Agent-Based Simulation of In Vitro Hepatic Drug Metabolism: In Silico Hepatic Intrinsic Clearance

Shahab Sheikh-Bahaei, Glen E. P. Ropella, C. Anthony Hunt
The BioSystems Group, Department of Biopharmaceutical Sciences
University of California, San Francisco, CA, 94143, USA
shahab@tempusdictum.com gepr@tempusdictum.com hunt@itsa.ucsf.edu

Keywords: hepatic drug clearance, stochastic modeling, biological systems

Abstract

The Functional Unit Representation Method (FURM) [1] initiates a new procedure for the design of in silico models that are flexible, adaptable, and evolvable. We conduct experiments using FURM and an agent-based experimental framework to generate validated in silico hepatocytes that are capable of clearing simulated test drugs (e.g. diltiazem, FK1052, and acetaminophen) in such a way that the experimental outputs are indistinguishable from the real in vitro data. We call this experiment in silico hepatic intrinsic clearance (IS-HIC). A similarity measure was used to automate the evolution process of in silico hepatocytes, by scoring the simulation output based on its similarity to the in vitro published data and/or accepted mathematical models. The final output of the simulation is indistinguishable from in vitro published data.

1. INTRODUCTION

Computer simulations can increase the effectiveness drug discovery and development Pharmacokinetics, tracking the time course of drug metabolism, distribution, and elimination, is an important aspect of that research. There are two main approaches used by researchers to perform pharmacokinetics simulations. The first, known as physiologically based pharmacokinetic (PBPK) modeling, uses mathematical equations similar to those used by engineers, i.e. it formulates the system based on chemical rate laws governing species concentration and locations, leading to a set of linear or non-linear differential equations. PBPK models are used to represent the flow of fluids and the chemicals that they contain through the "pipes and tanks" of the body [2-6]. The resulting simulations are deterministic, i.e., the trajectory of the system is along a fixed path from its initial state. Such simulations assume that the concentration (in the simulated biological system) is a continuous (real) value. This assumption is valid in many situations where the volumes are large and the contents are well-mixed and the number of molecules of interest is also large, which is not always the case. As a consequence the discrete nature of change at low concentrations needs to be taken into account [7].

The second approach uses *stochastic modeling* to keep track of every single or group of molecules or cells in the system. Concentrations are assumed to be discrete values, which are stochastically changed in response to reaction events. A limitation of stochastic algorithms is that they are computationally expensive. An overview of approaches used for modeling stochastic phenomena in cellular systems is presented in [7]. There are additional technologies to predict drug metabolism, including rule-based tools, electronic models and homology models (surveyed in [8]).

The Functional Unit Representation Method (FURM) is introduced in [1] as a method to simulate biological systems. It seeks more flexible, adaptable and evolvable biological models, whose base components are the modular functional units that are designed to be easily decomposed for reuse in other biological models. The first demonstration example of this class of models is the ISL [1].

The primary goal of this work is to show how easily one can decompose an existing ISL and reuse its component submodels to build new models. The consequential purpose of this report is to provide validated in silico hepatocytes for use in simulating in vitro experiments. They, in turn, can be reused in other models. We use FURM to implement flexible in silico biomimetic experiments that produce data that can be used to calculate a measure that corresponds directly to intrinsic drug clearance, the standard measure of drug metabolism. That measure is the in silico hepatic intrinsic clearance (IS-HIC). The experimental framework reuses the agents from our previous model, the in silico Liver (ISL) [1, 9], that were used to represent hepatocyte metabolic functions within that larger system. Using a stochastic approach, the ISL simulates the clearance of drugs in the liver, and is validated against data obtained from experiments on isolated perfused rat livers. The ISL is implemented in a discrete-event manner using the Swarm [10] multi-agent software platform. The primary functional unit of the ISL is a component that maps to the liver lobule, which is a network of interconnected tubes called sinusoids through which blood flows after entering the liver. In the model, the sinusoids are formalized as sinusoidal segments (SS) and are represented as nodes in a graph connected by edges that show their interconnections and directions of fluid flow. Each SS is further composed of lower level components that map to the endothelial layer, the space of Dissé, hepatocytes, and bile canaliculi. The walls of hepatic sinusoids consist of endothelial cells, the cell type that forms capillaries in all mammals. Behind that layer is the space of Dissé, and it is bounded on the other side by hepatocytes. Hepatocytes are responsible for most of the biochemical functions of the liver. They produce bile, which flows through the bile canaliculi and out of the liver. However, the primary functional unit of IS-HIC is the hepatocyte. In the simulation hepatocytes are represented using agents while compounds are mobile objects. The simulated hepatocytes interact with a simulated test drug within a 2D space. The IS-HIC framework measures the in silico clearance (Cl) of drug objects by the in silico hepatocytes.

