# A REVISED CATALOG OF ARIZONA LICHENS

Scott T. Bates University of Colorado at Boulder Rm. 318 CIRES Bldg. Boulder, CO 80309

Anne Barber
Edward Gilbert
Robin T. Schroeder
and
Thomas H. Nash III
School of Life Sciences
Arizona State University
P. O. Box 874601
Tempe, AZ 85282-4601

### **ABSTRACT**

This revised "catalog" of lichens includes 969 species of lichenized fungi (lichens) presented for the state of Arizona (USA), and updates the original catalog published in 1975 by Nash and Johnsen. These taxa are derived from within 5 classes and over 17 orders and 54 families (the latter two with 3 and 4 additional groups of uncertain taxonomic position, "Incertae sedis") in the phylum Ascomycota. The total number of species reported here represents approximately 20% of all species known from the North American lichen flora. The list was compiled by extracting Arizona records from the three volume published set of the Lichen Flora of the Greater Sonoran Desert Region, a collaborative, authoritative treatment of lichen groups for this region that encompasses the entire state of Arizona.

# INTRODUCTION

Nearly 80,000 species of fungi are known to science (Schmit and Mueller 2007). Of these, approximately 17% are lichenized, forming symbioses with green algae (*Chlorophyta*, *Viridiplantae*) or the so called blue-green algae (*Cyanobacteria*, *Bacteria*). These relationships produce symbiotic organisms commonly called lichens. The fungal partner (mycobiont) is thought to benefit by having access to photosynthates produced by the "algae" (photobiont), which, within the symbiosis, is thought to receive some form of protection (e.g., against desiccation or UV radiation); however, evidence for other interpretations (e.g., controlled parasitism) exist (see Nash 2008a). The perception of lichens as classic dual mutualistic symbionts is, perhaps, rather simplistic as a single lichen thallus may host complex communities of microbes, including lichenicolous (lichen-associated) or endolichenic (growing within the cells of the lichen mycobiont) fungi, both types of photobionts (tripartite lichens), and even non-photosynthetic bacteria (Lawrey & Diederich 2003, Diederich 2004, Suryanarayanan et al. 2005, Nash 2008a, Arnold et al. 2009, Grube et al. 2009, Hodkinson & Lutzoni 2009).

Despite this potential for lichens to act as multifaceted symbiotic systems, their taxonomy rests solely on the mycobiont, as lichen species names refer only to

the lichenized fungus. A full 98% of all lichen-forming fungi can be found within the phylum *Ascomycota*, although some are members of the *Basidiomycota* (e.g., *Lichenomphalia* spp.), and approximately 42% of all known ascomycetous fungi are lichenized, compared to only 0.3% for basidiomycetes (Honegger 2008). In total, lichen-forming fungi are found within 5 classes of the *Ascomycota*, the majority of species being found within the *Lecanorales* (*Lecanoromycetes*), and within 1 class (*Agaricomycetes*) of the *Basidiomycota* (Honegger 2008). In addition, a rather interesting borderline case of "lichenization" is the fungus *Geosiphon pyriforme*, which unlike "true" lichens does not have extracellular photobionts surrounded by hyphae. Instead, this species has an intracellular (e.g., contained within the hyphae) endosymbiotic cyanobacterium (*Nostoc*). Being found within the *Glomeromycota*, *G. pyriforme* is more closely related to arbuscular mycorrhizal fungi than all other lichen species (Gehrig et al. 1996).

In addition to their rich phylogenetic diversity, lichens also display a wonderful diversity of colors, which are the byproduct of the numerous secondary metabolites (ca. 700 organic compounds are known, with over 90% being specifically produced by lichens; Elix & Stocker-Wörgötter 2008), and growth forms (see Fig. 1A-I). For the latter, three principal growth forms are traditionally recognized: crustose (e.g., Fig. 1G), foliose (e.g., Fig. 1D), and fruticose (e.g., 1E). Representing the most rudimentary thallus structure, the crustose (crust-like) lichen has only an upper cortex, while the lower surface adheres to the substrate on which the lichen is growing. This type of thallus is desiccation resistant and less susceptible to disturbance (e.g., storms, wind, etc.), and as a result it is often able to survive under extreme conditions (e.g., at high altitude on rock surfaces) where other growth forms are not able to exist. In addition to this adaptive resilience, crustose lichens are able to thrive under a wide range of environmental conditions, and as a result are broadly distributed: from leaf surfaces in the tropics, to rock and wood surfaces in temperate areas, and even on desert soils. Foliose (leaf-like) lichens typically have upper and lower cortices and are more loosely attached to their substrate. This type of growth form is well represented and often quite conspicuous in more moderate climates, growing on rock, tree bark, or over soils. interesting variation of this growth form, the vagrant lichen (Fig. 1B), does not attach to a substrate, and is found over soils, being blown about freely by wind. Fruticose lichens represent the most dimensional lichen growth form, having beard-, hair-, shrub-, and strap-like forms, and their thalli typically have a radial arrangement (a single, fairly uniform cortex surrounding a central axis). There is quite a bit of variation to the fruticose lichen thallus, ranging from a single stem-like appendage which elevates the apothecium (e.g., the podetium of *Cladonia* spp.), to highly-branched (e.g., Usnea intermedia; Fig. 1E) or long pendulous (e.g., Usnea The fruticose growth form is preferentially distributed in longissima) forms. temperate rain forests or dry areas where precipitation is infrequent but fog or dew events are regular (e.g., along arid coastal zones; Büdel & Scheidegger 2008).

Lichens are an important, but sometimes overlooked, part of the ecosystems in which they are found: as components in nutrient and mineral cycling; contributors to pedogensis; providers of food, shelter, and nest building material for micro- and mega-fauna; and they can serve as bioindicators of ecosystem health or air quality

(reviewed in Nash 2008a). For example, lichens (particularly those with cyanobacterial photobionts that can fix atmospheric nitrogen) contribute, sometimes significantly, to total N inputs in many ecosystems (Nash 2008b). These numerous ecological roles are, perhaps, more fully appreciated in light of the fact that lichens are poikilohydric (their water status is passively dependent on environmental conditions) and that they also rely, almost entirely, on atmospheric deposition in order to obtain macro- and micro-nutrients required for life (see Nash 2008a).

The history of lichenology in Arizona is fairly extensive, dating back to the late seventeenth and early eighteenth centuries, with some of more prominent lichenologists of that time period citing records of lichens from Arizona or describing new species from specimens collected in the state. For example in the late 1800s, Edward Tuckerman, the pioneering American lichenologist, and Henry Willey described Omphalodium hottentottum var. arizonicum Tuck. ex Willey [= Omphalora arizonica (Tuck. ex Willey) T.H. Nash & Hafellner] based on a specimen collected in the Santa Rita Mountains (Willey 1881). The distinguished Austrian lichenologist Alexander Zahlbruckner (1908) published 6 new lichen species (and one novel variety) based on collections from the Carnegie Desert Botanical Laboratory near Tucson. The following year Bruce Fink (1909), another leading American lichenologist of this period and author of the Lichen Flora of the United States, noted additional species occurring in this same area and reported 33 species from the "desert lichen flora". Additional like examples can be found in Nash and Johnsen (1975); however, it was not until the mid-1990s that lichen floras which focused on specific regions in the state were published (Darrow 1950, Weber 1963, Johnsen 1965).

Focus on the Arizona lichen flora intensified in the latter-half of the 1900s. In 1971, the Arizona State University Herbarium (ASU) began to expand its collection of lichens under the directorship of T.H. Nash III (from an original ca. 100 specimens, the majority being from Sweden, to the nearly 110,000 specimens today, approximately 50,000 attributed to Nash, originating from countries around the globe but focusing primarily on the greater Sonoran Desert region, and including Arizona), that eventually required these specimens to be housed separately from the vascular plant collection. Nash (1973b) began reporting records for the Arizona lichen flora and publishing on new species from the state (e.g., Nash 1973a). In the mid-1980s, the late Bruce D. Ryan enrolled in the graduate program at Arizona State University and received his doctorate degree in 1989, focusing on *Lecanora* subgen. Placodium for his dissertation. Ryan later served as the Lichen Herbarium Associate Curator (from 1989–2004), and amassed ca. 20,000 personal collections, deposited at ASU, many of which are from Arizona. The effort to increase knowledge of the Arizona lichen flora and to collect more extensively in the state continued on into the 20th century and resulted in publication of several regional floras for the state (Nash 1975, Nash 1977, Nash & Sigal 1981, Nash 1991, Boykin & Nash 1994, Sweat et al., 2004, Jackson et al. 2005), the original catalog of Arizona lichens (Nash & Johnsen 1975), consecutive additions to the flora (Moberg and Weber 1974, Nash 1985, Nash et al. 1998), as well as a field guide to epiphytic macrolichens from the state (Bungartz et al. 2002).

Nash and Ryan's combined and comprehensive knowledge of the area's lichen flora, the popular lichen keys produced by Dr. Ryan, the growing body of literature, and a fruitful collecting trip with the International Association of Lichenologists (IAL) in 1989 to the Sonoran Desert, which piqued the interest of several European lichen taxonomists, provided the foundation, network, and synergy for what eventually became a National Science Foundation funded project, the *Greater Sonoran Desert Lichen Flora*. This floristic effort, which included 92 collaborators from 23 countries coordinated by Nash, culminated with the publication of a three volume monograph (Nash et al. 2002, 2004, 2007) that provided keys, authoritative taxonomic treatments, as well as a thorough introduction to the science of lichenology. These volumes not only profoundly increased the understanding of the lichen floras of the Sonoran Desert and surrounding environs (e.g., southern California) as well as Arizona, but they expanded the taxonomic knowledge of the North American lichen flora in general and made a considerable contribution to lichenology in Mexico.

# **METHODS**

The compilation of this lichen "catalog" follows the methods outlined previously in the checklists of Arizona macrofungi and slime molds (Bates 2006, Bates & Barber 2008), with one notable exception: the records published here are referenced almost exclusively to the *Lichen Flora of the Greater Sonoran Desert Region* (Nash et al. 2002, 2004, 2007) as it represents the authoritative account of Arizona lichens. Reference specimens for the lichen flora of the state are housed in the Arizona State University Lichen Herbarium (ASU), which represents one of the more significant lichen collections (including important historical components; see e.g., Nash 2002) in North America, if not the world, and is irreplaceable in its value as a living regional collection. Synonymy and currently accepted names follow the *Cumulative Checklist for the Lichen-Forming, Lichenicolous and Allied Fungi of the Continental United States and Canada* (Esslinger 2010), and the classification system used conforms to the CABI *Index Fungorum* (http://www.indexfungorum.org).

