

Computing with Sequences and Ranges

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Sequences: packages

Biostrings General purpose biological sequence representation.

BSgenome Whole-genome representation.

ShortRead High-throughput sequencing.

Sequences: representation

DNASTringSet: Vector of sequences, e.g., sequence of each exon in the UCSC knownGene track

A *DNASTringSet* instance of length 289969

width seq

```
[1]      354 CTTGCCGTCAGCCTTT...TCACAACCTAGGCCA
[2]      127 GCTCCTGTCTCCCCC...CCCAGTGTTGCAGAG
[3]      109 GTGTGTGGTGATGCCA...CCCAGTGTTGCAGAG
...      ...
[289968]    109 GTGTGTGGTGATGCCA...CCCAGTGTTGCAGAG
[289969]    354 CTTGCCGTCAGCCTTT...TGACAACCTAGGCCA
```

- ▶ Acts like a *vector*, e.g., `length()`, `[`, `[[`
- ▶ Many methods – `methods(class="DNASTringSet")` – e.g., `reverseComplement()`, `letterFrequency()`, ...

Sequences: common classes

- DNASTring** Single DNA sequence, e.g., chromosome
- DNASTringSet** Vector of DNA sequences. Actually, XString, XStringSet: X could be DNA, RNA, AA)
- BSgenome** Collection of (large) DNA sequences
- ShortReadQ** High-throughput reads & their qualities

Sequences: file references

`TwoBitFile`, `FaFile` .2bit (in *rtracklayer*) or .fa (in *Rsamtools*)
indexed genome-scale fasta files.

`FastqFile` , e.g., *FastqStreamer* (in *ShortRead*)

Use – effectively manage large data

- ▶ *Restrict* input to specific genomic locations (specified by `GRanges()`).
- ▶ *Iterate* through large files in chunks (see `GenomicFiles::reduceByYield()`)

Sequences: annotations

*BSgenome.** packages

- ▶ E.g., *BSgenome.Hsapiens.UCSC.hg19*
- ▶ Packages containing whole-genome sequences for model organisms

AnnotationHub resources

- ▶ e.g., Ensembl FASTA files in *FaFile* format

Ranges: packages

GenomicRanges Essential representation and operations

GenomicAlignments Aligned reads as genomic ranges

GenomicFeatures Annotations as genomic ranges

rtracklayer Annotation (e.g., BED, GTF) input

A little more advanced usage: *IRanges* (); *S4Vectors* (underling conceptual ideas)

Ranges: *GRanges* representation

```
> gr = exons(TxDb.Hsapiens.UCSC.hg19.knownGene); gr
```

GRanges with 289969 ranges and 1 metadata column:

	seqnames	ranges	strand	exon_id
	<Rle>	<IRanges>	<Rle>	<integer>
[1]	chr1	[11874, 12227]	+	1
[2]	chr1	[12595, 12721]	+	2
[3]	chr1	[12613, 12721]	+	3
...
[289967]	chrY	[59358329, 59359508]	-	277748
[289968]	chrY	[59360007, 59360115]	-	277749
[289969]	chrY	[59360501, 59360854]	-	277750

seqinfo: 93 sequences (1 circular) from hg19 genome

GRanges

```
length(gr); gr[1:5]  
seqnames(gr)  
start(gr)  
end(gr)  
width(gr)  
strand(gr)
```

DataFrame

```
mcols(gr)  
gr$exon_id
```

Seqinfo

```
seqlevels(gr)  
seqlengths(gr)  
genome(gr)
```

- Data: aligned reads, called peaks, SNP locations, CNVs, ...
- Annotation: gene models, variants, regulatory regions, ...

