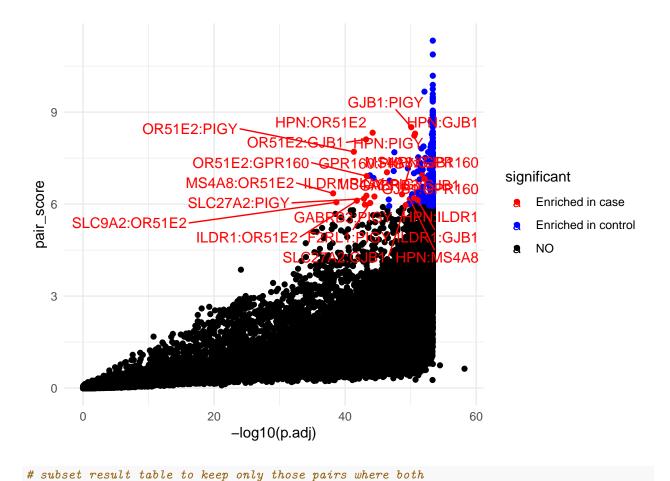
Bispecific_Antibody_Target_Selection_ProstateCancer

2024-02-21

```
library(edgeR)
## Loading required package: limma
library(bsabsfinder)
## Warning: replacing previous import 'ensembldb::filter' by 'stats::filter' when
## loading 'bsabsfinder'
library(cluster)
library(dplyr)
## Attaching package: 'dplyr'
## The following objects are masked from 'package:stats':
##
##
       filter, lag
## The following objects are masked from 'package:base':
##
##
       intersect, setdiff, setequal, union
library(ggplot2)
library(ggpubr)
suppressWarnings({
    case = subset(phenoDF, cancer == "prostate adenocarcinoma" &
       sample.type == "primary") #Select Prostate cancer as case
    case_id = case$sample.id #getting case IDS
    control = subset(phenoDF, sample.type == "normal" & biopsy.site ==
        "PROSTATE") #Select Normal Prostate samples
    control_id = control$sample.id #getting control IDS
   case_expr = loadOctadCounts(case_id, type = "tpm", file = "~/Downloads/octad.counts.and.tpm.h5")
    case_expr = as.data.frame(case_expr)
    control_expr = loadOctadCounts(control_id, type = "tpm",
       file = "~/Downloads/octad.counts.and.tpm.h5")
   control_expr = as.data.frame(control_expr)
    # final data
```

```
case_with_control_expr = cbind(case_expr, control_expr)
    # convert ensq to hqnc and select surface-expressed
    # genes according to compartments.jensenlab.org
    case_with_control_expr = ensg_to_hgnc(case_with_control_expr,
        select surface = TRUE)
    phenotype_vector = as.factor(c(rep("case", ncol(case_expr))),
        rep("control", ncol(control_expr))))
})
## [1] "loading 60498 TPM expression values for 494 samples"
## [1] "loading 60498 TPM expression values for 100 samples"
#Perform Differential Gene Expression to filter out non-significant genes to speed up the computation
annotation = data.frame(sample = c(colnames(case_expr), colnames(control_expr)),
    phenotype = c(rep("cancer", length(colnames(case_expr))),
        rep("control", length(colnames(control_expr)))))
annotation$phenotype = as.factor(annotation$phenotype)
expression = DGEList(counts = as.matrix(case with control expr),
    group = annotation$phenotype)
dim(expression)
## [1] 3736 594
keep <- rowSums(cpm(expression) > 100) >= 2
expression <- expression[keep, ]
dim(expression)
## [1] 3117 594
expression$samples$lib.size <- colSums(expression$counts)</pre>
suppressWarnings({
    expression <- calcNormFactors(expression)</pre>
})
expression_disp <- estimateCommonDisp(expression, verbose = T)</pre>
## Disp = 1e-04 , BCV = 0.01
expression_disp <- estimateTagwiseDisp(expression_disp)</pre>
DE <- exactTest(expression_disp, pair = c(1, 2)) # compare groups 1 and 2
DE = DE$table
DE$padj = p.adjust(DE$PValue, method = "BH")
DE1 = subset(DE, padj < 0.05 & abs(logFC) > 1) # The cutoff criteria can be changed
# filter out only surface-expressed DE genes. Just to speed
```

```
case_with_control_expr = case_with_control_expr[row.names(case_with_control_expr) %in%
   row.names(DE1), ]
dataframe_for_computation = as.data.frame(t(case_with_control_expr))
# this step takes a while
small_res = compute_bsabs(antigene_1 = colnames(dataframe_for_computation),
   data_input = dataframe_for_computation, pheno_input = phenotype_vector)
##
     1
                                                                                   ١
head(small_res)
     antigen_1 antigen_2 distance
                                   spread angle_cos pair_score
                                                                    p.value
## 1
        ITGA2B
                  SCN4A 1.266441 1.645612 0.7126953
