

Earlham Institute summer school on bioinformatics

25-29 July 2016

Basic Bioinformatics Sessions

Practical 4: Multiple Sequence Alignment

Wednesday 6 July 2016

Multiple Sequence Alignment

Here we will look at some software tools to align some protein sequences. Before we can do that, we need some sequences to align. I propose we try all the human **homeobox** domains from the well annotated section of **UniprotKB**. Getting the sequences is a trifle clumsy, so concentrate now! There used to be a much easier way, but that was made redundant by foolish people intent on making the future ever more tricky!!

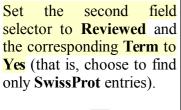
So, begin by going to the home of **Uniprot**:

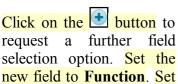
http://www.uniprot.org/

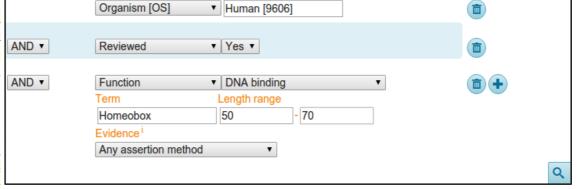
Choose the Advanced option of the Search button.

First specify that you are only interested in **Human** proteins. To do this, set the first field to **Organism [OS]** and

Term to Human [9606].



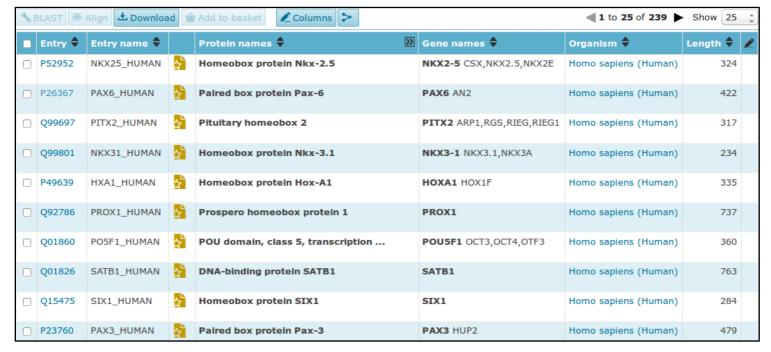




the type of Function to DNA binding. Set the Term selection to Homeobox.

From previous investigations, you should be aware that a **Homeobox** domain is **generally 60** amino acids in length. To avoid partial and/or really weird **Homeobox** proteins, set the **Length** range settings to recognise only **homeobox**s between **50** and **70** amino acids long.

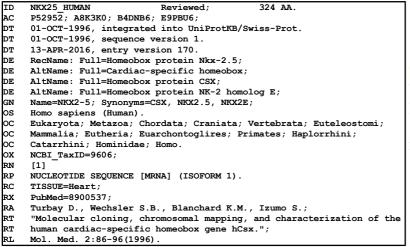
Leave the Evidence box as Any assertion method, one does not wish to be too fussy! Address the \(\text{\text{\text{\text{\text{o}}}}\) button with authority to get the search going.



A fine miscellany of sequences will assemble upon you screen. Most seem to declare themselves in possession of a **Homeobox** or two (including **PAX6 HUMAN**), so I suggest a declaration of success.

Course Description/Notes Now save the entire list into a file using the download to uncompressed. Make sure you have all sequences selected and that Text (i.e. EMBL or SwissProt) format selected. Press the Gobutton and do whatever it takes to ensure your results end up in a file residing on your Desktop called: Wednesday 6 July 2016 Download selected (0) Download selected (239) Format: Text Compressed Uncompressed

human homeobox proteins.emb



Take a swift look at the file you have just created. Your neat list of **Human Homeobox** sequences will have transformed into a flood of **many SwissProt** format **UniProtKB** entries. Ugly, but what is required.

Go

Preview first 101

Search (Control F) for the term DNA BIND.

It should occur many times (at least once per sequence) in the Feature Tables and most often refer to a **Homeobox** region.

In the **DNA_BIND** Feature Table entries, the position of the **Homeobox**es are recorded and will be used by the next program to isolate the sequence of the **Homeobox**es.

FT	CHAIN	1	430	Pre-B-cell leukemia transcription factor
FT				2.
FT				/FTId=PRO_0000049237.
FT	DNA BIND	244	306	Homeobox; TALE-type.
FT				{ECO:0000255 PROSITE-ProRule:PRU00108}.
FT	COMPBIAS	137	145	Poly-Ala.
FT	MOD RES	136	136	Phosphoserine.
FT	_			{ECO:0000269 PubMed:24275569}.
FT	MOD RES	151	151	Phosphoserine.
FT	_			{ECO:0000269 PubMed:18220336,
FT				ECO: 0000269 PubMed: 18669648,
FT				ECO: 0000269 PubMed: 20068231 } .
FT	MOD RES	330	330	Phosphoserine.
FT	_			{ECO:0000269 PubMed:18669648,
FT				ECO: 0000269 PubMed: 21406692 }.
FT	CONFLICT	393	393	M -> I (in Ref. 1; CAA42503).
FT				{ECO:0000305}.
SQ	SEQUENCE	430 AA;	45881 MW	; EF2FFA158C4DAF68 CRC64;
	MDERLLGPPP	PGGGRGGI	LGL VSGEPGO	GPGE PPGGGDPGGG SGGVPGGRGK QDIGDILQQI
	MTITDQSLDE	AQAKKHA I	LNC HRMKPA	LFSV LCEIKEKTGL SIRSSQEEEP VDPQLMRLDN
	MLLAEGVAGP	EKGGGSAZ	AAA AAAAAS	GGGV SPDNSIEHSD YRSKLAQIRH IYHSELEKYE
	QACNEFTTHV	MNLLREQS	SRT RPVAPKI	EMER MVSIIHRKFS AIQMQLKQST CEAVMILRSR
	FLDARRKRRN	FSKQATEV	VLN EYFYSH	LSNP YPSEEAKEEL AKKCGITVSQ VSNWFGNKRI
	RYKKNIGKFQ	EEANIYAV	VKT AVSVTQ	GGHS RTSSPTPPSS AGSGGSFNLS GSGDMFLGMP
	GLNGDSYSAS	QVESLRHS	SMG PGGYGDI	NLGG GQMYSPREMR ANGSWQEAVT PSSVTSPTEG
	PGSVHSDTSN			
//				

Now to extract from the whole protein sequences you have saved in a file, the sequences of just the **Homeobox** domains. One way of doing this (possibly not the best), is to use an **EMBOSS** package program called **extractfeat**. This can be found in many places, including the Bioinformatics server at **Wageningen** in the Netherlands. Go to:

http://emboss.bioinformatics.nl/

EDIT
aligncopy
aligncopypair
biosed
codcopy
cutseq
degapseq
descseq
entret
extractalign

extractfea

Find the program extractfeat (in the EDIT section), and set it going.

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	-Input section—
Use the Choose File button to upload the SwissProt format sequences from UniProtKB that you saved in the file human_homeobox_proteins.emb.	
	Additional section Amount of sequence before feature to extract
Set Type of feature to extract field to DNA_BIND (Make sure you remove the "*").	Amount of sequence after feature to extract Source of feature to display *
Set Value of feature tags to extract to Homeobox* (Make sure you append the "*" to ensure hits with, for example "homeoboxes").	Type of feature to extract DNA_BIND Sense of feature to extract (default is 0 - any sense, 1 - forward sense, -1 - reverse sense) Minimum score of feature to extract 0.0
	Tag of feature to extract * Value of feature tags to extract Homeobox*
Set the Output sequence format to SwissProt (Fasta would do, but SwissProt retains more annotation).	Output section Output introns etc. as one sequence? No Append type of feature to output sequence name? No V
Click on the Run extractFeat button to start	Feature tag names to add to the description Output sequence format SwissProt
extractfeat going. Many sequences of 60 amino acids (or so) in length will leap into view.	Run section Email address: If you are submitting a long job and would like to be informed by email when it finishes, enter your email address here
	Run extractfeat Reset
OUTPUT FILE outseq	
ID NKX25 HUMAN 138 197 Reviewed; 60 AA. DE [DNA contact] Homeobox protein Nkx-2.5 (Cardiac-specific homeobox) (Hi SQ SEQUENCE 60 AA; 7514 MW; 16EE564D071E5E8A CRC64; RRKPRVLFSQ AQVYELERRF KQQRYLSAPE RDQLASVLKL TSTQVKLWFQ NRRYKCKRQR // ID PAX6 HUMAN 210 269 Reviewed; 60 AA. DE [DNA contact] Paired box protein Pax-6 (Aniridia type II protein) (Oct SQ SEQUENCE 60 AA; 7447 MW; 075C194D089738D9 CRC64; LQRNRTSFTG EQIELEKEF ERTHYPDVFA RERLAKIDL PEARIQUWFS NRRAKWRREE	
// ID PITX2 HUMAN 85 144 Reviewed; 60 AA. DE [DNA_contact] Pituitary homeobox 2 (ALL1-responsive protein ARP1) (Homeobox 2 (ALL1-responsive protein ARP1)	meobox protein PITX2) (Paired-like homeodomain transcription factor 2) (RIEG bicoid-related homeobox transcription factor) (Solurshin