Clearance is one of the more useful pharmacokinetic parameters. It is a measure of the ability of a tissue or an organ such as liver or a kidney to irreversibly remove a compound from blood via metabolization or elimination [4]. In vivo, when that removal is due to metabolism, then the observed clearance is a function of blood flow rate to the tissue or organ and the intrinsic metabolic clearance, CL_{int} , a flow independent measure of the tissue or organ's ability to metabolize available compounds. It is the proportionality constant relating the rate of drug metabolism (V) and drug concentration at the enzyme site (C_E) [5]. The intrinsic metabolic clearance is the basis for the extrapolation of in vitro to in vivo measures of metabolism.

In the section that follows we briefly describe the implementation of the IS-HIC as a FURM framework, using components borrowed directly from the ISL and its framework. We then describe the experiments that were conducted to validate the in silico hepatocytes and the results of those experiments. Our concluding observations are presented in the Discussion.

2. METHODS

In the IS-HIC context FURM and its framework uses three models: an articulated*, functional unit model (ArtModel), an accepted mathematical model—the reference model (RefModel), and *in vitro* experimental data (DatModel) for validating the ArtModel. Each of the above three models is described below.

2.1 The Reference Model (RefModel)

RefModel is any accepted mathematical model that describes *in vitro* hepatic clearance under specific conditions. As long as there is a local equilibrium between drug and metabolic enzymes, metabolic elimination of that drug by metabolism generally follows first order kinetics. The metabolic rate decreases in proportion to the local drug concentration as given by equation (1) [12], where k is a first order rate constant.

$$\frac{dC}{dt} = \Box kC(t) \tag{1}$$

Intrinsic clearance is a measure of enzyme activity, and is independent of physiological factors such as the liver blood flow or drug binding in the blood. In vitro, the intrinsic clearance of a drug is commonly expressed by equation 2, where C_s is the concentration of the unbound drug at the enzyme site [12].

Rate of metabolism:
$$V = CL_{int} C_s$$
 (2)

Rate of metabolism is generally defined by the Michaelis-Mention enzyme kinetics relationship [12]. When the drug concentration is much smaller than K_m (the Michaelis-Mention constant), CL_{int} becomes:

$$CL_{int} = V/C_s$$
 (3)

What we need for our mathematical reference model is an equation that describes the concentration of the unchanged drug as a function of time. Researchers use the following equation to express the concentration of unchanged drug for in vitro experiments, when keeping the initial drug concentration much smaller than K_m [11-12]:

$$\frac{dC}{dt} = \Box CL \Box D \Box C(t) \tag{4}$$

where C(t) is the concentration, CL the in vitro clearance of the drug and D the cell density.

By solving differential equation (4) the amount of drug remaining after incubation time T is expressed as follows:

$$C(T) = C(0) \prod_{i=0}^{n} e^{\prod_{i=0}^{n} CL \prod_{i=0}^{n} T}$$
 (5)

The basic equations explained above cannot be applied to all drugs [4], but they are applicable to the drugs and conditions selected for this study, and so we use equation 5 to express *in vitro* clearance and to be the reference model for IS-HIC.

2.2. The Articulated Model (ArtModel)

The IS-HIC ArtModel consists of three basic components, solute objects, hepatocytes and a WanderSpace. The solute objects and hepatocyte agents

^{*} We use articulated to emphasize that that the IS-HIC model is assembled from components that are designed to fit together and, once assembled, to function as a whole.

are the exact same as the corresponding components used in the ISL [9].