                                                      1.485307 8.513137e-52
## 2
       ITGA2B
                   GIPR 2.655597 1.627037 0.9112219 3.937166 8.630330e-50
       ITGA2B
               NPC1L1 1.327407 1.654277 0.7319016 1.607182 1.249741e-51
## 3
       ITGA2B SLC11A1 2.383090 1.665728 0.9500795 3.771416 9.820476e-55
## 4
       ITGA2B CYP3A43 1.436237 1.646023 0.8889049 2.101442 2.291172e-51
## 5
## 6
       ITGA2B
                 SLC7A9 1.639132 1.656881 0.9921792 2.694607 1.994614e-52
           p.adj case_greater
## 1 3.597593e-51 FALSE_FALSE
## 2 2.706100e-49 FALSE_FALSE
## 3 5.124317e-51 FALSE FALSE
## 4 1.223267e-53 FALSE FALSE
## 5 8.991584e-51 FALSE FALSE
## 6 9.610274e-52 FALSE_FALSE
## PLOT FIG.2D
suppressWarnings({
   plot_bsabs(small_res, label = "case", pval_cut_off = 0.01,
       pair score cut off = quantile(small res$pair score, 0.99))
})
```



```
# markers have higher expression in case than control
small_res = small_res[small_res$case_greater == "TRUE_TRUE",
# ordering as per pair score , highest score should be at
# top
small_res = small_res[order(small_res$pair_score, decreasing = T),
    ]
# Subsetting top 20 pairs
small_res_selective = small_res[c(1:20), ]
# unique marker genes in top 20 pairs
marker_list = unique(c(small_res_selective$antigen_1, small_res_selective$antigen_2))
marker_list
    [1] "GJB1"
                                      "GPR160"
                                                                     "GABRB3"
                  "HPN"
                            "OR51E2"
                                                "MS4A8"
                                                           "ILDR1"
    [8] "SLC27A2" "SLC9A2"
                            "PIGY"
# Checking the expression of marker genes in healthy tissue
healthy_tissues = subset(phenoDF, sample.type == "normal")
healthy_tissues = subset(healthy_tissues, grepl("BRAIN", biopsy.site)
    biopsy.site == "LIVER" | biopsy.site == "LUNG" | grep1("HEART",
```

```
biopsy.site) | grepl("KIDNEY", biopsy.site))
healthy_tissues <- healthy_tissues %>%
   mutate(biopsy.site = ifelse(grepl("BRAIN", biopsy.site),
       "BRAIN", biopsy.site))
healthy_tissues <- healthy_tissues %>%
   mutate(biopsy.site = ifelse(grepl("HEART", biopsy.site),
       "HEART", biopsy.site))
healthy_tissues <- healthy_tissues %>%
   mutate(biopsy.site = ifelse(grepl("KIDNEY", biopsy.site),
       "KIDNEY", biopsy.site))
healthy tissues expr = loadOctadCounts(healthy tissues$sample.id,
   type = "tpm", file = "~/Downloads/octad.counts.and.tpm.h5")
## [1] "loading 60498 TPM expression values for 1950 samples"
healthy_tissues_expr = as.data.frame(healthy_tissues_expr)
healthy_tissues_expr = ensg_to_hgnc(healthy_tissues_expr, select_surface = FALSE)
healthy_tissues_expr = healthy_tissues_expr[row.names(healthy_tissues_expr) %in%
   marker_list, ]
healthy_tissues_expr = healthy_tissues_expr[order(rownames(healthy_tissues_expr)),
   1
healthy_tissues_expr = as.data.frame(t(healthy_tissues_expr))
healthy_tissues_expr$Sample = healthy_tissues$biopsy.site[match(rownames(healthy_tissues_expr),
   healthy_tissues$sample.id)]
case_with_control_expr2 = case_with_control_expr[row.names(case_with_control_expr) %in%
   marker_list, ]
case_with_control_expr2 = case_with_control_expr2[order(rownames(case_with_control_expr2)),
case_with_control_expr2 = as.data.frame(t(case_with_control_expr2))
case_with_control_expr2$Sample = ifelse(rownames(case_with_control_expr2) %in%
   case_id, "PROSTATE_CANCER", "PROSTATE")
colnames(case with control expr2) == colnames(healthy tissues expr)
marker_expr = rbind(case_with_control_expr2, healthy_tissues_expr)
table(marker_expr$Sample)
```

