

# R package glvm

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# R package gllvm

- R package **gllvm** fits Generalized linear latent variable models (GLLVM) for multivariate data (Niku et al., 2017).
- Developed by J. Niku, W.Brooks, R. Herliansyah, F.K.C. Hui, S. Taskinen, D.I. Warton, B. van der Veen.
- GitHub: <https://github.com/JenniNiku/gllvm>
- CRAN: <https://cran.r-project.org/web/packages/gllvm/index.html>

# R package glvm

- GLLVMs are computationally intensive to fit.
- Often such models have been fitted using MCMC approach, which is quite time consuming.
- **glvm** package overcomes computational problems by applying closed form approximations to log-likelihood and using automatic differentiation in C++ to accelerate computation times (**TMB**).
- Estimation is performed using either variational approximation (VA), extended variational approximation (EVA) or Laplace approximation (LA) method implemented via R package **TMB**.
- VA method is faster and more accurate than LA, but not applicable for all distributions and link functions. In such cases EVA can be used.

# R package gllvm

Using **gllvm** we can fit

- GLLVM without latent variables fits basic multivariate GLMs
- GLLVM without covariates gives model-based ordination and biplots, also constrained ordination is possible.
- GLLVM with environmental covariates for studying factors explaining species abundance
- Fourth corner models with latent variables for studying environmental-trait interactions

Additional tools: model checking, model selection, inference, visualization.

# Distributions

Response	Distribution	Method	Link
Counts	Poisson	VA/LA	log
	NB	VA/LA	log
	ZIP	LA	log
Binary	Bernoulli	VA/LA	probit
		EVA/LA	logit
Ordinal	Ordinal	VA	probit
Normal	Gaussian	VA/LA	identity
Positive continuous	Gamma	VA/LA	log
non-negative continuous	Exponential	VA/LA	log
Biomass	Tweedie	EVA/LA	log
Cover	Beta	EVA/LA	probit

# Data input

Main function of the **gllvm** package is `gllvm()`, which can be used to fit GLLVMs for multivariate data with the most important arguments listed in the following:

```
gllvm(y = NULL, X = NULL, TR = NULL, family, num.lv = NULL,  
      formula = NULL, method = "VA", row.eff = FALSE, n.init=1, ...)
```

- `y`: matrix of abundances
- `X`: matrix or data.frame of environmental variables
- `TR`: matrix or data.frame of trait variables
- `family`: distribution for responses
- `num.lv`: number of latent variables
- `method`: approximation used "VA", "EVA" or "LA"
- `row.eff`: type of row effects
- `n.init`: number of random starting points for latent variables

# Example: Spiders

- Abundances of 12 hunting spider species measured as a count at 28 sites.
- Six environmental variables measured at each site.
  - `soil.dry`: Soil dry mass
  - `bare.sand`: cover of bare sand
  - `fallen.leaves`: cover of fallen leaves/twigs
  - `moss`: cover of moss
  - `herb.layer`: cover of herb layer
  - `reflection`: reflection of the soil surface with a cloudless sky

# Data fitting

Fit GLLVM without covariates  $g(E(y_{ij})) = \beta_{0j} + \mathbf{u}_i' \boldsymbol{\theta}_j$  with `gllvm`, as a default 2 latent variables are used:

```
library(mvabund)
data("spider")
library(gllvm)
fitnb <- gllvm(y = spider$abund, family = "negative.binomial")
fitnb
## Call:
## gllvm(y = spider$abund, family = "negative.binomial")
## family:
## [1] "negative.binomial"
## method:
## [1] "VA"
##
## log-likelihood: -733.6806
## Residual degrees of freedom: 289
## AIC: 1561.361
## AICc: 1577.028
## BIC: 1623.975
```



