## EMBO Course in Population Genomics, practicals on Coalescent Theory

## Question 1. Group 1

Consider a diploid population of size N (with 2N chromosomes) that has two alleles, C and c, segregating in i and 2N-i copies respectively, at t=0. The population is evolving following the standard Wright-Fisher model.

Recall: Assuming Wright-Fisher, the probability that an allele with i copies in the present generation is found in j copies in the next generation is given by

$$P_{ij} = {2N \choose j} p_0^j (1 - p_0)^{2N - j},$$

 $p_0 = \frac{i}{2N}$  is the frequency of the allele in the present generation.  $P_{ij}$  is a binomial distribution.

(a) Calculate the probability that allele c will be present in k copies in the first generation (t=1).

Answer: For this case: probability of success  $p = (1 - \frac{i}{2N})$ , therefore, the required probability is:

$$\binom{2N}{k} (1 - \frac{i}{2N})^k \ (\frac{i}{2N})^{2N-k}.$$

(b) What is the probability of observing allele C in 2N copies in the first generation (t=1).

Answer: For this case: probability of success  $p = \frac{i}{2N}$ , therefore,

required probability =  $\binom{2N}{2N} (\frac{i}{2N})^{2N} (1 - \frac{i}{2N})^{2N-2N} = (\frac{i}{2N})^{2N}$ .

(c) What is the probability that a sample of size n = 2 taken from generation t share the same parent at t - 1?

Answer: Required probability = (probability that the  $1^{st}$  progeny chooses a parent from the 2N individuals present at t-1)\*(probability that the  $2^{nd}$  progeny chooses the same parent present at t-1)\*(number of parents present at t-1),

Required probability =  $(\frac{1}{2N}) * (\frac{1}{2N}) * 2N = \frac{1}{2N}$ .

(d) With what probability k offspring at generation t share the same parent at t-1?

Answer: Following the same logic, required probability =  $(\frac{1}{2N})^{k-1}$ .

## Question 2. Group 2

Answer:

The coalescence times  $T_i$  in the standard coalescent process are independent and exponentially distributed as

$$f_{T_i}(t) = \binom{i}{2} e^{-\binom{i}{2}t}$$

when time is measured appropriately.

(a) What is the distribution of  $T_2$ ,  $T_3$ , and  $T_4$ ? Answer:

$$f_{T_2}(t) = {2 \choose 2} e^{-{2 \choose 2} t} = e^{-t},$$

$$f_{T_3}(t) = {3 \choose 2} e^{-{3 \choose 2} t} = 3 e^{-3 t},$$

$$f_{T_4}(t) = {4 \choose 2} e^{-{4 \choose 2} t} = 6 e^{-6 t}.$$

(b) Write down the average and variance of waiting times  $T_2$ ,  $T_3$ , and  $T_4$ . Also, write down the value of  $E[T_2^2]$ ,  $E[T_3^2]$ , and  $E[T_4^2]$ .

$$E[T_2] = 1, \ E[T_3] = \frac{1}{3}, \ E[T_4] = \frac{1}{6}.$$
 
$$Var[T_2] = 1, \ Var[T_3] = \left(\frac{1}{3}\right)^2 = \frac{1}{9}, \ Var[T_4] = \left(\frac{1}{6}\right)^2 = \frac{1}{36}.$$

By definition,

$$E[i^2] = Var[i] + (E[i])^2$$
, therefore,  $E[T_2^2] = 2$ ,  $E[T_3^2] = \frac{2}{9}$ , and  $E[T_4^2] = \frac{2}{36}$ .

- (c) If one infers that  $T_3 > T_4 > T_2$  upon looking at a gene genealogy for a sample of size 4, would this be consistent with the standard coalescent process? Justify your answer. **Answer:** Yes, it would be consistent with the standard coalescent process.  $T_2$ ,  $T_3$ , and  $T_4$  are the random variables. So, for some genealogies,  $T_3 > T_4 > T_2$  holds. Though  $E[T_2] > E[T_3] > E[T_4]$  will always be true.
- (d) Does  $\binom{i}{2}$  in the expression given above have some meaning in standard coalescent process (Kingman's coalescent)? If yes, what is it?

**Answer:** This quantity represents the rate of coalescence events. Since only two lineages merge at the same in the standard coalesence, it is equal to  $\binom{i}{2}$  the number of pairs among i lineages.

## Question 3. Group 3

 $T_{MRCA}$  is the time to the most recent common ancestor of the entire sample and  $T_{total}$  is the total length of all the branches in the genealogy.

