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**FAIR in Circadian Practice**

**Day 2**

**Do you want to start the remaining two sessions at 13:00 and not half past**

NO:

YES:+1+1+1+1+1+1+1+1+1+1+1+1+1+1+1+1+1

**Your interests:**

- insects (Drosophila melanogaster, Triboilum castaneum, ??? ): +1

- Vertebrates (Danio rerio, Mus musculus, ???):+1+1+1

- Plants (Arabidopsis thaliana, Fragilariopsis cylindrus ???):+1+1

- Human: +1+1+1+1+1+1+1+1+1+1+1+1+1

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**Important notice:**

Before you begin today, please take some time to sign up for the following accounts:

1. Benchling     (the ELN we will use for one of today's lessons): <https://benchling.com/signup?pubref=pubref_zQlS6DPe>.
2. Protocols.io     (the protocol repository with PID, which we will also be using for today's     lessons): <https://www.protocols.io/create>
3. BioDare2 <https://biodare2.ed.ac.uk/>     (email us if your email is not recognised)

(do not worry if you cannot do it, right now... there will be some time to do this at the start of lesson 8)

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**(Meta)data in Excel part 2**

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**Exercise 2: Spotting problems**

Look at the following rows and columns in the problematic table:

·    Row 5

·    Row 2

·    Column C

·    Column E

·    Column L

Table image: <https://carpentries-incubator.github.io/fair-bio-practice/fig/bad-metadata.png>,

Excel file: <https://github.com/carpentries-incubator/fair-bio-practice/raw/gh-pages/files/04-bad-metadata.xlsx>

which of the problems discussed before can you spot in these rows and columns.

Here, we list them again:

1.     Using multiple tables

2.     Using multiple tabs

3.     Not filling in zeros

4.     Using problematic null values

5.     Using formatting to convey information and organizing data

6.     Placing comments or units in cells

7.     Entering more than one piece of information in a cell

8.     Inconsistency in used values

9.     Using problematic field names

10.  Using special characters in data

11.  Values without field labels

Type the problem number(s) next to the table elements

·    Row 5: 111111991919999191911 109199

·    Row 2: 7 66666866666

·    Column C:56685585558555558

·    Column E:1066855888888886886

·    Column L:333335533454333

DONE:+1+1+1+1+1+1++11+1+1+1+1+1+1+1

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**Exercise 3: Outsmarted by Excel**

Open Excel and type the following values into the cells:

A       B       C       D       E       F

Gene    Sept2   Sample  0013    Record  12/5/4

Mar/1   1March  Mar-1   1-3     14/3/20 43904

**Questions:**

* ·         Is what you see what typed? nonononononononoNoNoNononono

·         Can you force the above values? yesyes-format to numbers or textyes,, format cell adding 'yesYesYesyesyesyes

·         Do you know which year these dates represent?noNoNonononoNononononononono

DONE: +11+1+1+1+1+1+1+1+1+1+1

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**Exercise 4: Data tables Quiz:**

Which of the following statements is true/false (T or F):

·         Do’s and don’ts help in programmatic re-use:ttTTTTTTTTTTTTTT

·         Avoiding multiple tabs improves interoperability:TT tTTTTTTTTTTTTT

·         Having accompanying README file with a table description is not FAIR:FfF FFFFFFFFFFFFF

·         No ‘spaces’ in columns headers improve readability:TTtTTTTTTTTTTTT

·         2022-07-15 is ISO date format:TTTTTTTTTTTTTTT

·         20220715 date format is better for excel than 2022-07-15:FFFFFFFFFFFFFFF

·         “No data” is better than leaving cell “blank” for missing data: FFFFFFFFFFFFFFFF

DONE:+1+1+1+1+1+1+1+1+1+1+1+1+1+1+1

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**List of attendees**

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**Lesson 8: Laboratory Records**

**Which of these most accurately describes your record keeping experience?**

(type +1 next to the statement that best describes your situation)

- I have used hard copy lab notebooks before for research data records keeping.+1+1+1+1+1++11+1+1+1+1+1+1+1

- I have used electronic lab notebooks before for research data records keeping.+1 +1+1+1+1+1+1+1

- I have used Benchling before.+1

- I have used an online protocol database before.