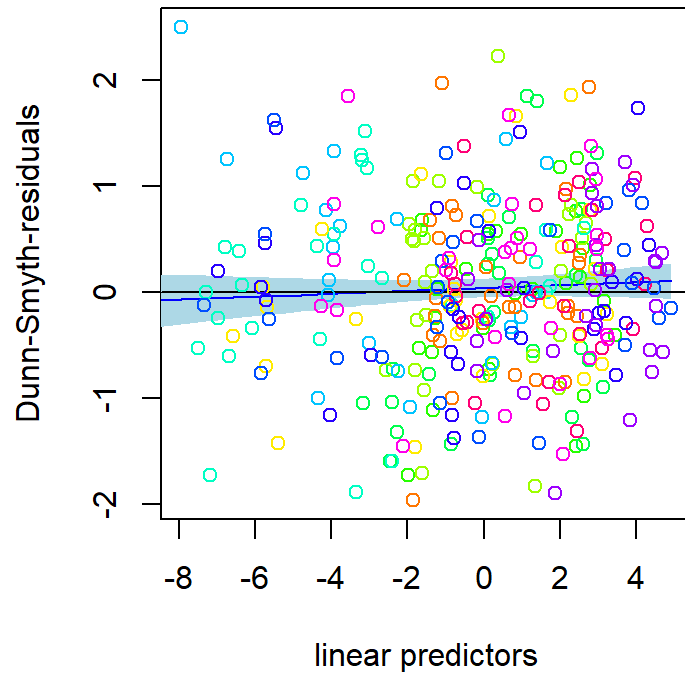
# Residual analysis

- Residual analysis can be used to assess the appropriateness of the fitted model (eg. in terms of mean-variance relationship).
- Function `residuals()` calculates randomized quantile residuals for the model, and `plot()` function provides residual plots.
- Randomized quantile/Dunn-Smyth residuals are used in the package, as they provide standard normal distributed residuals, even for discrete responses, in the case of a proper model.

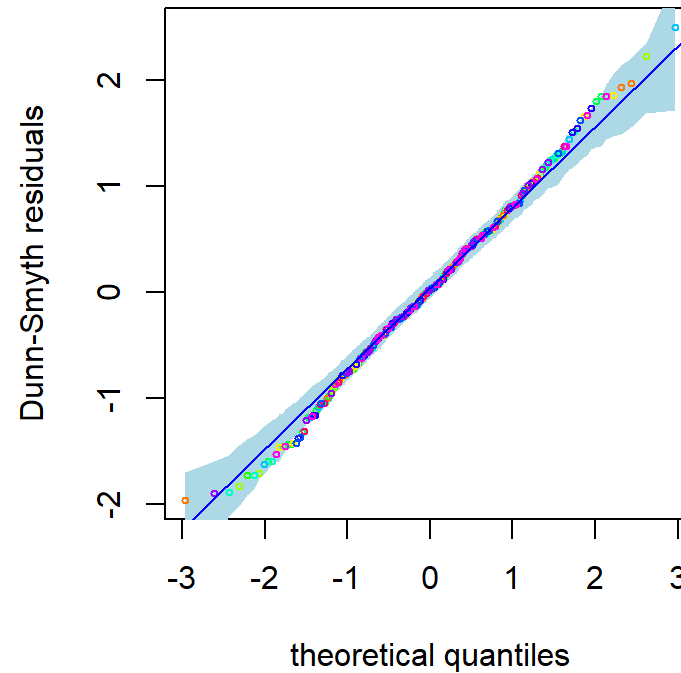
# Residual analysis

```
par(mfrow = c(1,2))  
plot(fitnb, which = 1:2)
```

**Residuals vs linear predictors**



**Normal Q-Q**



# Model selection

- Information criterias can be used for model selection.
- For example, compare distributions or choose suitable number of latent variables.

```
fitp <- gllvm(y = spider$abund, family = poisson())  
fitnb <- gllvm(y = spider$abund, family = "negative.binomial")  
AIC(fitp)  
## [1] 1761.655  
AIC(fitnb)  
## [1] 1561.361
```