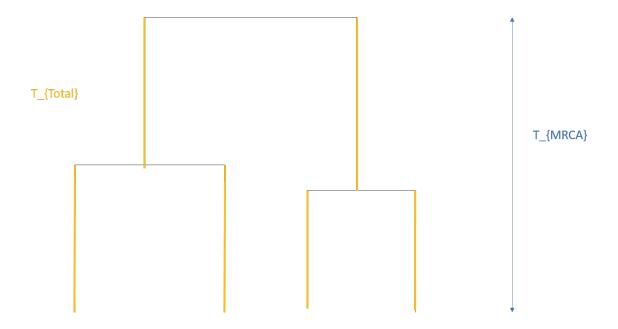
- (a) Draw a coalescent tree with sample size n=4 and indicate on the tree what  $T_{MRCA}$  and  $T_{total}$  correspond to.
- (b) Find  $E[T_{MRCA}]$  and  $E[T_{total}]$  for a sample of size n. Write down all the steps involved.

Answer:

$$T_{MRCA} = \sum_{i=2}^{n} T_i$$

where n is the sample size, since  $T_i$  are independent random variables, therefore,

$$E[T_{MRCA}] = \sum_{i=2}^{n} E[T_i],$$



$$E[T_{MRCA}] = \sum_{i=2}^{n} \frac{2}{i(i-1)} = 2\sum_{i=2}^{n} \left(\frac{1}{i-1} - \frac{1}{i}\right),$$

$$E[T_{MRCA}] = 2\left(\frac{1}{1} - \frac{1}{2} + \frac{1}{2} - \frac{1}{3} + \frac{1}{3} + \dots - \frac{1}{n-1} + \frac{1}{n-1} - \frac{1}{n}\right) = 2\left(1 - \frac{1}{n}\right).$$

$$T_{total} = \sum_{i=2}^{n} i T_i,$$

using E[cX] = c E[X],

$$E[T_{total}] = \sum_{i=2}^{n} i E[T_i],$$

$$E[T_{total}] = \sum_{i=2}^{n} \frac{2i}{i(i-1)} = \sum_{i=2}^{n} \frac{2}{(i-1)}.$$

(c) Calculate the value of  $E[T_{MRCA}]$  as n goes to infinity.

Answer:

$$\lim_{n \to \infty} E[T_{MRCA}] = 2$$

(d) Write down the  $E[T_{MRCA}]$  and  $E[T_{total}]$  when sample size is 4. Also, compute  $\frac{E[T_{MRCA}]}{E[T_2]}$  (definition of  $T_2$  is given in the previous question).

Answer:

$$E[T_{MRCA}] = 2\left(1 - \frac{1}{n}\right) = 2\left(1 - \frac{1}{4}\right) = \frac{3}{2} = 1.5,$$

$$E[T_{total}] = \sum_{i=2}^{4} \frac{2}{(i-1)} = \frac{11}{3} = 3.67.$$

$$\frac{E[T_{MRCA}]}{E[T_2]} = \frac{1.5}{1} = 1.5.$$

(e) Can  $\frac{E[T_{MRCA}]}{E[T_{total}]} > 1$  be true ? Justify your answer.

No, we have that  $E[T_{total}] < E[T_{MRCA}]$  cannot be true. The branches that are summed to get to  $T_{MRCA}$  are included in the sum of the  $T_{total}$  calculation.

Conclusion:  $E[T_{total}] > E[T_{MRCA}]$  will always be true.

Question 4. **Group 4** Consider the data below, where n=10 haploid sequences were simulated. Each line represents an individual and each column a segregating site S. The ancestral allele is denoted by "0" and the derived allele is denoted by "1." We observe a total of S=20 segregating sites.

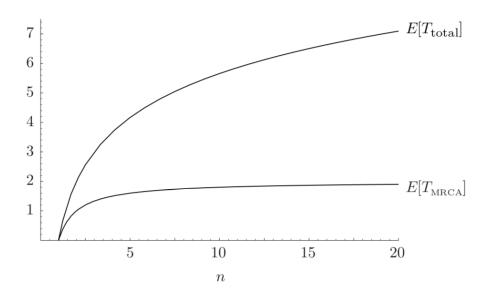


Figure 1: Reference: An Introduction to Coalescence Theory by John Wakeley.

(a) What is the frequency of the **derived** allele in each segregating site? Answer: 2 1 1 1 1 1 1 3 1 1 8 1 5 3 9 1 1 3 9

(b) Compute the SFS for the data above, using the allele frequencies you just computed.