- I have used Protocols.io before.+1

- I have not had any research record keeping experience in the past.+1+1+1+1+1

-

DONE:+1+1+!+1+1+1+1+1+1+1+1+1

Before you begin with lesson 8, please take some time to sign up for the following two accounts (if you haven't already done so):

1. Benchling     (the ELN we will use for one of today's lessons): <https://benchling.com/signup?pubref=pubref_zQlS6DPe>.
2. Protocols.io     (the protocol repository with PID, which we will also be using for today's     lessons): <https://www.protocols.io/create>

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**Exercise 1: 14:07**

**Differences between analog and digital record keeping**

Compare the electronic version of the tea protocol:

<https://www.protocols.io/view/how-to-make-a-cup-of-tea-buhknt4w>

with the paper one from the photo:

<https://github.com/carpentries-incubator/fair-bio-practice/blob/gh-pages/fig/06-handwritten-tea-protocol.jpg>

What are advantages and disadvantages of traditional analog records vs digital records? Try to find at least a handful of advantages and disadvantages for each. With all of these, which system do you think is most advantageous?

1 2 room

**Advantages of traditional analog records**

 not reliant on digit spaces/electricity/internet

easy to do quickly/in a rush whereas electronic labbook requires the time to sit down and do it

quickly accessible - can add things in easily/change easily

following thought process

ease of additional note taking

doesn't rely on internet/platform to be available

durable; difficult to accidentally delete...

**Advantages of digital records**

 dont have to read handwriting

 more widely accessible to wider community

versioning, save multiple copies

ease of sharing

More secured

Metadata

easy to edit

Can be linked to metadata/primary data

easier for non-native English speakers (or dyslectic)

easier to search

3 4 room

**Disadvantages of traditional analog records**

-people may not be able to read it

-Could be lost/destroyed/stolen

 - not easily shareable with others (in case you are not there)

 Require storage space

 Security issues

 Difficult to find information

 Not a uniform format

**Disadvantages of digital records**

 - need computer/tablet etc.

 - data corruption (potentiall

y lose everything)

-Company may go out of business

-needs access to internet

-Sensitive human data (NHS) may not be suitable to be on a cloud based platform

Security

DONE:+1+1+1+1

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Let's meet 14:30

Exercise 2:

**Re-using a published lab entry**

1.     Open Benchling (<https://benchling.com>) and log in.

2.     First within your own workspace click the big ‘+’ (Create Project) right next to Projects in your Benchling workspace

3.     Call the project ‘Breakfast’, and add an appropriate description, click ‘Create project’

4.     Click here <https://benchling.com/s/etr-SY8fi7L8ZIDSMCLCf92o> to access the public lab entry ‘Eggs Florentine in Portobello Mushrooms’.

5.     Select the clock symbol on the right-hand side underneath Share: Now you can see the history of the entry and changes that have previously been made to the document with a timestamp. If someone had tried to ‘manipulate’ data, you would be able to see this here. You also see the owner of the document.

6.     Click ‘Clone from version’.

7.     Select the ‘Breakfast’ folder to clone it to.

DONE:+1+1+1+1+1+1+1+1+1+1+1+1+1+1+1

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**Exercise 3:**

**Adapting a protocol to your needs**

1.     You have now accessed a digital record and want to reuse it to make your own breakfast. To show how reusable digital records are we will first navigate through the cloned file you made in your project.

2.     Navigate to your Project ‘Breakfast’, you can tell you are in your Project, if your initials show in a red circle next to entries in the side bar. You should see the lab entry ‘Eggs Florentine in Portobello Mushrooms’, and the top bar above the title and toolbar should read ‘Tea’, ‘Portobello Mushrooms and Spinach’, ‘Poached Egg and Hollandaise Sauce’, ‘Add Protocol’, ‘Notes’, and ‘Metadata’.