# Studying species associations

- Latent variables induce correlation across response variables, and so provide means of estimating correlation patterns across species, and the extent to which they can be explained by environmental variables.
- Information on correlation is stored in the LV loadings  $\boldsymbol{\theta}_j$ , so the residual covariance matrix, storing information on species co-occurrence that is not explained by environmental variables, can be calculated as  $\boldsymbol{\Sigma} = \mathbf{\Gamma}\mathbf{\Gamma}^\top$ , where  $\mathbf{\Gamma} = [\boldsymbol{\theta}_1 \dots \boldsymbol{\theta}_m]'$ .
- `getResidualCor` function can be used to estimate the correlation matrix of the linear predictor across species.

# Studying species associations

- Let's consider first the correlation matrix based on a model without predictors:  $g(E(y_{ij})) = \beta_{0j} + \mathbf{u}_i' \boldsymbol{\theta}_j$

```
fitnb <- gllvm(spider$abund, family = "negative.binomial")
```

- The obtained correlation matrix then does not take into account the environmental conditions driving species abundances at sites, and reflects only what has been observed.

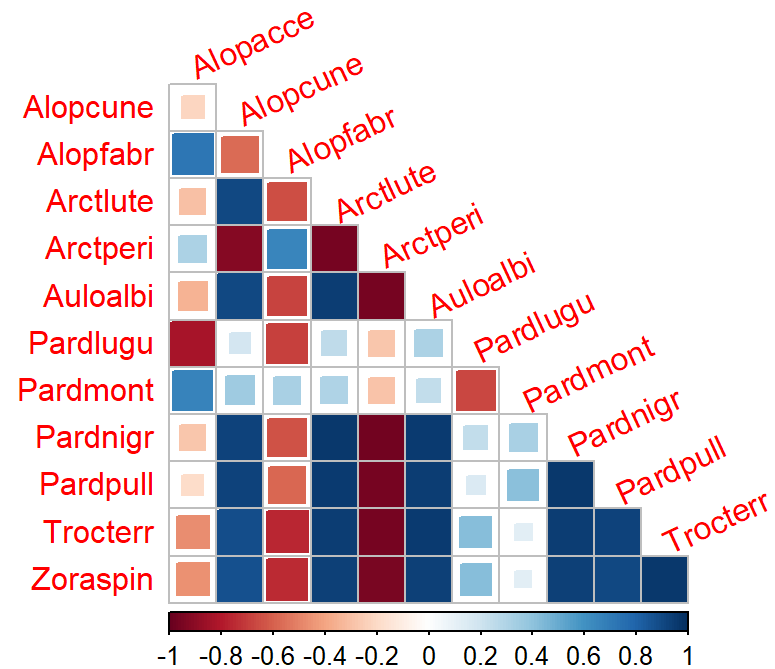
# Correlation matrix

The residual correlation matrix can be visualized using, eg., a `corrplot()` function from `corrplot` package:

```
cr <- getResidualCor(fitnb)
library(corrplot);
```

```
corrplot(cr, diag = FALSE, type = "lower", method = "square", tl.srt = 25)
```

# Correlation matrix



# GLLVM as a model based ordination method

- GLLVMs can be used as a model-based approach to unconstrained ordination by including two latent variables in the model:

