



asterics

A tool for the exploration and integration of omics data

ASTERICS is an online tool designed to help you to perform your statistical and integrative analyses in an interactive and easy-to-use way.

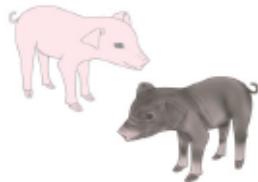
The illustration shows two scientists, one with red hair and one with dark skin, both wearing lab coats, interacting with a large digital screen displaying various data visualizations like bar charts, hexagonal grids, and circular plots. A magnifying glass highlights a specific area on the screen. The background is blue with abstract scientific icons like DNA helixes and chemical structures.

Quelques exemples avec des données multi-omiques et multi-tissus en néonatalité porcine pour illustrer les différentes fonctionnalités d'Asterics.

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Context: neonatal maturity and birth survival



Large White,
Meishan
and reciprocal
crosses

end of gestation

birth
0d 114d

1

```

graph TD
    Morphometry --> ClinicalBiology
    Morphometry --> Metabolome
    ClinicalBiology --> Metabolome
    
```

The diagram illustrates the relationship between Morphometry, Clinical biology, and Metabolome. Morphometry has two arrows pointing down to Clinical biology and Metabolome. Clinical biology also has an arrow pointing down to Metabolome.

```

    graph TD
        Place[Place] --> Adrenals[Adrenals]
        Place --> Blood[Blood]
        Place --> Jejunum[Jejunum]
        Place --> Liver[Liver]
        Adrenals --> Transcriptome[Transcriptome]
        Blood --> Transcriptome
        Jejunum --> Transcriptome
        Liver --> Transcriptome
        Transcriptome --> Pre[Pre]
        Transcriptome --> Post[Post]
    
```

The diagram illustrates the experimental workflow. Tissues (Adrenals, Blood, Jejunum, Liver) are sampled and their transcriptomes are analyzed. The transcriptome data is then used to compare pre- and post-treatment states.

A diagram showing a cross-section of bone tissue. A blue arrow points from the word "osteome" to a specific area within the bone structure.

A fluorescence microscopy image showing a bright blue band of staining against a dark background. A small white arrow points to a red fluorescent spot within this blue band. The bottom right corner contains the text "3 μm".

The diagram illustrates a biological workflow. A vertical line labeled "Serum" at the top points down to a horizontal line labeled "Metabolites". From this horizontal line, two arrows point to a chromatogram consisting of several vertical peaks.

```

graph TD
    A[Plasma membrane] --> B[Amino acids]
    B --> C[Polypeptide chain]
    C --> D[Protein]
  
```

The diagram illustrates the relationship between food, morphology, and the transcriptome. A green arrow points from 'Food' to 'Morphology'. Another green arrow points from 'Food' to 'Transcriptome'. The word 'Transcriptome' is written in blue. Below 'Transcriptome' is a graph showing four colored curves (red, orange, yellow, green) plotted against time (15, 18, 25, 30). To the left of the graph, the word 'Genes' is written in blue.

The image is a composite of two photographs. The left photograph shows a small piglet lying on a white surface, with a person's hands visible as they hold its front legs. A digital scale is positioned beneath the piglet, displaying the weight. The right photograph is a thermographic scan of a pig's body, showing heat distribution. A green outlined box highlights a specific area on the pig's side, and several small blue arrows point from these highlighted areas towards the top-left corner of the image, indicating the direction of heat flow or a specific measurement point.

3 stages: 90 or 110 days of gestation, birth
4 genotypes: LW, MS, LWMS, MSLW
N: 50 ... 600... 900

Voillet et al, 2014, 2018; Yao et al.2017 ; Gondret et al, 2018;
Marti-Marimon et al, 2018; Lefort et al, 2020,2021 ; Schmitt et al, 2021

ANR





asterics

A tool for the exploration
and integration of **omics** data

ASTERICS is an online tool designed to help you to perform
your statistical and integrative analyses in an interactive
and easy-to-use way.



[+ Create new workspace](#) [Load workspace](#)

Email

Load TCGA demo data into your workspace

I confirm that I have read and that I agree to Asterics' Privacy Policy.

