# The Bioconductor Project: Current Status

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### **Bioconductor**





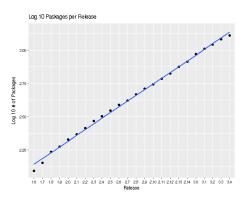


Analysis and comprehension of high-throughput genomic data.

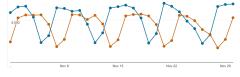
- Started 2002
- 1296 R packages developed by 'us' and user-contributed.

Well-used and respected.

- 43k unique IP downloads / month.
- 17,000 PubMedCentral citations.



- Packages
- Users
- Web & support sites
- Training & meetings
- Release & devel builders
- Funding
- Governance: (annual) Scientific Advisory Board; (monthly) Technical Advisory Board



1.	United States	<b>49,294</b> (30.24%)
2.	China	<b>16,450</b> (10.09%)
3.	United Kingdom	<b>12,456</b> (7.64%)
4.	Germany	10,095 (6.19%)
5.	[◆] Canada	<b>5,458</b> (3.35%)
6.	<ul><li>Japan</li></ul>	<b>5,264</b> (3.23%)
7.	France	<b>4,748</b> (2.91%)
8.	India	<b>4,643</b> (2.85%)
9.	Spain	<b>4,261</b> (2.61%)
10.	Australia	<b>4,180</b> (2.56%)

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https://bioconductor.org https://support.bioconductor.org

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## Recent developments

- New package submission
  - As github issues
  - Public; review participation welcome
- ExperimentHub and AnnotationHub
  - Similar to 'Annotation' and 'Experiment' data repositories
  - ExperimentHub often used as the 'data store' for experiment data packages, e.g., alpineData.
- Large data representation: *HDF5Array*
- (Sneak peak) Organism.dplyr

## HDF5Array

```
> library(HDF5Array) # available in release & devel
> library(h5vcData)
> h5file <- system.file("extdata", "example.tally.hfs5",
                        package="h5vcData")
> cov0 <- HDF5Array(h5file, "/ExampleStudy/16/Coverages")</pre>
> pcov <- t(drop(cov0[ , 1, ])) # coverage on plus strand</pre>
> mcov <- t(drop(cov0[ , 2, ])) # coverage on minus strand
> library(SummarizedExperiment)
> SummarizedExperiment(list(pcov=pcov, mcov=mcov))
class: SummarizedExperiment
dim: 90354753 6
metadata(0):
assays(2): pcov mcov
```

## Sneak peak: Organism.dplyr

```
> library(Organism.dplyr) # not yet publicly available
> src = src_ucsc("Homo sapiens") # any org.* + TxDb.*
using org. Hs.eg.db, TxDb. Hsapiens. UCSC.hg38.knownGene
> src
src: sqlite 3.8.6 [/home/mtmorgan/organism_dplyr.sqlite]
tbls: id, id_accession, id_go, id_go_all, id_omim_pm,
  id_protein, id_transcript, ranges_cds, ranges_exon,
 ranges_gene, ranges_tx
> tbl(src, 'id') %>% filter(symbol == 'BRCA1') %>%
    select(ensembl, symbol, genename)
> exons(src, filter=list(symobl='BRCA1'))
                                               # GRanges
> exons_tbl(src, filter=list(symbol='BRCA1'))
                                               # tibble
```

## Programming best practices

- Reuse & interoperability
  - GenomicRanges and SummarizedExperiment
  - rtracklayer::import() for BED, WIG, GTF, GFF, etc.
- Documentation: classic or roxygen2
- Testing: RUnit or testthat
- Correct, robust, efficient (vectorized) code; BiocParallel
- Classic, tidy, and semantically rich data

### Correct, robust, efficient...

```
f = function(n) {
                                f2 = function(n)
                                   vapply(1:n, c, integer(1))
  x = integer(0)
  for (i in 1:n)
      x = c(x, i)
                                f3 = function(n)
                                   seq_len(n)
  X
microbenchmark(f(1000),
                                 ## correct
  f(10000), f(100000))
                                 identical(f(100), f3(100))
f1 = function(n)  {
                                 ## robust!
                                f(0): f3(0)
  x = integer(n)
  for (i in 1:n)
    x[i] = i
                                 ## efficient
                                 system.time(f3(1e9)
  X
```

# Classic, tidy, rich: RNA-seq count data

#### Classic

• Sample x (phenotype + expression) Feature data.frame

### Tidy

 'Melt' expression values to two long columns, replicated phenotype columns. End result: long data frame.

Rich, e.g., SummarizedExperiment

 Phenotype and expression data manipulated in a coordinated fashion but stored separately.

# Classic, tidy, rich: RNA-seq count data

```
df0 <- as.data.frame(list(mean=colMeans(classic[, -(1:22)])))
df1 <- tidy %>% group_by(probeset) %>%
    summarize(mean=mean(exprs))
df2 <- as.data.frame(list(mean=rowMeans(assay(rich))))
ggplot(df1, aes(mean)) + geom_density()</pre>
```

# Classic, tidy, rich: RNA-seq count data

### Vocabulary

- Classic: extensive
- Tidy: restricted endomorphisms
- Rich: extensive, meaningful

### Constraints (e.g., probes & samples)

- Tidy: implicit
- Classic, Rich: explicit

### Flexibility

- Classic, tidy: general-purpose
- Rich: specialized

### Programming contract

- Classic, tidy: limited
- Rich: strict

### Lessons learned / best practices

- Considerable value in semantically rich structures
- Current implementations trade-off user and developer convenience
- Endomorphism, simple vocabulary, consistent paradigm aid use

## Future challenges

- Git
- Cloud. Possible visions:
  - As now, but 'in the cloud'
  - Integrated with 'third party' compute efforts, e.g., NCI, NIH in the United States

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