$$g(E(y_{ij})) = \beta_{0j} + \mathbf{u}_i' \boldsymbol{\theta}_j$$

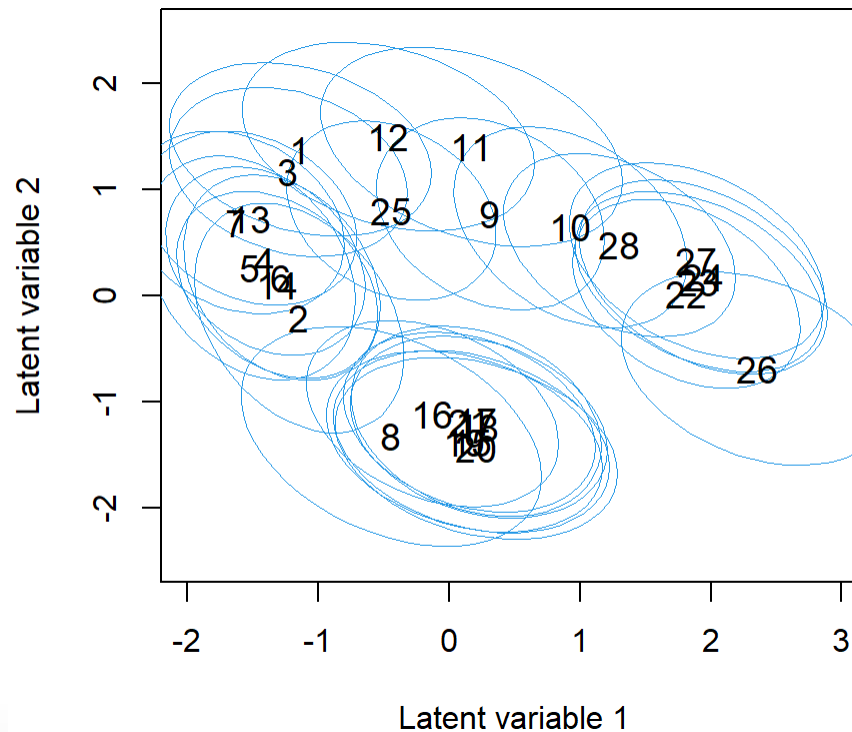
- The latent variable term try to capture the underlying factors driving species abundances at sites.
- Predictions for the two latent variables,  $\hat{\mathbf{u}}_i = (\hat{u}_{i1}, \hat{u}_{i2})$ , then provide coordinates for sites in the ordination plot and then provides a graphical representation of which sites are similar in terms of their species composition.



# Ordination plot

- `ordipLOT()` produces ordination plots based on fitted GLLVMs.
- Uncertainty of the ordination points in model based ordination can be assessed with prediction errors of latent variables.

```
ordipLOT(fitnb, predict.region = TRUE, ylim=c(-2.5,2.5), xlim=c(-2,3))
```