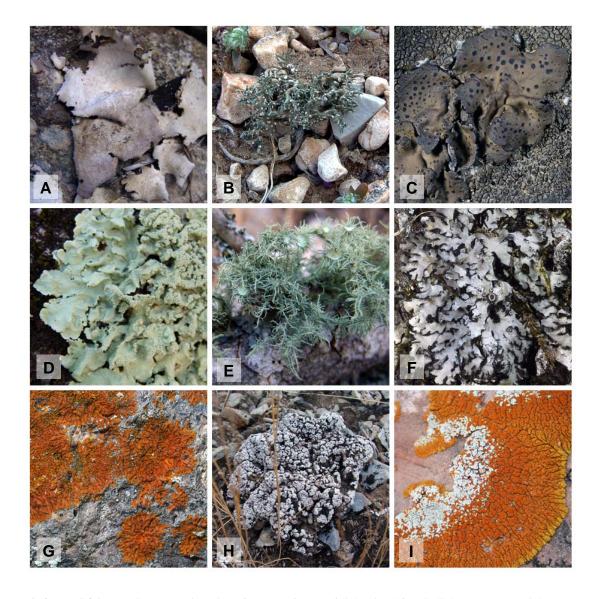
As with previous Arizona checklists (Bates 2006, Bates & Barber 2008), the data from this publication are available online (http://www.azfungi.org/checklist/), now as the *Checklist of Arizona Macrofungi, Lichens, and Slime Molds*, which includes approximately 7000 records of lichenized and non-lichenized fungi (and slime molds) from the state. In addition, many species in the online checklist are linked to images, which were generated primarily through the *Arizona Mycota Project* (www.azfungi.org/amp/). Perhaps more importantly, these data as well as those related to the *Lichen Flora of the Greater Sonoran Desert Region* and the ASU Lichen Herbarium are now accessible online in conjunction with data from several other lichen herbaria on the Symbiota (http://www.symbiota.org) lichen node, *Consortium of North American Lichen Herbaria* (CNALH; http://symbiota.org/nalichens/). Those data, representing a comprehensive coverage of North American lichens, are presented as a searchable database and as dynamic checklists (e.g., United States and Canada, Mexico, numerous U.S. National Parks,

as well as Arizona and specific regions within the state; http://symbiota.org/nalichens/checklists/) generated from the available herbaria records in real-time. Each checklist is also presented as a "user-friendly", interactive symbiota keying system (http://symbiota.org/nalichens/ident/), which lead the user through a series of progressive, multiple character decisions (based on a hierarchical ranking of character importance relevant to differing taxonomic levels). In addition, a large image database is also linked with species presented on the CNALH website, which include digitized photographs of herbarium specimens (including types) as well as from the field (including the extensive collection of Stephen Sharnoff; see Brodo et al. 2001).

# RESULTS, DISCUSSION, AND CONCLUSIONS

Records for nine hundred sixty-nine species of lichenized fungi (and 13 additional varieties or subspecies) from 54 families (and 3 additional groups of uncertain taxonomic position), 17 orders (and 4 additional groups of uncertain taxonomic position), and 5 classes (all of the classes known to contain lichenforming fungi), within the Ascomycota, are reported here. By far, the Lecanoromycetes represents the single largest group (~84% of all lichen species in the flora) of lichens at the class level, and this class also includes the most species-rich lichen families; the Parmeliaceae (~15% of all lichen species), the Lecanoraceae (~11% of all lichen species), and the *Physciaceae* (at 7.9% of all lichen species). Interestingly, the second largest class level group (Eurotiomycetes) represents only about 9% of all lichen species in the flora, yet it contains the fourth most species-rich lichen family (Verrucariaceae at 7.7% of all lichen species). When parsed by growth forms, crustose lichens (including subgroups such as endolithic, peltate, placodioid, and squamulose lichens) comprise the largest group, representing approximately 60% of all lichens in the flora, followed by foliose lichens (~30%), and then fruticose lichens (~10%). Two additional growth forms, leprose and calicioid, each account for roughly 1% of the flora.

Arizona has an exceptional variety of biomes which has resulted in an extensive diversity of macrofungi (see Bates 2006); likewise, the diversity of lichens present in the state is also considerably rich; representing approximately half of species recorded for the Lichen Flora of the Greater Sonoran Desert Region and nearly 20% of the known lichen flora of North America. The list of Arizona lichen species included here is noticeably larger, over a two-fold increase, than that presented in the original catalog (Nash and Johnsen 1975). This increased understanding of the Arizona lichen flora is the product of the tremendous effort and rigorous scientific study that was inherit in monographing the flora of the Greater Sonoran Desert Region, which resulted in the publication of over 400 taxa new to science and included the discovery of 2 novel genera (see the preface of Nash et al. 2007). These figures highlight the value of concentrated and continued work in monography as well as alpha-taxonomy, and point to the fact that, even for a "well studied" area such as the United States, a comprehensive understanding of the Earth's fungal biota (for both lichenized and non-lichenized groups) is far from being achieved.



Arizona Lichens Figure 1. Diversity of growth forms of lichenized fungi (lichens). (A) *Umbilicaria americana* (foliose/umbilicate, on rock); (B) *Aspicilia hispida* (fruticose, vagrant over soil); (C) *Umbilicaria phaea* (foliose/umbilicate, on rock); (D) *Flavoparmelia baltimorensis* (foliose, on rock); (E) *Usnea intermedia* (fruticose, on bark); (F) *Phaeophyscia hispidula* (small-foliose, on rock); (G) *Caloplaca brouardii* (crustose, on rock); (H) *Psora cerebriformis* (crustose/squamulose, on soil); (I) *Acarospora stapfiana* (crustose, parasitic on *Caloplaca* sp.) – (Photos C, F, G by F. Bungartz; A, B, D, E, H by S.T. Bates; I by J. Siminitus).

# A REVISED CATALOG OF ARIZONA LICHENS

□ FUNGI
• ASCOMYCOTA
† ARTHONIOMYCETES
ARTHONIOMYCETIDAE
ARTHONIALES

#### Arthoniaceae

Arthonia patellulata Nyl. [3]

#### Chrysothricaceae

Chrysothrix candelaris (L.) J.R. Laundon [2]

# † DOTHIDEOMYCETES DOTHIDEOMYCETIDAE INCERTAE SEDIS

#### Mycoporaceae

Mycoporum antecellens (Nyl.) R.C. Harris [1]\*

#### INCERTAE SEDIS INCERTAE SEDIS

#### Protothelenellaceae

Protothelenella sphinctrinoidella (Nyl.) H. Mayrhofer & Poelt [1]

#### Thelenellaceae

Thelenella inductula (Nyl.) H. Mayrhofer [1] Thelenella muscorum (Fr.) Vain. var. muscorum [1]\*

# PLEOSPOROMYCETIDAE PLEOSPORALES

#### Arthopyreniaceae

Arthopyrenia punctiformis (Schrank) A. Massal. [1] Arthopyrenia rhyponta (Ach.) A. Massal. [1] Leptorhaphis atomaria (Ach.) Szatala [1]

### † EUROTIOMYCETES CHAETOTHYRIOMYCETIDAE PYRENULALES

# Monoblastiaceae

Anisomeridium biforme (Borrer) R.C. Harris [1]

# VERRUCARIALES

# Incertae sedis

Staurothele areolata (Ach.) Lettau [1]
Staurothele clopimoides (Bagl. & Carestia) J. Steiner [1]
Staurothele drummondii (Tuck.) Tuck. [1]
Staurothele effigurata J.W. Thomson [1]
Staurothele elenkinii Oxner [1]
Staurothele lecideoides B. de Lesd. [1]
Staurothele monicae (Zahlbr.) Wetmore [1]
Staurothele polygonia B. de Lesd. [1]
Staurothele verruculosa J.W. Thomson [1]

Agonimia tristicula (Nyl.) Zahlbr. [1]

### Verrucariaceae

Bagliettoa baldensis (A. Massal.) Vězda [3]
Bagliettoa calciseda (DC.) Gueidan & Cl. Roux [3]\*
Catapyrenium psoromoides (Borrer) R. Sant. [1]
Catapyrenium squamellum (Nyl.) J.W. Thomson [1]
Dermatocarpon americanum Vain. [2]
Dermatocarpon bachmannii Anders [2]
Dermatocarpon leptophyllodes (Nyl.) Zahlbr. [2]
Dermatocarpon luridum (With.) J.R. Laundon [2]

Dermatocarpon moulinsii (Mont.) Zahlbr. [2]

Dermatocarpon polyphyllizum (Nyl.) Blomb. & Forssell [2]

Dermatocarpon reticulatum H. Magn. [2] Dermatocarpon schaechtelinii Werner [2] Dermatocarpon taminium Heiðm. [2]

Dermatocarpon tenue (Müll. Arg.) Heiðm. [2]

Digitothyrea polyglossa (Nyl.) P.P. Moreno & Egea [1]

Endocarpon loscosii Müll. Arg. [1] Endocarpon pallidulum (Nyl.) Nyl. [1] Endocarpon petrolepideum (Nyl.) Nyl. [1]

Endocarpon pusillum Hedw. [1]

Endocarpon schisticola (B. de Lesd.) Servít [1] Normandina pulchella (Borrer) Nyl. [2] Placidium acarosporoides (Zahlbr.) Breuss [1] Placidium andicola (Breuss) Breuss [1]

Placidium arboreum (Schwein. ex E. Michener)

Lendemer [1]\*

Placidium chilense (Räsänen) Breuss [1]
Placidium fingens (Breuss) Breuss [1]
Placidium lachneum (Ach.) B. de Lesd. [1]
Placidium lacinulatum var. atrans Breuss [1]
Placidium lacinulatum var. erythrostratum Breuss [1]
Placidium lacinulatum (Ach.) Breuss var. lacinulatum

Placidium lacinulatum (Ach.) Breuss var. lacinulatum [1] Placidium michelii A. Massal. [1]

Placidium pilosellum (Breuss) Breuss [1] Placidium rufescens (Ach.) A. Massal. [1] Placidium squamulosum (Ach.) Breuss [1]

Placidium umbrinum (Breuss) M. Prieto & Breuss [1]\* Placopyrenium caeruleopulvinum (J.W. Thomson)

Breuss [1]

Placopyrenium lecideoides (A. Massal.) Gueidan & Cl. Roux [3]\*

Placopyrenium stanfordii (Herre) K. Knudsen [1]\*

Verrucaria alutacea Wallr. [3]

Verrucaria americana (B. de Lesd.) Breuss [3]

Verrucaria amylacea Hepp [3]

Verrucaria beltraminiana (A. Massal.) Trevis. [3]

Verrucaria bernaicensis Malbr. [3] Verrucaria bernardinensis Breuss [3] Verrucaria calkinsiana Servít [3] Verrucaria cetera Breuss [3]

Verrucaria compacta (A. Massal.) Jatta [3] Verrucaria confluens A. Massal. [3] Verrucaria dolosa Hepp [3]

Verrucaria elaeina Borrer [3] Verrucaria endocarpoides Servít [3] Verrucaria falcata Breuss [3]

Verrucaria funckii (Spreng.) Zahlbr. [3] Verrucaria furfuracea (B. de Lesd.) Breuss [3]

Verrucaria fusca Pers. [3]

Verrucaria fuscoatroides Servít [3] Verrucaria glaucovirens Grummann [3] Verrucaria hydrela Ach. [3] Verrucaria incrassata Breuss [3]

