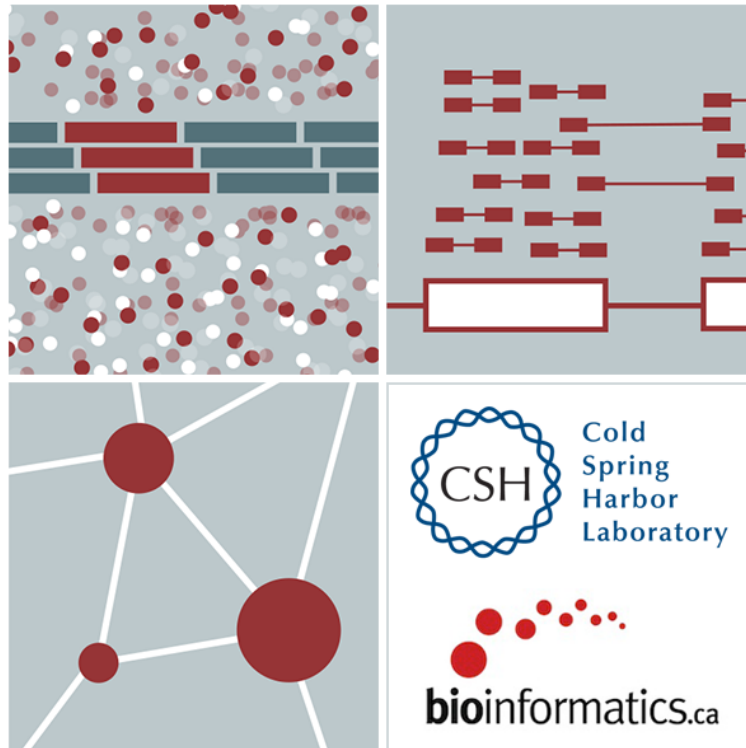


# RNA-Seq Module 3

## SAM/BAM/BED file formats

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# Example of SAM/BAM file format

## Example SAM/BAM/CRAM header section (abbreviated)

```
mqriffit@linux270 ~$ samtools view -H /gscmnt/gc13001/info/model_data/2891632684/build136494552/alignments/136080019.bam | grep -P "SN:[2\|HD\|RG\|PG"
```

```
@HD      VN:1.4    SO:coordinate  
@SQ      SN:12   LN:51304566     UR:ftp://ftp.ncbi.nih.gov/genbank/genomes/Eukaryotes/vertebrates_mammals/Homo_sapiens/GRCh37/special_requests/GRCh37-lite.fa.gz AS:GRCh37-lite M5:a718acaa6135fdca8357d5bfe9  
421118  SP:Homo sapiens  
@RG      ID:2888721359    PL:illumina    PU:D1B4ACXX.3 LB:H_KA-452198-0817007-cDNA-3-lib1 PI:365 DS:paired end DT:2012-10-03T19:00:00-0500 SM:H_KA-452198-0817007 ON:WUGSC  
@PG      ID:2888721359    VN:2.0.8      CL:tophat --library-type fr-secondstrand --bowtie-version=2.1.0  
@PG      ID:MarkDuplicates PN:MarkDuplicates PP:2888721359 VN:1.85(exported) CL:net.sf.picard.sam.MarkDuplicates INPUT=[/gscmnt/gc13001/info/build_merged_alignments/merged-alignment-blade10-2-5.gsc.wustl.edu-jwaker-15434-136080019/scrch-Ilg6v/H_KA-452198-0817007-cDNA-3-lib1-2888360300.bam] OUTPUT=/gscmnt/gc13001/info/build_merged_alignments/merged-alignment-blade10-2-5.gsc.wustl.edu-jwaker-15434-136080019/scrch-Ilg6v/H_KA-452198-0817007-cDNA-3-lib1-2888360300-post.dup.bam METRICS_FILE=/gscmnt/gc13001/info/build_merged_alignments/merged-alignment-blade10-2-5.gsc.wustl.edu-jwaker-15434-136080019/staging-ljuJS/H_KA-452198-0817007-cDNA-3-lib1-2888360300.metrics REMOVE_DUPLICATES=false ASSUME_SORTED=true MAX_FILE_HANDLES_FOR_READ_ENDS_MAP=9500 TMP_DIR=[/gscmnt/gc13001/info/build_merged_alignments/merged-alignment-blade10-2-5.gsc.wustl.edu-jwaker-15434-136080019/scrch-Ilg6v] VALIDATION_STRINGENCY=SILENT MAX_RECORDS_IN_RAM=500000 PROGRAM_RECORD_ID=MarkDuplicates PROGRAM_GROUP_NAME=Mark Duplicates MAX_SEQUENCES_FOR_DISK_READ_ENDS_MAP=50000 SORTING_COLLECTION_SIZE_RATIO=0.25 READ_NAME_REGEX=[a-zA-Z0-9+]+:(0-9)+:([0-9+]):([0-9+]).* OPTICAL_DUPLICATE_PIXEL_DISTANCE=100 VERBOSITY=INFO QUIT=false COMPRESSION_LEVEL=5 CREATE_INDEX=false CREATE_MD5_FILE=false
```

```
mqriffit@linux270 ~$
```