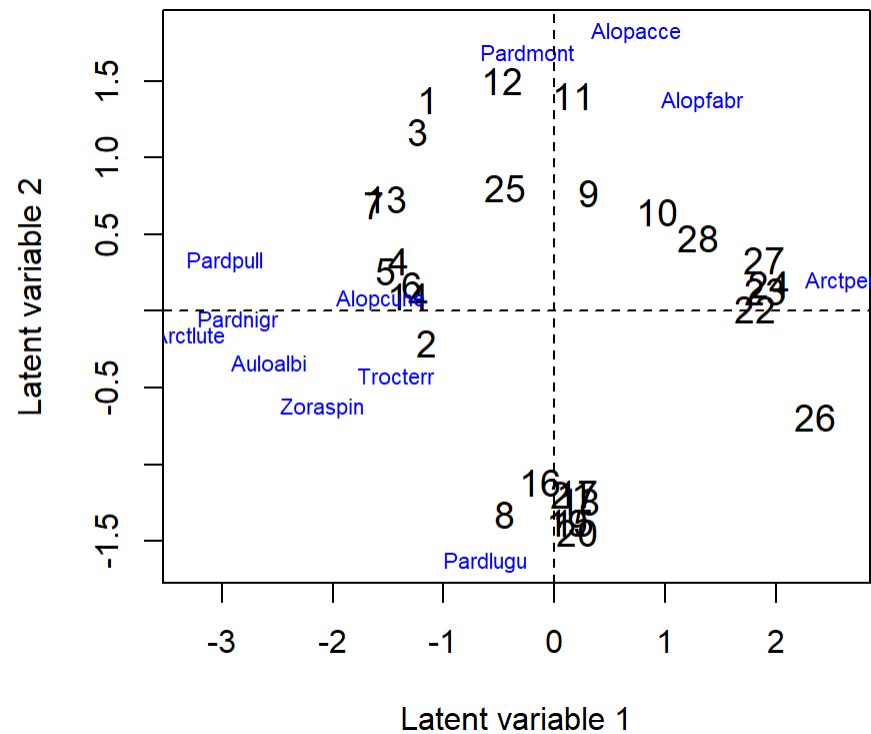


# Biplot

- The ordination can also be used for visualizing the species associations by producing a biplot, (argument `biplot = TRUE` in `ordiplot()`), that is, by adding latent variable loadings  $\theta_j$  to the ordination of sites.
- In a biplot latent variables and their loadings are rotated so that the LV loadings of the species are in the same direction with the sites where they are most abundant.
- The biplots can be used for finding groups of correlated species or finding indicator species common at specific sites.