Verrucaria inficiens Breuss [3] Verrucaria inornata Servít [3]

Verrucaria macrostoma Dufour ex DC. [3]

Verrucaria maculicarpa Breuss [3]

Verrucaria margacea (Wahlenb.) Wahlenb. [3]

Verrucaria minor Breuss [3] Verrucaria muralis Ach. [3]

Verrucaria murorum (Arnold) Lindau [3]

Verrucaria nigrescens Pers. [3] Verrucaria nigrofusca Servít [3] Verrucaria onegensis Vain. [3] Verrucaria papillosa Ach. [3]

Verrucaria prosoplectenchymatica Servít [3]

Verrucaria quercina Breuss [3] Verrucaria rubrocincta Breuss [3] Verrucaria rupicola (B. de Lesd.) Breuss [3] Verrucaria schindleri Servít [3] Verrucaria sphaerospora Anzi [3] Verrucaria trabicola Arnold [3] Verrucaria viridula (Schrad.) Ach. [3]

# † LECANOROMYCETES ACAROSPOROMYCETIDAE ACAROSPORALES

Acarosporaceae Acarospora affinis K. Knudsen [3] Acarospora badiofusca (Nyl.) Th. Fr. [3] Acarospora brouardii B. de Lesd. [3] Acarospora calcarea K. Knudsen [3] Acarospora chrysops (Tuck.) H. Magn. [3] Acarospora contigua H. Magn. [3] Acarospora dispersa H. Magn. [3] Acarospora erythrophora H. Magn. [3] Acarospora fuscata (Nyl.) Arnold [3] Acarospora glaucocarpa (Ach.) Körb. [3] Acarospora heufleriana Körb. [3] Acarospora interspersa H. Magn. [3] Acarospora macrospora (Hepp) A. Massal. ex Bagl. [3] Acarospora nevadensis H. Magn. [3] Acarospora nodulosa (Dufour) Hue [3] Acaospora novomexicana H. Magn. [3] Acarospora obnubila H. Magn. [3] Acarospora obpallens (Nyl.) Zahlbr. [3] Acarospora oligospora (Nyl.) Arnold [3] Acarospora peliocypha (Wahlenb.) Th. Fr. [3] Acarospora rhabarbarina Hue [3] Acarospora rouxii K. Knudsen, Elix & Reeb [3] Acarospora rosulata (Th. Fr.) H. Magn. [3]\* Acarospora scabrida Hedl. ex H. Magn. [3] Acarospora schleicheri (Ach.) A. Massal. [3] Acarospora scotica Hue [3] Acarospora socialis H. Magn. [3] Acarospora stapfiana (Müll. Arg.) Hue [3] Acarospora strigata (Nyl.) Jatta [3] Acarospora thamnina (Tuck.) Herre [3] Acarospora tuckerae K. Knudsen [3] Acarospora veronensis A. Massal. [3] Glypholecia scabra (Pers.) Müll. Arg. [1] Pleopsidium flavum (Bellardi) Körb. [3] Polysporina gyrocarpa (H. Magn.) N.S. Golubk. [3]\* Polysporina simplex (Davies) Vězda [3] Polysporina subfuscescens (Nyl.) K. Knudsen & Kocourk. [3]\* Polysporina urceolata (Anzi) Brodo [3] Sarcogyne clavus (DC.) Kremp. [3] Sarcogyne dakotensis H. Magn. [3] Sarcogyne desolata (H. Magn.) K. Knudsen & S. Standl. [3] Sarcogyne privigna (Ach.) A. Massal. [3]

### INCERTAE SEDIS **CANDELARIALES**

Sarcogyne regularis Körb. [3]

Sarcogyne similis H. Magn. [3]

# Candelariaceae

Candelaria concolor (Dicks.) Stein [1] Candelaria fibrosa (Fr.) Müll. Arg. [1] Candelaria pacifica M. Westb. & Arup [4] Candelariella antennaria Räsänen [2] Candelariella aurella (Hoffm.) Zahlbr. [2] Candelariella citrina B. de Lesd. [2] Candelariella complanata M. Westb. [2] Candelariella deppeanae M. Westb. [2] Candelariella efflorescens R.C. Harris & W.R. Buck [2] Candelariella kansuensis H. Magn. [3] Candelariella lutella (Vain.) Räsänen [2] Candelariella rosulans (Müll. Arg.) Zahlbr. [2] Candelariella subdeflexa (Nyl.) Lettau [2] Candelariella vitellina (Hoffm.) Müll. Arg. [2] Candelariella xanthostigma (Ach.) Lettau [2] Candelina mexicana (de Lesd.) Poelt [1] Candelina submexicana (de Lesd.) Poelt [1] Placomaronea mendozae (Räsänen) M. Westb. [2]

### INCERTAE SEDIS

Pycnora praestabilis (Nyl.) Hafellner [1] Pycnora sorophora (Vain.) Hafellner [1]

#### **UMBILICARIALES**

#### **Ophioparmaceae**

Hypocenomyce anthracophila (Nyl.) P. James & Gotth. Schneid. [1] Hypocenomyce castaneocinerea (Räsänen) Timdal [1] Hypocenomyce friesii (Ach.) P. James & Gotth. Schneid. [1] Hypocenomyce oligospora Timdal [1] Hypocenomyce scalaris (Ach. ex Lilj.) M. Choisy [1] Hypocenomyce sierrae Timdal [1]

#### Umbilicariaceae

Lasallia papulosa (Ach.) Llano [2] Lasallia pennsylvanica (Ach.) Llano [2] Umbilicaria americana Poelt & T.H. Nash [2] Umbilicaria cinereorufescens (Schaer.) Frey [2] Umbilicaria cylindrica (L.) Delise ex Duby [2] Umbilicaria decussata (Vill.) Zahlbr. [2] Umbilicaria deusta (L.) Baumg. [2] Umbilicaria hyperborea (Ach.) Hoffm. [2] Umbilicaria krascheninnikovii (Savicz) Zahlbr. [2] Umbilicaria muehlenbergii (Ach.) Tuck. [2] Umbilicaria nylanderiana (Zahlbr.) H. Magn. [2] Umbilicaria phaea Tuck. [2] Umbilicaria polyphylla (L.) Baumg. [2] Umbilicaria subglabra (Nyl.) Harm. [2] Umbilicaria torrefacta (Lightf.) Schrad. [2] Umbilicaria vellea (L.) Ach. [2] Umbilicaria virginis Schrad. [2]

# LECANOROMYCETIDAE INCERTAE SEDIS

#### Coniocybaceae

Chaenotheca chrysocephala (Turner ex Ach.) Th. Fr. [2] Chaenotheca furfuracea (L.) Tibell [2]

#### LECANORALES

# Aphanopsidaceae

Steinia geophana (Nyl.) Stein [2]

#### Biatorellaceae

Piccolia ochrophora (Nyl.) Hafellner [3]\*

#### Cladoniaceae

Cladonia acuminata (Ach.) Norrl. [1] Cladonia bacilliformis (Nyl.) Glück [1] Cladonia borealis S. Stenroos [1]

Cladonia cariosa (Ach.) Spreng. [1] Lecanora caesiorubella Ach. [2] Cladonia carneola (Fr.) Fr. [1] Lecanora campestris (Schaer.) Hue [2] Cladonia cenotea (Ach.) Schaer. [1] Lecanora carpinea (L.) Vain. [2] Cladonia cervicornis (Ach.) Flot. subsp. cervicornis [1] Lecanora cavicola Creveld [2] Cladonia chlorophaea (Flörke ex Sommerf.) Spreng. [1] Lecanora cenisia Ach. [2] Lecanora chlarotera Nyl. [2] Cladonia coniocraea (Flörke) Spreng. [1] Cladonia deformis (L.) Hoffm. [1] Lecanora circumborealis Brodo & Vitik. [2] Cladonia fimbriata (L.) Fr. [1] Lecanora comoduensis T.H. Nash & Hertel [2] Cladonia furcata (Huds.) Schrad. [1] Lecanora coniferarum Printzen [2] Cladonia gracilis (Ach.) Ahti subsp. turbinata [1] Lecanora crenulata (Dicks.) Hook. [2] Cladonia humilis (With.) J.R. Laundon [1] Lecanora densa (Śliwa & Wetmore) Printzen [2] Cladonia jaliscana Ahti & Guzm.-Dáv. [1] Lecanora dispersa (Pers.) Röhl. [2] Cladonia macilenta Hoffm. [1] Lecanora flowersiana H. Magn. [2] Cladonia macrophyllodes Nyl. [1] Lecanora gangaleoides Nyl. [2] Cladonia multiformis G. Merr. [1] Lecanora garovaglioi subsp. cascadensis (H. Magn.) Cladonia ochrochlora Flörke [1] B.D. Ryan & T.H. Nash [2] Lecanora garovaglioi (Körb.) Zahlbr. subsp. garovaglioi [2] Cladonia parasitica (Hoffm.) Hoffm. [1] Cladonia pleurota (Flörke) Schaer. [1] Lecanora geiserae B.D. Ryan [2] Cladonia pocillum (Ach.) O.J. Rich. [1] Lecanora hagenii (Ach.) Ach. [2] Cladonia pulvinella S. Hammer [1] Lecanora horiza (Ach.) Linds. [2] Cladonia pyxidata (L.) Hoffm. [1] Lecanora hybocarpa (Tuck.) Brodo [2] Cladonia subfimbriata Ahti [1] Lecanora aff. hypoptoides (Nyl.) Nyl. [2] Cladonia subradiata (Vain.) Sandst. [1] Lecanora impudens Degel. [2] Cladonia subulata (L.) Weber ex F.H. Wigg. [1] Lecanora intricata (Ach.) Ach. [2] Cladonia sulphurina (Michx.) Fr. [1] Lecanora juniperina Śliwa [2] Lecanora kofae B.D. Ryan & T.H. Nash [2] Cladonia symphycarpa (Flörke) Fr. [1] Lecanora laatokkensis (Räsänen) Poelt [2] Gynsonlacaceae Lecanora marginata (Schaer.) Hertel & Rambold [2] Gypsoplaca macrophylla (Zahlbr.) Timdal [3] Lecanora mazatzalensis B.D. Ryan & T.H. Nash [2] Lecanora melaena (Hedl.) Fink [2] Haematommataceae Lecanora meridionalis H. Magn. [2] Haematomma fenzlianum A. Massal. [2] Lecanora mughicola Nyl. [2] Lecanora muralis var. brunneola (Mereschk.) B.D. Ryan & Incertae sedis T.H. Nash [2] Botryolepraria lesdainii (Hue) Canals, Hern.-Mar., Lecanora muralis (Schreb.) Rabenh. var. muralis [2] Gómez-Bolea & Llimona [3] Lecanora nashii B.D. Ryan [2] Lecania arizonica B.D. Ryan & van den Boom [2] Lecanora neodegelii B.D. Ryan & T.H. Nash [2] Lecania coeruleorubella (Mudd) M. Mayrhofer [2] Lecanora novomexicana H. Magn. [2] Lecanora opiniconensis Brodo [2] Lecania polycycla (Anzi) Lettau [2] Leprocaulon albicans Nyl. [1] Lecanora oreinoides (Körb.) Hertel & Rambold [2] Leprocaulon gracilescens (Nyl.) I.M. Lamb & Lecanora orizabana Vain. [2] A.M. Ward [1] Lecanora pacifica Tuck. [2] Lecanora peltastictoides Hasse [2]\* Leprocaulon microscopicum (Vill.) Gams ex D. Hawksw. [1] Lecanora phaedrophthalma var. christoi (W.A. Weber) Myxobilimbia sabuletorum (Schreb.) Hafellner [2] B.D. Ryan [2] Psilolechia lucida (Ach.) M. Choisy [3] Lecanora phaedrophthalma Poelt var. phaedrophthalma [2] Scoliciosporum intrusum (Th. Fr.) Hafellner [2]\* Lecanora plumosa Müll. Arg. [2] Scoliciosporum umbrinum (Ach.) Arnold [2] Lecanora polytropa (Ehrh.) Rabenh. [2] Tremolecia atrata (Ach.) Hertel [2] Lecanora pringlei subsp. brandegei (Tuck.) B.D. Ryan [2] Vahliella californica (Tuck.) P.M. Jørg. [1]\* Lecanora pseudistera Nyl. [2] Vahliella hookerioides (P.M. Jørg.) P.M. Jørg. [1]\* Lecanora pulicaris (Pers.) Ach. [2] Vahliella leucophaea (Vahl) P.M. Jørg. [1]\* Lecanora rupicola (L.) Zahlbr. [2] Lecanora saligna (Schrad.) Zahlbr. [2] Lecanora semipallida H. Magn. [2]\* Carbonea latypizodes (Nyl.) Knoph & Rambold [2] Lecanora subcavicola B.D. Ryan [2] Carbonea vorticosa (Flörke) Hertel [2] Lecanora subimmergens Vain. [2] Clauzadeana macula (Taylor) Coppins & Rambold [2] Lecanora subintricata (Nyl.) Th. Fr. [2] Lecanora albella (Pers.) Ach. [2] Lecanora subrugosa Nyl. [2] Lecanora albellula (Nyl.) Th. Fr. [2] Lecanora swartzii (Ach.) Ach. [2] Lecanora allophana (Ach.) Nyl. [2] Lecanora symmicta (Ach.) Ach. [2] Lecanora arenisaxicola B.D. Ryan & T.H. Nash [2] Lecanora thallophila H. Magn. [2] Lecanora umbrosa Degel. [2] Lecanora argentata (Ach.) Malme [2] Lecanora argentea Oxner & Volkova [2] Lecanora valesiaca var. sibirica Poelt [2] Lecanora argopholis (Ach.) Ach. [2] Lecanora valesiaca (Müll. Arg.) Stizenb. var. valesiaca [2] Lecanora bicincta Ramond [2] Lecanora viridiflava B. de Lesd. [2] Lecanora bipruinosa Fink ex J. Hedrick [2] Lecanora weberi B.D. Ryan [2] Lecanora boligera (Th. Fr.) Hedl. [2] Lecanora wetmorei Śliwa [2]