3.     Click through those tabs and you will see that in your notes you have your lab entry describing how breakfast was made with embedded graphics and a shopping list and current prices. The other three tabs describe the protocols that were used, and you can add additional protocols with the ‘add protocol’ tab. We want you to adapt the ‘Tea’ protocol to suit your ingredients and methods.

4.     Once you have made appropriate changes in the Tea protocol, you should consider changing the order in which the breakfast and tea are made.

5.     Once you have made all suggested change have a look at the history of the record (clock button), you can see the changes you have recently made, and you can see it still relates to the original document. It tells you what record it has been cloned from and when.

6.     Click the link to the original record. As you can see digital record keeping allows provenance, crediting the original author, but also allowing you to keep track of your sources.

7.     Navigate back to your lab entry in your project (your initials are a sign that you are in the right place).

DONE: +1+1+1+1+1+1+1+1+1+1+1+1+1

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**Exercise 4:**

**Sharing your record**

1.     Click the info icon on the right-hand side underneath the clock symbol you used previously and select ‘Export entry’

2.     Your export is now running, you will receive an email when the export is complete

3.     Click the link in the email to download your protocol as a .zip

4.     Unzip the file and in your own time, print the protocol if you want to use the recipe in the kitchen, or share it with friends.

5.     You can share .pdf versions or click Share and generate a Share link of your lab entry. This makes your record interoperable as many users across many platforms across the world can access your entry if you make it public and share it on for example social media. If there is no IT access present, you always have the option to print the .pdf copy.

DONE: +1+1+1+1+1+1+1+1+1+1+1+1

Benchling demo:

<https://benchling.com/s/etr-D59zgfqSvefvhPmIxAne?m=slm-FOH9KYfv9n6eUZPJWbkt>

Plasmid example (plasmids private accessible for UoE BioRDM only)

<https://benchling.com/s/etr-Gje9DwS83aijjPlkmaH9?m=slm-0EcbtGdiD4I9xpcfOByr>

**Our Benchling tutorial:**

<https://www.wiki.ed.ac.uk/display/RDMS/Benchling+%28quick%29+tutorial>

**Further resources and tutorials from Benchling:**

·         Main help page, with access to several tutorials: <https://help.benchling.com/en/>

·         More molecular biology features: <https://help.benchling.com/en/collections/69523-molecular-biology>

·         Benchling training kit for academics: <https://help.benchling.com/en/collections/1608962-benchling-training-kit-for-academics>

**Materials on ELNs:**

<https://www.wiki.ed.ac.uk/pages/viewpage.action?pageId=463750271>

DONE: **+1+1**

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**Exercise/challenge 5:**

**Adapt a public protocol and retain its provenance**

Protocol link: <https://dx.doi.org/10.17504/protocols.io.buhknt4w>

*Fork the protocol, preserving the original for crediting*

1. Open the link to the above protocol, as you can see, we have assigned it its own DOI

2. First click on Metrics: Because we are FAIR, this shows you how many views over time this protocol has had, how many exports, how many individual steps it involves and how many times it has been forked.

3. Now click on the downwards arrow next to the title

4. Select 'Copy/Fork' and click 'make a fork'

5. Select the Folder you want the protocol to be forked to and click 'continue'

6. Your fork of "How to make a cup of tea" is ready now, click 'edit new fork'

7. On the right-hand tool bar, the clock icon, shows you the history of the protocol (as before in Benchling). Currently you should see no history as you have not made changes.

*Edit the forked protocol*

1. Go to 'Materials' in the top tool-bar: add or edit materials according to your preferences, e.g., change full-fat milk to oat-milk, or add honey, lemon etc

2. Go to 'Steps' in the top tool-bar: edit the protocol according to your preferences

3. You can edit the 'Description' and 'Guidelines & Warnings' if you would like to

4. As soon as you change anything, the timestamp and where in the protocol this change was made appears in the history.

5. Click 'View', you will now see the reader view of your protocol. It clearly states underneath the title 'Forked from How to make a cup of tea' and the original protocol is linked. This allows clear identification of your source.

