### Applied Genome Research

#### **Annotation**

205048 & 205049

# **QUESTION**

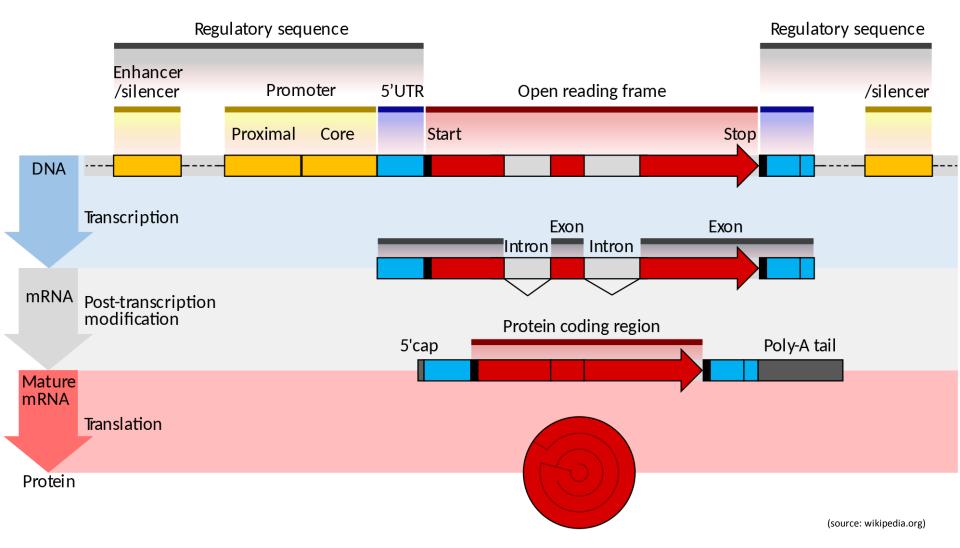
What is a gene?



### Prokaryotic gene structure

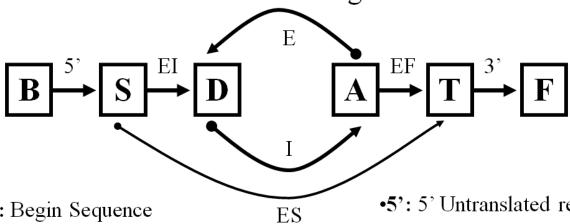
Gene = sequence from start codon to stop codon (without any gaps)

### Eukaryotic gene structure



### Gene prediction - theory

Basic Gene-finding HMM



- •B: Begin Sequence
- •S: Start translation
- •D: Donor splice site
- •A: Acceptor splice site
- •T: Stop translation
- •**F**: End sequence

•5': 5' Untranslated region

•EI: Initial exon

•ES: Single exon

•E: Exon

•I: Intron

•EF: Final exon

•3': 3' untranslated region

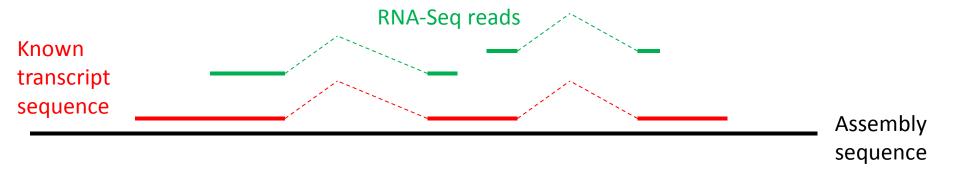
(source: blogspot.com)

### Gene prediction - different modi

- *ab initio* = gene prediction without any additional information
- hint guided = RNA-Seq data or similar information are used to support prediction
- Reference-based = map annotated sequences from a reference to a newly assembled sequence

### Hint-based gene prediction

RNA-Seq reads or known transcript sequences are mapped



 Due to a lack of RNA-Seq data for Nd-1 gene prediction will be computed ab initio.

#### Gene prediction - AUGUSTUS

```
$ augustus-3.2\
--species=<SPECIES> ... name of parameter set to use
--gff3=on ... output format is GFF3
--codingseq=on ... encoded protein sequences are written in output files
<FASTA_FILE_NAME>
> .... write output into file (instead of printing it to screen)
<outputfile>
```

Running ab initio gene prediction.

