

Single cell RNA - seq analysis for muscle samples

1. Results:

a. t-SNE :

In my analysis, the t-SNE plot (fig. 1)) closely aligns with the configuration observed in the reference paper. Specifically, the expected clustering of Endothelial Cells is evident, underscoring the reliability of the t-SNE analysis. Notably, T and B cells, alongside NK cells, exhibit close spatial affinity, reflecting their shared immune cell features. Furthermore, distinct separation is apparent among Satellite, LUM+FAP, and FBN1+FAP cells, delineating clear distinctions from both Endothelial and Immune cell groups. Importantly, the t-SNE plot unveils a distinct connection between Smooth Muscle Cells and Pericytes, akin to observations in the reference t-SNE plot. This congruence emphasizes the robustness of the t-SNE analysis in uncovering meaningful cellular relationships within the single-cell RNA sequencing dataset. The visualization details are derived from the t-SNE plot generated using `sc.pl.tsne` in Scanpy, with cell types indicated by color and a legend placed on data points for clarity.

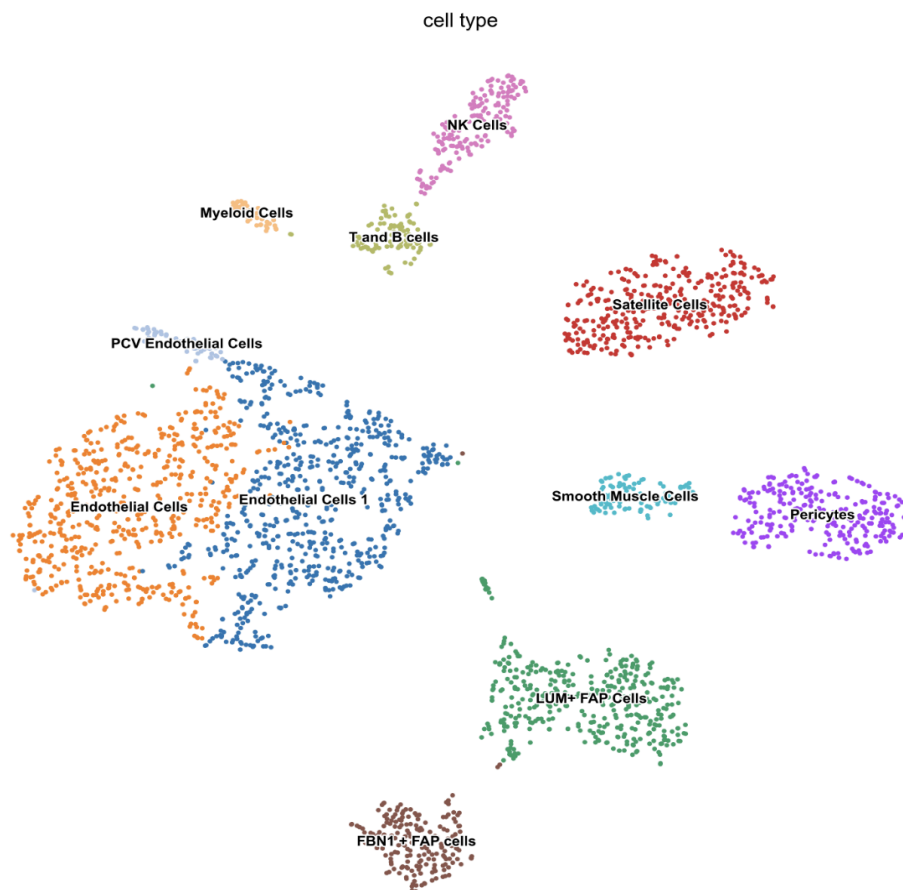


Figure. 1: t-SNE plot

b. Heatmap:

In my analysis using `sc.pl.heatmap` in Scanpy, the heatmap (fig. 2) provides a visual representation of marker gene expression patterns (`marker_genes_dict`) across diverse cell types (`groupby='cell_type'`). The color map is specifically tailored to capture varying expression levels, employing a 'Reds' palette (`cmap='Reds'`). Both row and column hierarchical clustering are incorporated into the heatmap, as indicated by the presence of dendrograms (`dendrogram=True`). Adjusting the figure size to 12 by 8 inches (`figsize=(12, 8)`) enhances the clarity of the heatmap, offering a comprehensive depiction of gene expression across different cell types. It's important to note that the Y-axis dendrogram orientation in Python reflects the convention of placing variables along columns, differing from the X-axis dendrogram used in R. Despite this distinction, the heatmap closely aligns with the expression patterns observed in the reference paper.