Right click the outsed button and select Save Link as... Do whatever it takes to save all your Homeobox domains into a file residing on your Desktop called:

homeobox_human.emb

Finally, we have some sequences with which to investigate the multiple sequence alignment programs.

Take a look at the file you have created. You should have many human homeobox domains in SwissProt format, looking rather as they did in your browser window. Happily ClustalX, the first multiple alignment program to be investigated, accepts multiple sequence SwissProt format files as input.

ClustalX is a part of the mostly widely known family of Multiple Sequence Alignments (MSA) programs, originating in the 1980s. Until relatively recently, it was the only real option. ClustalX still has merit, although it lacks some of the sophistication of more recent programs. ClustalX runs on effectively all workstations and has a

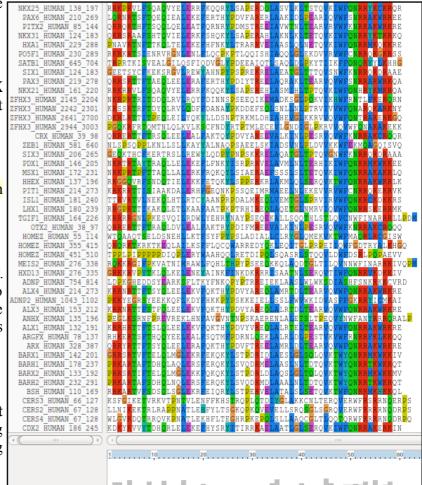
nice Graphical User Interface (GUI). A good place for us to start. It is installed on your workstations.

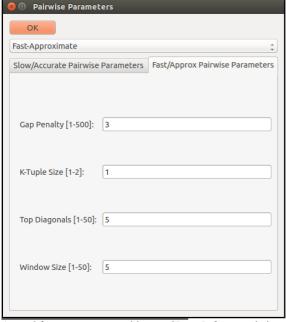
Start up the program ClustalX'. The ClustalX Graphical User Interface (GUI) will regally mount your screen.

Select Load Sequences from the File pull down menu and load your file of homeobox domains.

The sequences will arrange themselves colourfully. Many of the **homeoboxes** are similar enough to look convincing even before alignment. Note the "Manhattan skyline" under the sequences indicating the varying degrees of conservation.

You might like to increase the **Font** size from the minute default setting designed for Hawks and Eagles, to something more comfortable. **24** works tolerably well for me.





From the Alignment pull down menu, go to the Alignment parameters menu and select Pairwise Alignment Parameters. Just for a moment, change the setting from Slow-Accurate to Fast-Approximate. Bring the corresponding parameters into view by clicking on Fast/Approx Pairwise Parameters tab².

Hopefully, we will have discussed the way **ClustalX** (and similar multiple alignment tools) work. Intuitively, it should not make a lot of difference how the initial pairwise comparison stage is conducted. However, it very often does.

Specifically for this set of proteins, as well as generally, **ClustalX** will give a noticeably better alignment if the initial pairwise alignment stage is done carefully. Accordingly, reverse your whimsical setting change by moving back from **Fast-Approximate** to **Slow-Accurate**.

Of course, you could run Clustal from websites all over the world if you wished. Specifically, it is available both at the EBI and the Bioinformatics server at Wageningen. Try it if you have time. You get the same results but will, sadly, lose the pretty interface.

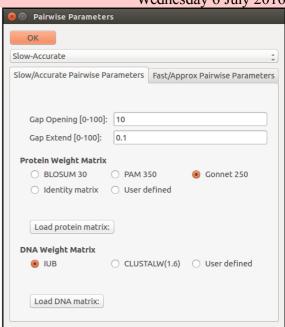
http://www.ebi.ac.uk/Tools/clustalw2/index.html

http://www.bioinformatics.nl/tools/clustalw.html

2 The **Fast-Approximate** algorithm is essential that which the database searching program **fasta** employs. Assuming we have discussed how **fasta** works, it should require no further explanation here.

Click on the Slow/Accurate Pairwise Parameters tab for a final look at the default parameters to be used. The Slow-Accurate option is essentially a version of Global Alignment algorithm we will have discussed previously. Hopefully, all the parameter options will therefore be familiar to you.

I will assume both sets of parameters at least seem familiar? If not please ask. The default **Slow/Accurate Pairwise Parameters** you now have in view are fine. Click the **OK** button to dismiss the **Pairwise Parameters** window.



Before proceeding, save the **homeobox** sequences in **FASTA** format, which will better suit the other **MSA** programs we will try. Do this by selecting **Save sequences as...** from the **File** pull down menu. **Deselect CLUSTAL format**, select **FASTA format**. Click **OK**. A file called **homeobox human.fasta** will be created. Take a look to check it is as you would expect.

Forma	at
	CLUSTAL format
	GCG/MSF format
	GDE format
\checkmark	FASTA format

Output Files	
CLUSTAL format	☐ NBRF/PIR format
☐ GCG/MSF format	☐ PHYLIP format
☐ GDE format	☐ NEXUS format
☐ FASTA format	

Strangely, saving your sequences in **FASTA** format convinces **clustalx** that it should now output its alignments in **FASTA** format. To prevent this, select **Output Format Options** from the **Alignments** pull down menu. Deselect **FASTA** format and select **CLUSTAL** format. Click **OK**.

From the **Alignment** pull down menu, select **Do Complete Alignment**. Accept the default names for output files and click on the **OK** button. **ClustalX** will start to think deeply and eventually come up with it view of how the **homeobox** domains should be aligned.

Not a bad first try. From an entirely non scientific, cosmetic viewpoint, the ragged ends offend a trifle, as does the gap just before position 30!



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In reality, these features might be very interesting, but here I go for pretty!

So, just to investigate what is possible, select all the **homeobox** sequences that are causing the gap around position **30** by clicking on their names (quite a lot of them I fear). Hold the **Ctrl** key down to allow multiple selection.

Once you have them all, go to the Edit pull down menu and select Cut Sequences. Then select Remove Gap-Only columns from the Edit pull down menu. Nasty gap gone ... along with all scientific credibility, but ... never mind.

You could recompute the alignment from scratch with the reduced sequence set, but you should end up with the same answer, of course. Just for the sake of it, select Select All Sequences from the Edit pull down menu. Then select Remove All gaps from the Edit pull down menu and confirm your intentions. You are now back where you started, but without the sequences that mess up the alignment intolerably!

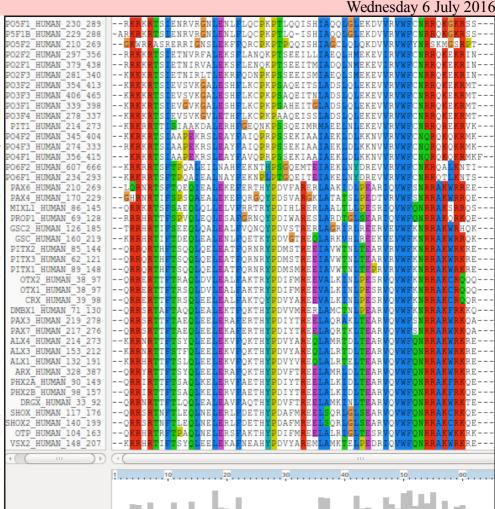
Save your filtered set of sequences in a file. From the **File** pull down menu select **Save Sequences as...** Choose **FASTA** format only.

