monaLisa MOtif aNAlysis with Lisa

European Bioconductor Meeting 2019

Dania Machlab Lukas Burger Michael Stadler

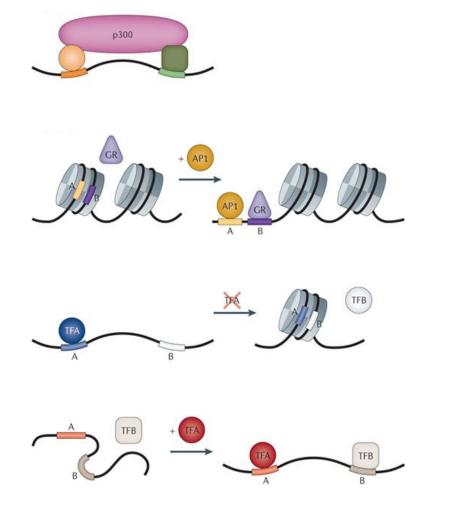






Friedrich Miescher Institute for Biomedical Research

Background and Motivation



Co-binding

Chromatin remodeling

Blocking repositioning

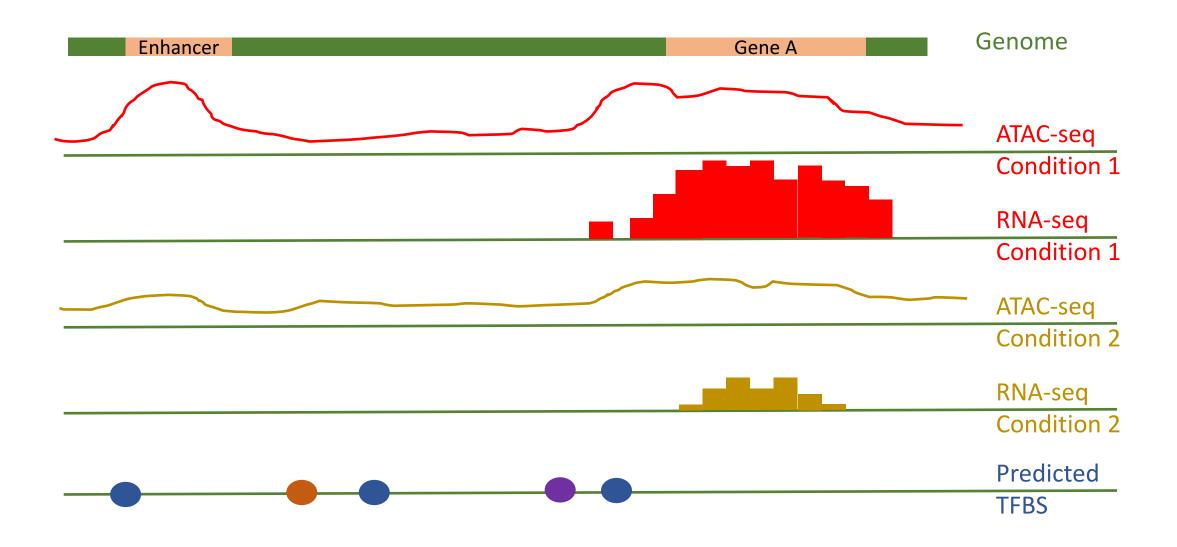
Architectural role



Use monaLisa to:

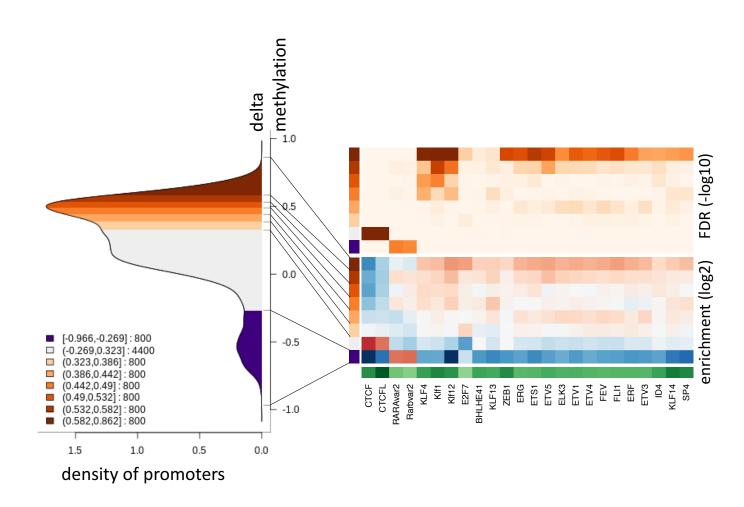
- Identify Enriched motifs
- Select motifs explaining observed changes

