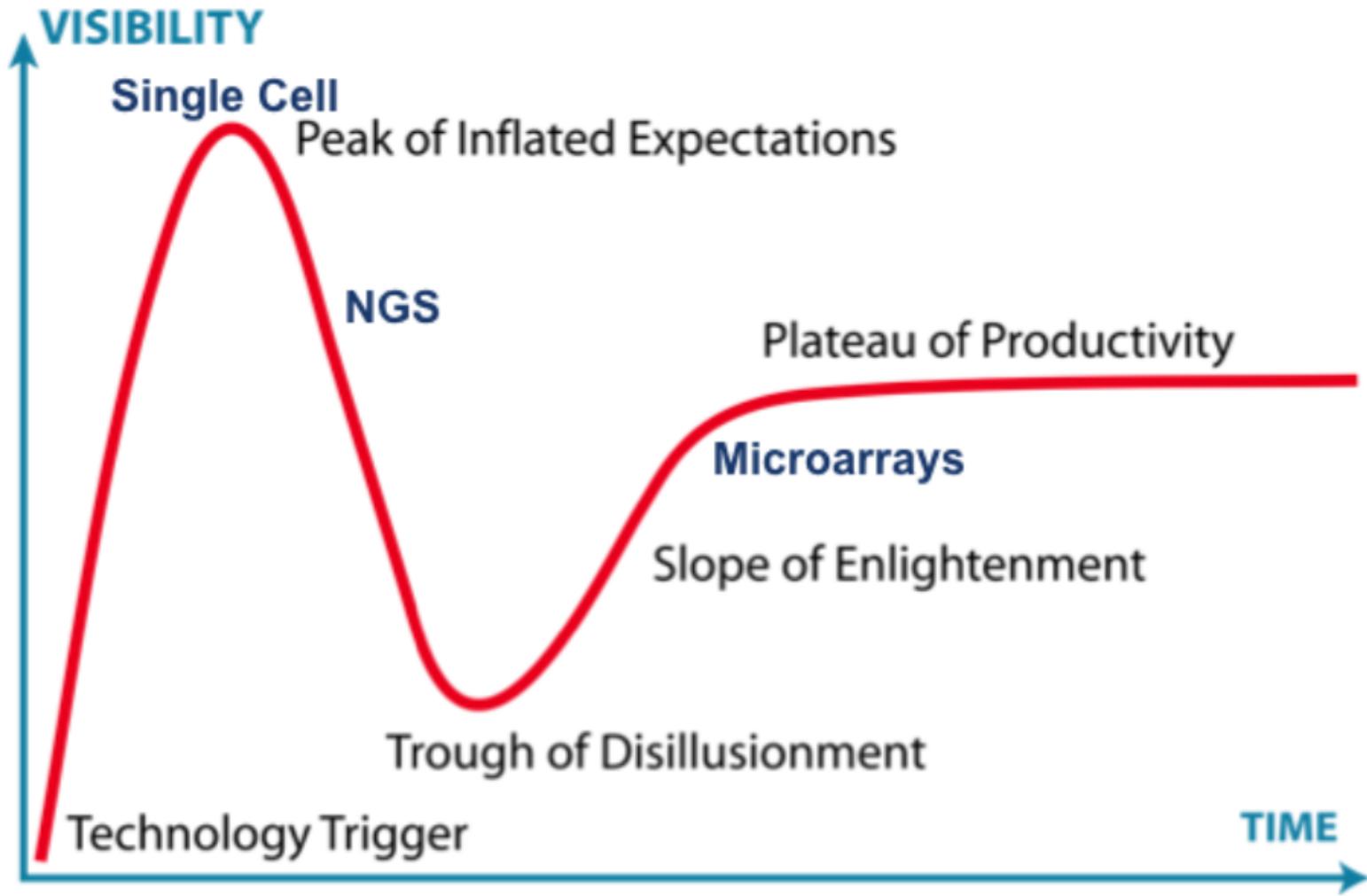


Single-cell transcriptomics overview

Alejandro Reyes
Huber group
CSAMA 2015



Rafael A Irizarry @rafalab · Feb 15

@Y_Gilad @joe_pickrell @davisjmcc @leonidkruglyak Are you not familiar with the "hype cycle" ?



Yoav Gilad @Y_Gilad · Feb 13

I guess single-cell data are so exciting that we all momentarily forgot everything we knew about study design, modeling, and multiple tests?



39



45

•••

NATURE BIOTECHNOLOGY | OPINION AND COMMENT | CORRESPONDENCE

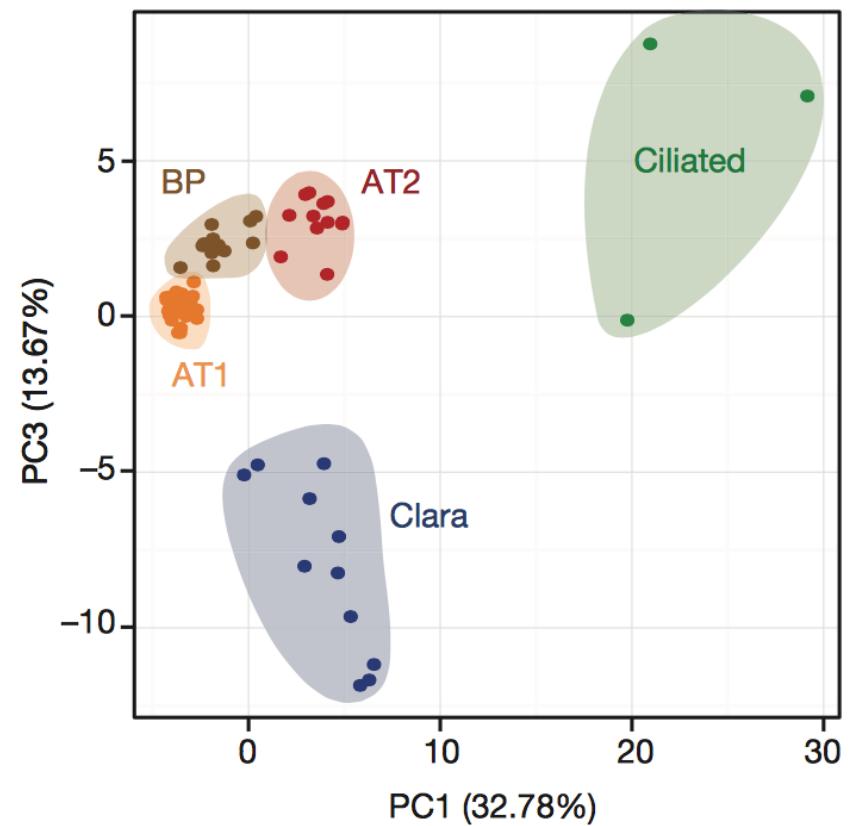
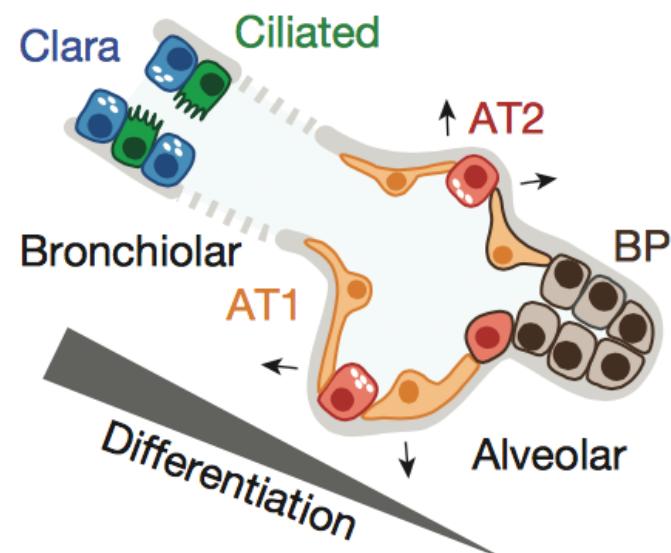
Sequencing technology does not eliminate biological variability

Kasper D Hansen, Zhijin Wu, Rafael A Irizarry & Jeffrey T Leek

Why?
(optimistic)

Research questions that can be answered with single-cell transcriptomics

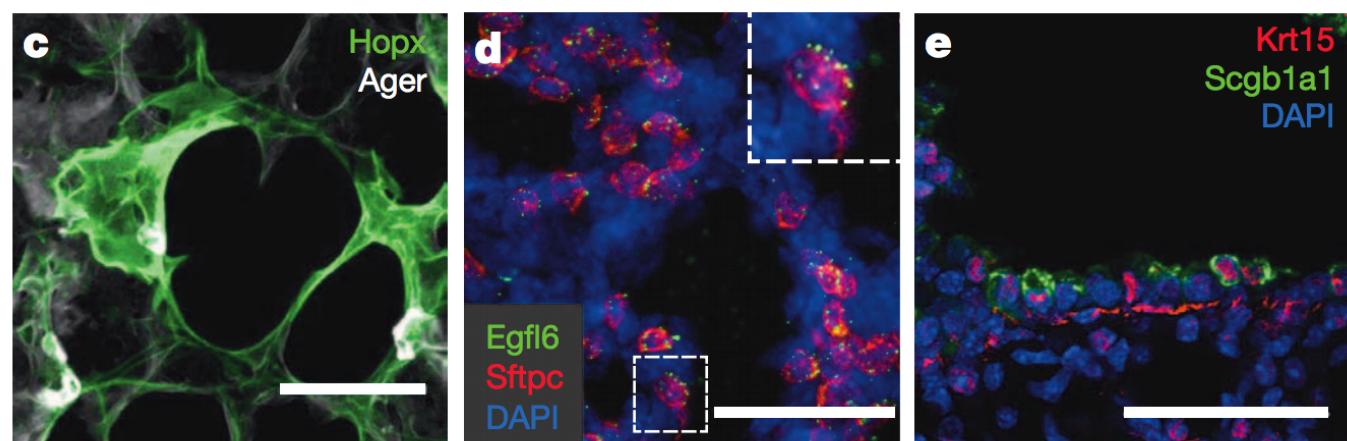
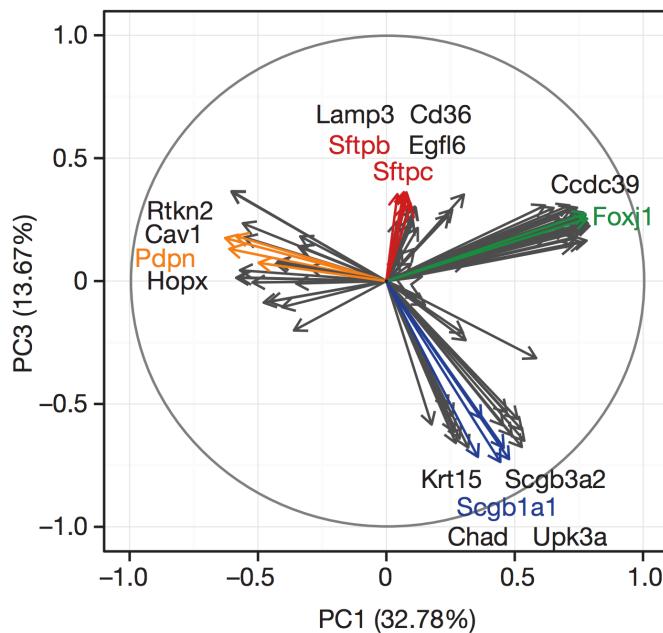
1) Identification of cell-types from heterogeneous samples