6. Click 'Edit'

*Optional: Export the forked protocol*

1. Click 'More' in the top tool-bar, select 'Export' > 'PDF' > 'To your computer' and click export (leave selections blank)

DONE: +1+1+1+1+1+1+1+1+1+1+1+1=1+1

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**Exercise 6:**

**Do you use an ELNs? Which one? What features do you like?**

·         HAVE YOU TRIED AN ELN and you gave up?:

·         Anyone switched from paper entirely to ELNs?:

·         Anyone only paper?:

Never used one, haven't needed to for my area of work+1+1

Which ELN do you use:

- LabArchives+1

-Google Docs+1+1+1

-Microsoft Notebook+1+1+1

-Labstep

DONE:+1+1+1+1+1+1+1+1+1+1+1+1+1+1

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**Quiz:**

Which of the following statement are true (T) / false (F)?

·         Good record keeping ensures transparency and reproducibility.TTTTTTTTTTTTTTtTTT

·         There are no advantages to using analog record keeping when compared to digital record keeping. FFFFFFFFFFFfFFFFF

·         Digital records help people view a protocol simultaneously.TTTTTTTTTtTTTTTTT

·         Digitally kept records can be quickly and easily edited.TTTTTTTTTTTtTTTTTT

·         On balance, digital record keeping is more advantageous than analog record keeping.TTTTTTTTTTTTTTtTTT

·         Digital records are easier to search (for and within) than analog records.TTTTTTTTtTTTTTTTTT

DONE:+1+1+1+1+1+1+1+1+1+1+1+1+1+1+1+1

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**Lesson 9: Files organisation**

**Exercise 1: Naming and sorting**

Have a look at the example files from a project, similar to the one from the previous metadata episode.

For example,

·         LD\_phyA\_off\_t04\_2020-08-12.norm.xlsx

is a file that contains normalized data (norm), from experiment in long day (LD) for genotype

phyA, with media off sucrose (off).

All the files have been sorted by name and demonstrate consequences of different naming strategies.

For your information, to encode experimental details the following conventions were taken

·         phyB/phyA are sample genotypes

·         sXX is the sample number

·         LD/SD are different photoperiodic conditions (long or short day)

·         on/off are different media (on sucrose, off sucrose)

·         measurement date

·         other details are timepoint and raw or normalized data

       2020-07-14\_s12\_phyB\_on\_SD\_t04.raw.xlsx     (1)

       2020-07-14\_s1\_phyA\_on\_LD\_t05.raw.xlsx       (2)

       2020-07-14\_s2\_phyB\_on\_SD\_t11.raw.xlsx       (3)

       2020-08-12\_s03\_phyA\_on\_LD\_t03.raw.xlsx     (4)

       2020-08-12\_s12\_phyB\_on\_LD\_t01.raw.xlsx     (5)

       2020-08-13\_s01\_phyB\_on\_SD\_t02.raw.xlsx     (6)

       2020-7-12\_s2\_phyB\_on\_SD\_t01.raw.xlsx         (7)

       AUG-13\_phyB\_on\_LD\_s1\_t11.raw.xlsx            (8)

       JUL-31\_phyB\_on\_LD\_s1\_t03.raw.xlsx             (9)

       LD\_phyA\_off\_t04\_2020-08-12.norm.xlsx         (10)

       LD\_phyA\_on\_t04\_2020-07-14.norm.xlsx          (11)

       LD\_phyB\_off\_t04\_2020-08-12.norm.xlsx         (12)

       LD\_phyB\_on\_t04\_2020-07-14.norm.xlsx          (13)

       SD\_phyB\_off\_t04\_2020-08-13.norm.xlsx          (14)

       SD\_phyB\_on\_t04\_2020-07-12.norm.xlsx          (15)

       SD\_phya\_off\_t04\_2020-08-13.norm.xlsx           (16)