Lecidella anomaloides (A. Massal.) Hertel & H. Kilias [2]

Lecidella asema (Nyl.) Knoph & Hertel [2]

Lecanora brodoana Lumbsch & T.H. Nash [2]

Lecanora cadubriae (A. Massal.) Hedl. [2]

Lecidella carpathica Körb. [2] Melanelixia subargentifera (Nyl.) O. Blanco, A. Crespo, Lecidella chiricahuana Knoph & Leuckert [2] Divakar, Essl., D. Hawksw. & Lumbsch [1]\* Lecidella effugiens (Nilson) Knoph & Hertel [2] Melanelixia villosella (Essl.) O. Blanco, A. Crespo, Divakar, Lecidella elaeochroma (Ach.) M. Choisy [2] Essl., D. Hawksw. & Lumbsch [1]\* Lecidella euphorea (Flörke) Hertel [2] Melanohalea elegantula (Zahlbr.) O. Blanco, A. Crespo, Lecidella granulosula (Nyl.) Knoph & Leuckert [2] Divakar, Essl., D. Hawksw. & Lumbsch [1] Lecidella latypiza (Nyl.) M. Choisy [2] Melanohalea exasperatula (Nyl.) O. Blanco, A. Crespo, Lecidella nashiana Knoph & Leuckert [2] Divakar, Essl., D. Hawksw. & Lumbsch [1]\* Lecidella patavina (A. Massal.) Knoph & Leuckert [2] Melanohalea subolivacea (Nyl.) O. Blanco, A. Crespo, Lecidella stigmatea (Ach.) Hertel & Leuckert [2] Divakar, Essl., D. Hawksw. & Lumbsch [1]\* Lecidella tumidula (A. Massal.) Knoph & Leuckert [2] Myelochroa aurulenta (Tuck.) Elix & Hale [1] Lecidella viridans (Flot.) Körb. [2] Neofuscelia ahtii (Essl.) Essl. [1] Lecidella wulfenii (Ach.) Körb. [2] Neofuscelia atticoides (Essl.) Essl. [1] Miriquidica garovaglioi (Schaer.) Hertel & Rambold [2] Neofuscelia brunella (Essl.) Essl. [1] Psorinia conglomerata (Ach.) Gotth. Schneid. [1] Neofuscelia chiricahuensis (R.A. Anderson & W.A. Weber) Ramboldia elabens (Fr.) Kantvilas & Elix [2] Essl. [1] Rhizoplaca chrysoleuca (Sm.) Zopf [1] Neofuscelia infrapallida (Essl.) Essl. [1] Rhizoplaca melanophthalma (DC.) Leuckert [1] Neofuscelia occidentalis (Essl.) Essl. [1] Rhizoplaca peltata (Ramond) Leuckert & Poelt [1] Omphalora arizonica (Tuck. ex Willey) T.H. Nash & Rhizoplaca subdiscrepans (Nyl.) R. Sant. [1] Hafellner [1] Strangospora microhaema (Norman) R.A. Anderson [3] Parmelia fraudans (Nyl.) Nyl. [1] Strangospora moriformis (Ach.) Stein [3] Parmelia glabra (Schaer.) Nyl. [1] Parmelia saxatilis L. (Ach.) [1] Parmelia sulcata Taylor [1] Parmeliaceae Ahtiana sphaerosporella (Müll. Arg.) Goward [1] Parmelinopsis horrescens (Taylor) Elix & Hale [1] Arctoparmelia centrifuga (Oxner) Hale [1] Parmelinopsis minarum (Vain.) Elix & Hale [1] Brodoa oroarctica (Krog) Goward [1] Parmeliopsis ambigua (Wulfen) Nyl. [1] Bryoria chalybeiformis (L.) Brodo & D. Hawksw. [1] Parmeliopsis hyperopta (Ach.) Vain. [1] Bryoria furcellata (Fr.) Brodo & D. Hawksw. [1] Parmotrema chinense (Osbeck) Hale & Ahti [1] Bryoria fuscescens (Gyeln.) Brodo & D. Hawksw. [1] Parmotrema crinitum (Ach.) M. Choisy [1] Bryoria lanestris (Ach.) Brodo & D. Hawksw. [1] Parmotrema eurysacum (Hue) Hale [1] Bryoria simplicior (Vain.) Brodo & D. Hawksw. [1] Parmotrema hababianum (Gyeln.) Hale [1] Canoparmelia crozalsiana (B. de Lesd.) Elix & Hale [1] Parmotrema mordenii (Hale) Hale [1] Canoparmelia texana (Tuck.) Elix & Hale [1] Parmotrema stuppeum (Taylor) Hale [1] Cetraria ericetorum subsp. reticulata (Räsänen) Parmotrema subtinctorium (Zahl.) Hale [1]\* Protoparmelia atriseda (Fr.) R. Sant. & V. Wirth [2] Kärnefelt [1] Evernia divaricata (L.) Ach. [1] Protoparmelia badia (Hoffm.) Hafellner [2] Flavoparmelia baltimorensis (Gyeln. & Fóriss) Hale [1] Protoparmelia cupreobadia (Nyl.) Poelt [2] Pseudephebe minuscula (Arnold) Brodo & D. Hawksw. [1] Flavoparmelia caperata (L.) Hale [1] Flavopunctelia darrowii (J.W. Thomson) Hale [2] Pseudevernia intensa (Nyl.) Hale & W.L. Culb. [1] Punctelia graminicola (B. de Lesd.) Egan [2] Flavopunctelia flaventior (Stirt.) Hale [2] Flavopunctelia praesignis (Nyl.) Hale [2] Punctelia hypoleucites (Nyl.) Krog [2] Punctelia perreticulata (Räsänen) G. Wilh. & Ladd [2] Flavopunctelia soredica (Nyl.) Hale [2] Hypogymnia austerodes (Nyl.) Räsänen [1] Punctelia rudecta (Ach.) Krog [2] Hypogymnia bitteri (Lynge) Ahti [1] Punctelia stictica (Delise ex Duby) Krog [2] Hypogymnia farinacea Zopf [1] Rimelia cetrata (Ach.) Hale & A. Fletcher [1] Hypogymnia physodes (L.) Nyl. [1] Rimelia reticulata (Taylor) Hale & A. Fletcher [1] Hypotrachyna dactylifera (Vain.) Hale [1] Rimelia simulans (Hale) Hale & A. Fletcher [1] Hypotrachyna laevigata (Sm.) Hale [1] Tuckermanella arizonica Essl. [2] Tuckermanella coralligera (W.A. Weber) Essl. [2] Hypotrachyna meridensis Hale & López-Fig. [1] Tuckermanella fendleri (Nyl.) Essl. [2] Hypotrachyna pulvinata (Fée) Hale [1] Hypotrachyna punoensis Kurok. & K.H. Moon [1] Tuckermanella weberi (Essl.) Essl. [2] Usnea amblyoclada (Müll. Arg.) Zahlbr. [3] Hypotrachyna pustulifera (Hale) Skorepa [1] Hypotrachyna revoluta (Flörke) Hale [1] Usnea cavernosa Tuck. [3] Hypotrachyna subsaxatilis (B. de Lesd.) Hale [1] Usnea ceratina Ach. [3] Imshaugia aleurites (Ach.) S.L.F. Mey. [1] Usnea cirrosa Motyka [3] Imshaugia placorodia (Ach.) S.L.F. Mey. [1] Usnea cornuta Körb. subsp. cornuta [3] Letharia columbiana (Nutt.) J.W. Thomson [1] Usnea diplotypus Vain. [3] Letharia vulpina (L.) Hue [1] Usnea halei P. Clerc [3] Melanelia disjuncta (Erichsen) Essl. [1] Usnea hirta (L.) Weber ex F.H. Wigg. subsp. hirta [3] Melanelia panniformis (Nyl.) Essl. [1] Usnea intermedia Jatta [3] Melanelia tominii (Oxner) Essl. [1] Usnea lapponica Vain. [3] Usnea myrmaiacaina P. Clerc [3] Melanelixia albertana (Ahti) O. Blanco, A. Crespo, Divakar, Essl., D. Hawksw. & Lumbsch [1]\* Usnea parvula Motyka [3] Melanelixia fuliginosa (Fr. ex Duby) O. Blanco, A. Crespo, Usnea praetervisa (Asahina) P. Clerc [3] Divakar, Essl., D. Hawksw. & Lumbsch [1]\* Usnea scabrata Nyl. [3] Melanelixia glabroides (Essl.) O. Blanco, A. Crespo, Usnea subfloridana Stirt. [3] Divakar, Essl., D. Hawksw. & Lumbsch [1]\* Usnea substerilis Motyka [3]