Solute objects: solute objects are the mobile objects representing a group of molecules of the chemical compound (the test drug) as they move around in the reaction mixture. These in silico objects can be parameterized to make their behavior similar to that of a drug in vitro. The more relevant parameters are as follows. SoluteBindingProb: The probability that a specific type of drug object will bind to a Binder within a hepatocyte. SoluteBindingCycles: The number of steps that a drug object remain attached to a Binder; embraneCrossing: indicates whether a compound can or cannot enter hepatocytes. A solute object is destroyed once it is "metabolized." Metabolites are not currently tracked.

Binder: Objects within hepatocyte agents that can both bind and metabolize compounds. Each Binder represents a fraction of all subcellular components that can bind or sequester a drug object, and includes a fraction of all metabolizing enzymes.

Hepatocytes: These objects are the agents representing the isolated hepatocytes suspended in the in vitro cell culture media. They are capable of taking in, binding and metabolizing drug objects. The parameters are: BindersPerCellMin, BindersPerCellMax: Minimum and maximum binders per cell, respectively. MetabolizationProb: The probability that the cell metabolizes an attached solute object at the end of its binding period.

WanderSpace is a fine-grained space, representing the *in vitro* cell culture, in which solute objects and hepatocytes interact with each other. SpaceWidth, SpaceLength: Width and length of WanderSpace. HepDensity: The density of hepatocytes in the space. TotalSoluteMass: The total compound mass initially added to WanderSpace.

Functioning of the IS-HIC ArtModel: First, hepatocyte agents are placed randomly in the WanderSpace as fixed objects. Drug (and other) mobile objects are then randomly placed within the space (external to all hepatocytes) based on an initial concentration (see Figure 1). The mobile objects move randomly, while the hepatocytes have the opportunity to "take up" and then metabolize nearby solute objects.

As the simulation progresses, the number of solute objects decreases as a consequence of metabolism. At intervals this number was counted, normalized and scaled to represent the concentration of the unchanged drug. Figure 2 illustrates how IS-HIC keeps track of each solute object in the space.

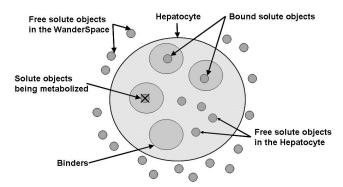


Figure 1. An illustration of in silico hepatocyte interaction with solute objects within the WanderSpace

Figure 3 is a block diagram of the whole simulation and the relationship of the primary modules within the simulation framework.

Figure 4 shows the three main components of the ArtModel: WanderSpace, simulated hepatocytes and solute objects, and their relationship. The simulation parameters (described in section 2.2) are the inputs to the ArtModel. The output is the simulated drug concentration, resulted from the in silico interaction of hepatocyte agents and solute objects in the WanderSpace.

