range from less than 10°C to more than 35°C for the mean temperature of warmest and from almost 0 mm to more than 150 mm for the mean precipitation of driest quarter. These 24 vegetation groups are distinguished by the two main climatic parameters (e.g., Bd-wb1 vs. Bd-wb3) or by edaphic parameters (e.g., Bd-wb 1 vs S-river). Sometimes it is secondary climatic parameters that explain the differences in vegetation groups. This is the case for the mean annual precipitation for Bd-wb 3 and Wcd, respectively from 446 mm to 919 mm and from 1013 mm to 1336 mm (column 26 in **Table S4**).

Our vegetation groups correspond to distinct physiognomic types of vegetation with different tree cover (**Figure 7**). With a 95% confidence interval, the estimated intervals of tree cover percentage are different and complementary for the 24 vegetation groups (e.g., Fa-sc 1 and Fa-sc 2). These estimated tree covers range from 0 to 82% depending on the vegetation group.

4.4. Morphologies and vegetation strip of rivers

This study differentiates several types of azonal and local vegetation, including seven river-related vegetation (S-river, Bd-river, Bd-rtf, We-river, Fb/Wd-river, Wcd-river 1, and Wcd-river2). These seven vegetations can each be associated with the morphology of the river (meandering or straight) and with a width of vegetation strip along the river banks (**Table 2**). For example, it is possible to say that a Bd-river pollen-inferred vegetation implies the presence of a meandriform and perennial river with a 50-100 m wide strip of open forest along its banks. The combination of these two parameters is specific to each group. Details are given for the seven vegetation groups in **Table 2**.

4.5. Most characteristic pollen types

For the sets of non-dynamic depositional environments (18 vegetation groups: **Table 2**), if we look at the most characteristic pollen types of each of these vegetation (**Table 3**) on the base of T-values with data in relative abundance scale (**Table S6**), we note that arboreal pollen types are more often used than non-arboreal pollen types for vegetation characterization. This is true for all vegetation groups except those related to the most open and xerophilic vegetation (S-river, S-xs, S-xds, D) or to Afromontane vegetation (E.A-e). For some vegetation types, such as Fa-w, the pollen taxa markers correspond to botanical markers identified by botanists (e.g., *Macaranga* or *Pouteria*) (Friis et al., 2010). Similarly, characteristic pollen types for riparian and azonal vegetation types were easily

identified as *Tamarix* for Bd-rft and *Hyphaene* for Bd-spring 1 et 2 in agreement with the relative abundance of these trees in riparian *Tamarix* forests and groundwater-fed woodlands. Overall, 10% to 40% of the pollen taxa identified as the most characteristic correspond to botanical markers identified by botanists.

For the sets of non-dynamic depositional environments (6 vegetation groups: **Table 2**), if we look at the most characteristic pollen types of each vegetation (**Table 3**) on the base of T-values with data in presence/absence (**Table S6**), correlations with the surrounding vegetation are not obvious, which is not surprising because the pollen spectra integrate pollen grains likely coming from vegetation much further upstream. Yet, six different river vegetation types were identified by our approach, which shows that riparian formations carry distinct pollen signals according to the vegetation zones crossed by the river .

5. Discussion

5.1. The distinction of vegetation

The $V_{\alpha,\beta}$ score presented in this paper effectively distinguishes 24 vegetation groups distributed in eight vegetation types (**Table 2**). Of these 24 vegetation types, 22 are subtypes of the regional vegetation, and of these 22, nine are local riparian and azonal vegetation. To be able to do this, an initial analysis must be carried out with the pollen data in relative abundance scale. This is to discriminate vegetation linked with non-dynamic depositional environments and for isolating the pollen assemblages from dynamic depositional environments (pollen assemblage from S-, Bd-, We-, Fb/Wd-, and the two Wcd-river vegetation group). A second analysis of pollen assemblages identified as pollen assemblages from dynamic depositional environments to specify their vegetation: S-river, Bd-river, We-river, Fb/Wd-river, Wcd-river 1, or Wcd-river 2. This second operation can be carried out either with presence/absence data (preferably) or in relative abundance scale.