       SD\_phya\_ons\_t04\_2020-07-12.norm.xlsx          (17)

       ld\_phyA\_ons\_t04\_2020-08-12.norm.xlsx           (18)

**1 & 2 room:**

Focus on the data with date first:

       2020-07-14\_s12\_phyB\_on\_SD\_t04.raw.xlsx     (1)

       2020-07-14\_s1\_phyA\_on\_LD\_t05.raw.xlsx       (2)

       2020-07-14\_s2\_phyB\_on\_SD\_t11.raw.xlsx       (3)

       2020-08-12\_s03\_phyA\_on\_LD\_t03.raw.xlsx     (4)

       2020-08-12\_s12\_phyB\_on\_LD\_t01.raw.xlsx     (5)

       2020-08-13\_s01\_phyB\_on\_SD\_t02.raw.xlsx     (6)

       2020-7-12\_s2\_phyB\_on\_SD\_t01.raw.xlsx         (7)

       AUG-13\_phyB\_on\_LD\_s1\_t11.raw.xlsx            (8)

       JUL-31\_phyB\_on\_LD\_s1\_t03.raw.xlsx             (9)

**Questions:**

a ) What are the problems with having the date first? Easy to miss the file description

Difficult to find what is needed if the date is far back in a long series

Not the most important thing that is clear

doesn't make file information most obvious

different file formats will cause ordering problems

many experiments done in a day so its messy - can get very long if you have to add hours, minutes etc

b) How do different date formats behave once sorted (eg 1,2 vs 8,9)?

sorting depends on date format

not in order in different formats

can't easily dort

Mixes up everything

c) Do you see what happens when you mix conventions?

 difficult to sort

 mixes the lists

 problems with sorting

d) Can you tell the importance of a leading 0 (zeros)?

will put file at the top of the list (before files without 0)

incorrect order can happen

**3 & 4 room:**

Focus on the other half of the files:

       LD\_phyA\_off\_t04\_2020-08-12.norm.xlsx         (10)

       LD\_phyA\_on\_t04\_2020-07-14.norm.xlsx          (11)

       LD\_phyB\_off\_t04\_2020-08-12.norm.xlsx         (12)

       LD\_phyB\_on\_t04\_2020-07-14.norm.xlsx          (13)

       SD\_phyB\_off\_t04\_2020-08-13.norm.xlsx          (14)

       SD\_phyB\_on\_t04\_2020-07-12.norm.xlsx          (15)

       SD\_phya\_off\_t04\_2020-08-13.norm.xlsx           (16)

       SD\_phya\_ons\_t04\_2020-07-12.norm.xlsx          (17)

       ld\_phyA\_ons\_t04\_2020-08-12.norm.xlsx           (18)

**Questions:**

a) Is it equally easy to find all data from LD conditions as ON media? No. NoNo

b) Can you spot the problem when using different cases (upper/lower) eg 15, 16,     17, 18? Not consistent - confusing implies different conditions + ordering of data affected

c) Do you see benefits of keeping consistent lengths of the naming conventions     (10-12 vs 16-17)?For coding purpose, having same characters number is better

 Easy to compare

d) Can you tell the importance of a leading 0 (zeros) (dated sample 1-3)?

 It respect the ISO convention

DONE:+1 +1+1++1++11+1+1+1+1

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**Exercise 2: A good name**

Select which file options adhere the best to the presented recommendations:

1.

a) analysis-20210906.xlsx+1 +1+1+1+1+1+1+1++11+1

b) rna-levels-by-site.v002.xlsx+1 +1+1+1++11+1+1+1

c) analysis of rna levels from 5Aug2021.xlsx

2.

a) 20210906-birds-count-EDI.csv+1+1+1+1+1+1+1+1+1+1++11+1+1+1+1

b) birds.csv+0

c) birds-count&diversity EDI 2021-09-06.csv+0

3.