Treutlein et al, 2014

Research questions that can be answered with single-cell transcriptomics

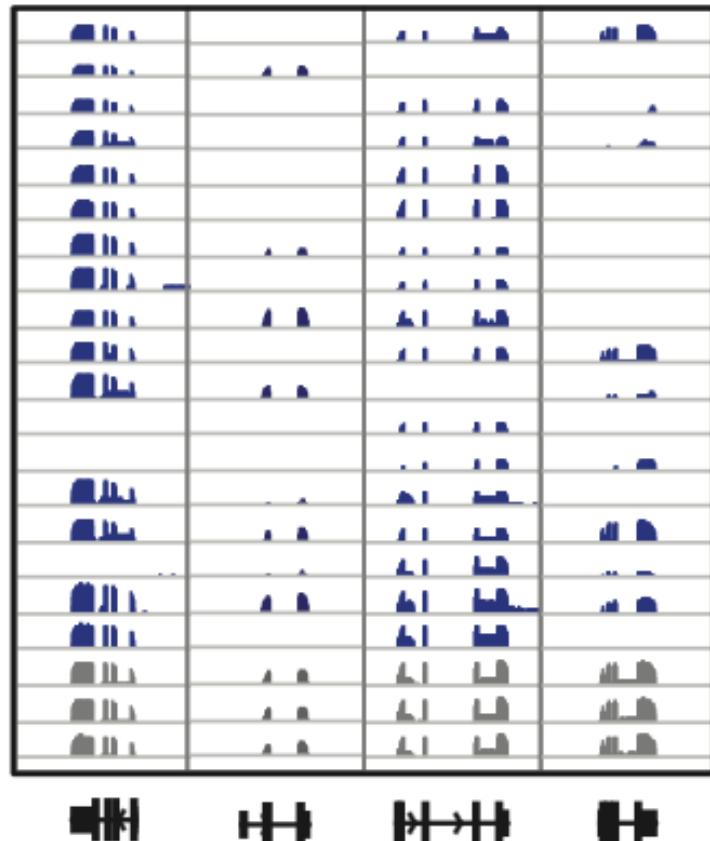
2) Identification of cell-type specific markers (genes, isoforms)



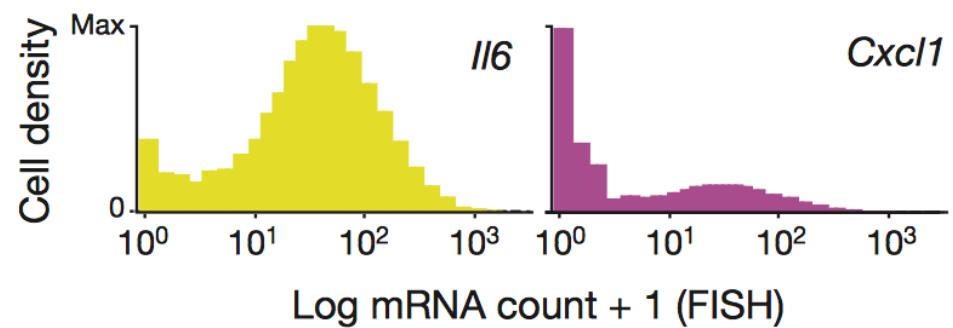
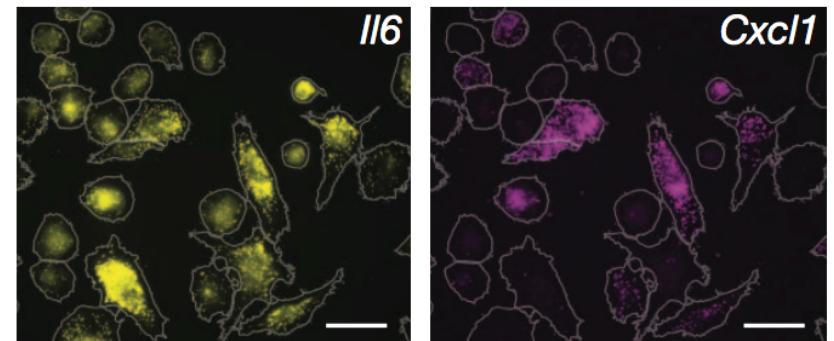
Treutlein et al, 2014

Research questions that can be answered with single-cell transcriptomics

3) Identifying highly varying genes across cells



Cxcl10 *Ifitm1* *Il6* *Cxcl1*

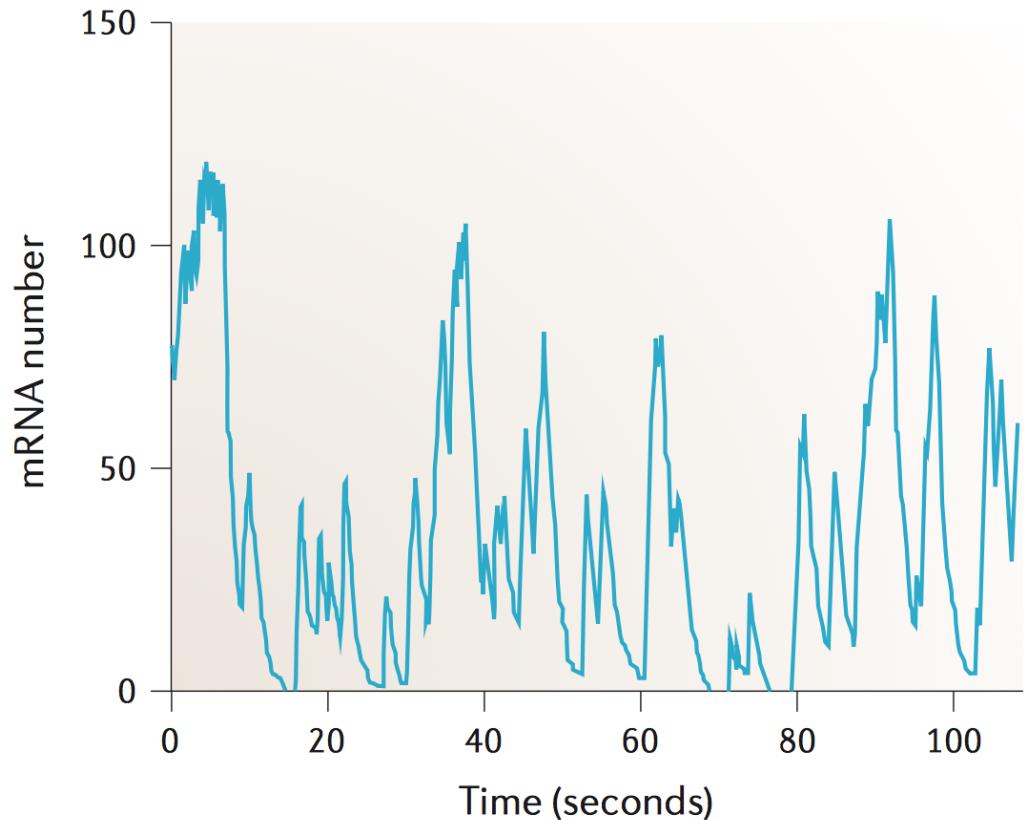


Shalek et al, 2013

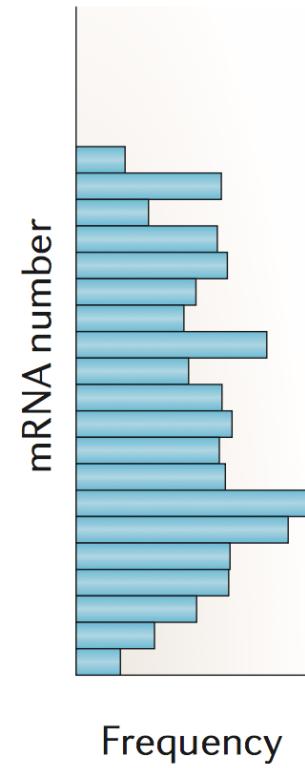
Research questions that can be answered with single-cell transcriptomics