[+ Create new workspace](#)

Your unique identifier will allow you to access your work and data during 30 days after the last activity on ASTERICS.

<https://asterics.miat.inrae.fr/>

Project coordinators



Funder



Partners



Dear user,

Your workspace has been successfully created at

<https://asterics.miat.inrae.fr/workspace/ba20fb6d-4196-45c0-999f-7ff6d522b109>.

Thank you for using ASTERICS,

The ASTERICS Team

Dear user,

your workspace <https://asterics.miat.inrae.fr/78f3057f-ed70-42c8-ae5c-8d8b7be83484> will be deleted in 3 days.

thanks for using Asterics,

The Asterics Team,

Datasets 0

Variables 0

Analyses 0

Remaining days 30

Copy url

Workflow

All datasets

Remove

Add

<input type="checkbox"/>	Name	Type	# Rows	# Columns	Log	Normalized
--------------------------	------	------	--------	-----------	-----	------------



Nothing's here...

All analyses

Remove

Export report

?

<input type="checkbox"/>	Name	Function
--------------------------	------	----------



Nothing's here...

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ASTERICS: User documentation

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2022-08-17

Section 1 Introduction

This document is the user guide for the web tool [ASTERICS](#). It is still a work-in-progress and mostly incomplete. Come back later for a better version. Thank you for your understanding.

Link to the application: <http://asterics.miat.inrae.fr/>

Support for [ASTERICS](#) can be obtained at asterics-tlse@inrae.fr.

Bugs can be reported [here](#) and suggestions can be made [here](#).



asterics

A tool for the exploration and integration of [omics](#) data

ASTERICS: User documentation

Section 16 Case studies

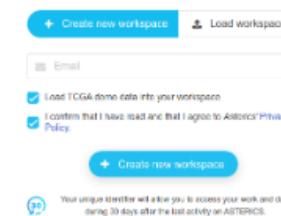
16.1 Breast cancer

This case study focuses on breast cancer, trying to link omics data to cancer subtypes. There are four breast cancer subtypes that somewhat represent sub-diseases, with different biological mechanisms. Their identification is particularly important when designing / selecting treatments. For this case study, we have mRNA and miRNA data on 988 individuals, as well as the associated subtypes. mRNA data is already normalized and is loaded in ASTERICS by default, whereas miRNA comes in the form of raw counts.

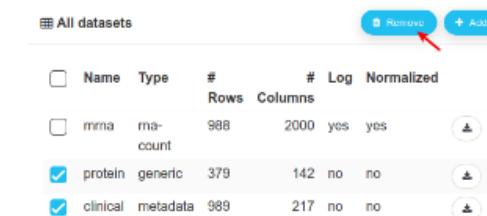
The data is available online ([Vialaneix 2021](#)).

16.1.1 Setup

Let us open an empty session and load the TCGA demo data.



Since we will only use the mRNA dataset, let us delete the other two.



Here is mRNA in the workspace:

mRNA
 rna-count
 #rows: 988 #col.: 2000

1
File upload**2**
Dataset structure

1. Select a file to upload

File

Select a file

Dataset name

Dataset name



The dataset name field is required

If you are not sure!

Dataset nature

Dataset nature



- generic
- sequencing data (RNAseq, miRNA, ...) counts
- microarray transcriptomics**
- metagenomics count data
- metagenomics composition (compositional data)
- metabolite/protein/lipid composition (compositional data)
- phenotypes
- metadata (or design)
- other

Before importation

 Have data been log-transformed? Have data been normalized?

Q Raw file

Select a file to get a preview

1. Select a file to upload

File

 TranscriptomeMuscle50.csv

Dataset name

 TranscriptomeMuscle50



Dataset nature

 microarray transcriptomics



If you don't know which nature to choose, use "generic".

Before importation

Have data been log-transformed?

Have data been normalized?