##

```
##
             BRAIN
                              HEART
                                              KIDNEY
                                                                LIVER
                                                                                  LUNG
                                                                  110
##
              1148
                                376
                                                  28
                                                                                   288
          PROSTATE PROSTATE CANCER
##
##
               100
```

```
sample_order <- c("PROSTATE_CANCER", "PROSTATE", "BRAIN", "HEART",
    "LUNG", "LIVER", "KIDNEY")

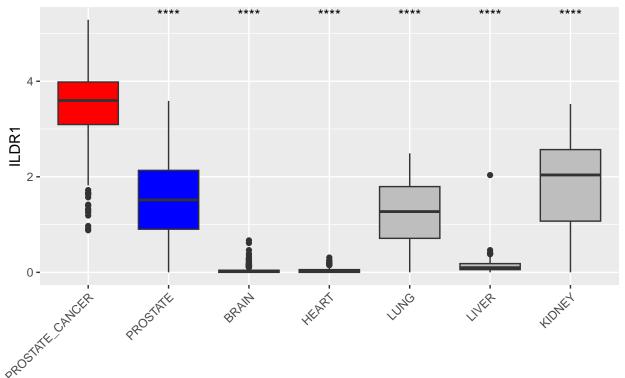
marker_expr$Sample <- factor(marker_expr$Sample, levels = sample_order)

site_comparisons = list(c("PROSTATE_CANCER", "PROSTATE"), c("PROSTATE_CANCER",
    "BRAIN"), c("PROSTATE_CANCER", "HEART"), c("PROSTATE_CANCER",
    "LUNG"), c("PROSTATE_CANCER", "LIVER"), c("PROSTATE_CANCER",
    "KIDNEY"))

# PLOT FIG. 2E with STATS

ggplot(marker_expr, aes(x = Sample, y = marker_expr[[5]], fill = Sample)) +
    geom_boxplot() + scale_fill_manual(values = c("red", "blue",
    rep("gray", length(sample_order) - 2)), guide = "none") +
    labs(x = "Biopsy Site", y = names(marker_expr)[5]), title = paste("Marker expression by names(marker_expr)[5])) + theme(axis.text.x = element_text(angle = 45,
    hjust = 1)) + guides(fill = "none") + stat_compare_means(method = "t.test",
    ref.group = "PROSTATE_CANCER", label = "p.signif")</pre>
```