4) Study kinetics of transcription

Pulse microscopy

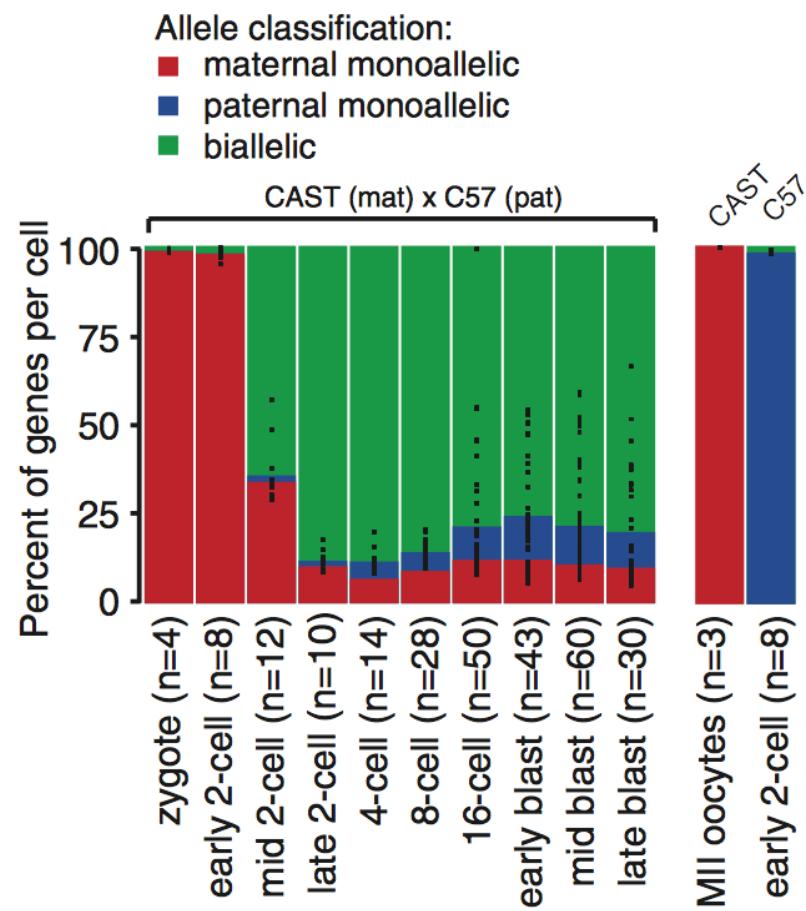


scRNA-seq



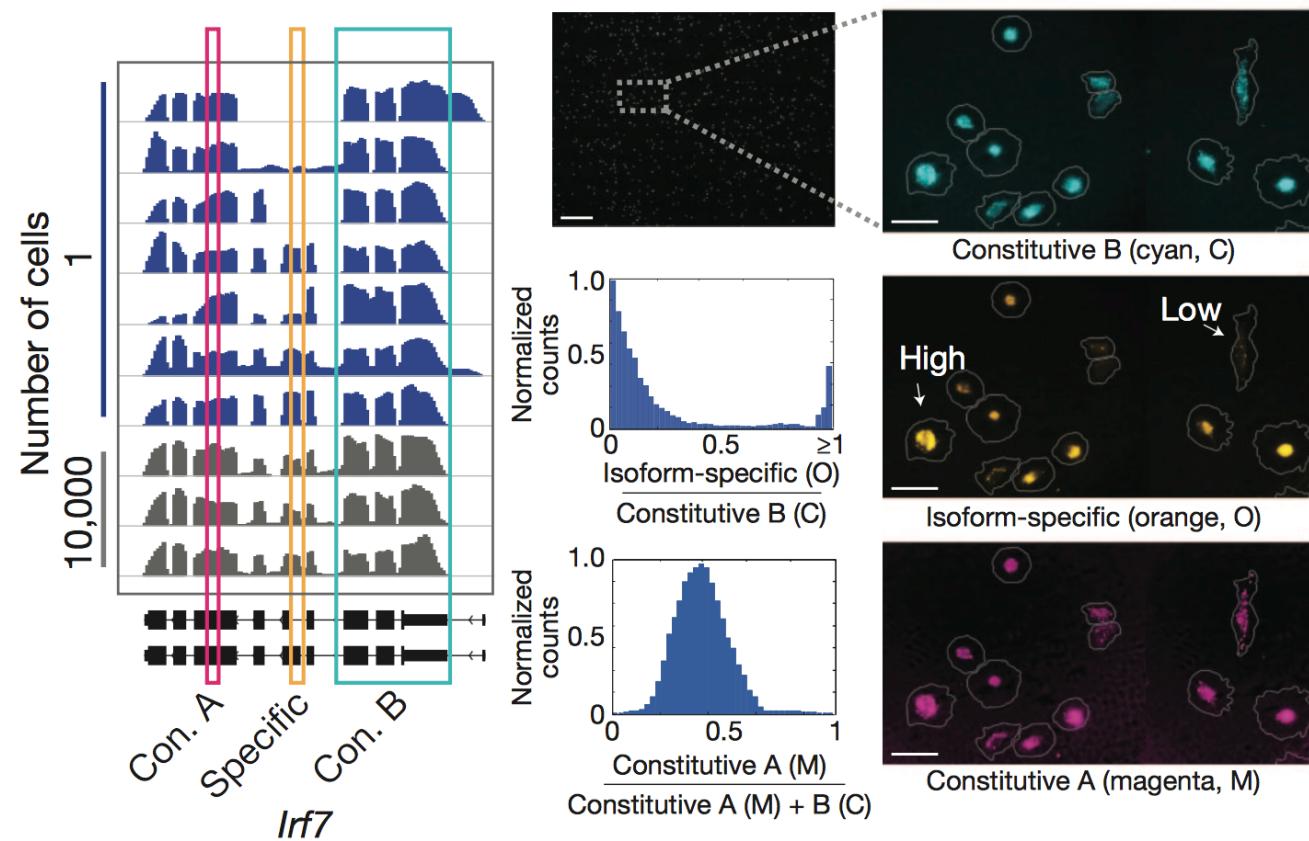
Research questions that can be answered with single-cell transcriptomics

5) Allelic expression heterogeneity



Research questions that can be answered with single-cell transcriptomics

5) Transcript isoform expression heterogeneity



Shalek et al, 2013

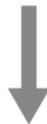
Protocols and noise (pesimistic)

Single-cell transcriptomics protocols overview

SINGLE CELL CAPTURE



SINGLE CELL LYSIS



REVERSE TRANSCRIPTION

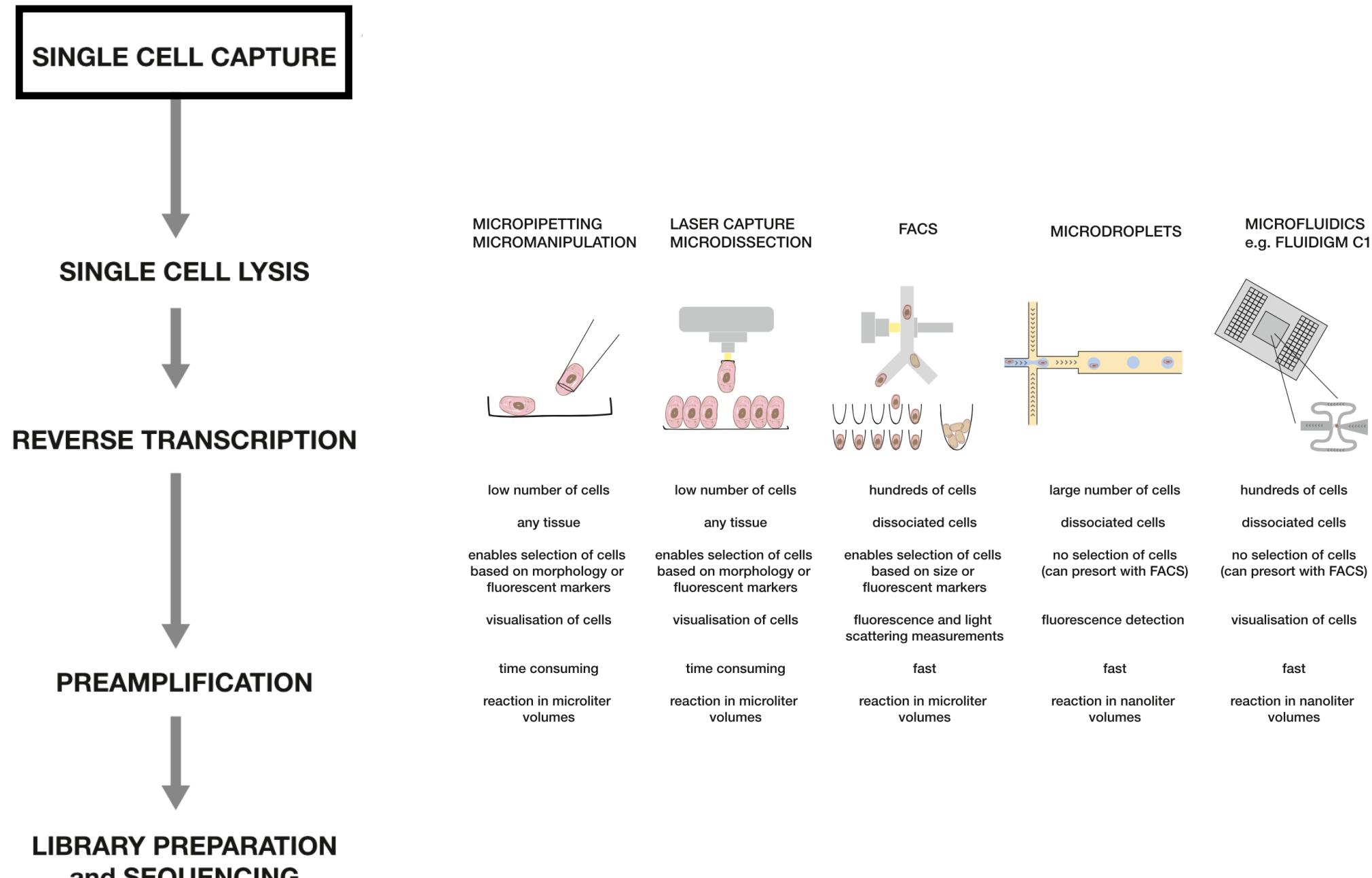


PREAMPLIFICATION



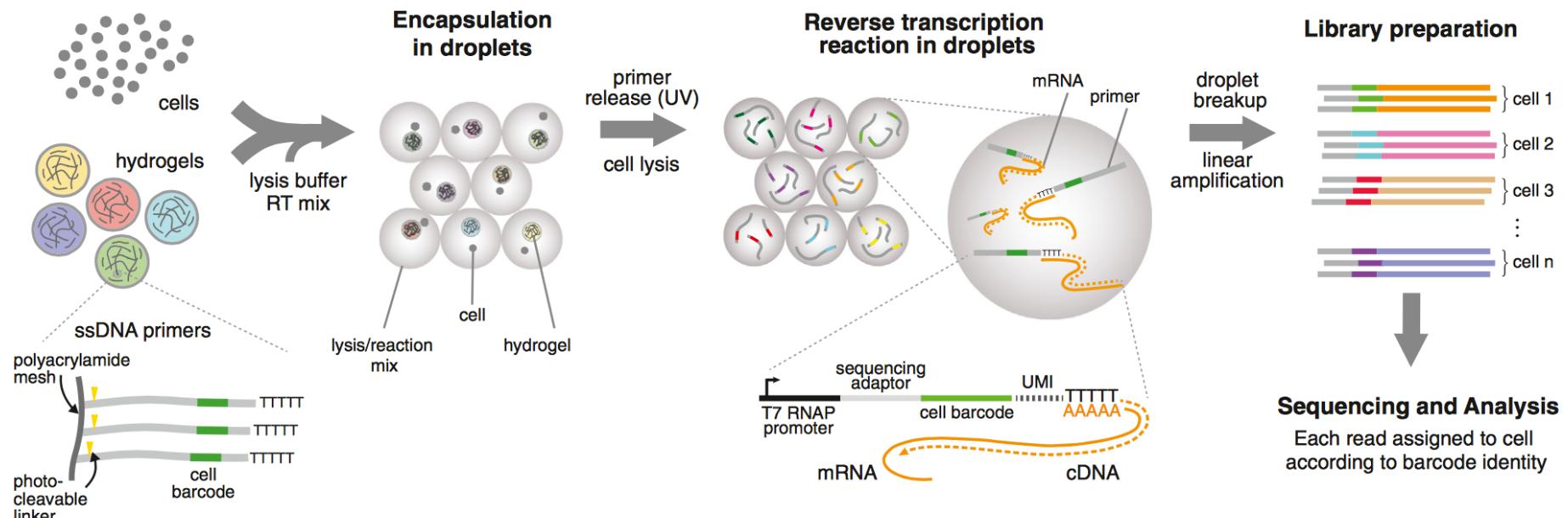
LIBRARY PREPARATION
and SEQUENCING

Single-cell transcriptomics protocols overview



Kolodziejczyk et al, 2013

Single cell protocols

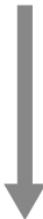


Thousands of cells!

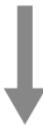
Klein et al, 2015

Single-cell transcriptomics protocols overview

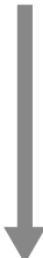
SINGLE CELL CAPTURE



SINGLE CELL LYSIS



REVERSE TRANSCRIPTION

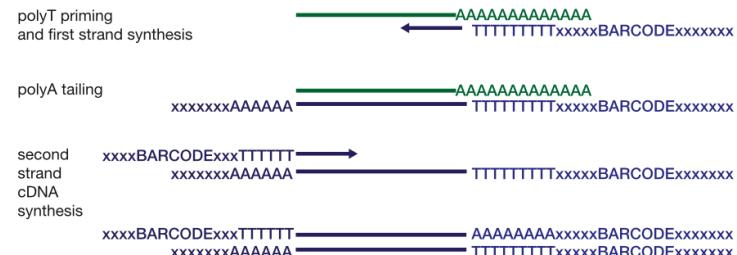


PREAMPLIFICATION



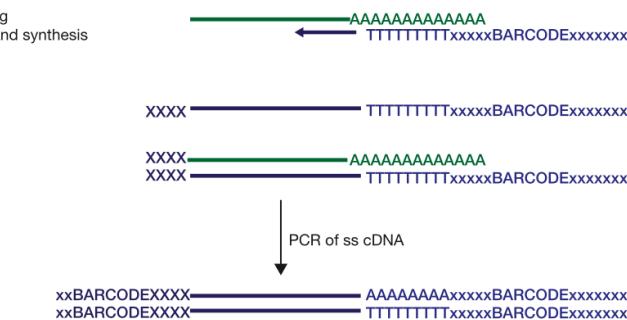
LIBRARY PREPARATION
and SEQUENCING

polyA tailing + second strand synthesis



Tang protocol (Tang et al 2009)
CELseq/MARSseq (Hashimony et al. 2013, Jaitin et al. 2014)
QuartzSeq (Sasagawa et al. 2013)

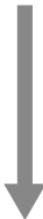
template switching



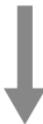
SmartSeq/SmartSeq2 (Ramskold et al. 2012, Deng et al. 2014)
STRT (Islam et al. 2011)

Single-cell transcriptomics protocols overview

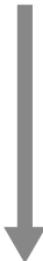
SINGLE CELL CAPTURE



SINGLE CELL LYSIS

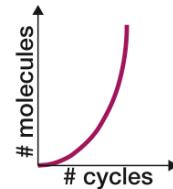


REVERSE TRANSCRIPTION



PREAMPLIFICATION

PCR

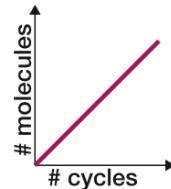


- exponential amplification
- PCR base specific biases

Tang protocol (Tang et al. 2009)
STRT (Islam et al. 2011)
SmartSeq/SmartSeq2 (Ramskold et al. 2012, Deng et al. 2014)