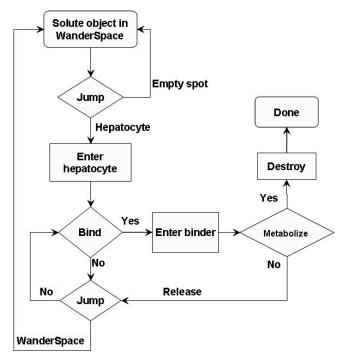


Figure 2. Movement of a solute object in IS-HIC

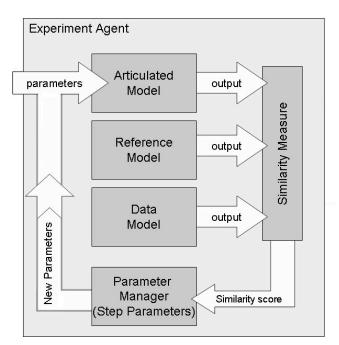


Figure 3. The block diagram of the simulation

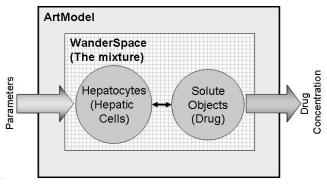


Figure 4. The components inside the ArtModel

2.3. The Data (DatModel)

The DatModel represents the real biological system. It contains the data obtained from *in vitro* experiments, and is used to validate the in silico model (ArtModel). The validation process involves measuring the output of the in silico model and comparing it with the data provided by DatModel. In this work the data was obtained from Fig. 2 of [11], which depicts the time course for each of nine unchanged compounds in cell culture media containing freshly isolated rat hepatocytes. The data were carefully obtained from the graphs using computer design tools. The DatModel interpolates the data points using a linear interpolation method to estimate the drug concentration at each time step of the simulation.

2.4. The Similarity Measure

After each in silico experiment a *similarity measure* algorithm compares the output of that experiment with the data provided by DatModel and assigns a score to the output based on the degree of their similarity. Several similarity measure algorithms are surveyed in [13]. The similarity measure used in this work was the "global standard deviation" [13] with a wider envelope: $upper_i = m_i (1 + sd) + 10$, $lower_i = m_i (1 - sd) - 10$, where m_i and sd are the nominal mean and standard deviation of the time series (for additional detail see [13]). The similarity score is calculated by counting the number of observations of the candidate time series that fall within the envelope and dividing that by the total observations in the series [13], as a result similarity- $score \sqcap [0,1]$.

3. EXPERIMENTS AND RESULTS

Three chemicals from [11] (Diltiazem, FK1050, and Acetaminophen) were chosen as our test drugs to be cleared in silico by simulated rat hepatocytes. *In vitro*, each compound (at a concentration of 1000 μ M) had been incubated for various time periods at 37°C with freshly isolated rat hepatocytes. The cell density in the reaction mixture is reported 0.5 \square 10⁶ (cells/ml) for FK1052 and Diltiazem, and 1 \square 10⁶ (cells/ml) for Acetaminophen. *Calculation of parameter*. Because the Hepatocytes and the test drug are in the same volume of mixture,

$$C_{1} = \frac{P_{1}}{V} C_{2} = \frac{P_{2}}{V} \qquad \frac{P_{1}}{P_{2}} = \frac{C_{1}}{C_{2}}$$
 (6)

For example consider FK1052:

$$\frac{P_1}{P_2} = \frac{1000(pmol/ml)}{0.5 \square 10^6 (cells/ml)} = \frac{1000(fmol)}{500(cells)}$$

To encompass 500 cells we need 0.001ml of the mixture (V = cells/cell-density = $500/.5 \Box 10^6$ = 0.001ml). Assuming each spot in the WanderSpace corresponds to $350 \Box 10^{-9}$ ml of the mixture, for 0.001ml we will need 2857 spots. So we chose the WanderSpace to be 53 by 54 (=2862). Other parameters were chosen as follows: HepDensity = 500/2857 = 0.175, TotalSoluteMass = 1000 fmol.

Other parameters (SoluteBindingProb,SoluteBinding-Cycles, BindersPerCe-llMin, BindersPerCellMax and MetabolizationProb) were iteratively swept to optimize the output—an acceptable similarity score was attained.

Figure 5 shows the output of the IS-HIC ArtModel using above parameterization along with *in vitro* clearance profiles of Diltizem, FK1052, and Acetaminophen. The simulation results are in a good agreement with both the mathematical model and the in vitro data.

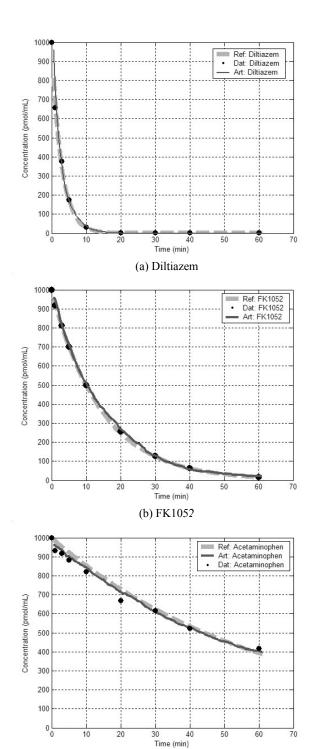


Figure 5. ArtModel outputs against RefModel and DatModel: (a) Diltizem, (b)FK1052, (c) Acetaminophen