Vulpicida pinastri (Scop.) J.-E. Mattsson [1]

Xanthoparmelia ahtii (Essl.) O. Blanco, A. Crespo, Elix, D. Hawksw. & Lumbsch [1]\* Xanthoparmelia ajoensis (T.H. Nash) Egan [2] Xanthoparmelia amableana (Gyeln.) Hale [2] Xanthoparmelia atticoides (Essl.) O. Blanco, A. Crespo, Elix, D. Hawksw. & Lumbsch [1] Xanthoparmelia australasica D.J. Galloway [2] Xanthoparmelia brunella O. Blanco, A. Crespo, Elix, D. Hawksw. & Lumbsch [1] Xanthoparmelia californica Hale [2] Xanthoparmelia chlorochroa (Tuck.) Hale [2] Xanthoparmelia coloradoensis (Gyeln.) Hale [2] Xanthoparmelia conspersa (Ehrh. ex Ach.) Hale [2] Xanthoparmelia cumberlandia (Gyeln.) Hale [2] Xanthoparmelia dierythra (Hale) Hale [2] Xanthoparmelia digitiformis (Elix & P.M. Armstr.) Filson [2] Xanthoparmelia dissensa (T.H. Nash) Egan [2] Xanthoparmelia eganii Elix & T.H. Nash [2] Xanthoparmelia huachucensis (T.H. Nash) Egan [2] Xanthoparmelia hypomelaena (Hale) Hale [2] Xanthoparmelia incerta (Kurok. & Filson) Elix & J. Johnst. [2] Xanthoparmelia isidiigera (Müll. Arg.) Elix & J. Johnst. [2] Xanthoparmelia lavicola (Gyeln.) Hale [2] Xanthoparmelia lineola (E.C. Berry) Hale [2] Xanthoparmelia lobulatella T.H. Nash & Elix [2] Xanthoparmelia maricopensis T.H. Nash & Elix [2] Xanthoparmelia mexicana (Gyeln.) Hale [2] Xanthoparmelia montanensis Hale [2] Xanthoparmelia monticola (J.P. Dey) Hale [2] Xanthoparmelia neocongensis (Hale) Hale [2] Xanthoparmelia neoconspersa (Gyeln.) Hale [2] Xanthoparmelia neorimalis (Elix & P.M. Armstr.) Elix & T.H. Nash [2] Xanthoparmelia neowyomingica Hale [2] Xanthoparmelia nigrolavicola T.H. Nash & Elix [2] Xanthoparmelia nigropsoromifera (T.H. Nash) Egan [2] Xanthoparmelia nigroweberi T.H. Nash & Elix [2] Xanthoparmelia novomexicana (Gyeln.) Hale [2] Xanthoparmelia oleosa (Elix & P.M. Armstr.) Elix & T.H. Nash [2] Xanthoparmelia planilobata (Gyeln.) Hale [2] Xanthoparmelia plittii (Gyeln.) Hale [2] Xanthoparmelia pseudocongensis Hale [2] Xanthoparmelia psoromifera (Hale) Hale [2] Xanthoparmelia standaertii (Gyeln.) Hale [2] Xanthoparmelia stenophylla (Ach.) Ahti & D. Hawksw. [2] Xanthoparmelia subcumberlandia Elix & T.H. Nash [2] Xanthoparmelia subdecipiens (Vain. ex Lynge) Hale [2] Xanthoparmelia subplittii Hale [2] Xanthoparmelia substenophylloides Hale [2] Xanthoparmelia subtasmanica Elix & T.H. Nash [2] Xanthoparmelia tasmanica (Hook. f. & Taylor) Hale [2] Xanthoparmelia tinctina (Maheu & A. Gillet) Hale [2] Xanthoparmelia tuberculata (Gyeln.) T.H. Nash & Elix [2] Xanthoparmelia tuckeriana Elix & T.H. Nash [2] Xanthoparmelia tucsonensis (T.H. Nash) Egan [2] Xanthoparmelia weberi (Hale) Hale [2] Xanthoparmelia wyomingica (Gyeln.) Hale [2]

#### Pilocarpaceae

Micarea denigrata (Fr.) Hedl. [3] Micarea micrococca (Körb.) Gams ex Coppins [3] Micarea misella (Nyl.) Hedl. [3]

#### Psoraceae

Protoblastenia rupestris (Scop.) J. Steiner [2] Psora cerebriformis W.A. Weber [1]

Psora crenata (Taylor) Reinke [1] Psora decipiens (Hedw.) Hoffm. [1] Psora globifera (Ach.) A. Massal. [1] Psora himalayana (C. Bab.) Timdal [1] Psora icterica (Mont.) Müll. Arg. [1] Psora luridella (Tuck.) Fink [1] Psora nipponica (Zahlbr.) Gotth. Schneid. [1] Psora pseudorussellii Timdal [1] Psora russellii (Tuck.) A. Schneid. [1] Psora tuckermanii R.A. Anderson ex Timdal [1] Psorula rufonigra (Tuck.) Gotth. Schneid. [1]

#### Ramalinaceae

Bacidia bagliettoana (A. Massal. & De Not.) Jatta [2] Bacidia beckhausii Körb. [2] Bacidia circumspecta (Norrl. & Nyl.) Malme [2] Bacidia subincompta (Nyl.) Arnold [2] Bacidia vermifera (Nyl.) Th. Fr. [2] Bacidina "dissecta" S. Ekman ad int. [2] Bacidina egenula (Nyl.) Vězda [2] Biatora chrysantha (Zahlbr.) Printzen [2] Biatora globulosa (Flörke) Fr. [2] Biatora meiocarpa (Nyl.) Arnold [2] Japewia tornoensis (Nyl.) Tønsberg [2] Mycobilimbia carneoalbida (Müll. Arg.) S. Ekman & Printzen [2] Mycobilimbia tetramera (De Not.) Vitik., Ahti, Kuusinen, Lommi & T. Ulvinen [2] Ramalina intermedia Delise ex Nyl. [2] Ramalina pollinaria (Westr.) Ach. [2] Ramalina sinensis Jatta [2]

#### Stereocaulaceae

Hertelidea botryosa (Fr.) Printzen & Kantvilas [2] Lepraria borealis Loht. & Tønsberg [2] Lepraria caesioalba (B. de Lesd.) J.R. Laundon [2] Lepraria elobata Tønsberg [2] Lepraria lecanorica Tønsberg [2] Lepraria lobificans Nyl. [2] Lepraria neglecta (Nyl.) Erichsen [2] Lepraria nivalis J.R. Laundon [2] Lepraria rigidula (B. de Lesd.) Tønsberg [2] Lepraria vouauxii (Hue) R.C. Harris [2] Squamarina lentigera (Weber) Poelt [1] Stereocaulon sasakii Zahlbr. [1]

# **Tephromelataceae**

Calvitimela armeniaca (DC.) Hafellner [2] Tephromela atra (Huds.) Hafellner [2]

#### Vězdaeaceae

Vězdaea rheocarpa Poelt & Dobbeler [2]

#### **LECIDEALES**

#### Lecideaceae

Lecidea atrobrunnea (Ramond) Schaer. subsp. atrobrun Lecidea atrobrunnea subsp. saxosa Hertel & Leuckert [2] Lecidea atrobrunnea subsp. stictica Hertel & Leuckert [2] Lecidea berengeriana (A. Massal.) Nyl. [2] Lecidea cruciaria Tuck. [2] Lecidea diducens Nyl. [2] Lecidea holopolia (Tuck.) Zahlbr. [2] Lecidea laboriosa Müll. Arg. [2] Lecidea leprarioides Tønsberg [2] Lecidea nylanderi (Anzi) Th. Fr. [2] Lecidea promiscens Nyl. [2] Lecidea pseudaglaea Hertel [2]

Lecidea sauteri Körb. [2] Lecidea tessellata Flörke [2] Lecidea trapelioides Printzen [2]

#### Porpidiaceae

Bellemerea alpina (Sommerf.) Clauzade & Cl. Roux [3] Bellemerea sanguinea (Kremp.) Hafellner & Cl. Roux [3] Immersaria athroocarpa (Ach.) Rambold & Pietschm. [2] Porpidia crustulata (Ach.) Hertel & Knoph [2] Porpidia macrocarpa (DC.) Hertel & A.J. Schwab [2] Romjularia lurida (Ach.) Timdal [3]

#### **PELTIGERALES**

#### Coccocarpiaceae

Coccocarpia erythroxyli (Spreng.) Swinscow & Krog [1] Coccocarpia palmicola (Spreng.) Arv. & D.J. Galloway [1] Spilonema revertens Nvl. [1]

#### Collemataceae

Collema callopismum A. Massal. [2] Collema coccophorum Tuck. [2] Collema conglomeratum Hoffm. [2] Collema crispum (Huds.) Weber ex F.H. Wigg. [2] Collema cristatum (L.) Weber ex F.H. Wigg. [2] Collema flaccidum (Ach.) Ach. [2] Collema furfuraceum Du Rietz [2] Collema fuscovirens (With.) J.R. Laundon [2] Collema occultatum var. populinum (Th. Fr.) Degel. [2] Collema polycarpon Hoffm. [2] Collema subflaccidum Degel. [2] Collema subnigrescens Degel. [2] Collema subparvum Degel. [2] Collema tenax (Sw.) Ach. [2] Collema texanum Tuck. [2] Collema undulatum var. granulosum Degel. [2] Leptogium arsenei Sierk [2] Leptogium austroamericanum (Malme) C.W. Dodge [2] Leptogium burgessii (L.) Mont. [2] Leptogium burnetii C.W. Dodge [2] Leptogium californicum Tuck. [2] Leptogium chloromelum (Ach.) Nyl. [2] Leptogium coralloideum (Meyen & Flot.) Vain. [2] Leptogium cyanescens (Pers.) Körb. [2] Leptogium "denticulatum" Nyl. Sensu Sierk [2] Leptogium digitatum (A. Massal.) Zahlbr. [2] Leptogium gelatinosum (With.) J.R. Laundon [2] Leptogium hypotrachynum Müll. Arg. [2] Leptogium juniperinum Tuck. [2] Leptogium laceroides B. de Lesd. [2] Leptogium lichenoides (L.) Zahlbr. [2]

#### Leptogium resupinans Nyl. [2] Leptogium rugosum Sierk [2]

A.K. Wallace [2]

Leptogium milligranum Sierk [2]

Leptogium plicatile (Ach.) Leight. [2] Leptogium pseudofurfuraceum P.M. Jørg. &

Leptogium saturninum (Dicks.) Nyl. [2]