IVT

- linear amplification
- 3' bias due to two rounds of reverse transcription



CELseq/MARSseq (Hashimony et al. 2013, Jaitin et al. 2014)

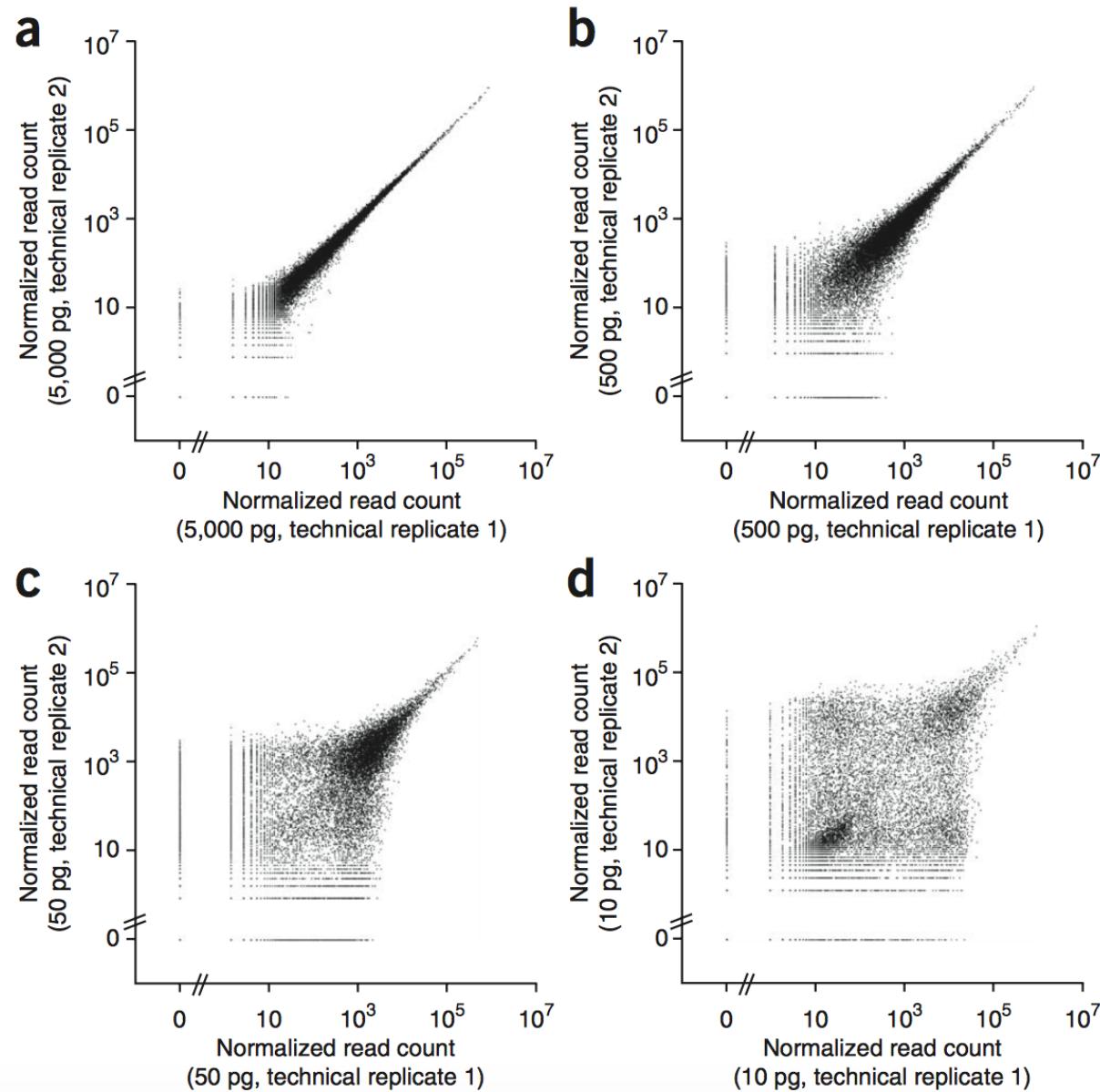
LIBRARY PREPARATION
and SEQUENCING

Analysis

Observed read counts are a combination of different factors

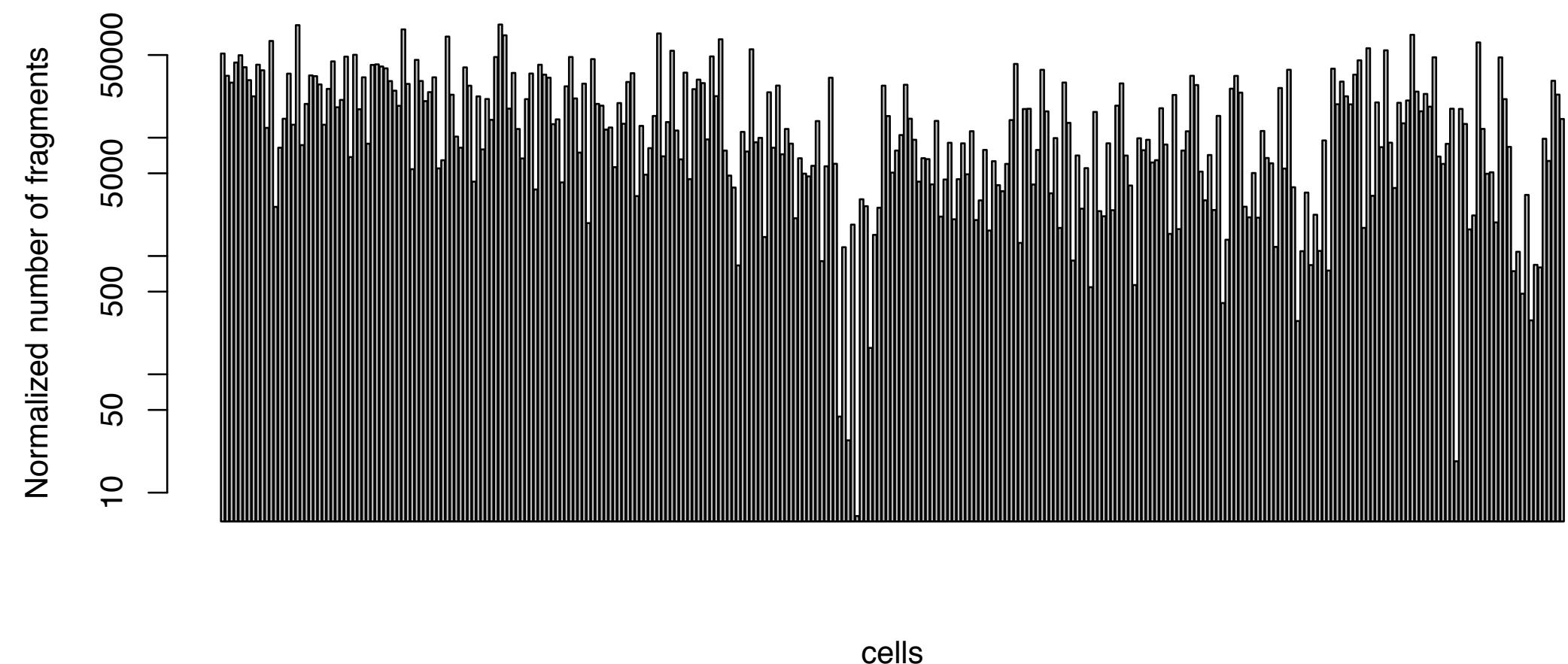
counts = cell state + cell cycle + cell size +
apoptosis + ...+ technical noise