a) 2020-7-12\_s2\_phyB\_+\_SD\_t01.raw.xlsx+0

b) ld\_phyA\_on\_s02-t01\_2020-07-12.norm.xlsx+1+1+1+1+1++11+1+1+1+1+1+1=1+1

c) ld\_phya\_ons\_02-01\_2020-07-12.norm.xlsx+1+0

DONE:+1+1+1+1+1+1+1+1+1+1+1+1+1+1

  +1+1

  SEE YOU 15.40

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**Exercise 3: Folders vs Files 15:40**

Have a look at these two different organization strategies:

(1) |-- Project

|-- |-- arab\_LD\_phyA\_off\_t04\_2020-08-12.metab.xlsx

(2) |-- Project

|-- |-- arabidopsis

|-- |-- |-- long\_day

|-- |-- |-- |-- phyA

|-- |-- |-- |-- |-- off\_sucrose\_2020-08-12

|-- |-- |-- |-- |-- |-- t04.metab.xlsx

Can you think of scenarios in which one is better suited than the other?

- 1if more than one file to compare

very few things to compare

long series of very similar experiments

sis

If the analBetter for coding/analyysis requires importing all the data togather (to analysis software).

Good for sharing with others without having to explain (and going through file tree)

if you need your data pretty quickly

- 2 if more species/genes/files/conditions, better for sharing the data

for dealing with more diverse conditions

more findable in a large project

good for a lot of conditions / assay types / n values

-

**Hint:** think of additional files that could be present in the folder.

DONE:+1+1+1+1

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**Exercise 4: Typical folder organizations 15:51**

Have a look at the four different folder structures A-D.

<https://github.com/carpentries-incubator/fair-bio-practice/blob/gh-pages/fig/07-file_organisation.png>

The first two” A) B) are recommended for computing, the other two: C) D) are for more wet/biological projects.

·         Which one is the most similar to your project structure

A   +1 +1      B)+1++1+1+1 +1  C)+1+1+1+1+1+1+1 +1 +1   D)

**1 & 2 room:**

When/why would you use A) and when/why B)

A)

- better sorting for publication potentially

- Python code in a separate folder - data and code may get lost

- when there is one main experiment going on (as opposed to several sub-projects)

 -you can add folders and structures along the progress

B)

 - raw experimental collection

 -easier to tell what is being measured

- more logical

-Code and data in same folder - easier to understand each folder

- when you have several sub-project running (e.g. migration, feeding, etc).

**3 & 4 room:**

When/why would you use C) and when/why D)

C)

-use after analysis

-good for small n size

Better for publication

Better for sharing the data

D)

- if more than one condition

- use before analysis

good for bigger n size (can put more numbers in each folder)

More suitable for a proposal

DONE:+1

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**Exercise 5. FAIR files: 16:11**

Choose 3 main benefits of a good strategy for folder organisation and naming conventions

\* Makes data more findable+1+1+1+1+1++1+111+1+1+1+1+1+1

\* Aids in making data more reproducible - projects can be copied easily+1

\* Raw data can be reanalysed multiple times+1+1+1+1+1+1+1+1+1+1+1+1+1

\* Naming conventions can be read automatically+1+1+1

\* Easy to understand content by name, less misunderstandings+1+1+1+1++11++11+1+1+1+1+1

\* Easier to find and share data with others +1+1+1+1+1+1++11++11+1+1+1

\* Easy inspection of the project progress (present files)+1+1+1+1

\* Fewer meetings required when sharing data

\* Time saving+1+1+1+1

DONE: +1+1+1+1+1+1++11+1+1+1+1+1+1+1+1+1+1

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**16:13**

**Period analysis with BioDare2**

<https://biodare2.ed.ac.uk/>

 Let's be back 16:30

**1. Creating experiment 16:30**

- Select **New Experiment** in the top menu

- Enter descriptive name

e.g. BioDare demo *WT vs PRR79 at long day*

- Purpose

*Testing periodic properties of the new LUC constructs in PRR79 after long day entrainment (18:6 LD)*

- Description

*Seedlings were grown under 24 light/dark cycles (18h of light and 6h of darkness), corresponding to physiological summer day lengths for 7 days. The entrained seedling were then moved into constant light conditions and their luminescence monitored with high sensitivity camera.*