# Gene prediction - AUGUSTUS

```
$ getAnnoFasta.pl\
--seqfile=<assembly_file>\
<GFF3_file>
```

Extraction of sequences of different predicted features.

#### **EXERCISE**

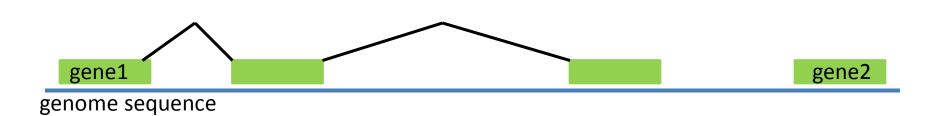
- Run AUGUSTUS on SOAPdenovo2 contigs!
- Run AUGUSTUS on SOAPdenovo2 scaffolds!
- Run AUGUSTUS on SSPACE scaffolds!
- Analyze the differences!
- Find 'getAnnoFasta.pl' (online), download it, and use it!

### 'Fragmented genes'

Short read assembly:



#### Reality:



#### **GFF** = **Generic Feature Format**

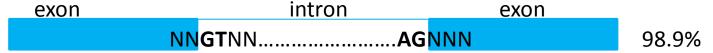
	Chr1»	TAIR10» chromosome»	1 >	30427671	>		.» .» ID=Chr1;Name=Chr1	
	Chr1»	TAIR10» gene» 3631»	5899»		+»	. >	ID=AT1G01010; Note=protein_coding_gene; Name=AT1G01010	
	Chr1»	TAIR10» mRNA» 3631»	5899»		+»	. »	ID=AT1G01010.1;Parent=AT1G01010;Name=AT1G01010.1;Index=1	
	Chr1»	TAIR10» protein»3760»	5630»		+»	. >	ID=AT1G01010.1-Protein; Name=AT1G01010.1; Derives_from=AT1G01010.1	
9	Chr1»	TAIR10 exon 3631	3913	. »	+»	. >	Parent=AT1G01010.1	
	Chr1»	TAIR10» five prime UTR		3759»	. >	+>	.» Parent=AT1G01010.1	O a l:4
	Chr1»	TAIR10 CDS 3760	3913»		+>	O>	Parent=AT1G01010.1,AT1G01010.1-Protein:	— Quality
•	Chr1»	TAIR10» exon» 3996»	4276»	. »	La		Parent-Arrigororo.i	/
Sequence	Chr1»	TAIR10 CDS 3996	4276»	, »	+>	2>	Parent=AT1G01010.1,AT1G01010.1-Protein;	
009000000	Chr1»	TAIR10 exon 4486	4605»	. »	+>	. >	Parent=AT1G01010.1	
nama	Chr1»	TAIR10 CDS 4486	4605»	. »	+>	0 »	Parent=AT1G01010.1,AT1G01010.1-Protein;	
name	Chr1»	TAIR10 exon 4706	5095»	. »	+>	. >	Parent=AT1G01010.1	
	Chr1»	AIR10» CDS» 4706»	5095»	. »	+>	0»	Parent=AT1G01010.1,AT1G01010.1-Protein;	0
	Chr1	TAIR10» exon» 5174»	5326	. »	+>	. >	Parent=AT1G01010.1	<ul><li>Orientation</li></ul>
	Chr 1»	TAIR10° CDS° 5174°	5326»	. »	+>	0»	Parent=AT1C01010.1,AT1801010.1-Protein;	
	Chr1»	TAIR10» exon» 5439»	5899»	. »	+>	. >	Parent=AT1G01010.1	
	Chr1»	TAIR10» CDS» 5439»	5630»		+»	0»	Parent=AT1G01010.1,AT1G01010.1-Protein;	
Source	Chr1»	TAIR10» three_prime_UTF		5899»	. »	+>	.» Parent=AT1G01010.1	
Jource	Chr1	TAIR10» gene» 5928»	8737	. >	- »	. >	ID=AT1G01020;Note=protein_coding_gene;Name=AT1G01020	
	Chr1»	TAIR10» mRNA» 5928»	8737	0.50	- »	. >	ID=AT1G01020.1;Parent=AT1G01020;Name=AT1G01020.1;Index=1	
	Chr1»	TAIR10 protein 6915 TAIR10 five_prime_UTR	8666		- »	. >	ID=AT1G01020.1-Protein;Name=AT1G01020.1;Derives_from=AT1G01020.1	
	Chr1»	TAIR10» five_prime_UTR:		8737»	. »	- >	.» Parent=AT1G01020.1	<ul><li>Comments</li></ul>
	Chr1	TAIR10» CDS» 8571»	8666»	. »	- >	0»	Parent=AT1G01020.1,AT1G01020.1-Protein;	- Comments
Footure *	chr1»	TAIR10» exon» 8571»	8737	. »	- »	. >	Parent=AT1G01020.1	
Feature 7	Chr1»	TAIR10» CDS» 8417»	8464»	-	- »	0»	Parent=AT1G01020.1,AT1G01020.1-Protein;	
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Start	Chr1» Chr1»	TAIR10» exo 7564» TAIR10» CDS» 7384»	7649» 7450»	. "	- >	1	Parent=AT1601020.1	
Start	Chr1»		7450°	-	- >	1 »	Parent=AT1G01020.1,AT1G01020.1-Protein; Parent=AT1G01020.1	
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	enr1»	TAIR10 CDS 6915	7069°	. *	- »	2×	Parent=AT1G01020.1, AT1G01020.1-Protein;	
	Chr1»	TAIR10 three prime UT		6914	- //	- »	.» Parent=AT1G01020.1	
End /	Chr1»	TAIR10° cm ee_prime_om	7069		- >	. >	Parent=AT1G01020.1	
LIIU	Chr1»	TAIR10 three prime UT		6263	7.0	- »	.» Parent=AT1G01020.1	
	Chr1»	TAIR10 exon 5928	6263		- »	.»	Parent=AT1G01020.1	
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				10.500		-		