Ranges: *GRangesList* representation

```
> grl = exonsBy(TxDb.Hsapiens.UCSC.hg19.knownGene, "tx", use.names=TRUE); grl
```

GRangesList of length 82960:

\$uc001aaa.3

GRanges with 3 ranges and 3 metadata columns:

	seqnames	ranges	strand	exon_id	exon_name	exon_rank
	<Rle>	<IRanges>	<Rle>	<integer>	<character>	<integer>
[1]	chr1	[11874, 12227]	+	1	<NA>	1
[2]	chr1	[12613, 12721]	+	3	<NA>	2
[3]	chr1	[13221, 14409]	+	5	<NA>	3

GRangesList
(list of *GRanges*)
length(grl)
grl[1:3]
shift(grl, 1)
range(grl)

\$uc010nxq.1

GRanges with 3 ranges and 3 metadata columns:

	seqnames	ranges	strand	exon_id	exon_name	exon_rank
[1]	chr1	[11874, 12227]	+	1	<NA>	1
[2]	chr1	[12595, 12721]	+	2	<NA>	2
[3]	chr1	[13403, 14409]	+	6	<NA>	3

GRanges
grl[[2]]
grl[["uc010nxq.1"]]

\$uc010nxr.1

GRanges with 3 ranges and 3 metadata columns:

	seqnames	ranges	strand	exon_id	exon_name	exon_rank
[1]	chr1	[11874, 12227]	+	1	<NA>	1
[2]	chr1	[12646, 12697]	+	4	<NA>	2
[3]	chr1	[13221, 14409]	+	5	<NA>	3

Two kinds of fun!
introns =
psetdiff(range(grl), grl)

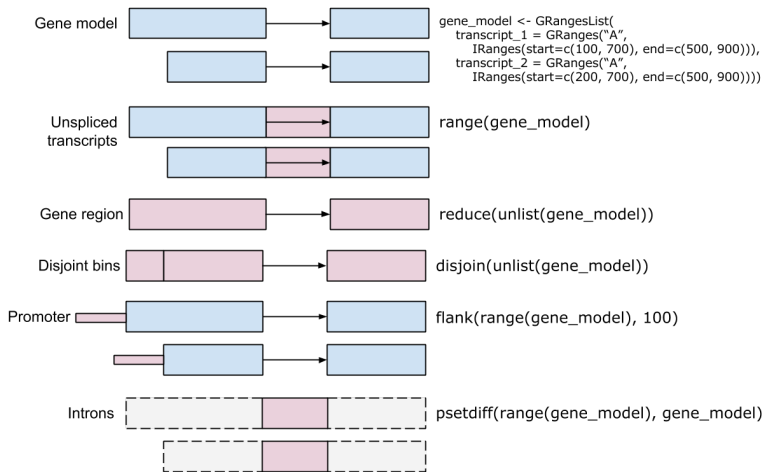
grr = unlist(grl)
transform grr, then...
grl = relist(grr, grl)

'flesh' 'skeleton'

...
<82957 more elements>

seqinfo: 93 sequences (1 circular) from hg19 genome

Ranges: operations



- Many more, e.g., `methods(class="GRanges")`

Ranges: `findOverlaps()`

- ▶ Overlaps between query and subject genomic ranges
- ▶ Different types of overlap, e.g., 'any', 'within', ...

```
> q <- GRanges("chr1", IRanges(10, 20))  
> s <- GRanges("chr1", IRanges(5, width=c(3, 6, 9)))  
> findOverlaps(q, s)
```

Hits object with 2 hits and 0 metadata columns:

	queryHits	subjectHits
	<integer>	<integer>
[1]	1	2
[2]	1	3

queryLength: 1
subjectLength: 3

- ▶ *Hits* object describing many-to-many relationship between overlapping ranges.

Ranges: working with files

`import` (*rtracklayer*) for BED, GTF, and other common web file import functions. *BEDFile*, *GTFFile*, etc.

`readGAlignments` / `readGAlignmentsList` (*GenomicAlignments*) for aligned reads in BAM files

`BamFile` (*Rsamtools*) for lower-level access to BAM files, e.g., restriction and iteration

Ranges: annotation

*TxDb.** packages

- ▶ E.g., *[TxDb.Hsapiens.UCSC.hg19.knownGene](#)*
- ▶ Genomic ranges for exons, transcripts, coding sequences, and how these are ordered into gene models, e.g., exons grouped by transcript

AnnotationHub resources

- ▶ Ensembl gene models
- ▶ Roadmap Epigenomics regulatory marks
- ▶ Many other range-based resources

Demos

See markdown document.

Other resources

- ▶ [Workflows](#) & package vignettes
- ▶ [GenomicRanges](#) and other ‘[cheat sheets](#)’
- ▶ [Course material](#)

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