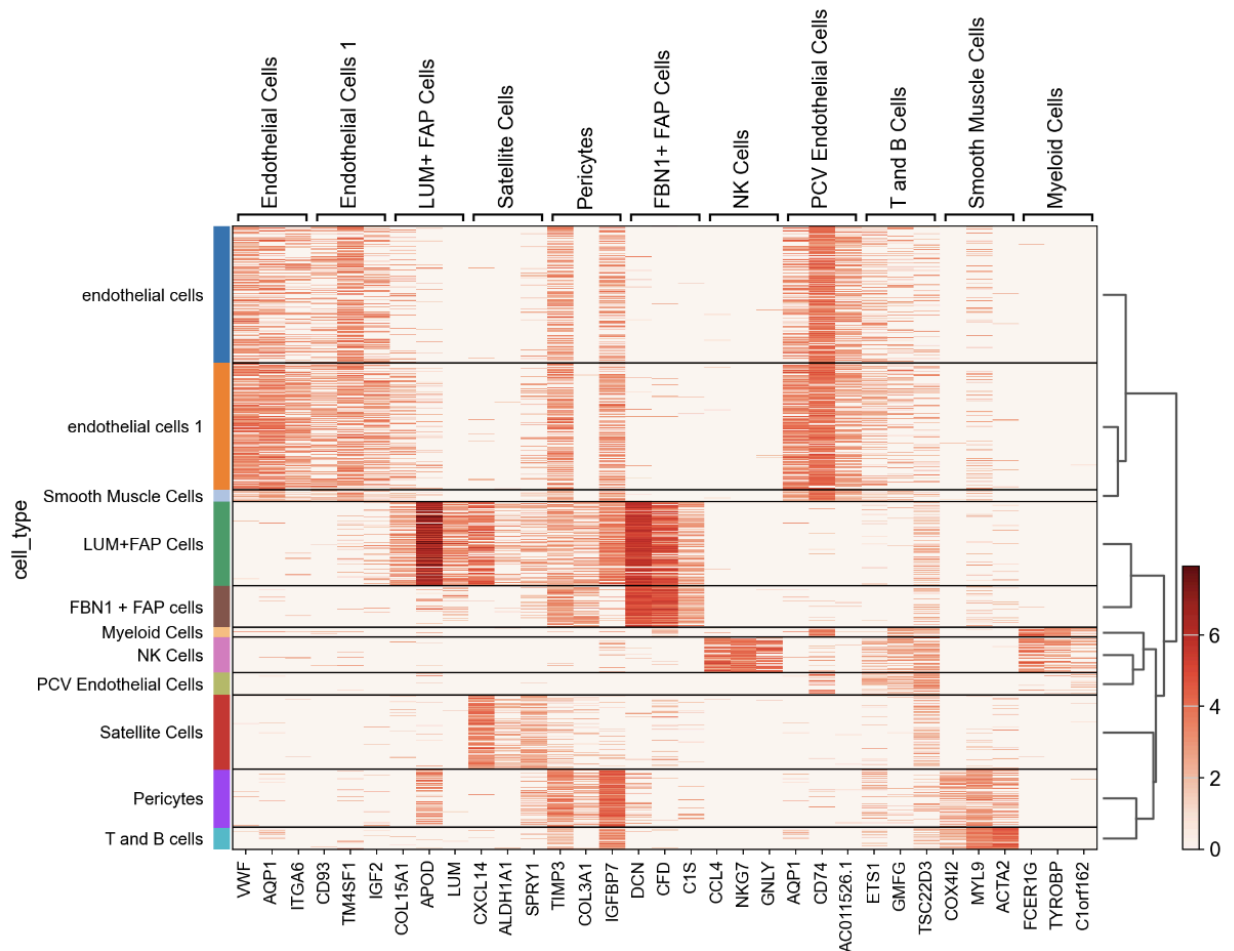


Figure. 2: Heatmap

2. References:

1. Rubenstein, A.B., Smith, G.R., Raue, U. *et al.* Single-cell transcriptional profiles in human skeletal muscle. *Sci Rep* 10, 229 (2020).
<https://doi.org/10.1038/s41598-019-57110-6>
2. Scanpy: Preprocessing and clustering 3k PBMCs
<https://scanpy-tutorials.readthedocs.io/en/latest/pbmc3k.html>
3. Scanpy: Core plotting functions
<https://scanpy-tutorials.readthedocs.io/en/latest/plotting/core.html>
4. Seurat's [guided clustering tutorial](#) (Satija et al., 2015)
https://satijalab.org/seurat/articles/pbmc3k_tutorial
5. anndata: Annotated data
Isaac Virshup, Sergei Rybakov, Fabian J. Theis, Philipp Angerer, F. Alexander Wolf
bioRxiv 2021 Dec 19. doi:[10.1101/2021.12.16.473007](https://doi.org/10.1101/2021.12.16.473007).
<https://anndata.readthedocs.io/en/latest/>
6. The scverse project provides a computational ecosystem for single-cell omics data analysis.
Isaac Virshup, Danila Bredikhin, Lukas Heumos, Giovanni Palla, Gregor Sturm, Adam Gayoso, Ilia Kats, Mikaela Koutrouli, Scverse Community, Bonnie Berger, Dana Pe'er, Aviv Regev, Sarah A. Teichmann, Francesca Finotello, F. Alexander Wolf, Nir Yosef, Oliver Stegle & Fabian J. Theis
Nat Biotechnol. 2023 Apr 10. doi: [10.1038/s41587-023-01733-8](https://doi.org/10.1038/s41587-023-01733-8).
<https://www.nature.com/articles/s41587-023-01733-8>