Q Raw file

;F405;F409;F411;F449;F455;F458;F469;F476;F481;F485;F498;F500;F524;F525;F53
A1CF_52154;5.6803695954516;5.45610320624039;5.74484272573927;5.92019267767
A2M_40313;5.50693239097091;5.76073759718855;5.5227941006065;5.663677602194
A2M_10645;5.41017364325691;5.79477650004684;6.04237760287688;5.68692117450
A2ML1_14639;5.4491082475774;5.36385961056552;5.17100665288747;5.6208513292
A4GALT_23662;8.36976136324847;7.80328185134206;8.55421636844331;7.22995624
A4GALT_9159;8.82297280814355;8.92285084244449;9.17229076620426;8.834796301
AAAS_61230;5.66491543991508;5.70809268946484;5.85667371223367;6.9757878359
AAAS_2894;5.89283524674721;5.6564212452558;5.66624190687793;6.090788421745
AAAS_18735;7.74349641324072;7.70023645818195;7.43741001836278;8.7585108190
AAAS_24237;6.41516988709249;6.40535808528583;6.82967565563798;7.0901431036

2. Set the dataset structure



Column names are included in the first row

Index of the column with row labels

1

Comment character

#

Switch rows and columns

For ASTERICS, individuals must be in rows.

Separator

, (comma) ; (semicolon) \t (tab) Space Custom :

Quote mark for text

" (double quote)

Decimal mark

. (dot) , (comma)

Encoding

Auto UTF-8 latin1

Missing values are encoded with these strings:

NA Add na.strings

Be patient: large files might take a long time to be uploaded and processed!



Import

Q Raw file

```
;F476;F481;F560;F578;F688;F455;F485;F498;F738;F899;F900;F469;F557;F559;F67
A1CF_52154;5.82599980707194;5.91662162096354;5.62770189995087;5.9748791178
A2M_40313;5.20290966528126;5.71662080795284;5.73529430299502;6.27994060037
A2M_10645;6.05043272137919;5.78608436449946;5.92397285829613;6.31840463378
A2ML1_14639;5.09454305674891;4.89392730422894;5.06610396900029;5.109927232
A4GALT_23662;7.44499238746051;7.88769803161222;7.75157714142366;7.85478806
A4GALT_9159;8.80734586555401;8.7439253300205;8.55935004010872;8.291677946
AAAS_61230;5.82599980707194;5.60366933006277;6.19088825000998;6.6908486193
AAAS_2894;5.59207508123505;5.29881778254516;5.87420980899597;5.93550088274
AAAS_18735;7.9656631095485;7.65139989082614;7.78415314320795;8.09455028881
AAAS_24237;6.38767805826733;6.53943474153246;6.73107621612731;6.5142754375
```