# Biplot

```
ordiplot(fitnb, biplot = TRUE)  
abline(h = 0, v = 0, lty=2)
```

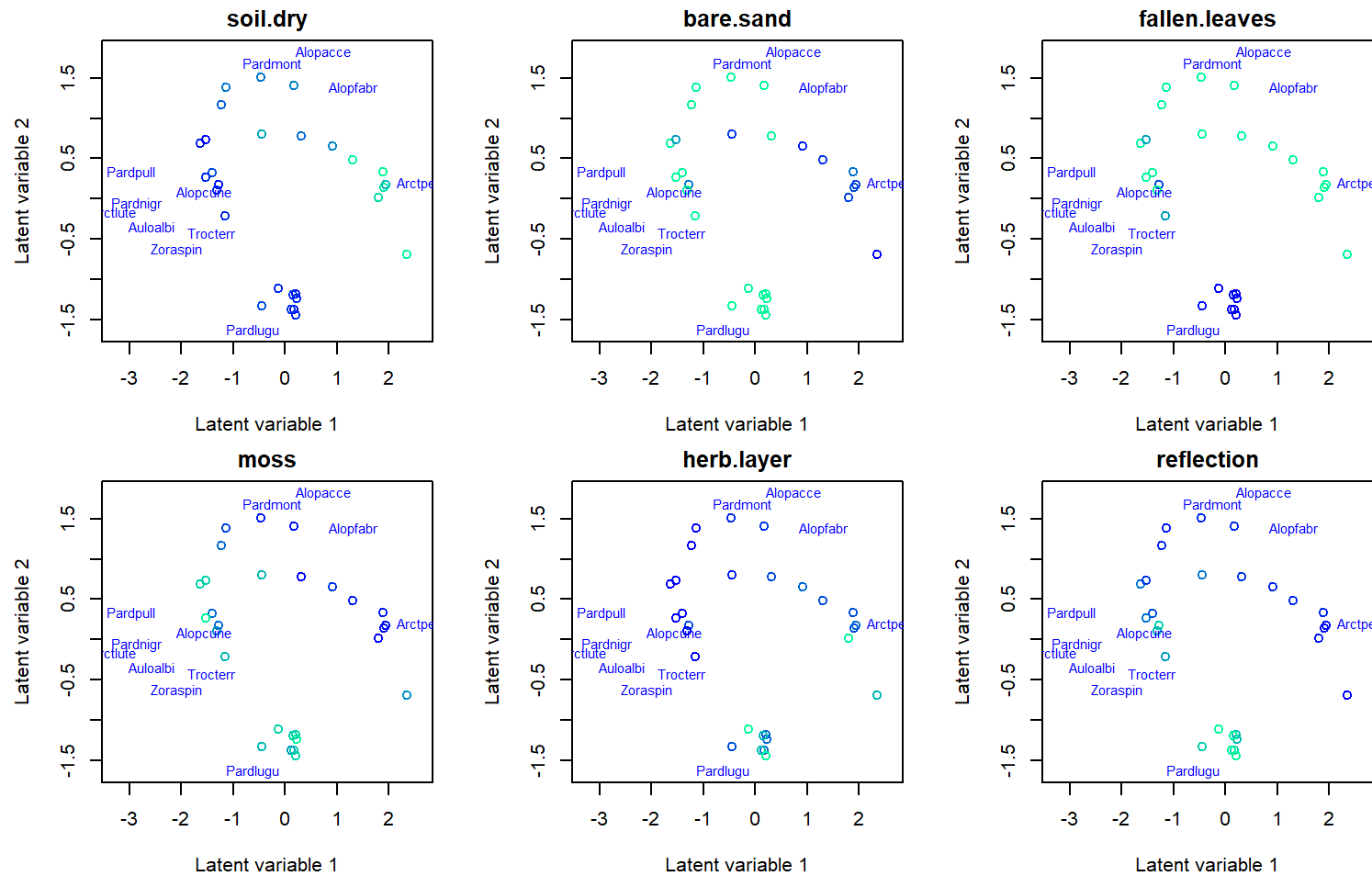


# Environmental gradients

The potential impact of environmental variables on species communities can be viewed by coloring ordination points according to the variables.

```
# Arbitrary color palette, a vector length of 20
rbPal <- c("#00FA9A", "#00EC9F", "#00DFA4", "#00D2A9", "#00C5AF", ...)
X <- spider$x
par(mfrow = c(2,3), mar=c(4,4,2,2))
for(i in 1:ncol(X)){
  Col <- rbPal[as.numeric(cut(X[,i], breaks = 20))]
  ordiplot(fitnb, symbols = T, s.colors = Col, main = colnames(X)[i],
           biplot = TRUE)
}
```

# Environmental gradients



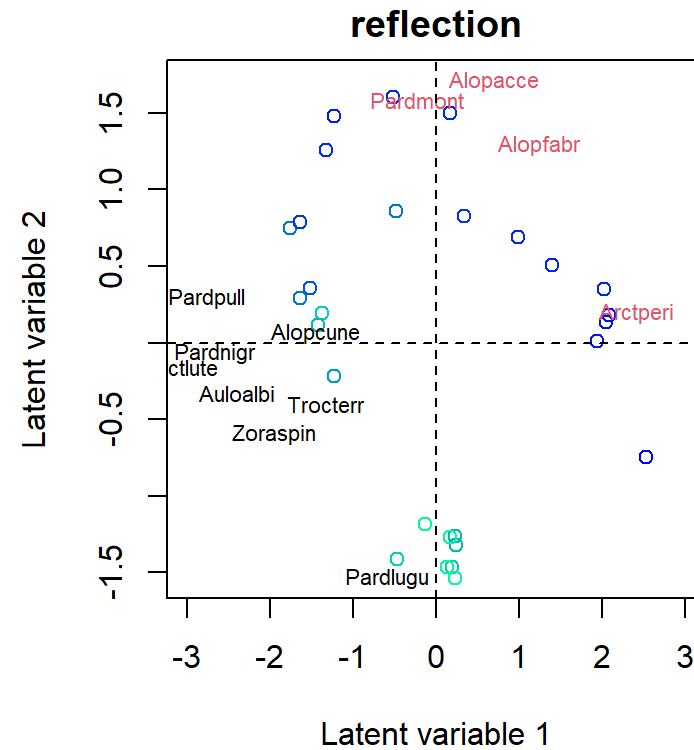
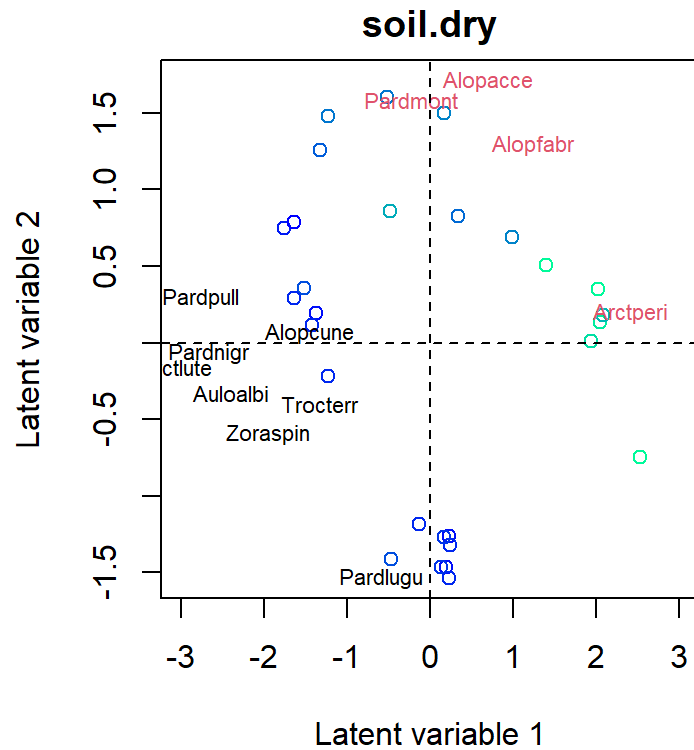
# Studying effects of environmental variables