## Example SAM/BAM/CRAM alignment section (only 10 alignments shown)

[illegible]

# Introduction to the SAM/BAM format

- The specification
  - <http://samtools.sourceforge.net/SAM1.pdf>
- SAM is uncompressed text data
- BAM is a compressed version of SAM
  - lossless BGZF format
- BAM files are usually 'indexed'
  - A '.bai' file will be found beside the '.bam' file
  - Indexing provides fast retrieval of alignments overlapping a specified region without going through all alignments.
  - BAM must be sorted by the reference ID and then the leftmost coordinate before indexing

# SAM/BAM header section

- Used to describe source of data, reference sequence, method of alignment, etc.
- Each section begins with character '@' followed by a two-letter record type code. These are followed by two-letter tags and values:
  - @HD The header line
    - VN: format version
    - SO: Sorting order of alignments
  - @SQ Reference sequence dictionary
    - SN: reference sequence name
    - LN: reference sequence length
    - SP: species
  - @RG Read group
    - ID: read group identifier
    - CN: name of sequencing center
    - SM: sample name
  - @PG Program
    - PN: program name
    - VN: program version

# SAM/BAM alignment section

Col	Field	Type	Regex/Range	Brief description
1	QNAME	String	[!-?A-~]{1,255}	Query template NAME
★ 2	FLAG	Int	[0,2 <sup>16</sup> -1]	bitwise FLAG
3	RNAME	String	\*  [!-( )+-<>-~] [!-~]*	Reference sequence NAME
4	POS	Int	[0,2 <sup>29</sup> -1]	1-based leftmost mapping POSition
5	MAPQ	Int	[0,2 <sup>8</sup> -1]	MAPping Quality
★ 6	CIGAR	String	\*  ([0-9]+[MIDNSHPX=])+	CIGAR string
7	RNEXT	String	\* =  [!-( )+-<>-~] [!-~]*	Ref. name of the mate/next segment
8	PNEXT	Int	[0,2 <sup>29</sup> -1]	Position of the mate/next segment
9	TLEN	Int	[-2 <sup>29</sup> +1,2 <sup>29</sup> -1]	observed Template LENgth
10	SEQ	String	\*  [A-Za-z=.]+	segment SEQUENCE
11	QUAL	String	[!-~]+	ASCII of Phred-scaled base QUALity+33

## Example values

1	QNAME	e.g.	HWI-ST495_129147882:1:2302:10269:12362
2	FLAG	e.g.	99
3	RNAME	e.g.	1
4	POS	e.g.	11623
5	MAPQ	e.g.	3
6	CIGAR	e.g.	100M
7	RNEXT	e.g.	=
8	PNEXT	e.g.	11740
9	TLEN	e.g.	217
10	SEQ	e.g.	CCTGTTTCTCCACAAAGTGTTTACTTTTGGATTTTTGCCAGTCTAACAGGTGAAGCCCTGGAGATTCTTATTAGTGATTTGGGCTGGGGCCTGGCCATGT
11	QUAL	e.g.	CCCFFFFFFHHHHHJJJIJFIJJJJJJJJJJJHIJJJJJJJIJJJJJGGHIJHIJJJJJJJJJGHGGIJJJJJJIJEEHHHHFFFFCDCDDDDDDDB@ACDD