*WildType and prr79 mutant were transformed with different LUC constructs driven by a clock gene promotor: LHY, TOC1, ZTL, PRR5, PRR7*

-          Comments

*Fungus contamination on prr79 TOC1 line*

Press **Accept**

In the section bellow: **Biological details**, enter the species and data category (Expression reporter)

In the **Measurements details**, enter technical information

*Seedling were sprayed with luciferin on the last day of the entrainment. Exposition time 10 minutes with 2 hours interval between the pictures. Recorded pictures were analysed using Metamorph. The standard region grid was used instead of manually selecting regions for analysis.*

**2.    Importing data**

Get the data file from: <https://biordm.github.io/fair-in-circadian-practice/files/biodare-demo-wt_prr.xlsx>

Go to import data in the dashboard menu.

Select data file (demo-wt-prr.xslx) leave the default File Format as Excell.

Next, Describe data layout (data in columns, data labels are present, no background noise records)

Next, Define time column

In the snapshot of the data table click on the first cell containing the first timepoint (A5).

Change the unit, to image nr, set time interval to 2 hours.

Next, Import label from the correct row (4th)

Next, click on the first data column (there may be other columns between the time column and the measurements)

Press **Import data**

Examine the plotting options (**Show data** and **Heatmap**).

DONE:  +1+1+1+1+1+1+1+1+1+1+1+1+1+1+1

**3.    Period analysis**

Navigate to period analysis

Define analysis parameters (data subset, detrending, analysis method and range of periods of interest.

Press **analyse**

The screen switches to the display of results of period analysis (almost instantaneously),

It shows box plots for period values by biological replicates.

If you see “X results needs attention” press Select Periods to see the results which were not included in the stats and why.

The phase plot has different view settings:

Phases by: FIT (by fitting a cos with the main period and using its peak time), by Method (as defined in original method), FIRST (by time of the first peak), Avg. (by the average times of all peaks). The parametes are describe under ? in the panel above.

There is a download icon on top to export all the results.

* DONE:+1+1++11+1+1+1+1+1+1

**4.    Rhythmicity test**

BioDare has implemented a classic JTK and eJTK methods for rhythmicity test of omics like data (short, infrequently sampled measurements).

To simulate such data, we will only analyse the last two days of data.

In the data window type 120 in “from”, leave “to” as 0 (which is the end).

Leave the rest of the presets and run the test.

DONE:  +1+1+1+1+1+1+1+1+1

**5.    BioDare2 as circadian resource**

What do you like about BioDare2

- Automatic and fast+1+1

-clear presentation of data+1+1+1+1+1

-Great platform to share data+1

 -options explained (question marks that pop up)+1+1

What is missing in BioDare2

-ability to edit graphs more i.e. colour / SEM / legend titles +1+1

-i wish it could find the best time window for the best rhythm since different time windows give different periods and errors+1

-

What would make BioDare2 a better community resource

- a place to post questions? +1+1

-

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 17:30 ENDED

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**Q&A:**

Do you have any questions about the topics discussed today? Please write them down here. Use +1 to upvote the ones you are interested in if someone already asked it. We will briefly discuss them before the following set of lessons.

 If some data gives no rhythmicity how do i locate it and remove it?

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**Feedback:**

1.      How do you feel about the presented topics after this session (type +1 next to the statement that best describes your feeling):

•       I am more confused:

•       I have a better understanding of them now:+1+1+1+1+1+1+1+1+1+1+1+1+1+1

•       My knowledge has not changed much:

2.      How was the pace of the lesson:

•       Too fast:

•       About right:+1+1+1+1+1+1+1+1+1+1

•       Too slow:

3. If the lessons could be 5 minutes longer, what would you add or spend more time on:

Explaining the different analysis methods used in biodare like NLLS....+1+1

4. What could be improved: More time for the demo/practical+1

5. What did you like: Good mix between individual work and breakout rooms+1+1+1+1+1