Multicolor fluorescence per bacterial particle calibration

Plate readers report fluorescence values in arbitrary units that vary widely from instrument to instrument. Therefore absolute fluorescence values cannot be directly compared from one instrument to another. In order to compare fluorescence output of biological devices, it is necessary to create a standard fluorescence curve. This variant of the protocol uses two replicates of three colors of dye, plus beads. Adapted from https://dx.doi.org/10.17504/protocols.io.bht7j6rn and https://dx.doi.org/10.17504/protocols.io.6zrhf56

Protocol Outputs:

Dataset: multicolor-particle-calibration.xlsx

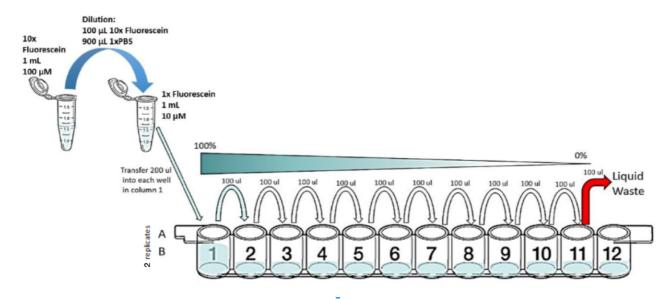
Protocol Materials:

- Water, sterile-filtered, BioReagent, suitable for cell culture
- NanoCym 950 nm monodisperse silica nanoparticles
- Phosphate Buffered Saline
- Fluorescein
- Cascade Blue
- Sulforhodamine
- stock reagent container (x 4)
- 96 well plate

Protocol Steps:

- 1. Provision the stock reagent container containing Fluorescein calibrant
- 2. Provision the stock reagent container containing Sulforhodamine 101 calibrant
- 3. Provision the stock reagent container containing Cascade blue calibrant
- 4. Provision the stock reagent container containing NanoCym 950 nm microspheres
- 5. Transfer 1.0 milliliter of Phosphate Buffered Saline sample to wells A1:H12 of stock reagent container Fluorescein calibrant. The reconstituted Fluorescein should have a final concentration of 10 uM in Phosphate Buffered Saline
- 6. Vortex Fluorescein calibrant
- 7. Transfer 1.0 milliliter of Phosphate Buffered Saline sample to wells A1:H12 of stock reagent container Sulforhodamine 101 calibrant. The reconstituted Sulforhodamine standard will have a final concentration of 2 uM in Phosphate Buffered Saline
- 8. Vortex Sulforhodamine 101 calibrant
- 9. Transfer 1.0 milliliter of Water, sterile-filtered, BioReagent, suitable for cell culture sample to wells A1:H12 of stock reagent container Cascade blue calibrant. The reconstituted Cascade Blue calibrant will have a final concentration of 10 uM in Water, sterile-filtered, BioReagent, suitable for cell culture.
- 10. Vortex Cascade blue calibrant

- 11. Transfer 1.0 milliliter of Water, sterile—filtered, BioReagent, suitable for cell culture sample to wells A1:H12 of stock reagent container NanoCym 950 nm microspheres. The resuspended NanoCym 950 nm monodisperse silica nanoparticles will have a final concentration of 3e9 microspheres/mL in Water, sterile—filtered, BioReagent, suitable for cell culture.
- 12. Vortex NanoCym 950 nm microspheres
- 13. Provision a container named calibration plate such as: NEST96WellPlate, Corning96WellPlate360uLFlat.
- 14. Transfer 100.0 microliter of Phosphate Buffered Saline sample to wells A1:H12 of 96 well plate calibration plate. These are blanks.
- 15. Transfer 100.0 microliter of Water, sterile-filtered, BioReagent, suitable for cell culture sample to wells A1:H12 of 96 well plate calibration plate. These are blanks.
- 16. Transfer 200.0 microliter of Fluorescein calibrant sample to wells A1:H12 of 96 well plate calibration plate.
- 17. Transfer 200.0 microliter of Fluorescein calibrant sample to wells A1:H12 of 96 well plate calibration plate.
- 18. Transfer 200.0 microliter of Sulforhodamine 101 calibrant sample to wells A1:H12 of 96 well plate calibration plate.
- 19. Transfer 200.0 microliter of Sulforhodamine 101 calibrant sample to wells A1:H12 of 96 well plate calibration plate.
- 20. Transfer 200.0 microliter of Cascade blue calibrant sample to wells A1:H12 of 96 well plate calibration plate.
- 21. Transfer 200.0 microliter of Cascade blue calibrant sample to wells A1:H12 of 96 well plate calibration plate.
- 22. Transfer 200.0 microliter of NanoCym 950 nm microspheres sample to wells A1:H12 of 96 well plate calibration plate.
- 23. Transfer 200.0 microliter of NanoCym 950 nm microspheres sample to wells A1:H12 of 96 well plate calibration plate.
- 24. Perform a series of 10 2-fold dilutions on A1 using Phosphate Buffered Saline as diluent to a final volume of 200.0 microliter in wells ['A1', 'A2', 'A4', 'A5', 'A6', 'A7', 'A8', 'A9', 'A10', 'A11'] of 96 well plate calibration plate. For each transfer, pipette up and down 3X to ensure the dilution is mixed homogeneously.