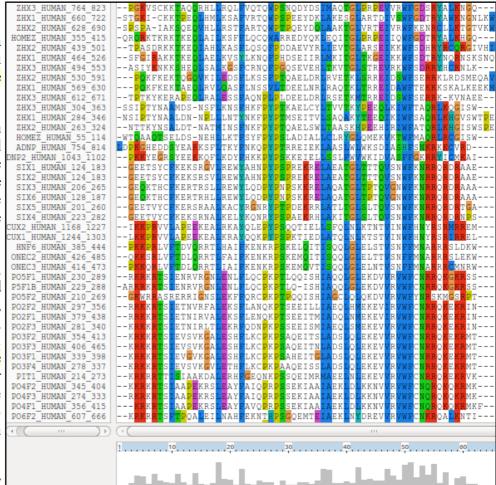
The default file name is OK, even though it will overwrite the original sequences. I am convinced the sequences eliminated would not be aligned convincingly with any of the tools we have at hand. Let us lose them! Press the **OK** button.

From the **Alignment** pull down menu, select **Output Format Options** and select **CLUSTAL format** only. Again, from the **Alignment** pull down menu, select **Do Complete Alignment**. Accept the default names for the output files. This will overwrite your previous efforts, but no matter. More deep thought. Well, I got back to where I was, no gaps around position **30** but still with ragged ends!

It is difficult to prove you have exactly

the same alignment as previously as the order of the MSA will be different. This order being determined by the pairwise comparison stage of the **clustalx MSA** computation.





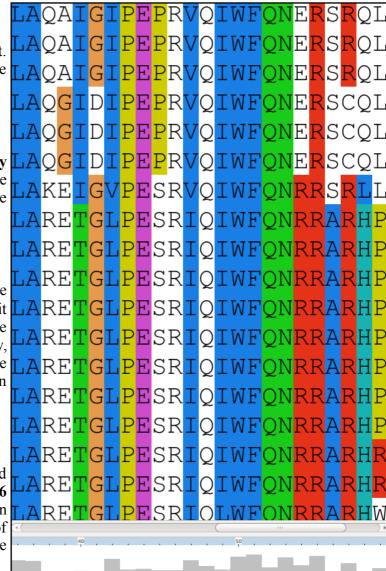
You will recall from earlier that the **Prosite** pattern for a **homeobox** is the ever memorable:

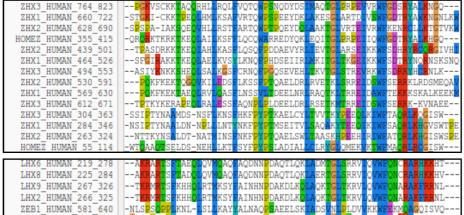
This corresponds to positions 36 to 59 in my alignment. See that the "Manhatten Skyline" is encouraging in the parts of this region that matter.

Note that the profile **Tryptophan**, in position **50**, is **very** CAQGI consistent, but not quite **100%** as suggested by the **Prosite** pattern³. The **W** was even conserved in the sequences that were cosmetically removed.

Position 52 is not conserved ("-x-") according to the Prosite pattern. In the alignment segment offered here, it looks like a pretty consistent Q. However, the "Manhattan skyline" at this position is very low, suggesting that the sequences in view might not be typical of the whole alignment set. Which, upon checking they are not!

Looking through this alignment, I get the feeling I could design a better, stricter pattern for the region between 36 and 59. Possibly true, but remember the pattern in Prosite aims to represent the conservation of Homeobox domains in ALL organisms. Here we have only sequences from Human.





AMNME<mark>P</mark>NSDELLK

YEEDLON

1 10 20 30 40 50 60

MTHK

Of course, things are not quite so convincing throughout. If you look at the top and bottom few sequences, you will see that **ClustalX** had its moments of uncertainty.

Note, however, the consistent **W** in position **50** despite the surrounding crumble.

ZEB2 HUMAN 644 703

ZHX3 HUMAN 835 894

ZHX1 HUMAN 777 832

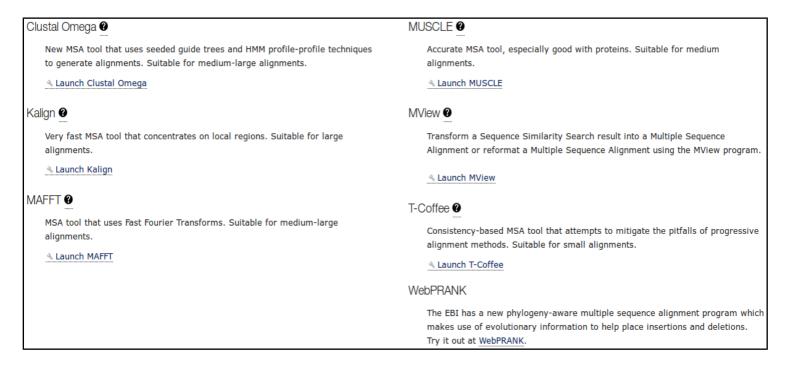
HOMEZ_HUMAN_451_510 NANGN_HUMAN_102_161

From the "Manhattan Skyline", you can see the conservation is less than 100%. Less conserved than the F that immediately follows in fact? Look at your alignment, the "Manhattan Skyline" does not seem to reflect reality? The W is very well conserved, although the scoring matrices would regard any deviation from W as serious? I need to find out more about how the Skyline is computed.

Now to show existence of some **msa** program options available on the web. There are many. They are available from a number of server sites. An obvious place to start has to be the **EBI** page dedicated to **MSA**. Go to:

http://www.ebi.ac.uk/Tools/msa/

Offered here is a selection of popular, current generation **MSA** tools. Each is accompanied by advice to guide the choice of tool to best fit the circumstances. Each tool is provided with a link to its **Launch** interface. All the **Launch** interfaces are very consistent. Once you have run one of the **MSA** options, you should have no trouble running any of the others.

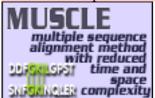


Here I intend to align again the human **homeboxes** with just one of the tools on offer. Then take a quick look at how the machine generated multiple alignment can be manually edited using **Jalview**, a program that is installed on your workstation and that you have already used as an alignment viewer when investigating **Pfam** and also **Jpred**.

Then I will invite you to try a few of the other options for yourself and see that they do not all produce the same alignment! Differences reflect not only the parameters selected, which we will have discussed, but also the particular objectives of the program selected. For example, a multiple protein sequence alignment optimal for investigating conservation of protein structure might well not be identical to one best representing protein evolution.

Used to align the **Homeobox** sequences used in this exercise, I do not expect you will see much difference between the outputs of any of these options. They will all work sufficiently on such a simple data set.

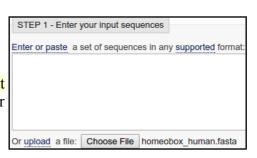
The program whose use I choose to describe carefully, leading on to a short **Jalview** exercise is **MUSCLE**. I choose thus as **MUSCLE** is now the first choice of most of the people with whom I work. Also popular are **Clustal Omega**, **MAFFT** and, for **phylogeny**, **WebPRANK**.



So the plan now is to use MUSCLE⁴ to align again the homeobox sequences previously aligned with clustalX. MUSCLE works in a way similar to clustalX but it takes rather more care in the generation of the Guide Tree used to control the order of pairwise construction of the final multiple alignment⁵. Particularly for more difficult alignments, MUSCLE should do a better job than clustalX. The alignment you will generate here will certainly be different. I leave you to judge for yourselves whether it is better.

Start by requesting to \(\lambda \) Launch MUSCLE

Use the **Choose File** button to upload the file containing the **FASTA** format **homeobox** sequences, **homeobox_human.fasta**. This file should no longer included the sequences with a mess around position **30**.



STEP 2 - Set your Parameters

OUTPUT FORMAT: ClustalW

The default settings will fulfill the needs of most users and, for that reason, are not visible.

More options... (Click here, if you want to view or change the default settings.)

Take a look at the **Set your Parameters** section of the page. I find the claim that "The default settings will fulfill the needs of most users and, for that reason, are not visible" a little strange? What about the users who are not in the category "most"? I want all the control

over the programs I use that their creators deemed it sensible to make available6?