- The effects of environmental variables on species can be studied by including environmental variables  $\mathbf{x}_i$  to GLLVM:

$$g(E(y_{ij})) = \beta_{0j} + \mathbf{x}_i' \boldsymbol{\beta}_j + \mathbf{u}_i' \boldsymbol{\theta}_j.$$

- $\boldsymbol{\beta}_j$  is a vector of species specific coefficients for environmental variables.
- Next consider for example two environmental variables, `soil.dry` (soil dry mass) and `reflection` (reflection of the soil surface with a cloudless sky), which shows different environmental gradients in ordination:

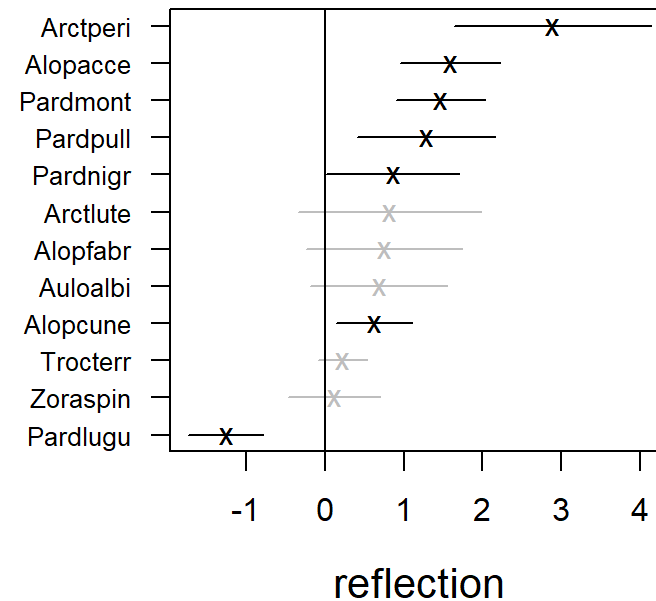
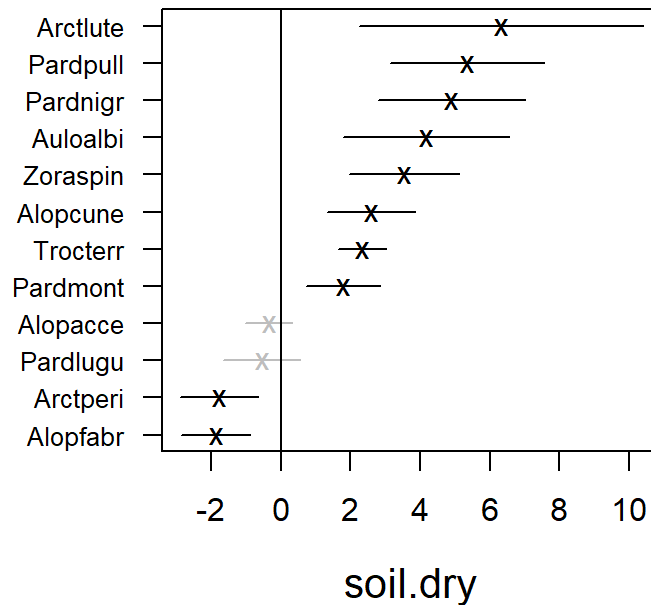
# Environmental gradients