(c) Acetaminophen

Table 1. The parameter values and the corresponding similarity scores

Test Drug Parameters	Diltiazem	FK1052	Acetami- nophen
SoluteBindingProb	0.14	0.022	0.0028
SoluteBindingCycles	1	2	3
MetabolizationProb	0.5	0.5	0.5
HepDensity	0.175	0.175	0.350
BindersPerCellMin	5	5	5
BindersPerCellMax	10	10	10
TotalSoluteMass	1000	1000	1000
WanderSpace size	53∏54	53∏54	53∏54
Similarity Score	0.885	0.893	0.984

Table 1 shows the final values of parameters which resulted in satisfactory similarity scores.

Figure 6 shows the similarity score as a function of two parameters *SoluteBindingCycles* and *SoluteBindingProb* for FK1052. The similarity score is maximum when *SoluteBindingCycles* = 2 and *SoluteBindingProb* = 0.022.

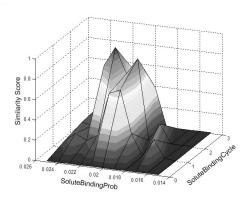


Figure 6. The similarity score as a function of two parameters: *SoluteBindingCycles* and *SoluteBindingProb* for FK1052.

4. CONCLUSION AND DISCUSSION

This study was done with two goals in mind: first, show how easily and conveniently the existing FURM-developed models could be reused to perform new in silico experiments; second, provide verified in silico FURM components representing hepatocytes and compound of interest that can be used in other in silico experiments. We used ISL components [9] to perform totally new in silico experiments that provided measures of IS-HIC (in silico Hepatic Intrinsic Clearance). The IS-HIC is much simpler and faster than the ISL, because it has only three main elements: hepatocytes, test drugs and the suspension media. We validated hepatocyte models

er1.pdf.

that are capable of clearing test drugs (Diltiazem, FK1052, and Acetaminophen) in such a way that the outputs are indistinguishable from actual *in vitro* data. The resulting IS-HIC provides a clean measure of in silico drug clearance which does not depend on the blood flow in the liver sinusoids. However, in silico hepatocytes that have been newly parameterized and validated as above for drug X can be used to replace the existing hepatocytes within a separately validated ISL producing a new ISL model that can be used to estimate the expected hepatic clearance of X.

The in silico model used here is not appropriate for simulating hepatic intrinsic clearance of compounds such as tolbutamide that are slowly metabolized because cell death is not accounted for in this work. In fact, even the *in vitro* clearance of such low extraction ratio drugs is not easy to predict [14]. Sophisticated *in vitro* methods have been proposed to measure the *in vitro* intrinsic clearance of slowly metabolized drugs [14-15].

A practical limitation may be Swarm [10] itself. It is not an easy-use platform for non-computer scientists. It is designed to serve as a generic platform for modeling and simulating complex space-time dynamics. It provides a set of classes for defining an agent's properties and behaviors using computer language objective C.

ACKNOWLEDGMENT

We are grateful for the funding (R21-CDH-00101) provided by the CDH Research Foundation.