# SAM/BAM flags explained

- 12 bitwise flags describing the alignment
- Stored as a binary string of length 11 instead of 11 columns of data
- Value of '1' indicates the flag is set. e.g. 00100000000
- All combinations can be represented as a number from 1 to 2048 (i.e.  $2^{11}-1$ ). This number is used in the BAM/SAM file.
- You can specify 'required' or 'filter' flags in samtools view using the '-f' and '-F' options respectively

Bit		Description
1	0x1	template having multiple segments in sequencing
2	0x2	each segment properly aligned according to the aligner
4	0x4	segment unmapped
8	0x8	next segment in the template unmapped
16	0x10	SEQ being reverse complemented
32	0x20	SEQ of the next segment in the template being reverse complemented
64	0x40	the first segment in the template
128	0x80	the last segment in the template
256	0x100	secondary alignment
512	0x200	not passing filters, such as platform/vendor quality controls
1024	0x400	PCR or optical duplicate
2048	0x800	supplementary alignment

Note that to maximize confusion, each bit is described in the SAM specification using its hexadecimal representation (i.e., '0x10' = 16 and '0x40' = 64).

<http://broadinstitute.github.io/picard/explain-flags.html>

# CIGAR strings explained

- The CIGAR string is a sequence of base lengths and associated ‘operations’ indicating which bases align to the reference (either a match or mismatch), are deleted, are inserted, represent introns, etc.

Op	BAM	Description	
M	0	alignment match (can be a sequence match or mismatch)	
I	1	insertion to the reference	
D	2	deletion from the reference	
N	3	skipped region from the reference	
S	4	soft clipping (clipped sequences present in SEQ)	
H	5	hard clipping (clipped sequences NOT present in SEQ)	;
P	6	padding (silent deletion from padded reference)	that
=	7	sequence match	
X	8	sequence mismatch	

•e.g. 81M859N19M

- A 100 bp read consists of: 81 bases of alignment to reference, 859 bases skipped (an intron), 19 bases of alignment



# CRAM files

- CRAM is an ultra-compressed version of a BAM file
  - Usually between 30-60% smaller than the corresponding BAM
- Stores “diffs” from the reference genome
  - requires the matching reference genome to restore original data!
- Base quality binning may be used as well
- Some tools still require conversion back to bam

Quality Score Bins	Example of Empirically Mapped Quality Scores*
N (no call)	N (no call)
2-9	6
10-19	15
20-24	22
25-29	27
30-34	33
35-39	37
≥ 40	40

By replacing the quality scores between 19 and 25 with a new score of 22, data storage space is conserved.

\*The mapped quality score of each bin (except “N”) is subject to change depending on individual Q-tables.



# Introduction to the BED format

- When working with BAM files, it is very common to want to examine a focused subset of the reference genome
  - e.g. the exons of a gene
- These subsets are commonly specified in 'BED' files
  - <https://genome.ucsc.edu/FAQ/FAQformat.html#format1>
- Many BAM manipulation tools accept regions of interest in BED format
- Basic BED format (tab separated):
  - Chromosome name, start position, end position
  - Coordinates in BED format are 0 based

# Manipulation of SAM/BAM and BED files

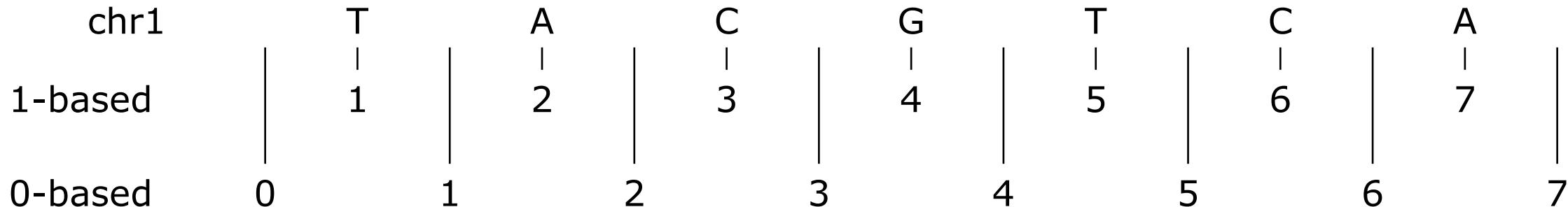
- Several tools are used ubiquitously in sequence analysis to manipulate these files
- SAM/BAM files
  - samtools
  - bamtools
  - Picard
- BED files
  - bedtools
  - bedops