# Coefficient plot

`coefplot()` plots point estimates of the species specific environmental coefficients  $\beta_j$  with confidence intervals.

```
fitx1 <- gllvm(spider$abund, X, formula = ~soil.dry + reflection, family = "negative.binomial")  
coefplot(fitx1, mfrow = c(1,2), cex.ylab = 0.8)
```

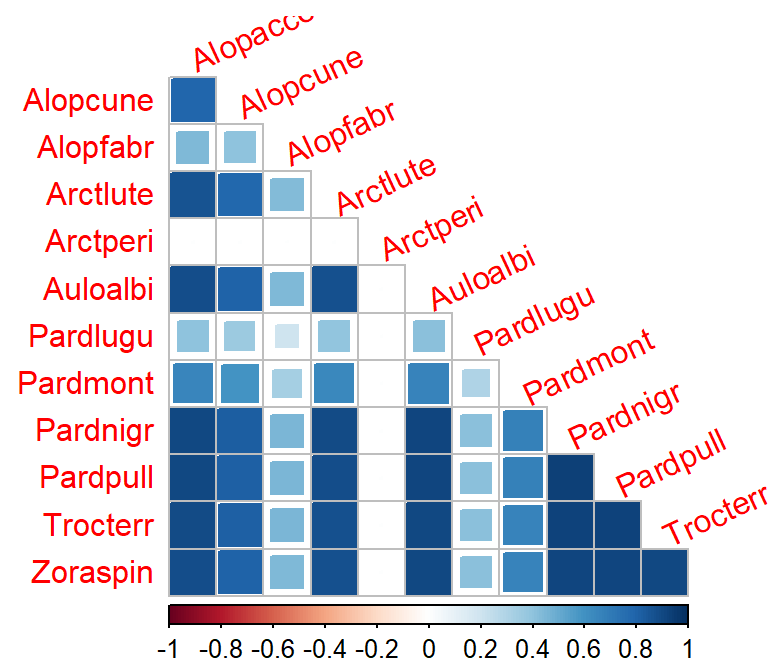




# Correlation matrix

Correlation matrix for model with predictors shows correlation patterns between species when the effect of the predictors are taken into account.

```
crx <- getResidualCor(fitx1)
corrplot(crx, diag = FALSE, type = "lower", method = "square", tl.srt = 25)
```



# Fourth corner models

- If species trait variables  $\mathbf{t}_j$ , measuring eg. species behaviour or physical appearance, would be available, fourth corner models should be considered:  $g(E(y_{ij})) = \beta_{0j} + \mathbf{x}'_i \boldsymbol{\beta}_j + \mathbf{x}'_i \mathbf{B}_I \mathbf{t}_j + \mathbf{u}'_i \boldsymbol{\theta}_j$
- Such models can also be fitted with `gllvm()` function by including a matrix of traits with argument `TR`.
- Examples can be found in the `gllvm` package's vignettes.

# More

More information and examples about the usage of the package can be found from the **gllvm** package's website: <https://jenniniku.github.io/gllvm/>

Github repository of the workshop:

<https://github.com/BertvanderVeen/IRSAE2020GLLVMworkshop>