Marker expression by biopsy site: ILDR1

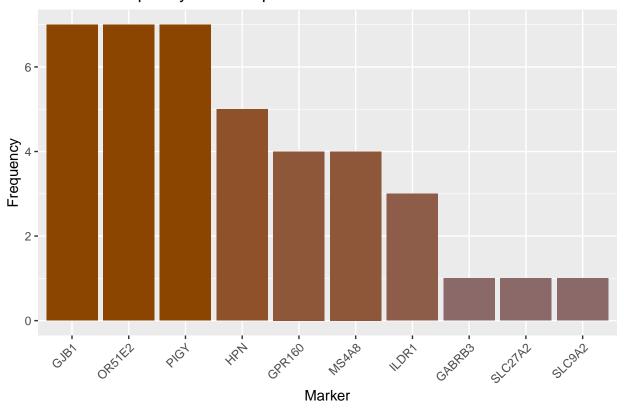


Biopsy Site

```
# Similar to above plot , this code will generate plots for
# all markers. You can specify the working directory where
# you would like to save the pdf.
pdf("~/Downloads/BSAB PROSTATE marker.pdf")
   for (i in 1:length(marker_list)) {
       gg <- ggplot(marker_expr, aes(x = Sample, y = marker_expr[[i]],
            fill = Sample)) + geom_boxplot() + scale_fill_manual(values = c("red",
            "blue", rep("gray", length(sample_order) - 2)), guide = "none") +
            labs(x = "Biopsy Site", y = names(marker_expr)[i],
                title = paste("Marker expression by biopsy site: ",
                  names(marker_expr)[i])) + theme(axis.text.x = element_text(angle = 45,
            hjust = 1)) + stat_compare_means(method = "t.test",
            ref.group = "PROSTATE_CANCER", label = "p.signif")
       print(gg)
   }
}
```

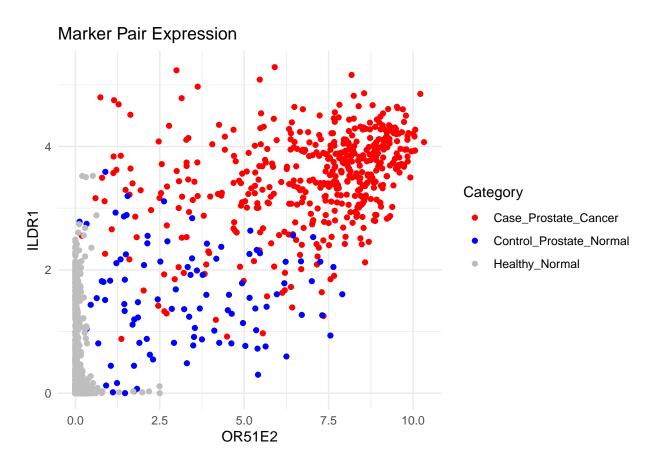
```
# PLOT FIG. 2C Marker Frequency plot
markers <- c(small_res_selective santigen_1, small_res_selective santigen_2)
# Count the occurrences of each antigen in the combined
# vector
marker_counts <- table(markers)</pre>
# Convert the result to a data frame
marker_counts_df <- as.data.frame(marker_counts)</pre>
names(marker_counts_df) <- c("Marker", "Frequency")</pre>
marker_counts_df <- marker_counts_df[order(-marker_counts_df$Frequency),</pre>
marker_counts_df$Marker <- factor(marker_counts_df$Marker, levels = marker_counts_df$Marker)
proxy_var <- as.numeric(marker_counts_df$Frequency)</pre>
gg_freq <- ggplot(marker_counts_df, aes(x = Marker, y = Frequency,</pre>
    fill = proxy_var)) + geom_bar(stat = "identity") + scale_y_continuous(breaks = seq(0,
    max(marker_counts_df$Frequency) + 2, by = 2), limits = c(0,
    max(marker_counts_df$Frequency))) + theme(axis.text.x = element_text(angle = 45,
    hjust = 1)) + labs(x = "Marker", y = "Frequency", title = "Mareker Frequency in BSAB pairs") +
    scale_fill_gradient(low = "rosybrown4", high = "darkorange4",
        guide = "none")
print(gg_freq)
```

Mareker Frequency in BSAB pairs



```
# PLOT FIG.2B Marker Frequency plot
marker_expr$Category = NA
unique(marker_expr$Sample)
```

```
## [1] PROSTATE_CANCER PROSTATE HEART BRAIN
## [5] LUNG LIVER KIDNEY
## Levels: PROSTATE_CANCER PROSTATE BRAIN HEART LUNG LIVER KIDNEY
```



Save all important generated results

 $\textit{\# save(list=c('case_expr','control_expr','case_with_control_expr','case_with_control_expr','DE','DE1', } \\$