Leptogium subaridum P.M. Jørg. & Goward [2]

Leptogium papillosum (B. de Lesd.) C.W. Dodge [2] Leptogium phyllocarpum (Pers.) Mont. [2]

Leptogium subtile (Schrad.) Torss. [2]

Leptogium tenuissimum (Hoffm.) Körb. [2]

#### Lobariaceae

Sticta beauvoisii Delise [2] Sticta fuliginosa (Dicks.) Ach. [2] Sticta leucoblephara (Müll. Arg.) D.J. Galloway [2] Sticta sylvatica (Huds.) Ach. [2]

Sticta xanthotropa (Kremp.) D.J. Galloway [2]

#### Massalongiaceae

Leptochidium albociliatum (Desm.) M. Choisy [1] Massalongia carnosa (Dicks.) Körb. [1]

#### Nephromataceae

Nephroma bellum (Spreng.) Tuck. [1] Nephroma helveticum Ach. [1] Nephroma parile (Ach.) Ach. [1] Nephroma resupinatum (L.) Ach. [1]

#### Pannariaceae

Fuscopannaria mediterranea (Tav.) P.M. Jørg. [1] Fuscopannaria praetermissa (Nyl.) P.M. Jørg. [1] Pannaria conoplea (Pers.) Bory [1] Pannaria subfusca P.M. Jørg. [1] Pannaria tavaresii P.M. Jørg. [1] Parmeliella triptophylla (Ach.) Müll. Arg. [1] Protopannaria pezizoides (Weber ex F.H. Wigg.) P.M. Jørg. & S. Ekman [1] Psoroma hypnorum (Vahl) Gray. [1] Psoroma tenue var. boreale Henssen [1]

#### Peltigeraceae

Peltigera collina (Ach.) Röhl. [2] Peltigera didactyla (With.) J.R. Laundon [2] Peltigera elisabethae Gyeln. [2] Peltigera extenuata (Nyl.) Vain. [2] Peltigera fibrilloides (Gyeln.) Vitik. [2] Peltigera horizontalis (Huds.) Baumg. [2] Peltigera lepidophora (Nyl.) Bitter [2] Peltigera leucophlebia (Nyl.) Gyeln. [2] Peltigera malacea (Ach.) Funck [2] Peltigera membranacea (Ach.) Nyl. [2] Peltigera monticola Vitik. [2] Peltigera neckeri Hepp ex Müll. Arg. [2] Peltigera neopolydactyla (Gyeln.) Gyeln. [2] Peltigera polydactylon (Neck.) Hoffm. [2] Peltigera ponojensis Gyeln. [2] Peltigera praetextata (Flörke ex Sommerf.) Vain. [2] Peltigera rufescens (Weiss) Humb. [2] Peltigera venosa (L.) Hoffm. [2] Solorina spongiosa (Huds.) Anzi [2]

# Placynthiaceae

Koerberia biformis A. Massal. [1] Koerberia sonomensis (Tuck.) Henssen [1] Placynthium asperellum (Ach.) Trevis. [1] Placynthium nigrum (Huds.) Gray [1] Placynthium stenophyllum var.isidiatum Henssen [1] Placynthium subradiatum (Nyl.) Arnold [1]

#### RHIZOCARPALES

#### Catillariaceae

Catillaria chalybeia (Borrer) A. Massal. [3] Catillaria glauconigrans (Tuck.) Hasse [3] Catillaria lenticularis (Ach.) Th. Fr. [3] Catillaria nigroclavata (Nyl.) Schuler [3] Halecania australis Lumbsch [2] Sporastatia testudinea (Ach.) A. Massal. [2] Toninia candida (Weber) Th. Fr. [1] Toninia cinereovirens (Schaer.) A. Massal. [1] Toninia lutosa (Ach.) Timdal [1] Toninia massata (Tuck.) Herre [1] Toninia philippea (Mont.) Timdal [1] Toninia ruginosa (Tuck.) Herre subsp. ruginosa [1] Toninia sculpturata (H. Magn.) Timdal [1]

Toninia sedifolia (Scop.) Timdal [1] Gassicurtia coccinea Fée [3]\* Toninia squalida (Ach.) A. Massal. [1] Pyxine cocoes (Sw.) Nyl. [1] Toninia subdiffracta Timdal [1] Pyxine petricola Nyl. [1] Toninia submexicana B. de Lesd. [1] Pyxine sorediata (Ach.) Mont. [1] Toninia tristis subsp. arizonica Timdal [1] Pyxine subcinerea Stirt. [1] Toninia tristis subsp. asiae-centralis (H. Magn.) Timdal [1] Tetramelas chloroleucus (Körb.) A. Nordin [3]\* Toninia tristis (Th. Fr.) Th. Fr. subsp. tristis [1] Tetramelas triphragmioides (Anzi) A. Nordin & Tibell [3]\* Toninia weberi Timdal [1] Physciaceae Anaptychia elbursiana (Szatala) Poelt [1] Rhizocarpaceae Rhizocarpon arctogenum Gelting [2] Culbersonia nubila (Moberg) Essl. [1] Rhizocarpon badioatrum (Flörke ex Spreng.) Th. Fr. [2] Heterodermia albicans (Pers.) Swinscow & Krog [1] Rhizocarpon dimelaenae Timdal [2] Heterodermia appalachensis (Kurok.) W.L. Culb. [1] Rhizocarpon disporum (Nägeli ex Hepp) Müll. Arg. [2] Heterodermia granulifera (Ach.) W.L. Culb. [1] Rhizocarpon distinctum Th. Fr. [2] Heterodermia hypoleuca (Mühl.) Trevis. [1] Heterodermia japonica (M. Satô) Swinscow & Krog [1] Rhizocarpon effiguratum (Anzi) Th. Fr. [2] Rhizocarpon eupetraeum (Nyl.) Arnold [2] Heterodermia leucomela (L.) Poelt [1] Rhizocarpon geminatum Körb. [2] Heterodermia obscurata (Nyl.) Trevis. [1] Rhizocarpon geographicum (L.) DC. [2] Heterodermia podocarpa (Bél.) D.D. Awasthi [1] Rhizocarpon grande (Flörke ex Flot.) Arnold [2] Heterodermia pseudospeciosa (Kurok.) W.L. Culb. [1] Rhizocarpon macrosporum Räsänen [2] Heterodermia rugulosa (Kurok.) Trass [1] Rhizocarpon cf. obscuratum (Ach.) A. Massal. [2] Heterodermia speciosa (Wulfen) Trevis. [1] Rhizocarpon polycarpum (Hepp) Th. Fr. [2] Heterodermia tropica (Kurok.) Kurok. [1] Rhizocarpon riparium Räsänen [2] Hyperphyscia adglutinata (Flörke) H. Mayrhofer & Poelt [1] Rhizocarpon simillimum (Anzi) Lettau [2] Phaeophyscia adiastola (Essl.) Essl. [2] Rhizocarpon superficiale (Schaer.) Malme [2] Phaeophyscia ciliata (Hoffm.) Moberg [2] Rhizocarpon viridiatrum (Wulfen) Körb. [2] Phaeophyscia decolor (Kashiw.) Essl. [2] Phaeophyscia endococcinoides (Poelt) Essl. [2] TELOSCHISTALES Phaeophyscia hirsuta (Mereschk.) Moberg [2] Phaeophyscia hispidula (Ach.) Moberg [2] Caliciaceae Phaeophyscia insignis (Mereschk.) Moberg [2] Amandinea dakotensis (H. Magn.) P.F. May & Sheard [3]\* Phaeophyscia kairamoi (Vain.) Moberg [2] Amandinea polyspora (Willey) E. Lay & P.F. May [3]\* Phaeophyscia nigricans (Flörke) Moberg [2] Amandinea punctata (Hoffm.) Coppins & Scheid. [3]\* Phaeophyscia orbicularis (Neck.) Moberg [2] Buellia aethalea (Ach.) Th. Fr. [3] Phaeophyscia pusilloides (Zahlbr.) Essl. [2] Buellia badia (Fr.) A. Massal. [3] Phaeophyscia sciastra (Ach.) Moberg [2] Buellia cedricola Werner [3] Physcia adscendens (Th. Fr.) H. Olivier [1] Physcia aipolia (Ehrh. ex Humb.) Fürnr. [1] Buellia disciformis (Fr.) Mudd [3] Buellia dispersa A. Massal. [3] Physcia biziana (A. Massal.) Zahlbr. [1] Buellia eganii Bungartz [3] Physcia caesia (Hoffm.) Hampe ex Fürnr. [1] Buellia erubescens Arnold [3] Physcia convexa Müll. Arg. [1] Buellia lacteoidea B. de Lesd. [3] Physcia dimidiata (Arnold) Nyl. [1] Buellia mamillana (Tuck.) W.A. Weber [3] Physcia dubia (Hoffm.) Lettau [1] Buellia mexicana J. Steiner [3] Physcia erumpens Moberg [1] Physcia halei J.W. Thomson [1] Buellia nashii Bungartz [3] Buellia navajoensis Bungartz [3] Physcia nashii Moberg [1] Physcia phaea (Tuck.) J.W. Thomson [1] Buellia pullata Tuck. [3] Buellia sequax (Nyl.) Zahlbr. [3] Physcia poncinsii Hue [1] Buellia spuria (Schaer.) Anzi [3] Physcia sinuosa Moberg [1] Buellia subaethalea B. de Lesd. [3] Physcia stellaris (L.) Nyl. [1] Buellia subdispersa Mig. [3] Physcia tribacia (Ach.) Nyl. [1] Buellia tesserata Körb. [3] Physcia undulata Moberg [1] Buellia triseptata A. Nordin [3] Physciella chloantha (Ach.) Essl. [2] Buellia tyrolensis Körb. [3] Physciella melanchra (Hue) Essl. [2] Buellia uberior Anzi [3] Physciella nepalensis (Poelt) Essl. [2] Buellia vilis Th. Fr. [3] Physconia detersa (Nyl.) Poelt [1] Calicium abietinum Pers. [2] Physconia elegantula Essl. [1] Calicium corynellum (Ach.) Ach. [2] Physconia enteroxantha (Nyl.) Poelt [1] Calicium montanum Tibell [2] Physconia isidiomuscigena Essl. [1]

Physconia leucoleiptes (Tuck.) Essl. [1]

Physconia perisidiosa (Erichsen) Moberg [1] Rinodina archaea (Ach.) Arnold [2]

Rinodina bischoffii (Hepp) A. Massal. [2]

Rinodina castanomela (Nyl.) Arnold [2]

Physconia muscigena (Ach.) Poelt [1]

Rinodina athallina H. Magn [2]

Rinodina aurantiaca Sheard [2]

Rinodina boulderensis Sheard [2]

Rinodina capensis Hampe [2]

Cyphelium tigillare (Ach.) Ach. [2]
Dimelaena oreina (Ach.) Norman [2]
Dimelaena thysanota (Tuck.) Hale & W.L. Culb. [2]
Diplotomma alboatrum (Hoffm.) Flot. [3]\*
Diplotomma venustum (Körb.) Körb. [3]\*