Background and Motivation

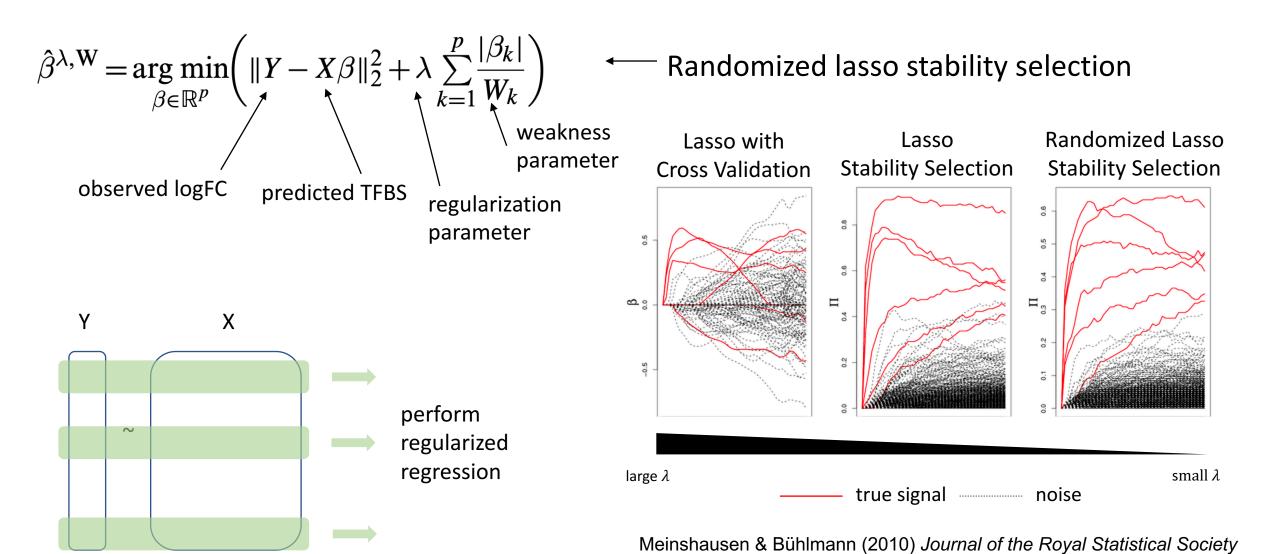


Identify Enriched Motifs

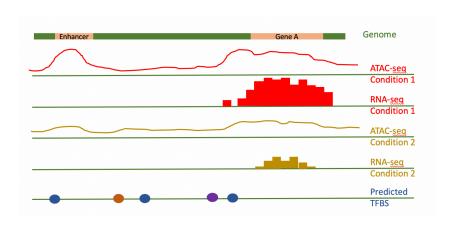
```
# bin regions by delta methylation
bins <- bin(x = lmrsel$deltaMeth, binmode = "equalN",</pre>
            nElement = 800, minAbsX = 0.3
# dump motifs into file for use by Homer
motiffile <- tempfile(fileext = ".motif")</pre>
dumpJaspar(motiffile, pkg = "JASPAR2018")
# find Homer (findMotifsGenome.pl)
homerfile <- findHomer(dirs = "/work/gbioinfo/Appz</pre>
                        /Homer/Homer-4.10.4/bin/")
# run analysis
outdir <- tempfile(fileext = ".output")</pre>
se <- runHomer(gr = lmrsel, b = bins,</pre>
                genomedir = "/work/gbioinfo/DB/genomes/mm9",
                outdir = outdir, motifFile = motiffile,
                homerfile = homerfile,
                regionsize = "given", Ncpu = 20L)
```

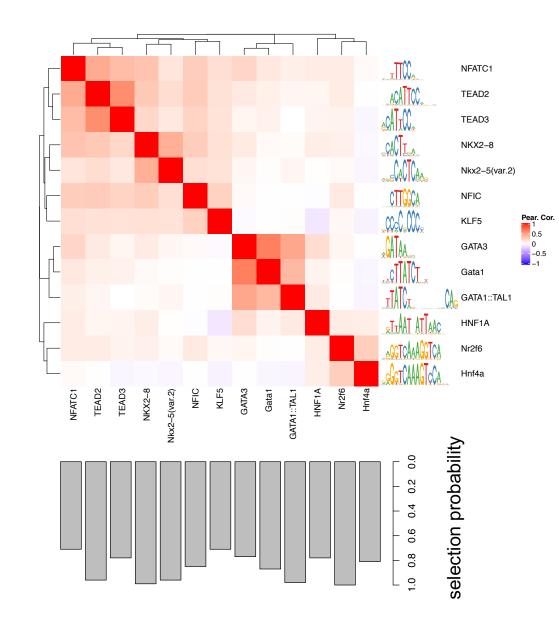


Select Motifs using Stability Selection



Select Motifs Explaining Observed Changes in Accessibility





Summary and Outlook

- We can identify TFs enriched in regions of interest that display certain log-fold changes
- We can select TFs that are likely to explain the observed log-fold changes using stability selection
- We can be use any fold-change defined on regions of interest (ATAC-seq, methylation, expression, ChIP-seq ...) to select motifs explaining the observed logFC
- We want to look at motif enrichment without using existing databases (unbiased view)
- Enriched k-mers, grouping them, aligning them to predict the motif
- Submit to Bioconductor
- https://github.com/fmicompbio/monaLisa