The default settings behind the More options... button are not those that affect the computation of the MSA. I confess myself confused at the lack of any meaningful options to consider? I was expecting at least the gap open and gap extension penalty options (available elsewhere, including Wageningen), plus a way to change the scoring matrix. I have inquired why things are as they are



(most recently **2016.04.17**). No practical issue here, as I intended to suggest the defaults whatever they were. Look at the range of settings for the **OUTPUT TREE** parameter. **none** is indeed the thinking persons choice, but ... one or the other (but not both?) of the **Guide Trees** that **MUSCLE** will compute can be saved if you wish⁷. You may also set the **OUTPUT ORDER** to **aligned** or ... **aligned**?



There are a number of **OUTPUT FORMATS** offered. For a quick glance at your results, both **ClustalW** or **HTML** are fine. Here I suggest it would be nice to generate an output that can be downloaded and viewed in **Jalview**⁸. The default **ClustalW** or **Pearson/FASTA** serve for this purpose. As **ClustalW** looks more like an alignment in the web page, I choose **ClustalW**⁹.

How do the options for the **OUTPUT TREE** relate to the output files of **ClustalX** and the difference between the way that **ClustalX** and **muscle** work?

Comment on how one might choose between the range of options offered for the aligned parameter?

⁴ Available from a variety of websites in addition to the **EBI**, including the Bioinformatics server at **Wageningen**: http://www.bioinformatics.nl/tools/muscle.html

⁵ As discussed, superficially at least, previously. I hope.

I have asked the EBI about their policy (the same for all the locally provided MSA options). Discussion is ongoing (2016.04.20).

A useful option if you thought it possible you might want to rerun **MUSCLE** with different parameter setting for the stages after the **Guide**Tree(s) are generated. The same possibilities exist for **ClustalX**. Of course, utterly pointless if it is impossible to control the relevant parameters
.... so I really cannot see the point of any of the **More options** section? I am open to elucidation from all/any sources.

The java alignment editor and viewer you used to look at the **Pfam** and **Jpred** alignments earlier.

⁹ But feel free to try the others. **HTML** is the default at **Wageningen**. The **Phylip** formats are the best if you are going to analyse your output further with the phylogeny programs of the **PHYLIP** package.

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After considering these enigmas, or before if you prefer, Click on the sit back to admire muscle in action.	LHAS_HUNAN_207_320	QRRYR-TTFTSYQLEELERAFQKTHYPDVFTREELAMRLDLTEARVQVWFQNRRAKWRKRRHR-TTFTSYQLEELEKVFQKTHYPDVYVREQLALRTELTEARVQVWFQNRRAKWRKRRNR-TTFTSYQLEELEKVFQKTHYPDVYAREQLAMRTDLTEARVQVWFQNRRAKWRKRRNR-TTFSTFQLEELEKVFQKTHYPDVYAREQLAMRTDLTEARVQVWFQNRRAKWRKRRNR-TTFSTFQLEELEKVFQKTHYPDVYAREQLAMRTDLTEARVQVWFQNRRAKWRTTRVR-TVLNEKQLHTLRTCYAANPRPDALMKEQLVEMTGLSPRVIRVWFQNKRCKDKTTRVR-TVLNEKQLHTLRTCYAANPRPDALMKEQLVEMTGLSPRVIRVWFQNKRCKDKTTRVR-TSFKHHQLRTMKSYFAINHNPDAKDLKQLAQKTGLTKRVLQVWFQNARAKFRTKRMR-TSFKHHQLRTMKSYFAINHNPDAKDLKQLAQKTGLTKRVLQVWFQNARAKFRKRMR-TSFKHQLQVMQAQFAQDNNPDAQTLQKLAQMTGLSRRVIQVWFQNARAKFRAKRAR-TSFTAEQLQVMQAQFAQDNNPDAQTLQKLADMTGLSRRVIQVWFQNCRARHKDKRLR-TTITPEQLEILYQKYLLDSNPTRKMLDHIAHEVGLKKRVVQVWFQNTRARERDKRLR-TTITPEQLEILYQKYLLDSNPTRKMLDHIAREVGLKKRVVQVWFQNTRARERDKRLR-TTITPEQLEILYRYMYMQDSNPTRKMLDTISEEVGLKKRVVQVWFQNTRARERDKRLR-TTITPEQLEILYRYMYMQDSNPTRKMLDCISEEVGLKKRVVQVWFQNTRARERDKRLR-TTITPEQLEILYRYMYMGDSNPTRKMLDCISEEVGLKKRVVQVWFQNARAKEKGRRYR-TQMSSLQLKIMKACYEAYRTPTMQECEVLGBEIGLPKRVIQVWFQNARAKEKHKRFR-TQMSNLQLKVLKACFSDYRTPTMLECEVLGNDIGLPKRVVQVWFQNARAKEKPKRPR-TILTTQQRRAFKASFEVSSKPCRKVRETLAAETGLSVRVVQVWFQNARAKEKPKRPR-TILTTQQRRAFKASFEVSSKPCRKVRETLAAETGLSVRVVQVWFQNQRAKMK
The alignment that is computed is, superficially at least, similar to that offered by ClustalX .	LMX1B HUMAN 219 278 LHX1 HUMAN 180 239 LHX5-HUMAN 180 239 LHX3-HUMAN 157 216 LHX4-HUMAN 157 216 HOMEZ HUMAN 451 510	PKRPR-TILTTQQRRAFKASFEVSSKPCRKVRETLAAETGLSVRVVQVWFQNQRAMKRRGPR-TTIKAKQLETLKAAFAATPKPTRHIREQLAQETGLNMRVIQVWFQNRRSKERRRGPR-TTIKAKQLETLKAAFAATPKPTRHIREQLAQETGLNMRVIQVWFQNRRSKERAKRPR-TTITAKQLETLKSAYNTSPKPARHVREQLSSETGLDMRVVQVWFQNRRAKEKAKRPR-TTITAKQLETLKNAYKNSPKPARHVREQLSSETGLDMRVVQVWFQNRRAKEK : : :
The alignment is irritatingly split into two sections. A nice extra parameter might have been "How wide would you like your alignment to be"? A problem with the format rather than the program, to be fair.	ZHX3_HUMAN_835_894 ZHX1_HUMAN_777_832	RAV LGIELF ISW ISWSPE ISW VSWTPE SNS SVQ KEM LKEEKM SMEQAV AEE RGIVHI NLK NSKSNQ HGQ

At the very bottom of the page, **muscle** whines:

PLEASE NOTE: Showing colors on large alignments is slow.

```
So click the Show Colors button at the top of the page and try to live with the pain of such gross Trans-Atlantic inept spelling in a European site!!! Good Grief! They get everywhere!!
```