This is the first time that pollen score has been successfully developed to discriminate vegetation with a spatial resolution lower than the regional scale. In East Africa, vegetation discriminations using pollen score calculations on modern pollen assemblages (and by extension on fossil pollen assemblages) were limited to the supra-regional or regional scale for Ethiopia, Kenya, Tanzania and Uganda. (Bonnefille et al., 2004; Vincens et al., 2006b). This is also the case for Central and West Africa (Hély et al., 2006; Izumi and Lézine, 2016; Lebamba et al., 2009). The difficulty of separating the local and regional pollen signal has been known for a long time (Bonnefille, 1984). Until now, no solution to this problem had been found, either by traditional palynological approaches or by calculation methods.

We also show that it is possible to distinguish nine azonal and riparian vegetation very well via pollen assemblages from dynamic or non-dynamic environments. These are also new results because these vegetation are generally neglected in the palynological studies of our study area (Bonnefille et al., 2004; Hély et al., 2006; Izumi and Lézine, 2016; Lebamba et al., 2009; Vincens et al., 2006b), and aquatic taxa are usually removed from the analyses as originally proposed by Jolly et al. (1998). Our results are a step forward for future application in the study of Hominini sites in East Africa, because they are very frequently associated with riparian or spring contexts (Barboni et al., 2019; Behrensmeyer and Reed, 2013; Campisano and Feibel, 2008; Levin, 2015). The reconstructions of these local and azonal environments where the Hominini fossils are located, are not clearly defined yet by palynological or faunal studies, which normally define them as "swamp", "wetland" or "floodplain" (Bonnefille et al., 1987, 2004; Bonnefille, 2010; Reed, 2008).

We have not been able to separate the Afroalpine vegetation (A) and Ericaceous belt (E) pollen signal. The samples of these two vegetation are in the A.E-e vegetation sub-type. An explanation is that all samples of A are in the Sanetti plateau (Ethiopia) and correspond to a particular area with isolated *Erica* shrubs (Mohammed et al., 2007). Miehe and Miehe (1994) suggested that this area represents the potential extent of *Erica*-dominated vegetation in the absence of natural or anthropic fires, hence our difficulty in the separation of A and E samples. Another difficulty is at the lower limit of the Fa where Friis et al. (2010) observe a "Moist evergreen Afromontane forest, bushland, woodland and wooded grassland". Although several Fa-sc 1, Fa-sc 2, and Fa-w samples are available in this area, our analyses did not reveal any specific pollen composition for this "Moist evergreen Afromontane forest, bushland, woodland and wooded grassland". The potential samples related to this vegetation are mainly in 3 sample sets: E201 to E209, E210-E235 and E268 to E282. The first set consists of unpublished data in APD and for the third set the authors do not mention this sub-type of vegetation (Mohammed, 1992). For the second set, the palynologists refer to this vegetation as "Intermediate forest" (cf., next paragraph) (Bonnefille et al., 1993). With their 25 pollen samples from South West Ethiopia, they show that the "Moist evergreen Afromontane forest, bushland, woodland and wooded grassland" is well distinguished from the other samples of Fa. Our results suggest that this distinction may not be valid on a larger scale. For Fa-w, Bonnefille et al. (1993) indicate that the Fa-w samples collected near Gore (western Ethiopia) come from the "Intermediate forest" (corresponds to the Transitional rain forest type of Friis et al. (2010)). However, Friis et al. (2010) describe the vegetation at the location of the sampling sites of Fa-w samples (near Gore) as "Primary or mature secondary moist evergreen Afromontane forest", a sub-type of Fa type. Normally, the elevation of Fa-w samples (lowest sample: 1020 m a.s.l.) is too low for Fa (low limit of Fa: 1550-1450 m a.s.l.). But in this part of Western Ethiopia, the "Primary or mature secondary moist evergreen Afromontane forest" has the

particularity of occurring below the calculated lower elevation limit of the Fa, and grows at the same altitude as the Transitional rain forest (Friis et al., 2010). In addition, the present plants described by Bonnefille et al. (1993) corresponds to vegetation of "Primary or mature secondary moist evergreen Afromontane forest" (Friis et al., 2010). This is why we preferred to define Fa-w as Secondary moist evergreen Afromontane forest rather than as Transitional rainforest

Finally, our results show that the pollen type *Hyphaene*-type is a main marker of spring as suggested by Barboni et al. (2019). This is an important step because it is supposed that springs have an important role in the hominin evolution in East Africa (Ashley et al., 2009; Cuthbert et al., 2017; Cuthbert and Ashley, 2014). It is interesting to note that there is clear distinction between Ethiopian (Bd-spring 1) and Tanzanian (Bd-spring 2) springs related to the regional differences in vegetation, groundwater (alkaline water in Ethiopia and fresh water in Tanzania: personal observation) and mean annual temperature.