Small amounts of starting material impact on technical noise

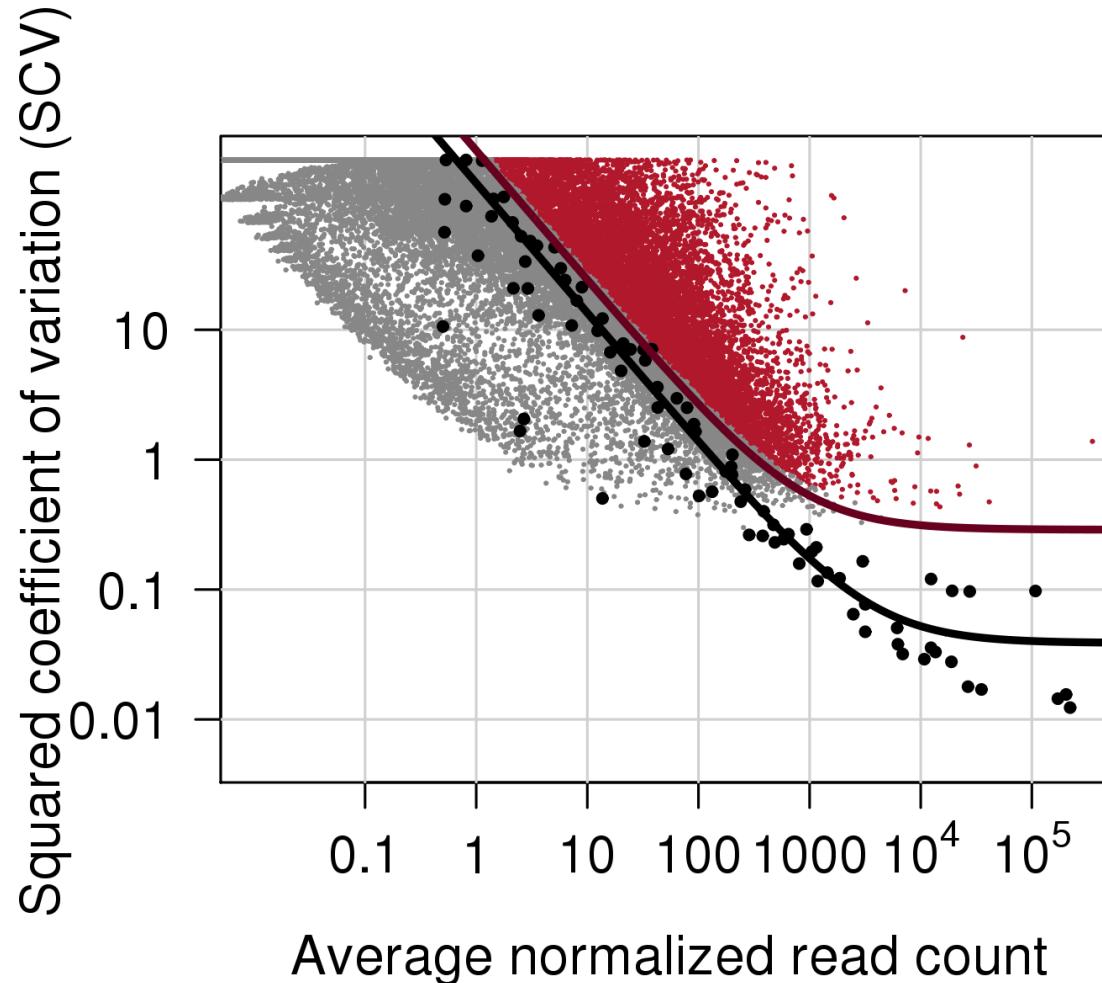


Brennecke, Anders et al, 2013

Detection problems

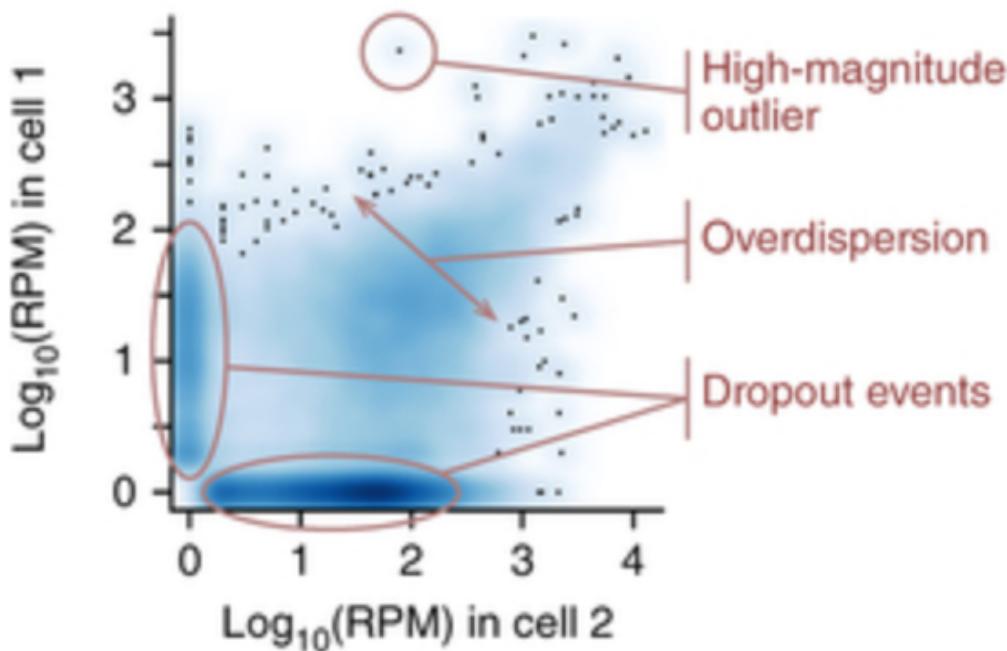


Accounting for technical noise using spike-in sequences



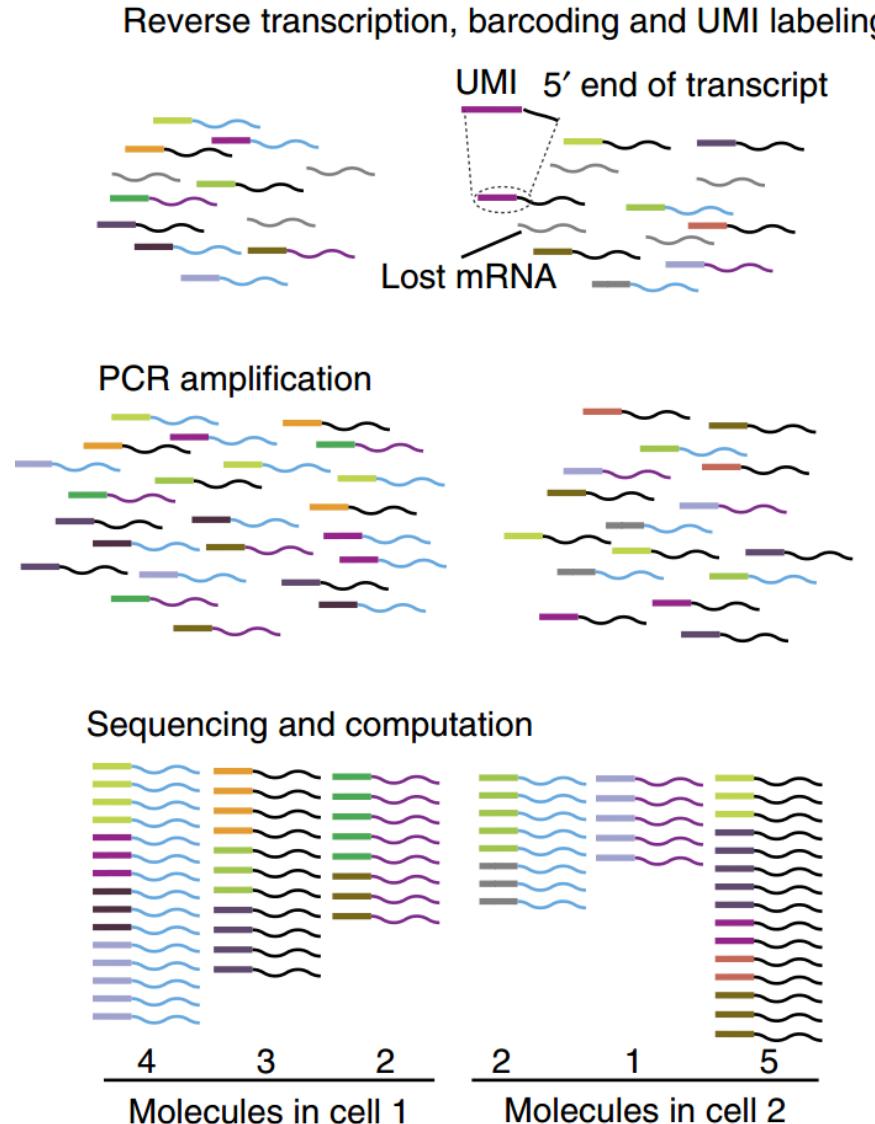
Method by Brennecke, Anders et al, 2013
Data from Brennecke, Reyes et al, 2015

Accounting for technical noise by considering “dropout” events

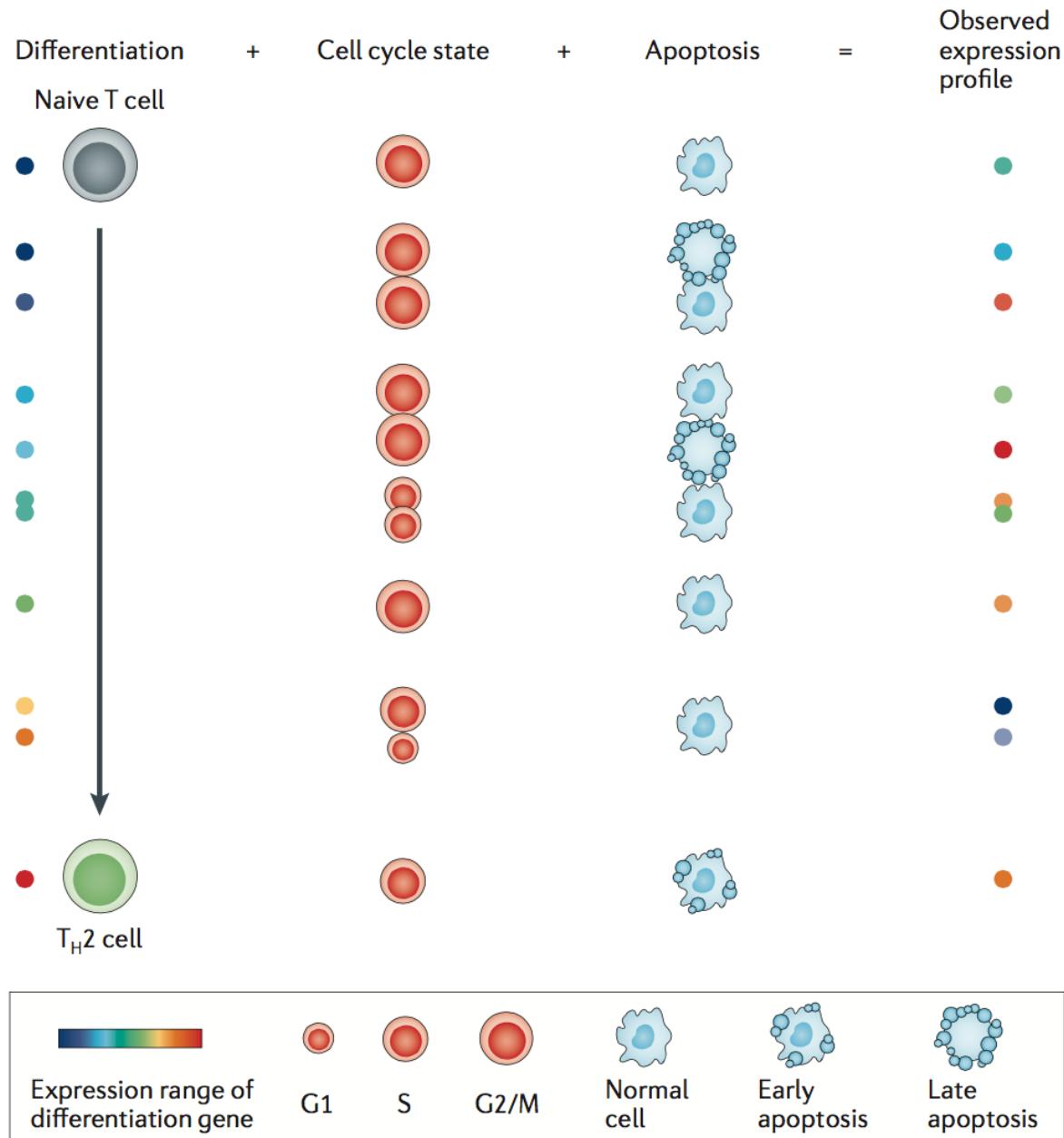


$$\begin{cases} r_1 \approx \text{Poisson}(\lambda_0) & \text{Dropout in } c_1 \\ \begin{cases} r_1 \approx NB(r_2) \\ r_2 \approx NB(r_1) \end{cases} & \text{Amplified} \\ r_2 \approx \text{Poisson}(\lambda_0) & \text{Dropout in } c_2 \end{cases}$$

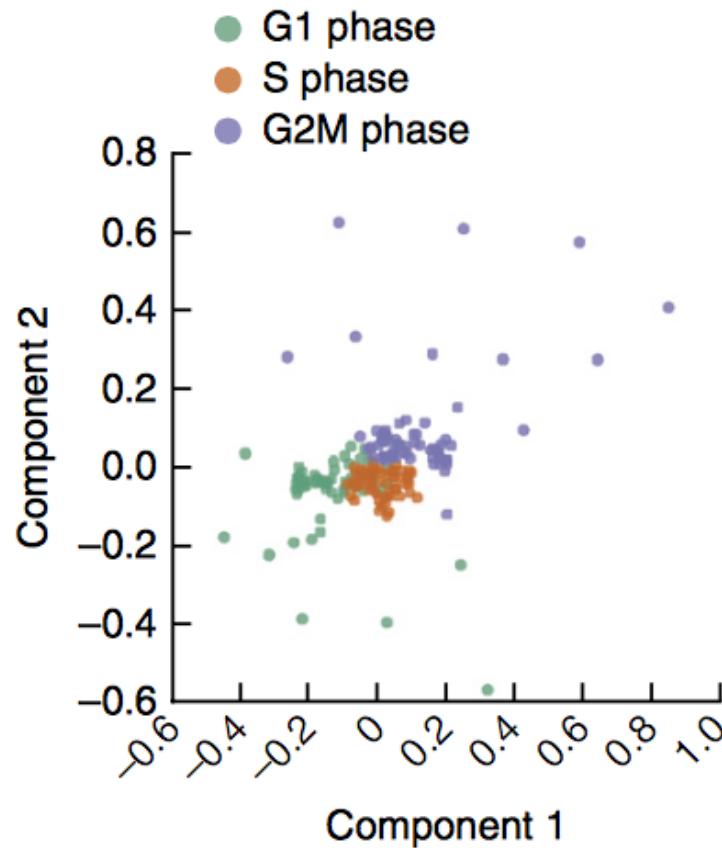
Accounting for technical noise using unique molecular identifiers



Accounting for “biological” confounders



scLVM is useful to regress out variation explained by latent variables



scLVM is useful to regress out variation explained by latent variables

Given H hidden factors,

$$\mathbf{y}_g \sim \mathcal{N} \left(\mu_g \mathbf{1}, \sum_{h=1}^H \sigma_{gh}^2 \Sigma_h + \nu_g^2 \mathbf{I} + \delta_g^2 \mathbf{I} \right)$$