Well, an improvement I suppose? Colours are very useful (even slow ones) in the interpretation of alignments. Various colour schemes are used to clarify the message of alignments. Colouring can indicate shared amino acid properties not immediately evident when the letter representations differ.

```
ARX HUMAN 328 387
                              -- QRRYR-TTFTSYQLEELERAFQKTHYPDVFTREELAMRLDLTEARVQVWFQN
ALXĪ HUMAN 132 191
                              -- KRRHR-TTFTSLOLEELEKVFOKTHYPDVYVREOLALRTELTEARVOVWFONF
                              --KRRNR-TTFTSYQLEELEKVFQKTHYPDVYAREQLAMRTDLTEARVQVWFQNRRAKW
ALX4 HUMAN 214 273
                              -- KRRNR-TTFSTFQLEELEKVFQKTHYPDVYAREQLALRTDLTEARVQVWFQNRRAKW
ALX3 HUMAN 153 212
ISL1 HUMAN 181 240
                              --TTRVR-TVLNEKQLHTLRTCYAANPRPDALMKEQLVEMTGLSPRVIRVWFQNK
ISL2 HUMAN 191 250
                              --TTRVR-TVLNEKQLHTLRTCYAANPRPDALMKEQLVEMTGLSPRVIRVWFQNKRCKD
LHX9 HUMAN 267 326
                              --TKRMR-TSFKHHOLRTMKSYFAINHNPDAKDLKQLAQKTGLTKRVLQVWFQNARAKF
LHX2 HUMAN 266 325
                              --TKRMR-TSFKHHQLRTMKSYFAINHNPDAKDLKQLAQKTGLTKRVLQVWFQNARAKF
LHX6 HUMAN 219 278
                              --AKRAR-TSFTAEQLQVMQAQFAQDNNPDAQTLQKLADMTGLSRRVIQVWFQNCRARH
LHX8_HUMAN_225
                              --AKRAR-TSFTADQLQVMQAQFAQDNNPDAQTLQKLAERTGLSR
                                                                                 RVIQVWFQNCRARH
ZFHX3_HUMAN_2641_2700
ZFHX4_HUMAN_2560_2619
                              -- DKRLR-TTITPEOLEILYOKYLLDSNPTRKMLDHIAHEVGLKKRVVOVWFONTRARE
                              -- DKRLR-TTITPEQLEILYEKYLLDSNPTRKMLDHIAREVGLK
                                                                                 RVVOVWFONTRARE
                              -- DKRLR-TTILPEQLEILYRWYMQDSNPTRKMLDCISEEVGL
                              -- QRRYR-TQMSSLQLKIMKACYEAYRTPTMQECEVLGEEIGLP
                                                                               KRVIOVWFONARAKE
7FHX3 HUMAN 2944 3003
                              PGOKRER-TOMTNI OLKVI KSCENDYRTPTMI ECEVI GNDTGI PKRVVOVWEONARAKE
ZFHX4 HUMAN 2884 2943
                              -- HKRFR-TQMSNLQLKVLKACFSDYRTPTMQECEMLGNEIGLPKRVVQVWFQNARAKE
LMX1A_HUMAN_195_254
                              --PKRPR-TILTTQQRRAFKASFEVSSKPCRKVRETLAAETGLSVRVVQVWFQNQRAKM
                              --PKRPR-TILTTQQRRAFKASFEVSSKPCRKVRETLAAETGLSVRVVQVWFQNQRAKM
--RRGPR-TTIKAKQLETLKAAFAATPKPTRHIREQLAQETGLNMRVIQVWFQNRRSKEI
LMX1B HUMAN 219 278
LHX1_HUMAN_180_239
LHX5_HUMAN_180_239
                              -- RRGPR-TTIKAKQLETLKAAFAATPKPTRHIREQLAQETGLNMRVIQVWFQNRRSKE
 HX3 HUMAN 157
                              -- AKRPR-TTITAKQLETLKSAYNTSPKPARHVREQLSSETGLDMRVVQVWFQNRRAKE
LHX4_HUMAN_157_216
                              --AKRPR-TTITAKQLETLKNAYKNSPKPARHVREQLSSETGLDMRVVQVWFQNRRAKE
HOMEZ_HUMAN_451 510
                              EVV - - -
                              RAV---
ZHX1_HUMAN_777_832
                              LGIELF
HOMEZ HUMAN 55 114
                              ISW-
ZHX2_HUMAN_263_324
ZHX3_HUMAN_304_363
                              ISWSPE
ZHX1_HUMAN_284_346
                              VSWTPE
ZEB2_HUMAN_644_703
ZEB1_HUMAN_581_640
                              SNS--
                              SV0---
NANGN HUMAN 102
ZHX1_HUMAN_569_630
                              LKEEKM
ZHX2 HUMAN 530 591
                              SMEQAV
ZHX3 HUMAN 612 671
                              AEE-
ZHX2 HUMAN 439 501
                              RGIVHI
7HX3 HUMAN 494 553
                              MLK
7HX1 HUMAN 464 526
                              NSKSNO
HOMEZ HUMAN 355 415
```

But any decoration available here is far short of what can be achieved with **Jalview**, so click on the **Download Alignment File button** to save you alignment in a file on your **Desktop** called:

homeobox human muscle.aln

Jalview can be easily installed under all commonly used operating systems and run locally. For these exercises, I attempt to use services available freely from the INTERNET wherever possible, so let us run Jalview from the web here by first going to:

http://www.jalview.org/

and selecting the Launch Jalview Desktop link at the top of the page. Close down all the example outputs

Jalview sees fit to show you on start up. From the File pull down menu choose from File from the Input Alignment option. Locate and load the file homeobox_human_muscle.aln (you must adjust the file name

filter to included .aln files).

The default view is a trifle bland.

HXC12_HUMAN_214_273/1-60

HXD12_HUMAN_202_2617.60

HXD12_HUMAN_202_2617.60

HXD12_HUMAN_202_2617.60

HXD12_HUMAN_111_230/1-60

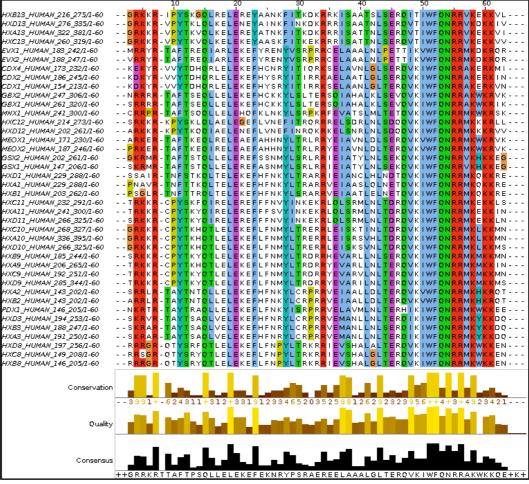
MEOX2_HUMAN_187_246/1-60

SX82_HUMAN_202_2617.60

You could try the default colour scheme used by **ClustalX**, for example.

HXA10_HUMAN_336_335;1-60
HXB10_HUMAN_266_325;1-60
HXB1_HUMAN_266_265;1-60
HXB2_HUMAN_192_25;17-60
HXB2_HUMAN_193_25;344;1-60
HXB2_HUMAN_193_26_365.60

Now the **MUSCLE** and massaged **ClustalX** alignments look even more similar! In the nicely aligned regions at least.



There are many **Jalview** features that merit investigation. Have a look around if you have time. In particular, **Jalview** will compute simple phylogenetic trees for you employing a number of methods (**Calculate Tree** from the **Calculate** pull down menu). Try it, but be aware this is only sensible if you were very sure of your alignment (and have a few less sequences maybe?).

Jalview is made by the same group as produce JPred. You could send your alignment for Secondary Structure Prediction via the Web Service pull down menu, if you wished.

A very important purpose of **Jalview** is to allow users to edit alignments as well as just to view them. For example, hold down the **Shift** key, click and hold on any amino acid at the edge of a gap, slide left and right and see that you can introduce and/or alter the position of gaps. It is very important to be able to edit alignments generated by even the best of programs. As I hope has been made clear, the alignment algorithms are crude. If you know something about the sequences you are aligning it is very reasonable to suppose you can improve upon the computer's alignments. **Jalview** tries to make this possibility easy. Look through some of the other **Edit** pull down menu options, it does not matter how much you mangle your alignment, you can always make another one.

Finally, take a look at the **Jalview** "**Manhattan Skyline**" for the highly conserved **W** at position **51**. This seems better quality than **clustalX** managed? I am not sure how one can make further comment without knowing what parameters were used. Is there really an improvement? If so, is it due to the improved alogorithm or more appropriate choice of parameters?

In my alignment, the **W** at position **51** was at position **50**, according to **clustalx**. This slippage to the right is due to **MUSCLE** introducing an extra gap, inspired by just one sequence at position **8**. Is this sensible? No idea ... exactly when it might be good idea to investigate the effect of lighter gap penalties? Not possible, but fear not children!! The sages of the **EBI** assure us that they have selected the correct setting for our sequences that they have never seen??? Sing praises and have faith brothers and sisters for we are in the safe hands of the blessed!!!