5.2. The extrapolated information from the inferred vegetation

In the East African Rift, the variations in river vegetation are not only linked to the variations in climatic conditions and surrounding vegetation (Friis et al., 2010), but also to the variations in river morphology (Carr, 1998). This relationship allowed us to associate these seven types of river vegetation with river morphologies (**Table 2**). One of the most interesting cases in this study concerns the pollen assemblages from Wcd where the subtypes Wcd-river 1 and Wcd-river 2 are associated respectively with straight rivers and meandering rivers. In the same way, it is possible to associate the seven river-related vegetation groups with the width of the vegetative strip along the rivers (**Table 2**). This agrees with the observations of Carr (1998). Showing that it is possible to do this type of discrimination with pollen scores is interesting because the structure of the river and the width of the vegetation strip are important elements as they influence the type of fauna present (Geheb and Abebe, 2003).

Like previous studies in East Africa (Bonnefille et al., 2004; Peyron et al., 2000) or in other parts of the world (Barboni and Bonnefille, 2001), our vegetation groups can be linked to several parameters, mainly climatic. It is the mean temperature of warmest quarter and the mean precipitation of driest quarter are the climate parameters that best explain our vegetation groups. This is not surprising because in our study area, the limiting parameters for vegetation are high temperatures and low precipitation (Friis et al., 2010). The new contributions of this study are the ability to strongly associate our 24 vegetation groups, and by extension our pollen assemblages, with ranges of percent tree cover. We show that pollen can be a strong proxy for tree cover and that they can help to quantify it. Pollen-

inferred tree cover will be interesting to compare with paleo-tree cover estimates based on $\delta^{13}\text{C}$ at hominin sites (e.g., Cerling et al., 2011).

5.3. Importance of data type

With $V_{\alpha,\beta}$ score, the relative abundance scale data allowed a very good discrimination of vegetation related to non-dynamic depositional environments and effectively isolated the set of vegetation related to dynamic depositional environments (**Figure 5**). This was not possible with the relative abundance data and was partially possible with the presence/absence data. The reason for this result is that for these depositional environments, the abundance scale minimizes the magnitude of variations in the pollen type relative abundance within the vegetation group while avoiding the loss of information of variation between vegetation groups (Borcard et al., 2018). The relative abundance scale data minimizes also the problems of over-representation (e.g., Cyperaceae, Poaceae) or under-representation (e.g., arboreal pollen) of pollen types and has the same role as the square root (Prentice et al., 1996; Vincens et al., 2006b), but is more relevant because our relative abundance scale are specifically calculated for the pollen counts of East Africa.

For all 6 vegetation groups related to dynamic depositional environments, discrimination is poor with relative abundance data, good with relative abundance scale data and very good with presence/absence data. (**Figure 6**). We assumed that this was due to the depositional environments, which created very large the magnitude of variations in the pollen type abundance (relative or scaled) within the vegetation group, caused by taphonomic bias (transport and weathering) and by large influx of pollen from upstream vegetation. The variation in abundance of pollen types within each vegetation group then becomes so large that these variations between vegetation groups loses all informative character. This is why the use of presence/absence is most effective.

We emphasize here that, contrary to what one might think, discrimination of vegetation from pollen is more effective when the data are in presence/absence or abundance scale than when they are in relative abundance. It is also important to match the type of data to the deposition environment. These are important results because currently, in our study area, pollen studies tend to focus on relative abundance data and analyze pollen assemblages from different depositional environments in the same way (Bonnefille et al., 2004; Peyron et al., 2000; Saylor et al., 2019). Finally, for future use on fossil data, the use of the abundance scale and the presence/absence makes more sense in our study areas where taphonomic biases are assumed to be important (Bonnefille, 1984; El-Moslimany, 1983).

5.4. A more objective approach

Some authors such as Izumi and Lézine (2016) have used thresholds for Poaceae that can be compared to our relative abundance classes, but these thresholds are defined subjectively for each pollen type. We have seen that it is possible to overcome this by using relative abundance scale defined by statistical analysis, or simple observations such as presence/absence. Similarly, the choice to attribute a pollen taxon to a vegetation type is often arbitrary, especially for herbaceous species (cf., Poaceae in Izumi and Lézine, 2016). This choice is based on botanical observations (Bonnefille, 1984), however, the statistical analyses of this study show that the most characteristic pollen types of a vegetation are not always those associated with the plants used by botanists to describe this vegetation (**Table 3**). We have shown here that it is possible to solve these problems of vegetation attributions types by using statistical analyses to define the weight of each pollen type for each vegetation.