Serial Dilution

- 25. Perform a series of 10 2-fold dilutions on **B1** using **Phosphate Buffered Saline** as diluent to a final volume of 200.0 microliter in wells ['B1', 'B2', 'B3', 'B4', 'B5', 'B6', 'B7', 'B8', 'B9', 'B10', 'B11'] of 96 well plate **calibration plate**. For each transfer, pipette up and down 3X to ensure the dilution is mixed homogeneously.
- 26. Perform a series of 10 2-fold dilutions on C1 using Phosphate Buffered Saline as diluent to a final volume of 200.0 microliter in wells ['C1', 'C2', 'C3', 'C4', 'C5', 'C6', 'C7', 'C8', 'C9', 'C10', 'C11'] of 96 well plate calibration plate. For each transfer, pipette up and down 3X to ensure the dilution is mixed homogeneously.
- 27. Perform a series of 10 2-fold dilutions on D1 using Phosphate Buffered Saline as diluent to a final volume of 200.0 microliter in wells ['D1', 'D2', 'D3', 'D4', 'D5', 'D6', 'D7', 'D8', 'D9', 'D10', 'D11'] of 96 well plate calibration plate. For each transfer, pipette up and down 3X to ensure the dilution is mixed homogeneously.
- 28. Perform a series of 10 2-fold dilutions on E1 using Water, sterile-filtered, BioReagent, suitable for cell culture as diluent to a final volume of 200.0 microliter in wells ['E1', 'E2', 'E3', 'E4', 'E5', 'E6', 'E7', 'E8', 'E9', 'E10', 'E11'] of 96 well plate calibration plate. For each transfer, pipette up and down 3X to ensure the dilution is mixed homogeneously.
- 29. Perform a series of 10 2-fold dilutions on F1 using Water, sterile-filtered, BioReagent, suitable for cell culture as diluent to a final volume of 200.0 microliter in wells ['F1', 'F2', 'F3', 'F4', 'F5', 'F6', 'F7', 'F8', 'F9', 'F10', 'F11'] of 96 well plate calibration plate. For each transfer, pipette up and down 3X to ensure the dilution is mixed homogeneously.
- 30. Perform a series of 10 2-fold dilutions on G1 using Water, sterile-filtered, BioReagent, suitable for cell culture as diluent to a final volume of 200.0 microliter in wells ['G1', 'G2', 'G3', 'G4', 'G5', 'G6', 'G7', 'G8', 'G9', 'G10', 'G11'] of 96 well plate calibration plate. For each transfer, pipette up and down 3X to ensure the dilution is mixed homogeneously.
- 31. Perform a series of 10 2-fold dilutions on H1 using Water, sterile-filtered, BioReagent, suitable for cell culture as diluent to a final volume of 200.0 microliter in wells ['H1', 'H2', 'H3', 'H4', 'H5', 'H6', 'H7', 'H8', 'H9', 'H10', 'H11'] of 96 well plate calibration plate. For each transfer, pipette up and down 3X to ensure the dilution is mixed homogeneously.
- 32. Discard 100.0 microliter from wells ['A11', 'B11', 'C11', 'D11', 'E11', 'F11', 'G11', 'H11'] of 96 well plate calibration plate. This step ensures that all wells contain an equivalent volume. Be sure to change pipette tips for every well to avoid cross-contamination
- 33. Transfer 100.0 microliter of Phosphate Buffered Saline sample to wells A1:H12 of 96 well plate calibration plate. This will bring all wells to volume 200 microliter.
- 34. Transfer 100.0 microliter of Water, sterile-filtered, BioReagent, suitable for cell culture sample to wells A1:H12 of 96 well plate calibration plate. This will bring all wells to volume 200 microliter.
- 35. Measure fluorescein and bead fluorescence of calibration plate with excitation wavelength of 488.0 nanometer and emission filter of 530.0 nanometer and 30.0 nanometer bandpass.
- 36. Measure sulforhodamine 101 fluorescence of calibration plate with excitation wavelength of 561.0 nanometer and emission filter of 610.0 nanometer and 20.0 nanometer bandpass.
- 37. Measure cascade blue fluorescence of calibration plate with excitation wavelength of 405.0 nanometer and emission filter of 450.0 nanometer and 50.0 nanometer bandpass.
- 38. Measure absorbance of calibration plate at 600.0 nanometer.
- 39. Import data into the provided Excel file: Dataset: multicolor-particle-calibration.xlsx.

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