10

ZHX3_HUMAN - - S S I PT - Y N A A

ZHX1_HUMAN - - N S I PT - Y N A A

ZEB2_HUMAN - - GMT S P - I N P Y

ZEB1_HUMAN - - N L S P S - Q P P L

NANGN_HUM, - - Q Y P E K - R L V S

ZHX1_HUMAN - - P Q K F K - - E K T

ZHX2_HUMAN - - T P T K Y - K E R A

ZHX3_HUMAN - - T P T K Y - K E R A

ZHX3_HUMAN - - T P A S D - R K K T

ZHX3_HUMAN - - S F G I R - A K K T

HOMEZ_HUMAN - - S P S P A - I A K S

ZHX1_HUMAN - - S P S P A - I A K S

ZHX1_HUMAN - - S T G K I - C K K T

ZHX1_HUMAN - - S T G K I - C K K T

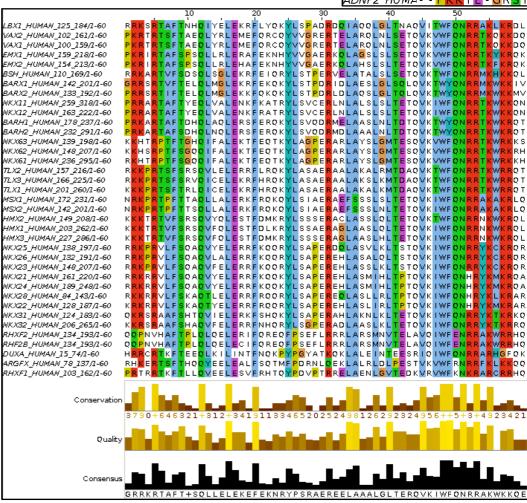
ZHX1_HUMAN - S T G K I - C K K T

ADNP_HUMAN L D P K G H E - D D S Y

ADNP2_HUMA - - P K K Y E - G R S Y

You can also **Select** and **Cut** wax1_Human_100_159R_160 sequences in a way similar to that you employed with **clustalx**. I could not resist it! I removed all the ugly sequences that caused the gaps at the start and finish of the alignment (just select their names and then select **Cut** or **Delete** from the **Edit** menu). I achieved the gap-free beautiful alignment illustrated.

Of course, **Jalview** does not compute alignments, so once I had removed all the unfortunate proteins, I had to use an **Edit** option to tidy up my meddling. I used **Remove Empty Columns** to get rid of the gap columns at the start of the alignment. The gaps at the end just melted away once the sequences that supported their presence were removed.



Science is easy! Once you remove the need for honesty that is.

If it could be done slightly more meaningfully, I would suggest you might try some of the other **MSA** tools offered by the **EBI**, to investigate the differences in the alignments computed. Any differences might be due to different parameter selection or differences in the alignment. Whilst the **EBI** refuse to share with their users how they are running the programs, however, it is hardly worth the effort. Maybe I look for a more obliging service for next time?

PSI-BLAST

This program is used to find a comprehensive set of relatives of a protein. First, **BLAST** is used to find closely related proteins. From an alignment of these proteins a general "profile" (a Position Specific Scoring Matrix -**PSSM**) is computed. A **PSSM** is very similar in concept and purpose to an **HMM** profile in that it summarises significant features present in the sequences it represents.

A query against the protein database is then run using the **PSSM**, and a larger more widely associated group of proteins is found. This larger group is used to construct another PSSM, and the process is repeated until no more significantly matching new sequences can be detected, or the user tires of the whole process.

You have used PSI-BLAST integrated into Jpred already and similar ideas were used to create the PFAM alignments. Here we will use PSI-BLAST explicitly at the NCBI on the Paired DOMAIN of the PAX6 protein that you saved in a file earlier. It should be possible to detect a large family of PAX domains and to eventually multiply align them generating something like the Full alignment from the PFAM database viewed earlier¹⁰.

To investigate **PSI-BLAST** go first to the **NCBI** Home page at:

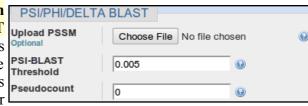
http://www.ncbi.nlm.nih.gov/

Click on the BLAST option. Select protein **BLAST** from the **Basic BLAST** section. Upload the PAX6 paired box domain sequence (stored in the file pax domain.fasta) using the appropriate **Browse** button.

Enter Query Sequence Enter accession number(s), gi(s), or FASTA sequence(s) 🤢 Query subrange @ From Or, upload file Choose File pax_domain.fasta Job Title Enter a descriptive title for your BLAST search (g) Align two or more sequences Choose Search Set Database Non-redundant protein sequences (nr) ▼ (a) Organism Exclude + Enter organism common name, binomial, or tax id. Only 20 top taxa will be shown. Exclude ■ Models (XM/XP) ■ Uncultured/environmental sample sequences **Entrez Query** Enter an Entrez query to limit search @ Program Selection Algorithm blastp (protein-protein BLAST) PSI-BLAST (Position-Specific Iterated BLAST) PHI-BLAST (Pattern Hit Initiated BLAST) O DELTA-BLAST (Domain Enhanced Lookup Time Accelerated BLAST)

Select PSI-BLAST from the **Program Selection** section. Leave all the others options at their default settings, particularly the option to search all the proteins available.

Before you set **PSI-BLAST** going, click on the **Algorithm** parameters link and take a look at the PSI/PHI/DELTA BLAST Upload PSSM section. Here is offered the option to use a **PSSM** from a previous run PSI-BLAST, potentially on a different database (but with the Threshold same query sequence). Accept the default that database entries Pseudocount scoring better than an Expect Threshold of 0.005 be offered for inclusion into the **PSSM** of each successive **PSI-BLAST** iteration. Remember the buttons.

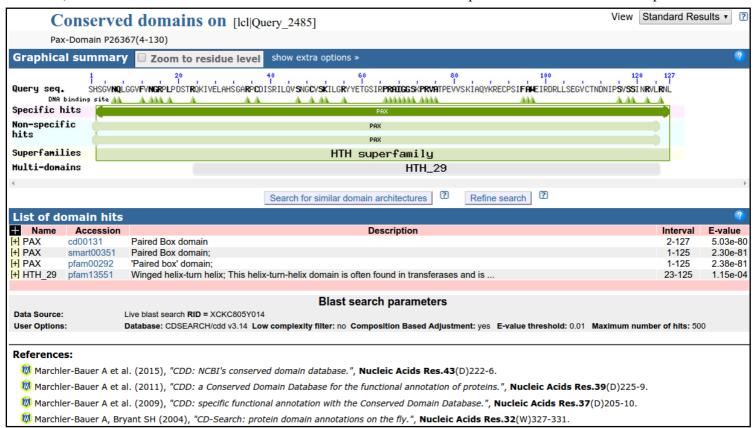


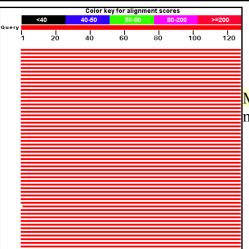
What do you suppose the choice of **Pseudocount** might influence?

Elect to Show results in a new window and then click on the button. After several moments of deep thought, **PSI-BLAST** will come back with its first set of results, at the top of which is a report that (unsurprisingly) matches have been detected between the query sequence and several domain databases. For more detail, click on the Conserved Domains graphic

Putative conserved domains have been detected, click on the image below for detailed results.										
	20	40	60	80	100	120 127				
Query seq.	HSGV NQ LGGV FVNGRPL PĎST R ÓKIVÉL									
DNA binding	site AA AAAA A	A_AA_A_/	64 A A66	4444 4444	444	A 44 A A				
Specific hits			PAX							
Superfamilies			HTH superf	family						
Multi-domains				HTH_29						
10 But hopefully	a mite more credible!									

SMART, Pfam and the NCBI Conserved Domains database hits are reported. None should be a surprise.

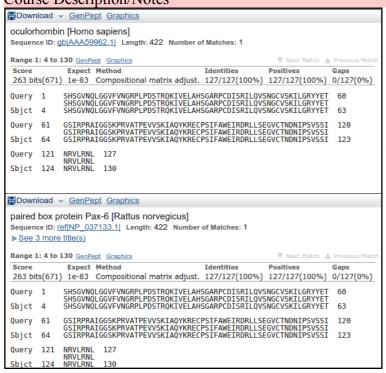




Moving back to the main PSI-BLAST results, you will see that there are many high quality hits covering the whole length of the query sequence.