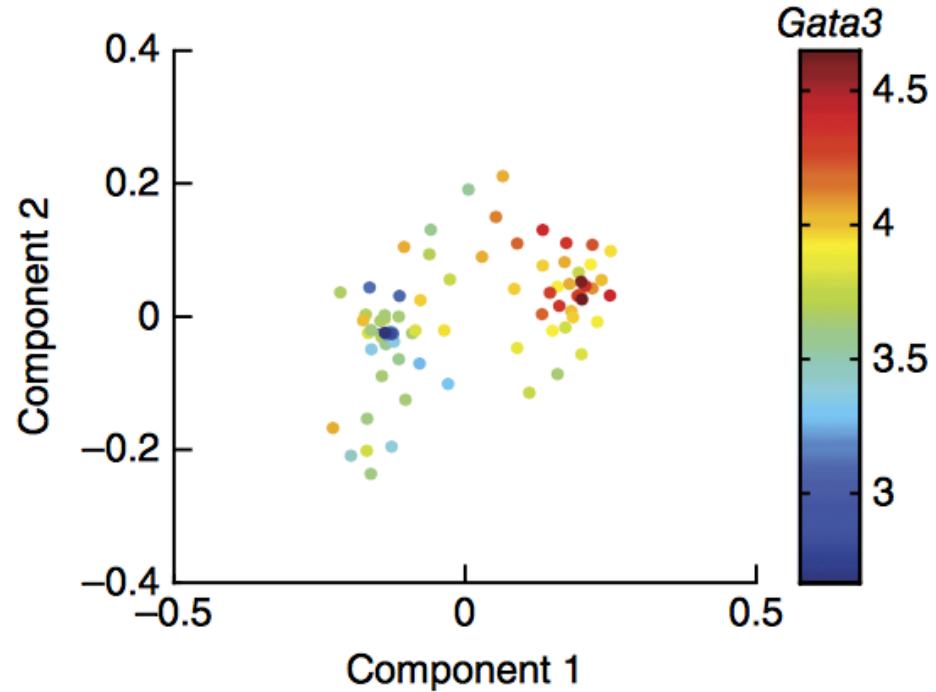
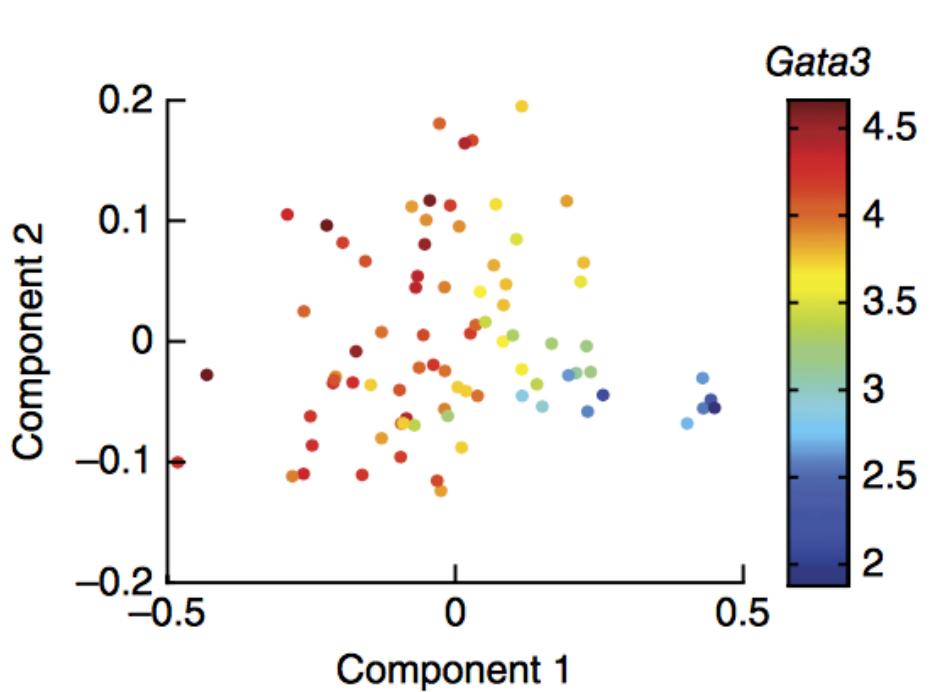
Observed expression for gene g

Variance attributed to hidden factors

Residual “biological” variance

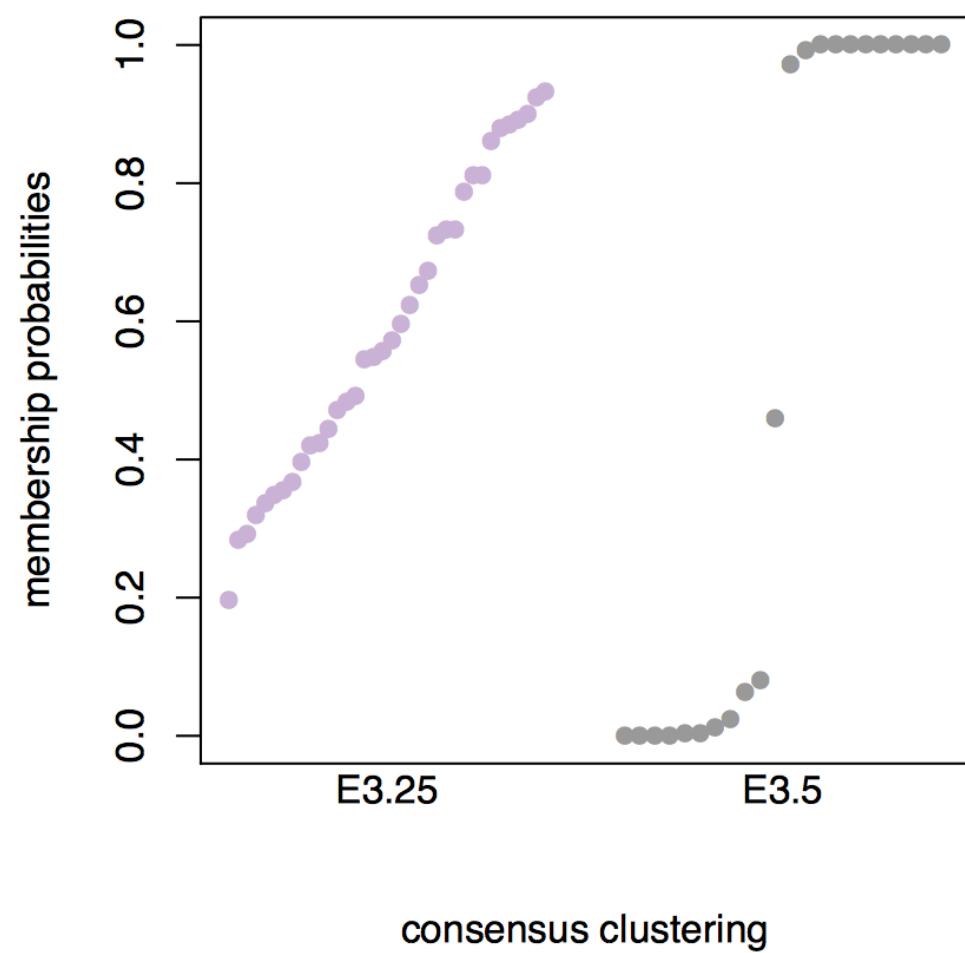
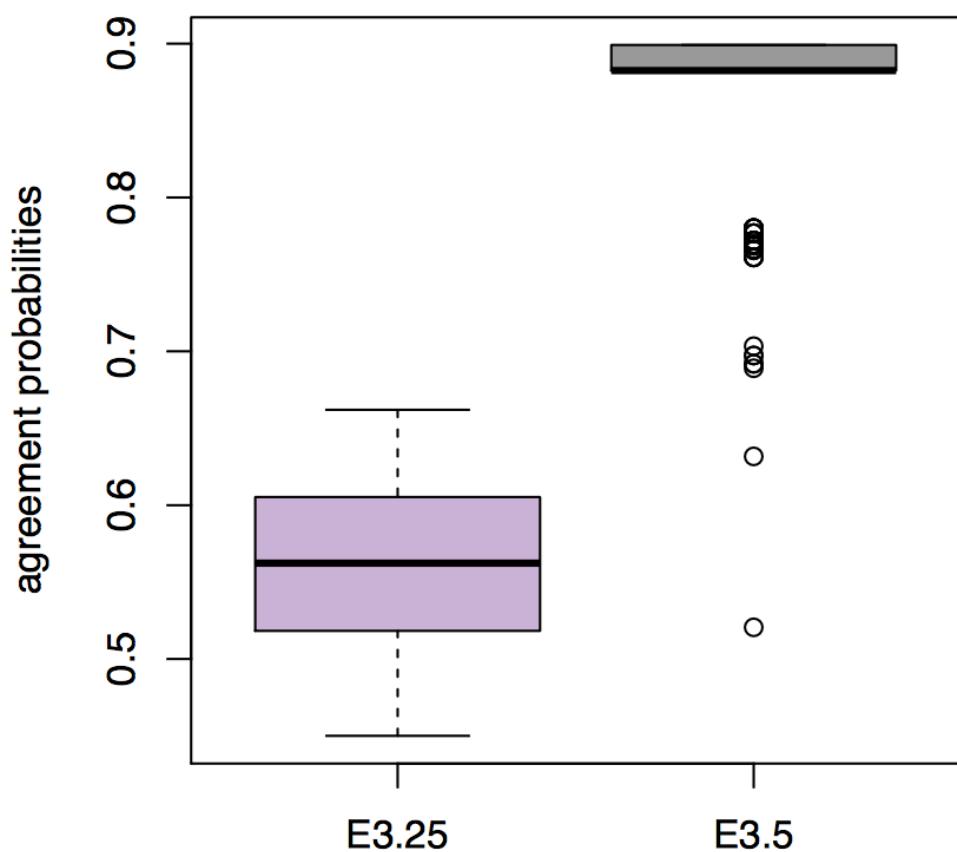
Technical variance

