

Use the NEBuilder® Protocol Calculator to calculate the optimal amounts of input DNA sequences given the length and concentration of each input fragment.

ENTER FRAGMENTS FOR ASSEMBLY

Vector ?	Name <input type="text" value="ID/Name"/>	Length ? basepairs <input type="text" value="2682"/>	Concentration ? ng/μl <input type="text" value="100"/>	Add Fragment
<input checked="" type="checkbox"/>	backbone	<input type="text" value="2682"/>	<input type="text" value="100"/>	Delete
<input type="checkbox"/>	insert	<input type="text" value="905"/>	<input type="text" value="10"/>	Delete

[Clear All Fragments](#)

SUGGESTED PROTOCOL

- Set up the reaction on ice (see table).

Component	Volume	Amount
backbone	0.8 μl	0.050 pmoles
insert	5.6 μl	0.100 pmoles
Deionized H ₂ O	3.6 μl	---
NEBuilder HiFi DNA Assembly Master Mix	10.0 μl	---
Total	20.0 μl	0.150 pmoles

[Maximize](#) [?](#)

- Incubate samples in a thermocycler at 50°C for 15 minutes. Following incubation, store samples on ice or at -20°C for subsequent transformation.

Note: Extended incubation up to 60 minutes may help to improve assembly efficiency in some cases.

- Transform NEB® 5-alpha, NEB Stable or NEB 10-beta Competent *E. coli* cells (provided in the cloning kit, bundle or purchased separately from NEB) with 2.0 μl (10%) of the chilled assembled product, following the transformation protocol.

Related Resources



[NEBuilder® HiFi DNA Assembly Reaction Protocol](#)
[NEBuilder® HiFi DNA Assembly](#)

Related Products



[NEBuilder® HiFi DNA Assembly Master Mix](#)
[NEBuilder® HiFi DNA Assembly Cloning Kit](#)
[NEBuilder® HiFi DNA Assembly Bundle for Large Fragments](#)

Related Tools



[NEBuilder® Assembly Tool](#)