### Gene prediction – reference-based

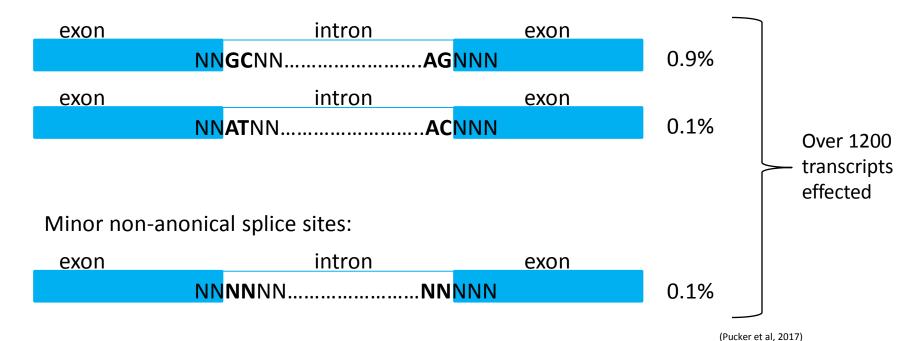
- Arabidopsis thaliana Col-0 reference sequence is very well annotated
- Mapping of Col-0 sequences to Nd-1 assembly might be useful
- Annotation data are available via TAIR10 and Araport11

### Non-canonical splice sites



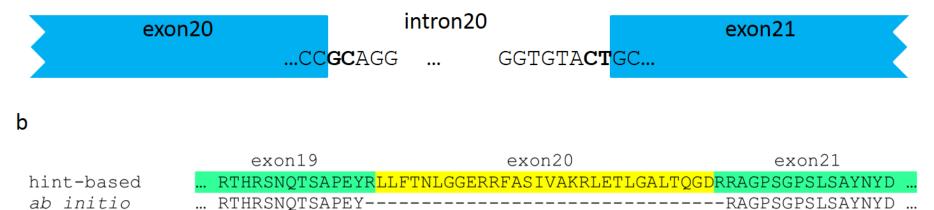


#### Major non-anonical splice sites:



#### Example: non-canonical splice sites





(source: Pucker et al., 2017)

- Only canonical splice sites (GT-AG) can be predicted ab initio
- Hint-based prediction of gene structures is more accurate

### QUESTION/EXERCISE

- Have a look at TAIR10 and Araport11!
- How many protein-coding genes are annotated in Araport11?
- How can you find publications describing certain genes?
- Are there websites for other model organisms?
- List some databases for (model) organisms!