scLVM is useful to regress out variation explained by latent variables



Buettner et al, 2015

Cluster stability analysis



Ohnishi, Huber, et al, 2013

Dimensionality reduction

Multidimensional scaling*

Isomap*

t-SNE*

Diffusion maps

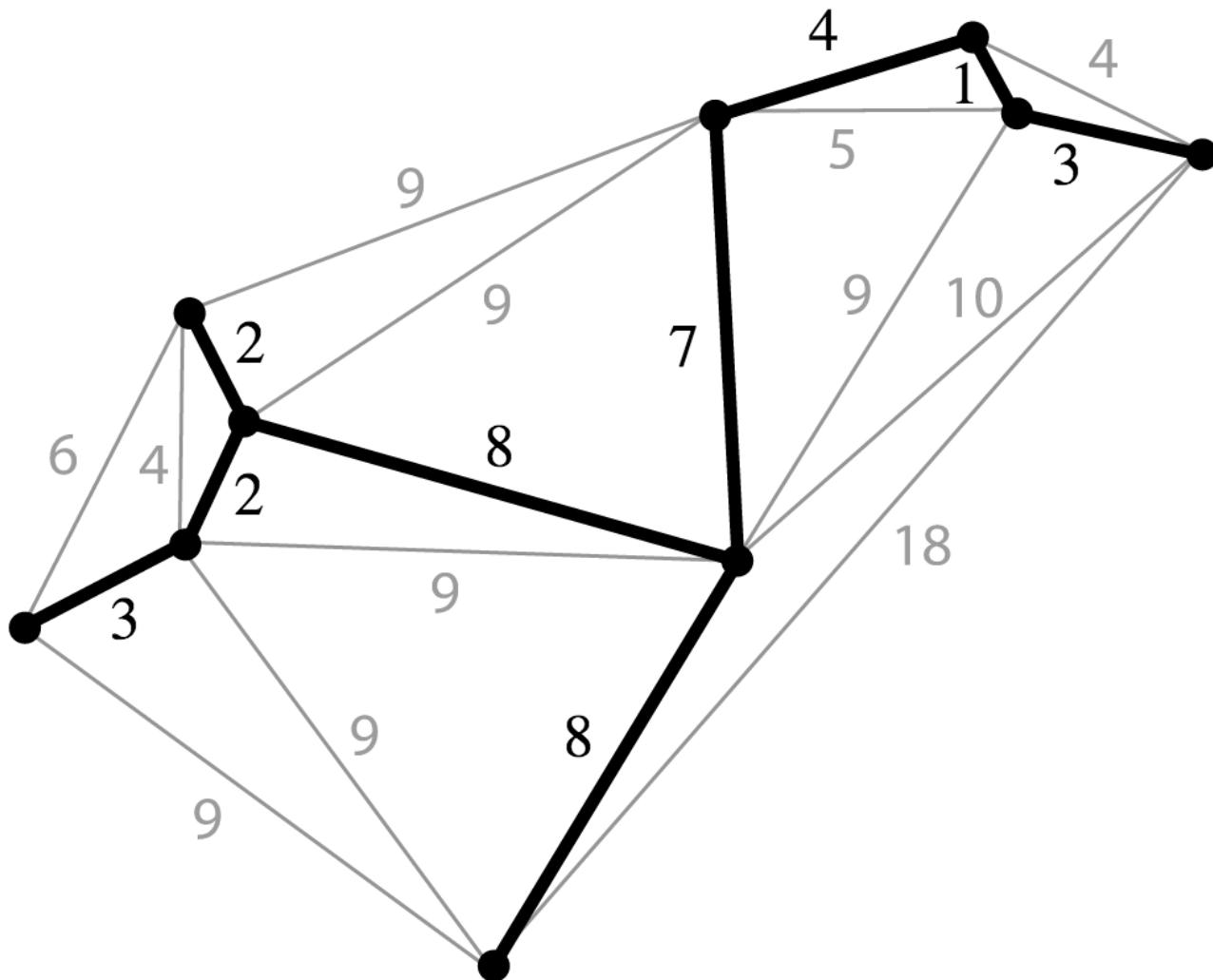
*Bioc Package: `scinCell`

Cell hierarchy reconstruction

Minimum Spanning Tree (MST)*
Maximum Similarity Spanning Tree (SST)*
Iterative Mutual Clustering Graph (IMC)*
Wanderlust

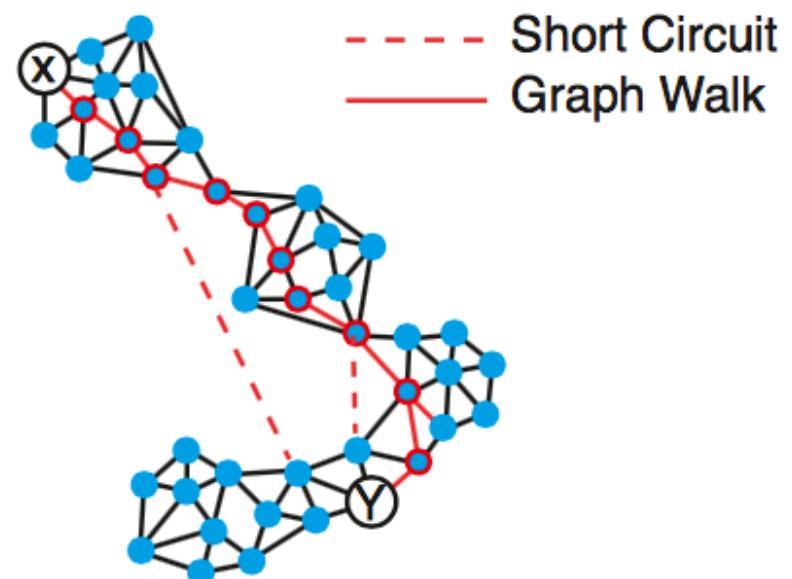
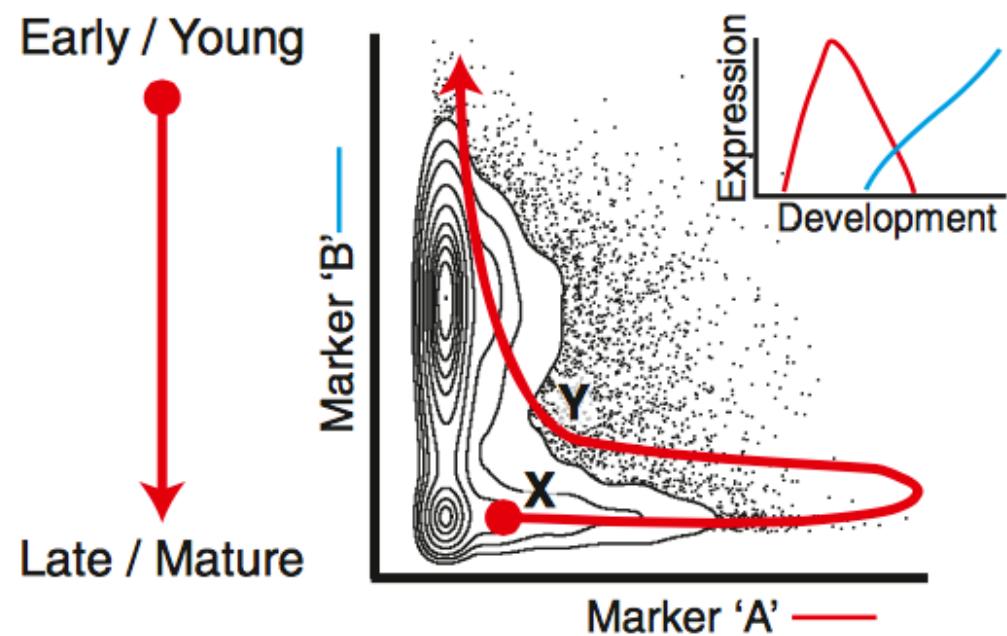
*Bioc Package: `sincell`

Construction of cell state hierarchies



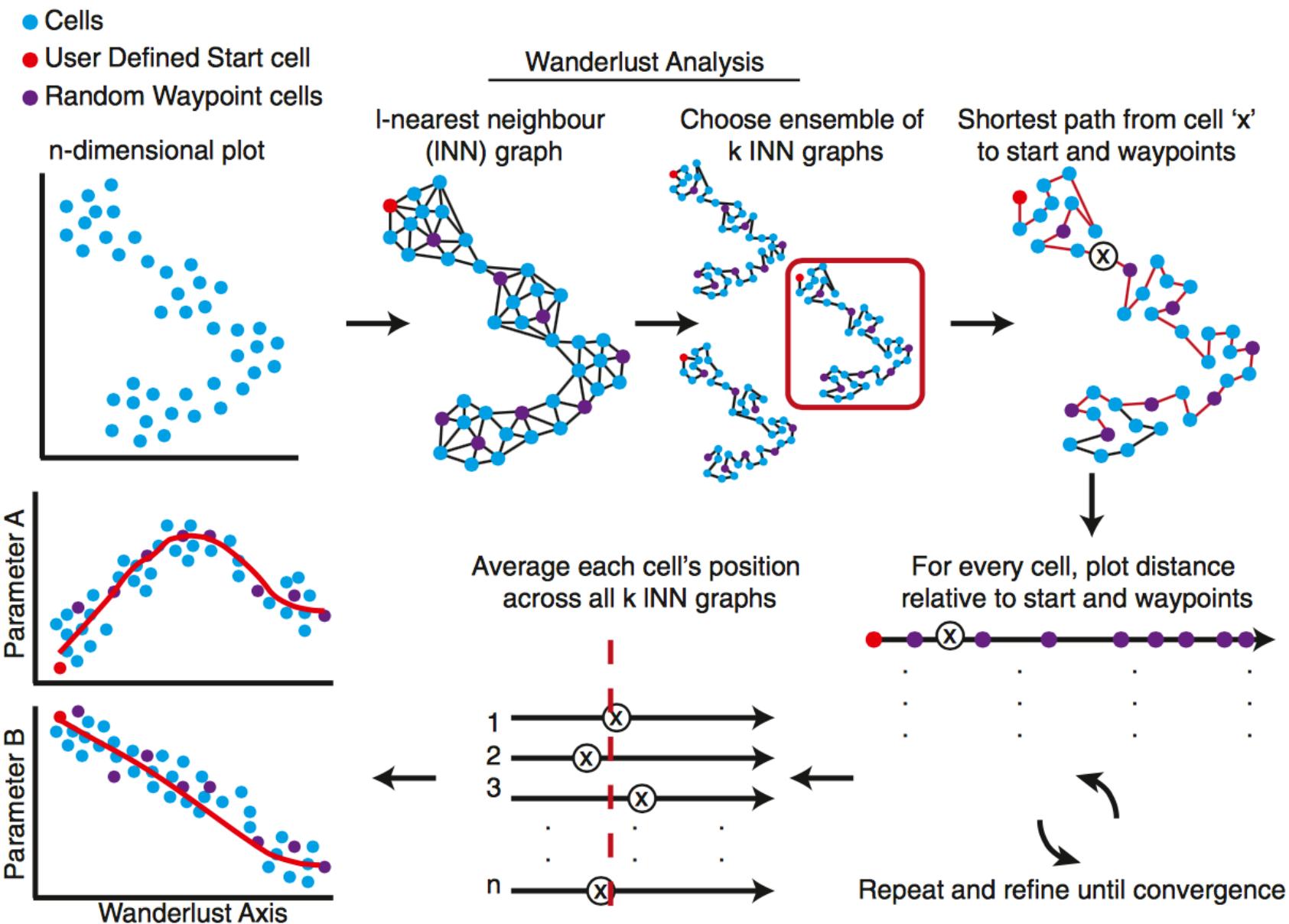
Minimum Spanning Tree Concept (wikipedia)