To reduce the noise due to occasional long-transport pollen grains, the current value used in East Africa is 0.5% and it is based on Jolly et al. (1998) and Prentice and Webb III (1998). But, these two studies transpose the Prentice et al. (1996) value used for Europe without justifying this transposition to Africa. Based on our graphical analyses (**Figure 3**), we believe that a threshold of 0.2% is more appropriate for Africa. Izumi and Lézine (2016) also suggest this with empirical observations. This changes a lot for pollen score calculations in Africa. For our study, for example, using a threshold of 0.2% removes only 9.5% of total pollen occurrences compared to 43.5% for a threshold of 0.5% (**Figure 3**).

6. Conclusions

Using the $V_{\alpha,\beta}$ score presented in this paper and based on a dataset of 283 modern pollen assemblages, we show that it is possible to discriminate 24 vegetation groups distributed in eight regional vegetation types and including 22 subtypes. This is a solution to the difficulties of separating the local and regional pollen signal for the East African Rift. This highlights the possibility of using pollen to inform vegetation at a scale smaller than the region. It was shown that riparian vegetation in north part of East African Rift carry distinct pollen and environmental signals, which was largely ignored before. Our new approach allows identifying nine riparian vegetation (two of springs and seven of rivers) which not only relate to regional vegetation composition and climate, but also to the type of wet edaphic conditions.

Except for the removal of non-native taxa from the analysis, it was possible to develop a functional and standardized approach, without preconditions on the pollen types to be considered or

the importance to be attributed to the different taxa. As in previous studies, it was possible to link the pollen signal of modern assemblages to climatic parameters. The ability to quantitatively define tree cover via the pollen signal was also highlighted, which is new. Also in the novelty, the pollen signal has been associated with the river morphology (meandering or straight) and type of vegetation strip along the river banks (notably the width).

Finally, this study highlighted several elements for optimizing palynological analyses in the East African Rift. It was shown that for this study area, the use of the 0.5% threshold was not optimal and that a 0.2% threshold was preferable. Variations in relative abundance of pollen taxa within the vegetation group was found to be a limiting parameter for vegetation discrimination below the regional scale using pollen data in relative abundance. In order to discriminate between vegetations at this geographic scale, it is necessary to use relative abundance scale or presence/absence pollen data. We highlight the importance of adapting the type of data used according to the variations in relative abundance of pollen taxa within the vegetation group, which is itself related to the depositional environment of pollen assemblages: relative abundance scale data for non-dynamic depositional environments and presence/absence for dynamic depositional environments.

These results provide food for thought for the processing and analysis of pollen data in East Africa. It is also a conclusive proof of concept of our pollen score approach for modern pollen assemblages that legitimizes its test on the fossil record. This is both to interpret new fossil data and to reflect on previous pollen analyses. The results of these future works will help to confirm or not the ability of the $V_{\alpha,\beta}$ score to be an alternative to the PFT method.

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870

871 **Table captions:**

872 **Table 1:** Comparison of nomenclatures for the main regional vegetation types in Ethiopia.

873 **Table 2:** Regional vegetation types identified by botanists, number of pollen samples, and types of
874 depositional environments of samples for each vegetation group inferred. 0: van Breugel et al. (2015).
875 Non-dynamic depositional environments: soils, muds of lake edges, and soils/muds of swamps;
876 dynamic depositional environments: muds of river beds or edges. Notes: a: observation using Google
877 Earth images (1984 to 2020) ; b: in situ observations ; 1:Barboni et al. (2019) ; 2: Bonnefille and Riollot
878 (1988) ; 3:Bonnefille et al. (1987) ; 4: Buchet (1982) ; 5: Carr (1998) ; 6: Frey and Kürschner (1989) ; 7:
879 Friis et al. (2010) ; 8: Kobayashi et al. (2017) ; 9: Mohammed et al. (2007) ; 10: van Breugel et al. (2015)
880 ; 11: El-Moslimany (1983).

Table 3: The 10 most characteristic pollen types of each vegetation group inferred based on the calculated T-values). Taxa are listed from the most to the least important. For abbreviations of vegetation groups, please refer to Table 2.