	Seq	uences producing significant alignments with E-value BETT	ER tha	n thres	hold					
	Select: All None Selected:0									
	Alignments Download GenPept Graphics Distance tree of results Multiple alignment									
									Select	Used
		Description				_	Ident	Accession	for	to
The best 500 of these are			Score	Score	cover	value				
listed.		PREDICTED: paired box protein Pax-6 isoform X4 [Macaca nemestrina]	262	262	100%	1e-83	100%	XP 011722295.1	✓	
		PREDICTED: paired box protein Pax-6 isoform X2 [Ursus maritimus]	263	263	100%	1e-83	100%	XP 008685073.1	•	
		Description								
		paired box protein Pax-6 [Rattus norvegicus]	263	263	100%	1e-83	100%	NP 037133.1	✓	
All the listed hits are		PREDICTED: paired box protein Pax-6 isoform X2 [Fukomys damarensis]	263	263	100%	1e-83	100%	XP 010638711.1	•	
selected for inclusion into		PREDICTED: paired box protein Pax-6 isoform X2 [Cavia porcellus]	263	263	100%	1e-83	100%	XP 003464531.1	•	
the PSSM for the next		PREDICTED: paired box protein Pax-6 isoform X2 [Aotus nancymaae]	263	263	100%	1e-83	100%	XP 012307699.1	•	
iteration. Unless you feel		PREDICTED: paired box protein Pax-6 isoform X2 [Callorhinchus milii]	263	263	100%	1e-83	100%	XP 007885973.1	•	
strongly about any		PREDICTED: paired box protein Pax-6 isoform X2 [Heterocephalus glaber]	263	263	100%	1e-83	100%	XP 004851665.1	•	
particular entry, leave them		PREDICTED: paired box protein Pax-6 isoform X2 [Octodon degus]	263	263	100%	1e-83	100%	XP 004638029.1	•	
all selected.		PREDICTED: paired box protein Pax-6 [Poecilia reticulata]	261	261	100%	1e-83	98%		•	
Basic Bioinformatics - A Practical	Us	er Introduction 17 of 1 (25)						07	1:09:4	18 PM

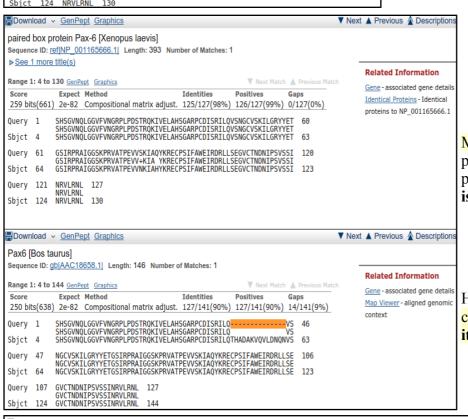
Course Description/Notes



Move down to the **Alignments** section of the results and you will see that many of the top hits match the query exactly.

Note that many of the top hits come from the **GenPept** database (roughly equivalent to the **TrEMBL** section of **UniProtKB**).

How might the inclusion of relatively poor quality sequences and the presence of so much duplication have been minimised?



Move down far enough and you will see less perfect matches, some of which involve proteins with the extra 14 amino acids of **isoform 5a** of **PAX6_HUMAN**.

Having browsed your results sufficiently, click on the Go button to Run PSI-Blast iteration 2 It is at the bottom of the hit list

paired box 6 [Monodelphis domestica] 238 238 94% 7e-76 90% ACZ54379.1 PREDICTED: paired box protein Pax-6-like isoform X1 [Acromyrmex ec 246 246 8e-76 XP 011063177.1 putative paired box protein pax-6 [Schistosoma mansoni] 254 99% 1e-75 90% CCD79466.1 putative Paired box protein Pax-6 [Operophtera brumata] 232 232 90% 1e-75 97% KOB68243.1 234 234 1e-75 89% NP 001189460.1 4 twin of eyeless [Bombyx mori] 242 242 99% 91% 4 PREDICTED: eyeless isoform X3 [Tribolium castaneum] 2e-75 XP 008192001.1 PREDICTED: eyeless isoform X2 [Tribolium castaneum] 242 242 99% 2e-75 91% XP 008192000.1 99% 245 94% 1 PREDICTED: paired box protein Pax-6-like isoform X1 [Megachile rotur] 245 2e-75 XP 012148240.1 Hypothetical protein CBG04481 [Caenorhabditis briggsae] 239 239 99% 2e-75 82% XP 002644124.1 pax6-like protein [Euperipatoides kanangrensis] 233 233 92% 3e-75 95% AGC51117.1 1 paired box protein Pax-6 [Clonorchis sinensis] 251 99% 251 3e-75 90% GAA48050.1 XP_013196296.1 4 PREDICTED: paired box protein Pax-6-like [Amyelois transitella] 231 231 91% 3e-75 92% hypothetical protein T265 09221 [Opisthorchis viverrini

After a few moments, **PSI-BLAST** will return with the results of searching through the database again using the **PSSM** derived from the hits of the first iteration(ded). This time the top of the list will be predominantly filled with hits that have already been incorporated into the **PSI-BLAST PSSM**. However, look far enough down the list and you will find some new ones, highlighted yellow.

Once more, click on the button to **Run PSI-Blast iteration 3**. That is probably enough! It took 4 iterations before there were no more new sequences suggested for inclusion into the **PSMM** when I ran this last, so if you really want to take things to their logical conclusion, it should not detain you long.

Next, move to the top of the **Descriptions** list and **Select All**. Click on the **Multiple Alignment** button. You have elected to use the **NCBI** multiple alignment program **Cobalt** to align all the **PAX** domain sequences of your final **PSI-BLAST** iteration that match with an **Expect** score better than **0.001**. In an impressively short time, your alignment will appear.

Multiple Alignment Results - Pax-Domain P26367(4-130) (127 letters)

Move past the long list of proteins that have been aligned (the easiest way is to hide the **Descriptions** view).

At the top of the actual alignment, set View Format to Plain Text (.... and then hide the Descriptions again??), this being the easiest format to understand in a hurry. The alignment will have very ragged ends, but the important region of 120 or so amino acids representing the PAX domain is really quite impressive. In particular, the isoform 5a insertion is very convincing¹¹.