Answer: We consider the frequencies of the derived allele (1) and count the number of derived alleles at each site to get the following:

Derived allele count	1	2	3	4	5	6	7	8	9
# of sites	12	1	3	0	1	0	0	1	2

Table 1: Unfolded SFS

The Figure 2 below shows two Site Frequency Spectra (SFS). The red distribution is the expected SFS for a given value of  $\theta$  for a population of constant size, not subdivided (geographically or otherwise), and where mutations have no effect in fitness. Mutation rate per base pair is constant and loci are not linked. The blue distribution is the observed SFS for a population simulated under neutrality using ms (Hudson 1990).

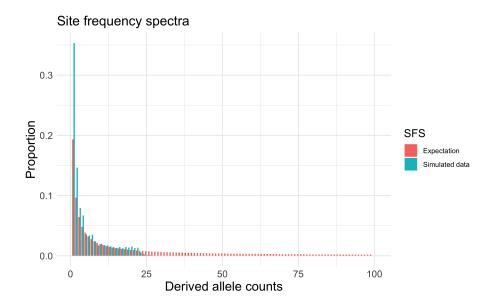


Figure 2: Site frequency spectra for two scenarios: a simulated dataset, and the expected SFS for a population with constant population size.

(c) For the red curve (expectation with constant  $N_e$ ), we see 800 singletons. What is the expected number of doubletons?

Answer:  $E[S_1] = \theta = 800$ .  $E[S_2] = \theta/2 = 400$ . We expect to see 400 doubletons.

(d) We can see that the SFS for the simulations (blue) departs from the expectation (red).

Besides some statistical noise, what is the most striking difference between these two distributions? Please use maximum two sentences to answer this question.

Answer: The most striking difference is that the blue curve has more than 50% singletons than the red curve. In other words, there seems to be an excess of singletons in the simulations relative to the expectation.

(e) Given that the same value of  $\theta$  was used to generate both distributions, can you think of an explanation for the observed differences? Please use maximum three sentences to answer this question.

Answer: The explanation is that the simulated population went through some sort of demographic change. Here, the population was simulated with a bottleneck and exponential growth.

OPTIONAL. **Group 1+2+3+4** Run the small program to simulate allele frequencies through time under a Wright-Fisher model for the simplest case, e.g. neutral alleles, constant population size. TIf you still have time, try the assignments below as well.

```
# WFsim.r simple simulations of the Wright-Fisher model.
# Code used by Andy Clark in 2018.
# Initialize popsize (N), number of samples (samp), number of
# generations (ngen) and starting frequency (startfreq)
# Note this is for a haploid population of size N.
N < -200
nsamp<-8
ngen < -200
startfreq < -.5
pcur <- matrix (1:N)
#Wright-Fisher is simply recurrent binomial sampling over generations
x<-rbinom (nsamp, N, startfreq)
p < -x/N
pcur<-p
for (i in 1:ngen){
x < -rbinom(nsamp, N, p)
p < -x/N
pcur<-rbind(pcur,p)</pre>
}
#Now plot these trajectories
gen < -seq(1, ngen + 1)
for (k in 1:nsamp) {
plot (gen, pcur [, k], type="l", ylim=c(0,1))
par (new=TRUE)
# ASSIGNMENT 1: Conduct runs of the above simulation, and
```

# calculate the variance in allele frequencies across 20

```
# independent sample populations for each of generations 20
# through 200 in steps of 20. Plot the results.
# Hint: the variance of allele freq at gen i is var(pcur[i,])
# ASSIGNMENT 2: Plot the expected time to fixation (when allele
# frequency is either 0 or 1) for 20 populations with population
\# sizes 50, 100, 150 and 200, starting from p = 0.5.
# ANSWER 1:
gen < -seq(1:200)
varvec < -seq(1,200)
for (i in 1:ngen){
  varvec[i]<-var(pcur[i,])
plot (gen, varvec, type="b", xlab="Generation", ylab="Variance among reps")
# You can see that the variance increases. It is non-linear in
# the plot because of the fact that subpopulations are
# going to fixation, when variance of course can no longer
# ANSWER 2:
fixtime <-matrix (seq (1,80),4,20)
for (i in 1:4){
  for (irep in 1:20)
     N < -i * 50
     p < -.5
     igen < -0
     while (abs(p-.5)<.499)
       x \leftarrow rbinom(1, N, p)
       p < -x/N
       igen < -igen + 1
   fixtime [i, irep]<-igen
}
```

```
# The mean fixation times for the populations of size
# 50,100,150, and 200 are:

mean(fixtime[1,])
mean(fixtime[2,])
mean(fixtime[3,])
mean(fixtime[4,])

# Note that the variance in fixation time is huge, so 20 replicates
# is not enough to give very consistent results.
# increase.
```