Figure captions:

Figure 1: Distribution of regional vegetation types and pollen assemblages from this study. Red labels represent samples collected prior to 2009 and available in the APD or the literature, and white labels represent new (unpublished) samples collected in 2009 and 2017. Vegetation distribution is based on (van Breugel et al., 2015) for Africa and on Global Maps - Land Cover (year 2013, 0.46×0.46 km: (Kobayashi et al., 2017) for Arabian Peninsula. For abbreviations of vegetation types, please refer to Table 1. NA: not available.

Figure 2: Present-day distribution of the vegetation types and pollen samples in the climate domain of Ethiopia (grey shading) as defined by mean annual precipitation (mm/yr) and mean annual temperature (WorldClim2, 1970-2000, Fick and Hijmans, 2017). The grey shading is made of 53381 dots, each representing 4.63×4.63 km. A-G, Climatic domain occupied by the main vegetation types of Ethiopia obtained using the georeferenced map of vegetation types and sub-types over Eastern Africa produce under the Vegetation and Climate change in Eastern Africa (VECEA) project (van Breugel et al., 2015). H, Position of our 283 surface samples within the present-day climatic domain of Ethiopia and according to the vegetation identified at the time of sampling (Table S5). For abbreviations of vegetation types, refer to Table 1.

Figure 3: Occurrence of each pollen type for each pollen assemblage accorded to its relative abundance.

Figure 4: Comparison according to the type of pollen data (relative abundance, relative abundance scale and presence/absence) of $V_{\alpha,\beta}$ scores (the vegetation score of pollen assemblage β for the vegetation group α) applied to the 262 pollen assemblages for each vegetation group (α). (1) The range of scores of pollen assemblages that belong to vegetation α ($V_{\alpha,\beta \in \alpha}$) represented by the black bars (this is the range of significant values of each vegetation α) and the range of scores of pollen assemblages that do not belong to vegetation α ($V_{\alpha,\beta \notin \alpha}$) represented by the grey bars (this is the range of non-significant values of each vegetation α). (2) For each vegetation α , the vegetation groups from which pollen assemblages β originated with $V_{\alpha,\beta}$ scores intersecting the range of significant values ($V_{\alpha,\beta} \in V_{\alpha,\beta \in \alpha}$). For abbreviations of vegetation groups, refer to Table 2.

Figure 5: Comparison according to the type of pollen data (relative abundance, relative abundance scale and presence/absence) of $V_{\alpha,\beta}$ score (the vegetation score of pollen assemblage β for the vegetation group α) applied to pollen assemblages from dynamic depositional environments for the each vegetation group α linked with dynamic depositional environments. (1) The range of scores of pollen assemblages that belong to vegetation α ($V_{\alpha,\beta \in \alpha}$) represented by the black bars (this is the range of significant values of each vegetation α) and the range of scores of pollen assemblages that do not belong to vegetation α ($V_{\alpha,\beta \notin \alpha}$) represented by the grey bars (this is the range of non-significant values of each vegetation α). (2) For each vegetation α , the vegetation groups from which pollen assemblages β originated with $V_{\alpha,\beta}$ scores intersecting the range of significant values ($V_{\alpha,\beta} \in V_{\alpha,\beta \in \alpha}$). For abbreviations of vegetation groups, please refer to Table 2.

Figure 6: Modern range of mean temperature of warmest quarter by mean precipitation of driest quarter for pollen assemblages grouped by vegetation group (WorldClim2, 1970-2000, 0.93x0.93 km, Fick and Hijmans, 2017), with 95% confidence intervals. For abbreviations of vegetation groups, please refer to Table 2.

Figure 7: Percentage of tree cover associated with pollen assemblages for each vegetation group (Global Maps, year 2008, 0.46x0.46 km, Kobayashi et al., 2017), with a confidence interval of 95%. For abbreviations of vegetation groups, please refer to Table 2.

Supplementary Online Material

Table S1: List of accepted pollen types and their synonyms.

Table S2: Pollen count with harmonized pollen types according to the table S1.

Table S3: Metadata of pollen samples.

Table S4: Climatic data, edaphic data, net primary production value, and % of tree cover for each pollen sample.

Table S5: vegetation information for each pollen sample.

Table S6: list of Value Test for pollen data in relative abundance (sheet n°1), relative abundance scale (sheet n°2), and presence/absence (sheet n°3).

Figure S1: Comparison of K-means partitions for the number of abundance classes with the simple structure index (ssi) criterion.

941 **Figure S2:** Mean Decrease Accuracy for the 38 parameters used for reconstruct our 20 k-mean groups
942 with Random Forest.