✓ XP_003977912	52	TRQKIVELAHSGARPCDISRILQTHDAVQVLDSEKVSNGCVSKILGRYYETGSIRPRAIGGSK	114
✓ XP_009296159	26	TRQKIVELAHSGARPCDISRILQTHADAKVQVLDNENVSNGCVSKILGRYYETGSIRPRAIGGSK	90
✓ XP_003246075	54	TRQKIVELAHSGARPCDISRILQVSNGCVSKILGRYYETGSIKPRAIGGSK	104
✓ XP_012793883	41	TRQRIIELAHSGARPCDISRILQVSNGCVSKILCRYYETGSIRPKAIGGSK	91
✓ XP_005991286	25	TRQKIVELAHSGARPCDISRILQTHADAKVQVLDIQNVSNGCVSKILGRYYETGSIRPRAIGGSK	89
✓ EFX75780	37	TRQKIVELAHSGARPCDISRILQVSNGCVSKILGRYYETGSIRPRAIGGSK	87
✓ ABB43131	25	TRQKIVELAHSGARPCDISRILQVSNGCVSKILGRYYETGSIRPRAIGGSK	75
✓ ETN66652	41	TRQKIVELAHSGARPCDISRILQVSNGCVSKILGRYYETGSIKPRAIGGSK	91
✓ XP_006128959	56	TRQKIVELAHSGARPCDISRILQ <mark>THADAKVQVLDNQN</mark> VSNGCVSKILGRYYETGSIRPRAIGGSK	120
☑ XP_010874560	44	TRQKIVELAHSGARPCDISRILQTHDDSKVQVLDNENVSNGCVSKILGRYYETGSIRPRAIGGSK	108
✓ AFJ24746	53	TRQRIVELAHSGARPCDISRILQVSNGCVSKILCRYYETGSIRPKAIGGSK	103
☑ XP_007885968	25	TRQKIVELAHSGARPCDISRILQTHADAKVQVVDNRKVSNGCVSKILGRYYETGSIRPRAIGGSK	89
☑ BAA24024	42	TRQKIVELAHSGARPCDISRILQTHADAKVQVLDSQNVSNGCVSKILGRYYETGSIRPRAIGGSK	106
✓ XP_012307695	25	TRQKIVELAHSGARPCDISRILQ <mark>THADAKVQVLDNQN</mark> VSNGCVSKILGRYYETGSIRPRAIGGSK	89
✓ CBY09679	55	TRQKIVELAHSGARPCDISRILQVSNGCVSKILARYYETGSIKPRAIGGSK	105
✓ XP_007181079	82	TRQKIVELAHSGARPCDISRILQ <mark>THADAKVQVLDNQN</mark> VSNGCVSKILGRYYETGSIRPRAIGGSK	146
✓ CAF29075	25	TRQKIVELAHSGARPCDISRILQTHADAKVQVLDSENVSNGCVSKILGRYYETGSIRPRAIGGSK	89
✓ XP_004264009	25	TRQKIVELAHSGARPCDISRILQ <mark>THADAKVQVLDNQN</mark> VSNGCVSKILGRYYETGSIRPRAIGGSK	89
☑ XP_009184622	25	TRQKIVELAHSGARPCDISRILQ <mark>THADAKVQVLDNQN</mark> VSNGCVSKILGRYYETGSIRPRAIGGSK	89
✓ AAW24017	55	TRQKIVELAHSGARPCDISRILQVSNGCVSKILARYYETGSIKPRAIGGSK	105
✓ XP_008547741	26	TRQKIVELAHSGARPCDISRILQVSNGCVSKILGRYYETGSIRPRAIGGSK	76
☑ XP_012162452	50	TRQKIVELAHSGARPCDISRILQVSNGCVSKILGRYYETGSIKPRAIGGSK	100
✓ XP_006975926	25	TRQKIVELAHSGARPCDISRILQTHADAKVQVLDNENVSNGCVSKILGRYYETGSIRPRAIGGSK	89
✓ KDR14710	21	TRQKIVELAHSGARPCDISRILQVSNGCVSKILGRYYETGSIRPRAIGGSK	71
✓ XP_005530321	25	TRQKIVELAHSGARPCDISRILQ <mark>THADAKVQVLDNQN</mark> VSNGCVSKILGRYYETGSIRPRAIGGSK	89
■ ABI98847	25	TRQKIVELAHSGARPCDISRILQ <mark>THADAKVQVLDNQN</mark> VSNGCVSKILGRYYETGSIRPRAIGGSK	89
☑ XP_010794780	44	TRQKIVELAHSGARPCDISRILQTHDEVQVLDSEKVSNGCVSKILGRYYETGSIRPRAIGGSK	106
■ NP_001103907	26	TRQKIVELAHSGARPCDISRILQVSNGCVSKILGRYYETGSIRPRAIGGSK	76
✓ XP_010356630	25	TRQKIVELAHSGARPCDISRILQ <mark>THADAKVQVLDNQN</mark> VSNGCVSKILGRYYETGSIRPRAIGGSK	89
✓ XP 010638709	25	TRQKIVELAHSGARPCDISRILQ <mark>THADAKVQVLDNQN</mark> VSNGCVSKILGRYYETGSIRPRAIGGSK	89
✓ XP_005064878	25	TRQKIVELAHSGARPCDISRILQTHADAKVQVLDNENVSNGCVSKILGRYYETGSIRPRAIGGSK	89
№ NP 038655	25	TRQKIVELAHSGARPCDISRILQTHADAKVQVLDNENVSNGCVSKILGRYYETGSIRPRAIGGSK	89
✓ XP 005401829	25	TRQKIVELAHSGARPCDISRILQ <mark>THADAKVQVLDNQN</mark> VSNGCVSKILGRYYETGSIRPRAIGGSK	89
✓ XP 004638028	25	TRQKIVELAHSGARPCDISRILQ <mark>THADAKVQVLDNQN</mark> VSNGCVSKILGRYYETGSIRPRAIGGSK	89
■ BAM74254	32	TRQRIVELAHSGARPCDISRILQVSNGCVSKILGRYYETGSIRPRAIGGSK	82

Much more so than the **Full** alignment offered by **PFAM**, I would contend. Although, it has to be admitted, the **Pfam** alignment included more sequences and I suspect they would have gone for a less closely homologous set of sequences. Even so ... I think the alignment illustrated here is **MUCH** more beautiful!!

Model Answers Wednesday 6 July 2016

Model Answers to Questions in the Instructions Text.

Notes:

For the most part, these "**Model Answers**" just provide the reactions/solutions I hoped you would work out for yourselves. However, sometime I have tried to offer a bit moer back ground and material for thought? Occasionally, I have rambled off into some rather self indulgent investigations that even I would not want to try and justify as pertenent to the objective of these exercises. I like to keep these meanders, as they help and entertain me, but I wish to warn you to only take regard of them if you are feeling particularly strong and have time to burn. Certainly not a good idea to indulge here during a time constrained course event!

Where things have got extreme, I am going to make two versions of the answer. One starting:

Summary:

Which has the answer with only a reasonably digestible volume of deep thought. Read this one.

The other will start:

Full Answer:

Beware of entering here! I do not hold back. Nothing complicated, but it will be long and full of pedantry.

This makes the Model answers section very big. **BUT**, it is not intended for printing or for reading serially, so I submit, being long and wordy does not matter. Feel free to disagree.

Model Answers Wednesday 6 July 2016

From your investigations of Multiple Sequence Alignment

Clustalx

How might you have saved the need to recompute the alignment by selecting an **Edit** option other than **Remove All Gaps**?

Pretty sure using **Remove Gap-Only columns** rather than **Remove All Gaps** would have got to the second alignment in one step. I struggle to see how this could fail, but cannot convince myself it would be absolutely certain to work. Maybe some of the sequences that were removed would subtly alter the alignment calculations? Doubtful, given how crude the whole thing is. Anyway, it would have been good enough for me in "Real Life" for this data. I did not suggest this approach as we had to make a **Fasta** format file with the entire sequence set to be aligned by the alternative software as a by product of working with **clustalx**. This would not be easily possible if the **Remove Gap-Only columns** short cut was taken.

Muscle

How do the options for the **OUTPUT TREE** relate to the output files of **ClustalX** and the difference between the way that **ClustalX** and **muscle** work?

I leave this question here in the hope that one day I will be able to offer a sensible answer. First draft answer below.

Essentially, both ClustalX and MUSCLE work in two stages. First they create Guide Tree(s) (ClustalX just one, MUSCLE creates 2). Then from the Guide Tree(s) they compute the final MSA. The purpose of saving the Guide Tree(s) to a file is to enable a rerun of the second phase with new parameter settings without having to rerun the first Guide Tree generation stage.

Of course, as mentioned above, utterly pointless if there is no way to change the parameters? but that is the theory.

More investigation by me and expansion of this answer required. Discussion with EBI current (2016.04.20).

Comment on how one might choose between the range of options offered for the aligned parameter?

I cannot ... beyond suggesting it simply does not make sense? Going by what is offered at Wageningen, the choice should be between **aligned** and **input order**. i.e. the order of the original set of sequences to be aligned or the order after they have all been compared with each other and arranged into a **Guide Tree** ... or two.

What do you suppose the choice of **Pseudocount** might influence?

I clicked with confidences upon the link to the help. It opined as illustrated.



I suppose the next step is to read **PMID 19088134**? There is most certainly no elucidation amongst the strangle of words offered here?

The article **Abstract** says:

"Position specific score matrices (PSSMs) are derived from multiple sequence alignments to aid in the recognition of distant protein sequence relationships. The PSI-BLAST protein database search program derives the column scores of its PSSMs with the aid of pseudocounts, added to the observed amino acid counts in a multiple alignment column. In the absence of theory, the number of pseudocounts used has been a completely empirical parameter. This article argues that the minimum description length principle can motivate the choice of this parameter. Specifically, for realistic alignments, the principle supports the practice of using a number of pseudocounts essentially independent of alignment size. However, it also implies that more highly conserved columns should use fewer pseudocounts, increasing the inter-column contrast of the implied PSSMs. A new method for calculating pseudocounts that significantly improves PSI-BLAST's; retrieval accuracy is now employed by default."

The article itself, continues in like vein how about we close our eyes and accept the defaults? I would just wonder why the whole thing does not commence with, at least an attempt, to answer the question in the forefront of my inquiry, which is .. "WHAT, in the current context, IS a pseudocount?". I do not believe it is as tricky as they appear to wish us to believe. I will try again later, when my view of the world is less storm infested.

DPJ - 2015.09.14