Dirinaria neotropica Kalb [2]

Calicium salicinum Pers. [2]

Calicium viride Pers. [2]

Calicium trabinellum (Ach.) Ach. [2]

Cyphelium lucidum (Th. Fr.) Th. Fr. [2]

Rinodina coloradiana H. Magn. [2] Rinodina confragosa (Ach.) Körb. [2] Rinodina efflorescens Malme [3] Rinodina glauca Ropin [2] Rinodina grandilocularis Sheard [2] Rinodina guzzinii Jatta [2] Rinodina herrei H. Magn. [2] Rinodina intermedia Bagl. [2] Rinodina juniperina Sheard [2] Rinodina metaboliza Vain. [2] Rinodina milvina (Wahlenb.) Th. Fr. [2] Rinodina mniaraea (Ach.) Körb. [2] Rinodina oxydata (A. Massal.) A. Massal. [2] Rinodina parasitica H. Mayrhofer & Poelt [2] Rinodina perreagens Sheard [2] Rinodina trevisanii (Hepp) Körb. [2] Rinodina verruciformis Sheard [2] Rinodina zwackhiana (Kremp.) Körb. [2]

#### Teloschistaceae

Caloplaca albovariegata (B. de Lesd.) Wetmore [3] Caloplaca ammiospila (Wahlenb.) H. Olivier [3] Caloplaca arenaria (Pers.) Müll. Arg. [3] Caloplaca arizonica H. Magn. [3] Caloplaca atroalba (Tuck.) Zahlbr. [3] Caloplaca atroflava (Turner) Mong. [3] Caloplaca brouardii (B. de Lesd.) Zahlbr. [3] Caloplaca cerina (Ehrh. ex Hedw.) Th. Fr. [3] Caloplaca chlorina (Flot.) H. Olivier [3] Caloplaca chrysodeta (Vain. ex Räsänen) Dombr. [3] Caloplaca cinnabarina (Ach.) Zahlbr. [3] Caloplaca citrina (Hoffm.) Th. Fr. [3] Caloplaca cladodes (Tuck.) Zahlbr. [3] Caloplaca conversa (Kremp.) Jatta [3] Caloplaca crenulatella (Nyl.) H. Olivier [3] Caloplaca dakotensis Wetmore [3] Caloplaca decipiens (Arnold) Blomb. & Forssell [3] Caloplaca demissa (Körb.) Arup & Grube [3] Caloplaca durietzii H. Magn. [3] Caloplaca epithallina Lynge [3] Caloplaca ferruginea (Huds.) Th. Fr. [3] Caloplaca flavovirescens (Wulfen) Dalla Torre & Sarnth. [3] Caloplaca furfuracea H. Magn. [3] Caloplaca grimmiae (Nyl.) H. Olivier [3] Caloplaca holocarpa (Hoffm.) A.E. Wade [3] Caloplaca microthallina Wedd. [3] Caloplaca parviloba Wetmore [3] Caloplaca pellodella (Nyl.) Hasse [3] Caloplaca persimilis Wetmore [3] Caloplaca phyllidizans Wetmore [3] Caloplaca pinicola H. Magn. [3] Caloplaca saxicola (Hoffm.) Nordin [3] Caloplaca schoeferi Poelt [3] Caloplaca sideritis (Tuck.) Zahlbr. [3] Caloplaca sinapisperma (Lam. & DC.) Maheu & A. Gillet [3] Caloplaca sonorae Wetmore [3] Caloplaca squamosa (B. de Lesd.) Zahlbr. [3] Caloplaca stellata Wetmore & Kärnefelt [3] Caloplaca subsoluta (Nyl.) Zahlbr. [3] Caloplaca tetraspora (Nyl.) H. Olivier [3] Caloplaca tiroliensis Zahlbr. [3] Caloplaca tominii Savicz [3] Caloplaca trachyphylla (Tuck.) Zahlbr. [3] Caloplaca variabilis (Pers.) Müll. Arg. [3] Caloplaca wetmorei Nimis, Poelt & Tretiach [3] Fulgensia desertorum (Tomin) Poelt [2] Fulgensia subbracteata (Nyl.) Poelt [2]

Seirophora contortuplicata (Ach.) Frödén [2]\*

Xanthomendoza fallax (Hepp) Søchting, Kärnefelt & S.Y. Kondr. [2]
Xanthomendoza montana (L. Lindblom) Søchting, Kärnefelt & S.Y. Kondr. [2]
Xanthomendoza ulophyllodes (Räsänen) Søchting, Kärnefelt & S.Y. Kondr. [2]
Xanthoria candelaria (L.) Th. Fr. [2]
Xanthoria elegans (Link) Th. Fr. [2]
Xanthoria sorediata (Vain.) Poelt [2]

# OSTROPOMYCETIDAE AGYRIALES

#### Agyriaceae

Lignoscripta atroalba B.D. Ryan & T.H. Nash [2] Trapeliopsis flexuosa (Fr.) Coppins & P. James [2] Trapeliopsis granulosa (Hoffm.) Lumbsch [2] Xylographa crassithallia B.D. Ryan & T.H. Nash [2] Xylographa parallela (Ach.) Fr. [2] Xylographa pruinodisca B.D. Ryan & T.H. Nash [2] Xylographa vitiligo (Ach.) J.R. Laundon [2]

#### Schaereriaceae

Schaereria dolodes (Nyl.) Schmull & T. Sprib. [3] Schaereria fuscocinerea (Nyl.) Clauzade & Cl. Roux [2]

### Trapeliaceae

Placynthiella icmalea (Ach.) Coppins & P. James [2]
Placynthiella oligotropha (J.R. Laundon) Coppins &
P. James [2]
Placynthiella uliginosa (Schrad.) Coppins & P. James [2]
Rimularia insularis (Nyl.) Rambold & Hertel [2]
Trapelia coarctata (Turner ex Sm.) M. Choisy [2]
Trapelia glebulosa (Sm.) J.R. Laundon [2]\*

# BAEOMYCETALES

# Baeomycetaceae

Baeomyces rufus (Huds.) Rebent. [1]

# OSTROPALES

# Gyalectaceae

Gyalecta foveolaris (Ach.) Schaer. [2]

# Stictidaceae

Absconditella lignicola Vězda & Pišút [2]

# The lot rematace a

Diploschistes actinostomus (Ach.) Zahlbr. [1]
Diploschistes aeneus (Müll. Arg.) Lumbsch [1]
Diploschistes arabiensis Lumbsch Elix [1]
Diploschistes badius Lumbsch & Elix [1]
Diploschistes caesioplumbeus (Nyl.) Vain. [1]
Diploschistes diacapsis (Ach.) Lumbsch [1]
Diploschistes muscorum (Scop.) R. Sant. [1]
Diploschistes scruposus (Schreb.) Norman [1]

#### **PERTUSARIALES**

### Megasporaceae

Aspicilia americana B. de Lesd. [3]
Aspicilia aquatica Körb. [3]
Aspicilia arizonica Owe-Larss. & A. Nordin [3]
Aspicilia boykinii Owe-Larss. & A. Nordin [3]
Aspicilia cinerea (L.) Körb. [3]
Aspicilia contorta (Hoffm.) Kremp. [3]
Aspicilia desertorum (Kremp.) Mereschk. [3]
Aspicilia determinata (H. Magn.) J.C. Wei [3]

Aspicilia hispida Mereschk. [3]
Aspicilia olivaceobrunnea Owe-Larss. & A. Nordin [3]
Aspicilia substictica Owe-Larss. & A. Nordin [3]
Lobothallia alphoplaca (Wahlenb.) Hafellner [2]
Lobothallia praeradiosa (Nyl.) Hafellner [2]
Lobothallia radiosa (Hoffm.) Hafellner [2]
Megaspora verrucosa var. mutabilis (Ach.) Nimis & Cl. Roux [3]
Megaspora verrucosa (Ach.) Hafellner & V. Wirth var. verrucosa [3]

#### Ochrolechiaceae

Ochrolechia africana Vain. [2]
Ochrolechia androgyna (Hoffm.) Arnold [2]
Ochrolechia mexicana Vain. [2]
Ochrolechia pseudopallescens Brodo [2]
Ochrolechia splendens Lumbsch & Messuti [2]
Ochrolechia subisidiata Brodo [2]
Ochrolechia subpallescens Verseghy [2]

#### Pertusariaceae

Pertusaria amara (Ach.) Nyl. [1]
Pertusaria azulensis B. de Lesd. [1]
Pertusaria flavicunda Tuck. [1]
Pertusaria hymenea (Ach.) Schaer. [1]
Pertusaria mariae B. de Lesd. [1]
Pertusaria moreliensis B. de Lesd. [1]
Pertusaria ophthalmiza (Nyl.) Nyl. [1]
Pertusaria saximontana Wetmore [1]
Pertusaria sommerfeltii (Sommerf.) Fr. [1]
Pertusaria stenhammarii Hellb. [1]
Pertusaria tejocotensis B. de Lesd. [1]
Pertusaria wulfenioides B. de Lesd. [1]

# † LICHINOMYCETES LICHINOMYCETIDAE LICHINALES

# Gloeoheppiaceae

Gloeoheppia polyspora Henssen [1] Gloeoheppia squamulosa (Zahlbr.) M. Schultz [3]

#### Lichinaceae

Anema progidulum (Nyl.) Henssen [1]
Ephebe ocellata Henssen [2]
Ephebe perspinulosa Nyl. [2]
Heppia adglutinata (Kremp.) A. Massal. [1]
Heppia conchiloba Zahlbr. [1]
Heppia despreauxii (Mont.) Tuck. [1]
Heppia lutosa (Ach.) Nyl. [1]
Lemmopsis arnoldiana (Hepp) Zahlbr. [1]

Lempholemma cladodes (Tuck.) Zahlbr. [2] Lempholemma polyanthes (Bernh.) Malme [2] Lichinella americana Henssen [3] Lichinella cribellifera (Nyl.) P.P. Moreno & Egea [3] Lichinella flexa Henssen, Büdel & T.H. Nash [3] Lichinella granulosa M. Schultz [3] Lichinella intermedia Henssen, Büdel & T.H. Nash [3] Lichinella iodopulchra (Couderc ex Croz.) P.P. Moreno & Egea [3] Lichinella minnesotensis (Fink) Essl. [3] Lichinella myriospora (Zahlbr.) P.P.Moreno & Egea ex Lichinella nigritella (Lettau) P.P. Moreno & Egea [3] Lichinella sinaica (Galun & Marton) P.P. Moreno & Egea [3] Lichinella stipatula Nyl. [3] Metamelanea melambola (Tuck.) Henssen [2] Peccania arizonica Tuck. ex Herre [3] Peccania subnigra (B. de Lesd.) Wetmore [3] Peccania tiruncula (Nyl.) Henssen [3] Phloeopeccania pulvinulina J. Steiner [3] Porocyphus coccodes (Flot.) Körb. [1] Psorotichia hassei Fink ex J. Hedrick [3] Psorotichia montinii (A. Massal.) Forssell [3] Psorotichia murorum A. Massal. [3] Psorotichia schaereri (A. Massal.) Arnold [3] Psorotichia taurica (Nyl.) Vain. [3] Pterygiopsis cava M. Schultz [3] Stromatella bermudana (Riddle) Henssen [1] Synalissa mattogrossensis (Malme) Henssen [1] Thyrea confusa Henssen [1]