REFERENCES

- [1] Hunt, C.A.; G.E.P. Ropella; M.S. Roberts; L. Yan. 2004. "Biomimetic In Silico Devices. Computational Methods in Systems Biology." *Second International Workshop, CMSB 2004 (Paris, France, May 26-28) Proceedings. Lecture Notes in Bioinformatics*, Springer (in press); available at http://biosystems.ucsf.edu/Researc/RecentPapers/HuntCMS B04b.pdf.
- [2] Leahy, D.E. 2004. "Drug Discovery Information Integration: Virtual Humans for Pharmacokinetics." DDT: *Biosilico 2*, No. 2:78-84.
- [3] Lipscomb, J.C.; M. Meek; K. Krishnan; G.L. Kedderis; H. Clewell; L. Haber. 2004. "Incorporation of Pharmacokinetic and Pharmacodynamic Data Into Risk Assessments." *Toxicology Mechanisms and Methods* 14, No. 3:145-158.
- [4] Gunaratna, C. 2001 "Drug Metabolism and Pharmacokinetics in Drug Discovery: A Primer for Bioanalytical Chemists, Part II." Current Separations 19, No. 3 (www.currentseparations.com/issues/19-3/19-3e.pdf.
- [5] Venkatakrishnan, K.; L.L. von Moltke; D.J. Greenblatt. 2001. "Human Drug Metabolism and the Cytochromes P450: Application and Relevance of In Vitro Models." *Journal of Clinical Pharmacology* 41, No. 11:1149-1179.

- [6] Takahashi, K.; K. Kaizu; B. Hu; M. Tomita. 2004. "A Multi-algorithm Multi-timescale Method for Cell Simulation." *Bioinformatics* 20, No. 4:538-46.
- [7] Meng, T.C.; S. Somani; P. Dhar. 2004. "Modeling and Simulation of Biological Systems with Stochasticity." In Silico Biology 4:0024.
- [8] S. Eklins. 2003. "In Silico Approaches to Predicting Drug Metabolism, Toxicology and Beyond." *Biochemical Society Transactions* 31(Pt 3):611-4.
- [9] Ropella, G.E.P. and C.A. Hunt. 2003. "Prerequisites for Effective Experimentation in Computational Biology." 25th Annual Conference of the IEEE Engineering in Medicine and Biology Society (Cancun, September 17-21); available at http://128.218.188.153:8080/~gepr/furm/docs/EMBC03Pap
- [10] Daniels, M. 1999. "Integrating Simulation Technologies with Swarm." *Agent Simulation: Applications, Models and Tools Conference* (University of Chicago, October); available at http://www.santafe.edu/~mgd/anl/anlchicago.html.
- [11] Naritomi, Y.; S. Terashita; A. Kagayama; Y. Sugiyama. 2003. "Utility of Hepatocytes in Predicting Drug Metabolism: Comparison of Hepatic Intrinsic Clearance in Rats and Humans In Vivo and In Vitro." *Drug Metabolism* and Disposition 31, No. 5:580-588.
- [12] Shibata, Y.; H. Takahashi; Y. Ishii. 2000. "A Convenient In Vitro Screening Method for Predicting In Vivo Drug Metabolic Clearance Using Isolated Hepatocytes Suspended in Serum." Drug Metabolism and Disposition 28, No. 12:1518-1523.
- [13] Ropella, G.E.P.; D.A. Nag; C.A. Hunt. 2000. "Similarity Measures for Automated Comparison of In Silico and In Vitro Experimental Results." Ibid; available at http://128.218.188.153:8080/~gepr/furm/docs/EMBC03Pap er2.pdf.
- [14] Treijtel N.; A. Barendregt; A.P. Freidig; B.J. Blaauboer; J.C.H. van Eijkeren. 2004. "Modeling the In Vitro Intrinsic Clearance of the Slowly Metabolized Compound Tolbutamide Determined in Sandwich-Cultured Rat Hepatocytes." *Drug Metabolism and Disposition 32*, No. 8:884-891.
- [15] Haenen, B.; C. Rompelberg; K. Van Twillert; M. Hamzink; J. Dormans; J. Van Eijkeren. 2002. "Utility of Rat Liver Slices to Estimate Hepatic Clearance for Application in Physiologically Based Pharmacokinetic Modeling: A Study With Tolbutamide, a Compound with Low Extraction Efficiency." *Drug Metabolism and Disposition 30*, No. 3:307-313.
- [16] Neddermeijer, H.G.; van Oortmarssen, G,H.; Piersma, N.; Dekker, R. 2000. "Adaptive extensions of the Nelder and Mead Simplex Method for optimization of stochastic simulation models." Econometric Institute Report 199. Econometric Institute, Erasmus University, Rotterdam.

[☐] For more information about Swarm please visit: wiki.swarm.org.