#### BLAST – sequence comparison

- Very efficient tool for sequence comparison
- Very highly cited publication by Altschul et al., 1990 (indicates importance)
- Comparison is based on matching words (seeds) which are than extended to final alignments
- NCBI offers web-based service
- Command line version of BLAST is much more efficient (for large data sets)
- Submitting data to the NCBI is not always allowed (e.g. clinical data)

#### BLAST – command line

```
$makeblastdb –in <fasta_file> -out <some_name> -dbtype 'nucl' $ blastn\ .... There are different verions of BLAST (n/p/x...) -query <query_file> ... file with sequences to search for -subject <subject_file> ... file with sequences to search in (-db <some_name>) -out <output_file> ... file for results -outfmt 6 ... set output format to table -evalue 0.001 ... set e-value cutoff (-num_threads 4 ... set number of threads to use for search)
```

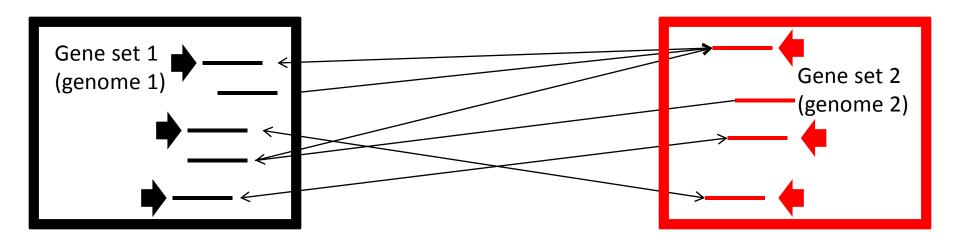
#### **EXERCISE**

- Run BLASTn search of Col-0 exons vs. SOAPdenovo2 contigs of Nd-1!
- How many hits?
- What is the next step?

#### BLAST – annotation transfer

- High sequence similarity indicates a common function of two genes/proteins
- Identification of Reciprocal Best Hits (RBHs) can be used to transfer functional annotations
  - Two data sets:
    - SeqA, SeqB, SeqC, ....
    - Seq1, Seq2, Seq3, ...
  - Best hit of SeqA is Seq2 and best hit of Seq2 is SeqA => RBH

#### Gene set comparison

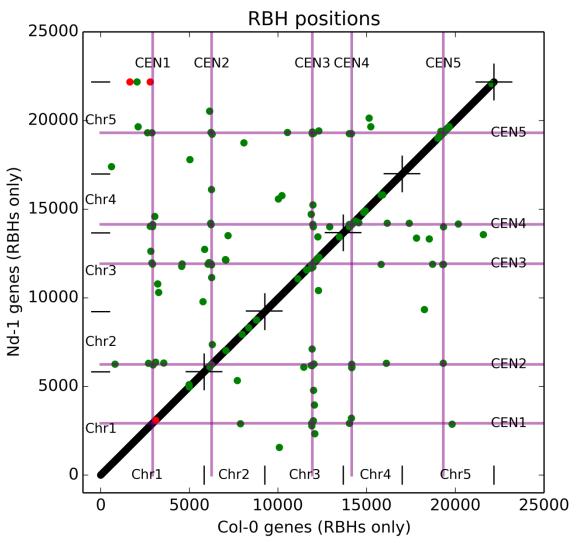


- RBH = <u>Reciprocal Best</u> (BLAST) <u>Hit</u>
- Comparison can be done on DNA or peptide sequence level
- Broad range of applications!

#### **EXERCISE**

- Identify RBHs between Col-0 and Nd-1 on peptide sequence level via identify\_RBHs.py!
- How many RBHs are there?
- Map TAIR10 annotation to the identified RBHs via map\_annotation.py!
- Select one gene and collect additional (functional) information!
- Collect information about all these candidate genes and find a systematic pattern!

### Application of RBHs



(source: Pucker et al., 2016)