Lempholemma chalazanum (Ach.) B. de Lesd. [2]

#### Peltulaceae

Peltula bolanderi (Tuck.) Wetmore [1]
Peltula clavata (Kremp.) Wetmore [1]
Peltula euploca (Ach.) Poelt ex Ozenda & Clauzade [1]
Peltula farinosa Büdel [1]
Peltula michoacanensis (B. de Lesd.) Wetmore [1]
Peltula obscurans var. deserticola (Zahlbr.) Wetmore [1]
Peltula obscurans var. hassei (Zahlbr.) Wetmore [1]
Peltula obscurans (Nyl.) Gyeln. var. obscurans [1]
Peltula omphaliza (Nyl.) Wetmore [1]
Peltula patellata (Bagl.) Swinscow & Krog [1]
Peltula placodizans (Zahlbr.) Wetmore [1]
Peltula psammophila (Nyl.) Egea [1]
Peltula richardsii (Herre) Wetmore [1]
Peltula tortuosa (Nees) Wetmore [1]
Peltula zahlbruckneri (Hasse) Wetmore [1]

**Annotation Key**. Annotations [in brackets] follow each taxon, and those with an asterisk '\*' have been updated according to the *North American Lichen Checklist* of Esslinger 2010. Each record cites the source with a number (see *Literature Cited*):

1 – Nash, Ryan, Gries, and Bungartz 2002; 2 – Nash, Ryan, Diederich, Gries, and Bungartz 2004; 3 – Nash, Gries, and Bungartz 2007; 4 – Westberg and Arup 2010

# **ACKNOWLEDGMENTS**

Although not directly associated, we would like to acknowledge the work accomplished under the NSF award #0103738. Furthermore, we thank those involved in producing the *Lichen Flora of the Greater Sonoran Desert Region* (see the preface of Nash et al. 2007 for a list of contributors); without the exceptional involvement of these individuals this revised "catalog" would not have been possible. We also thank Dr. Frank Bungartz and Jade Siminitus for kindly providing photographs for the figure. The constructive comments of the anonymous reviewers also helped considerably in improving the quality of this publication.

# LITERATURE CITED

ARNOLD, A.E., J. MIADLIKOWSKA, K.L. HIGGINS, S.D. SARVATE, P. GUGGER, A. WAY, V. HOFSTETTER, F. KAUFF and F. LUTZONI. 2009. A phylogenetic estimation of trophic transition networks for ascomycetous fungi: Are lichens cradles of symbiotrophic fungal diversification? *Systematic Biology* 58: 283–297.

BATES, S.T. 2006. A preliminary checklist of Arizona macrofungi. *Canotia* 2: 47–78.

BATES, S.T. and A. BARBER. 2008. A preliminary checklist of Arizona slime molds. *Canotia* 4: 8–19.

BOYKIN, M.A. and T.H. NASH III. 1994. The lichen flora of Grand Canyon National Park, Arizona. *Journal of the Arizona-Nevada Academy of Science* 28: 59–69.

BRODO, I.M., S.D. SHARNOFF and S. SHARNOFF. 2001. *Lichens of North America*. Yale University Press, New Haven.

BÜDEL, B. and C. SCHEIDEGGER. 2008. Thallus morphology and anatomy. Pp. 40–68. *In*: T.H. Nash III (ed.). *Lichen Biology*. 2nd edn. Cambridge University Press, Cambridge.

BUNGARTZ, F., R. ROSENTRETER and T.H. NASH III. 2002. Field Guide to Common Epiphytic Macrolichens in Arizona. Arizona State University Lichen Herbarium, Tempe.

DARROW, R.A. 1950. The arboreal lichen flora of southeastern Arizona. *The American Midland Naturalist* 43: 484–503.

DIEDERICH, P. 2004. Lichenicolous fungi. Pp. 617–714. *In*: T.H. Nash III, B.D. Ryan, P. Diederich, C. Gries and F. Bungartz (eds.). *Lichen Flora of the Greater Sonoran Desert Region*. Vol. II. Lichens Unlimited, Tempe.

ELIX, J.A. and E. STOCKER-WÖRGÖTTER. 2008. Biochemistry and secondary metabolites. Pp. 104–133. *In*: T.H. Nash III (ed.). *Lichen Biology*. 2nd edn. Cambridge University Press, Cambridge.

ESSLINGER, T.L. 2010. Cumulative Checklist for the Lichen-Forming, Lichenicolous and Allied Fungi of the Continental United States and Canada. http://www.ndsu.edu/pubweb/~esslinge/chcklst/chcklst7.htm#X (Accessed 2010 August 21).

FINK, B. 1909. The composition of a desert lichen flora. Mycologia 1: 87–103.

GEHRIG, H., A. SCHÜSSLER and M. KLUGE. 1996. *Geosiphon pyriforme*, a fungus forming endocytobiosis with *Nostoc* (cyanobacteria), is an ancestral member of the *Glomales*: evidence by SSU rRNA analysis. *Journal of Molecular Evolution* 43: 71–81.

GRUBE, M., M. CARDINALE, J. VIEIRA DE CASTRO, H. MÜLLER and G. BERG. 2009. Species-specific structural and functional diversity of bacterial communities in lichen symbioses. *The ISME Journal* 3(9): 1105–1115.

HODKINSON, B.P. and F. LUTZONI. 2009. A microbiotic survey of lichen-associated bacteria reveals a new lineage from the *Rhizobiales*. *Symbiosis* 49(3): 163–180.

HONEGGER, R. 2008. Mycobionts. Pp. 27–39. *In*: T.H. Nash III (ed.). *Lichen Biology*. 2nd edn. Cambridge University Press, Cambridge.

JACKSON, H.B., S.D. LEAVITT, T. KREBS and L.L. ST. CLAIR. 2005. Lichen flora of the eastern Mojave Desert: Blackrock Arizona, Mojave County, Arizona, USA. *Evansia* 22: 30–38.

JOHNSEN, A.B. 1965. Some lichens from West Fork, Coconino County, Arizona. *The Bryologist* 68: 241–243.

LAWREY, J.D. and P. DIEDERICH. 2003. Lichenicolous fungi: interactions, evolution, and biodiversity. *The Bryologist* 106: 80–120.

MOBERG, R. and W.A. WEBER. 1974. Additions to the lichen flora of Arizona II. *The Bryologist* 77: 472–474.

NASH III, T.H. 1973a. Two new species of *Xanthoparmelia* from Arizona. *The Bryologist* 76: 214–216.

NASH III, T.H. 1973b. Additions to the lichen flora of Arizona I. *The Bryologist* 76: 545–548.

NASH III, T.H. 1975. Lichens of Maricopa County, Arizona. *Journal of the Arizona-Nevada Academy of Science* 10: 119–125.

NASH III, T.H. 1977. Lichens of the White Mountains, Arizona. *Journal of the Arizona-Nevada Academy of Science* 12: 53–56.

NASH III, T.H. 1985. Additions to the lichen flora of Arizona III. The Bryologist 88: 19–22.

NASH III, T.H. 1991. Preliminary study of the lichens of Mesa Verde National Park. *Journal of the Arizona-Nevada Academy of Science* 23: 97–105.

NASH III, T.H. (ed.). 1996. *Lichen Biology*. 1st edn. Cambridge University Press, Cambridge.

NASH III, T.H. 2002. Arsène historical collection from New Mexico. Evansia 19: 161.

NASH III, T.H. (ed.). 2008a. *Lichen Biology*. 2nd edn. Cambridge University Press, Cambridge.

NASH III, T.H. 2008b. Nitrogen, its metabolism and potential contribution to ecosystems. Pp. 216–233. *In*: T.H. Nash III (ed.). *Lichen Biology*. 2nd edn. Cambridge University Press, Cambridge.

NASH III, T.H. and J.A. ELIX. 1986. A new species and new records in the lichen genus *Xanthoparmelia* (Vain.) Hale (*Ascomycotina*, *Parmeliaceae*) for North America. *Mycotaxon* 26: 453–455.

NASH III, T.H., C. GRIES and F. BUNGARTZ (eds.). 2007. *Lichen Flora of the Greater Sonoran Desert Region*. Vol. III. Lichens Unlimited, Tempe.

NASH III, T.H. and A.B. JOHNSEN. 1975. Catalog of the lichens of Arizona. *The Bryologist* 78: 7–24.

NASH III, T.H., B.D. RYAN, W.C. DAVIS, O. BREUSS, J. HAFELLNER, H.T. LUMBSCH, L. TIBELL and T. FEUERER. 1998. Additions to the lichen flora of Arizona IV. *The Bryologist* 101: 93–99.

NASH III, T.H., B.D. RYAN, C. GRIES and F. BUNGARTZ (eds.). 2002. *Lichen Flora of the Greater Sonoran Desert Region*. Vol. I. Lichens Unlimited, Tempe.

NASH III, T.H., B.D. RYAN, P. DIEDERICH, C. GRIES and F. BUNGARTZ (eds.). 2004. *Lichen Flora of the Greater Sonoran Desert Region*. Vol. II. Lichens Unlimited, Tempe.

NASH III, T.H. and L.L. SIGAL. 1981. Preliminary study of the lichens of Zion National Park, Utah. *Journal of the Arizona-Nevada Academy of Science* 16: 46–50.

SCHMIT, J.P. and G.M. MUELLER. 2007. An estimate of the lower limit of global fungal diversity. *Biodiversity and Conservation* 16: 99–111.

SURYANARAYANAN, T.S., N. THIRUNAVUKKARASU, G.N. HARIHARAN and P.Q. BALAJI. 2005. Occurrence of non-obligate microfungi inside lichen thalli. *Sydowia* 57: 120–130.

SWEAT, K.G., W.A. ISELIN, S.T. BATES and T.H. NASH III. 2004. The lichens of Parashant National Monument, Arizona: A preliminary study. *Journal of the Arizona-Nevada Academy of Science* 37: 85–90.

TRIEBEL, D., G. RAMBOLD and T.H. NASH III. 1991. On lichenicolous fungi from continental North America. *Mycotaxon* 42: 263–296.

WEBER, W.A. 1963. Lichens of the Chiricahua Mountains, Arizona. *University of Colorado Studies Series in Biology* No. 10: 1–27.

WESTBERG, M. and ARUP, U. 2010. *Candelaria pacifica* sp. nova (Ascomycota, Candelariales) and the identity of *C. vulgaris*. Bibliotheca Lichenologica 106: 347–359.

WILLEY, H. 1881. A new North American lichen. *Bulletin of the Torrey Botanical Club* 8: 140–141.

ZAHLBRUCKNER, A. 1908. New North American Lichens. *Bulletin of the Torrey Botanical Club* 35: 297–300.