Construction of cell state hierarchies



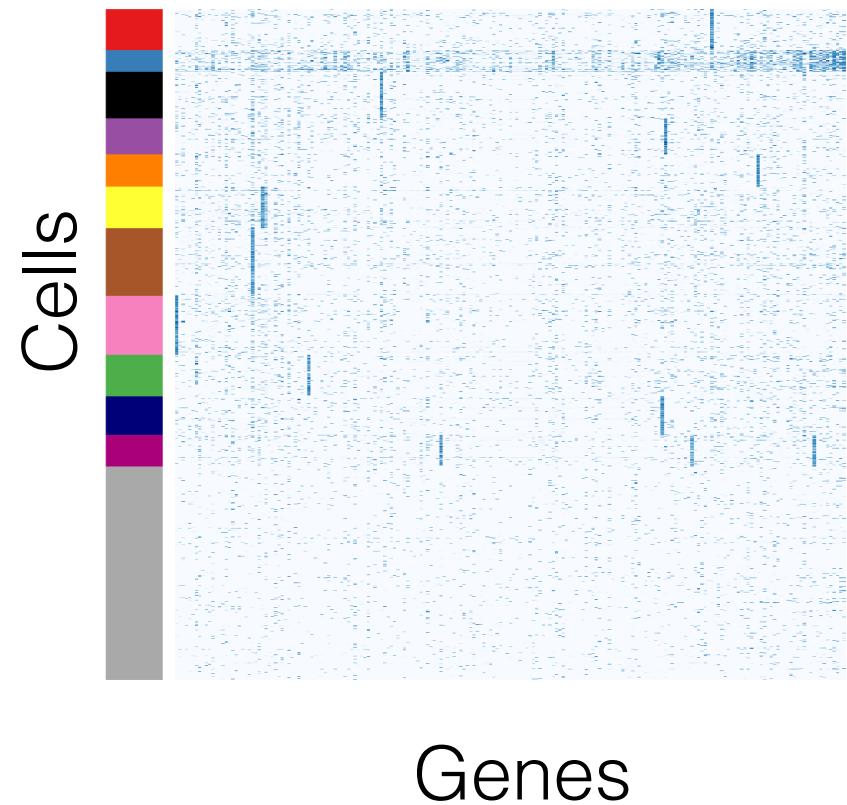
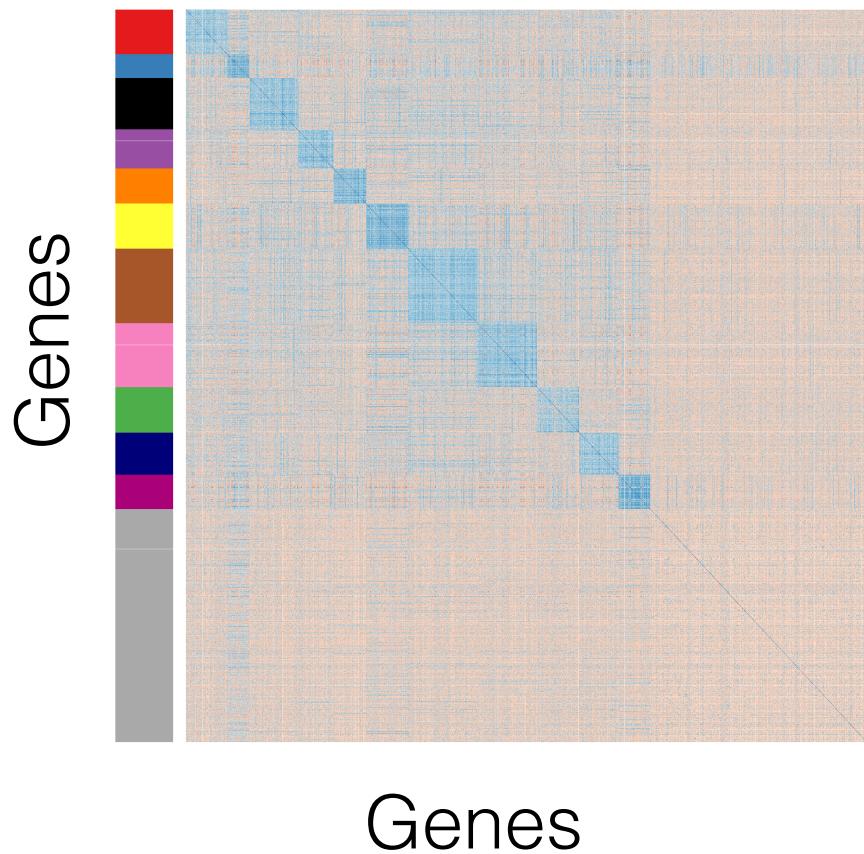
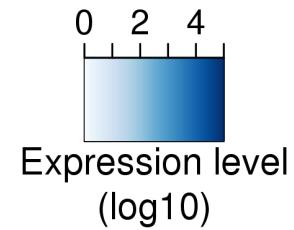
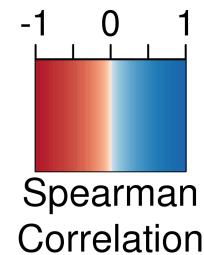
Bendall et al, 2014

Wanderlust algorithm

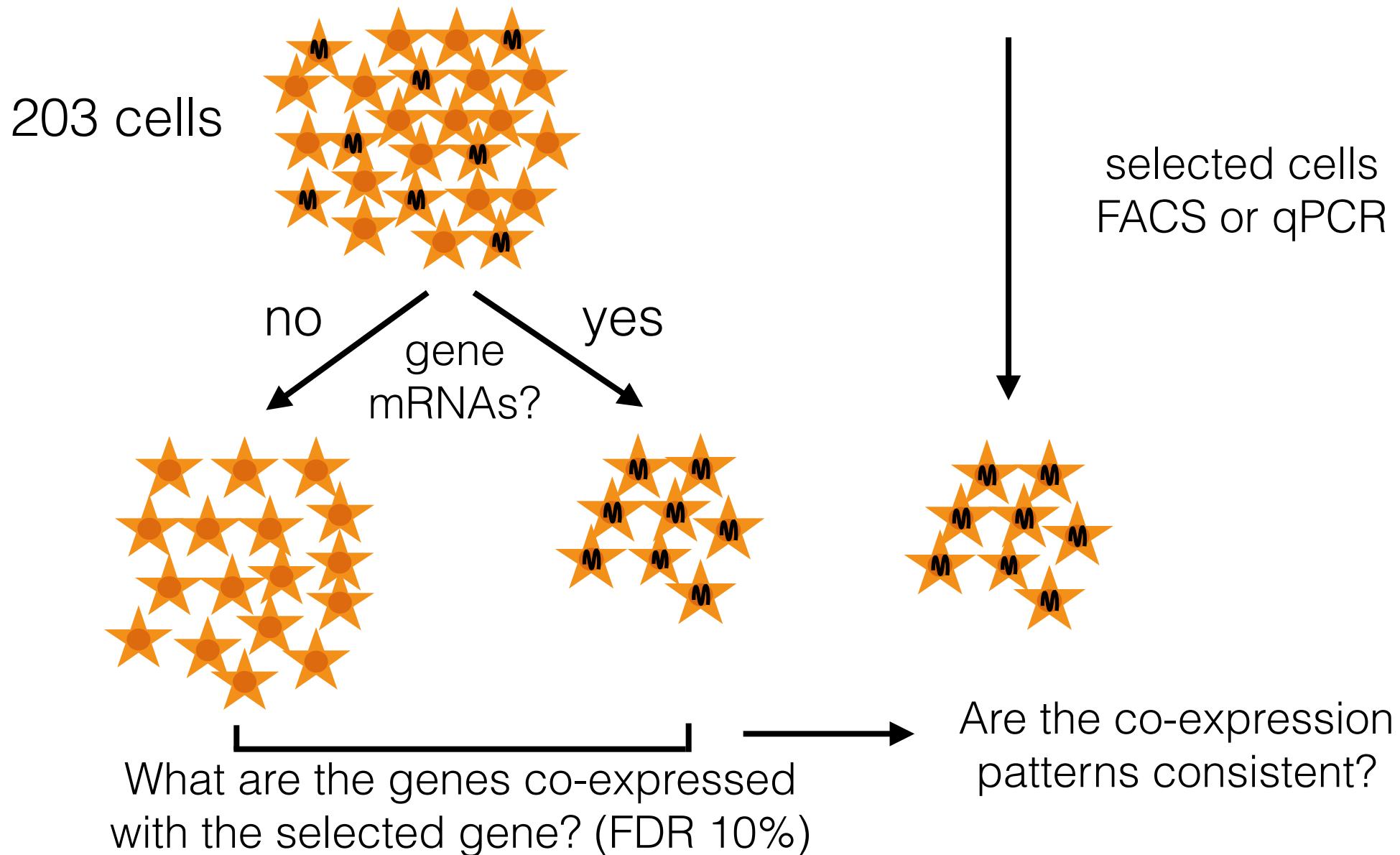


Validate!

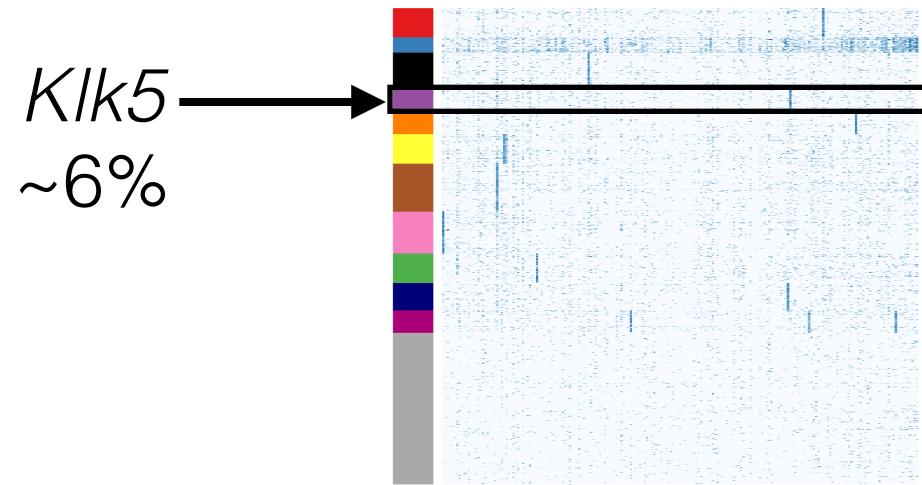
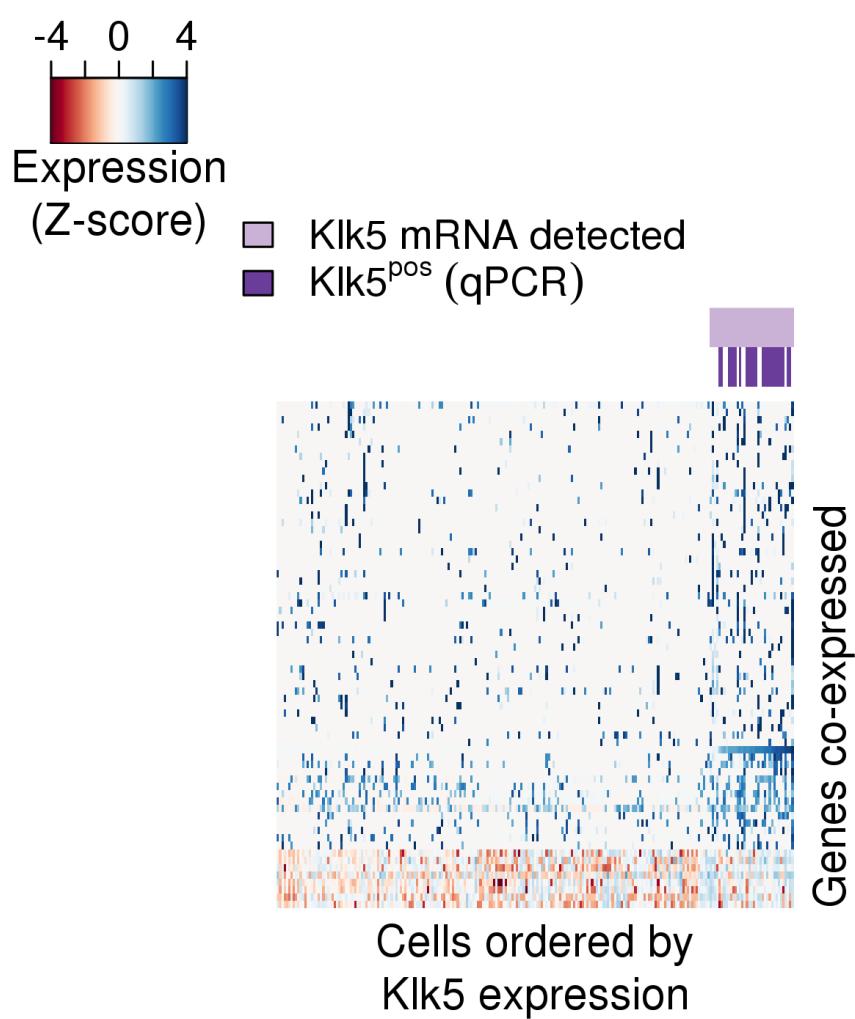
K-medoids clustering suggested non-random gene expression patterns



Co-expression of genes was confirmed using independent analytical and experimental validations



Co-expression of genes was confirmed using independent analytical and experimental validations



Thanks!