# Common sources of confusion

- Genomic coordinate systems
- Genome builds
- Variant representation

# Genomic coordinates – 1 vs 0 based



	1-based	0-based
Indicate a single nucleotide	chr1:4-4 G	chr1:3-4 G
Indicate a range of nucleotides	chr1:2-4 ACG	chr1:1-4 ACG
Indicate a single nucleotide variant	chr1:5-5 T/A	chr1:4-5 T/A

- 1-based : Single nucleotides, variant positions, or ranges are specified directly by their corresponding nucleotide numbers
  - GFF, SAM, VCF, Ensembl browser, ...
- 0-based: Single nucleotides, variant positions, or ranges are specified by the coordinates that flank them
  - BED, BAM, UCSC browser, ...

# Genome builds

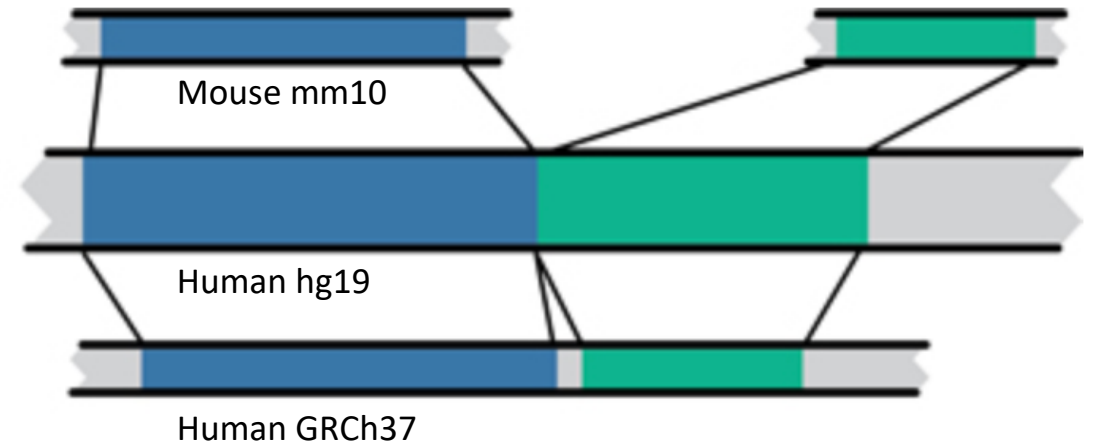
## Reference Genome builds

Current human: GRCh38, hg38, b38  
alternate: GRCh38v2\_ccdg

Previous human: GRCh37, hg19, b37

Current mouse: GRCm38, mm10

## Lift-over



# Variant shifting (alignment) and parsimony/trimming

Reference and alternative alleles of a CA short tandem repeat (STR)	REF	GGGCACACACAGGG			
	ALT	GGGCACACAGGG			
				← CA deletion from the reference	
	Genome Reference	Variant Call Format			
	GGGCACACACAGGG	POS	REF	ALT	
REF	CA	8	CA	.	Not left aligned and alternate allele is empty
ALT	.				
REF	CAC	6	CAC	C	Not left aligned but parsimonious
ALT	C				
REF	GCACA	3	GCACA	GCA	Not right trimmed
ALT	GCA				
REF	GGCA	2	GGCA	GG	Not left trimmed
ALT	GG				
REF	GCA	3	GCA	G	Normalized (left aligned & parsimonious)
ALT	G				
Alleles represented against the human genome reference. Allele pairs are colored the same, all are representations of the same variant.		Alleles represented in Variant Call Format, all are representations of the same variant.			

**Parsimony:** representing variant in as few nucleotides as possible without reducing the length of any allele to 0

**Left (right) aligning =** shifting the start position of a variant as far to the left (right) as possible

# How should I sort my SAM/BAM file?

- Generally BAM files are sorted by position
  - This is for performance reasons
    - When sorted and indexed, arbitrary positions in a massive BAM file can be accessed rapidly
- Certain tools require a BAM sorted by read name
  - Usually this is when we need to easily identify both reads of a pair
    - The insert size between two reads may be large
    - In fusion detection we are interested in read pairs that map to different chromosomes