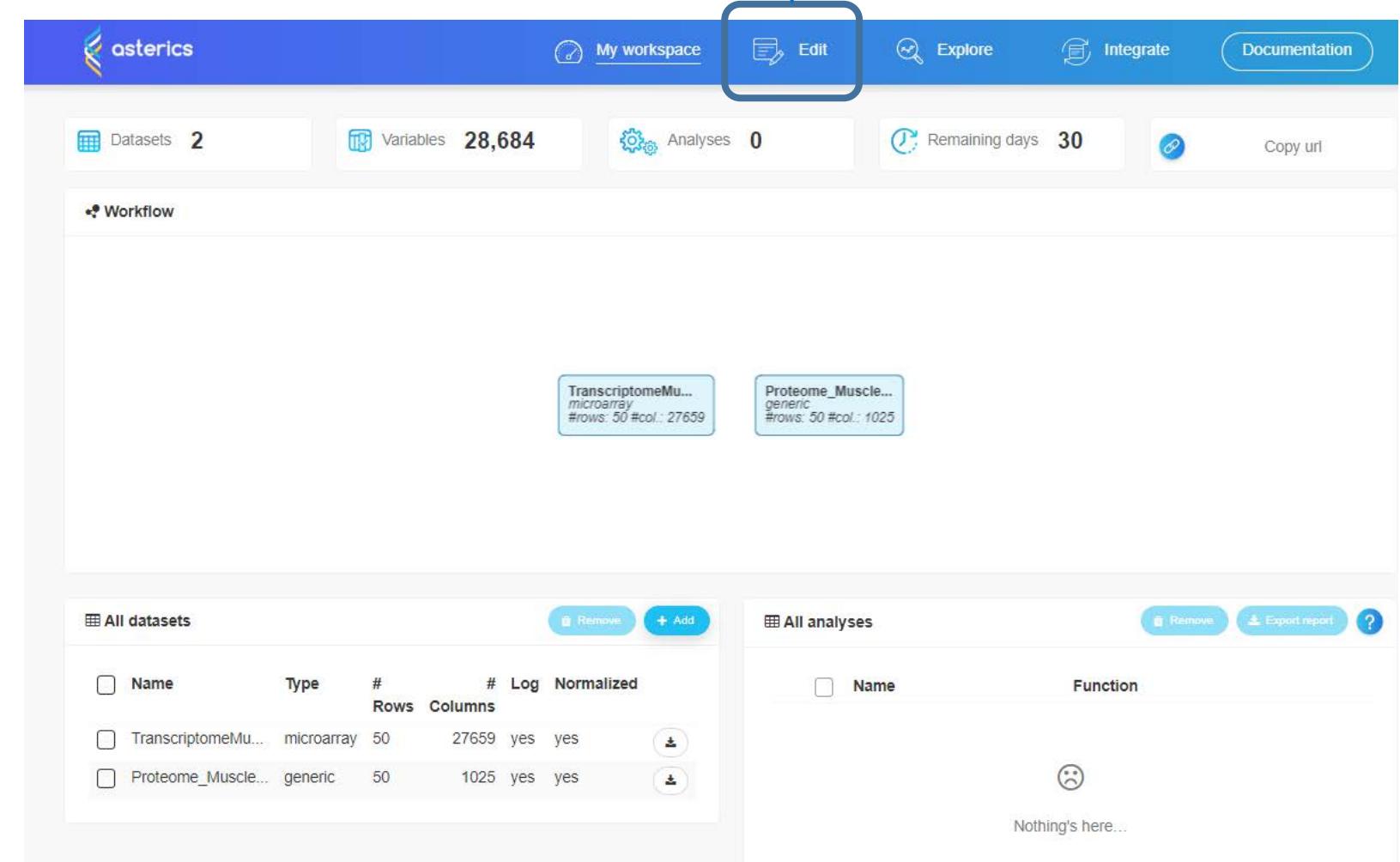
Switch rows and columns !

Q Dataset preview

	A1CF_52154 numeric	A2M_40313 numeric	A2M_10645 numeric	A2ML1_14639 numeric	A4GALT_23662 numeric	A4GALT_9159 numeric
F476	5.826	5.2029	6.0504	5.0945	7.445	5.60366933006277
F481	5.9166	5.7166	5.7861	4.8939	7.8877	6.19088825000998
F560	5.6277	5.7353	5.924	5.0661	7.7516	5.29881778254516
F578	5.9749	6.2799	6.3184	5.1099	7.8548	5.87420980899597
F688	5.7889	6.434	6.5869	4.7516	8.0703	5.93550088274
F455	5.5397	5.5826	5.8344	5.648	7.4885	6.53943474153246
F485	5.8868	6.1856	6.3453	5.661	7.7702	6.73107621612731
F498	5.6354	5.81	5.7552	5.8236	7.6827	6.5142754375
F738	5.6221	5.1073	5.5711	5.6786	7.1388	5.9748791178
F899	5.5986	5.5457	5.8591	5.0093	7.3409	5.06610396900029
F900	5.781	5.9127	6.1895	5.8663	7.7117	5.109927232
F469	6.1845	5.7959	5.7232	5.5926	8.0633	5.09454305674891
F557	5.9749	5.0005	5.051	5.5000	7.0404	5.20290966528126

1 unique identifier per variable, so arrange the name to have the most relevant information !

Edit



The screenshot shows the asterics workspace interface. At the top, there is a navigation bar with the following items: 'My workspace' (with a user icon), 'Edit' (which is highlighted with a blue box and has a blue arrow pointing down to it), 'Explore', 'Integrate', and 'Documentation'. Below the navigation bar, there are summary statistics: 'Datasets 2', 'Variables 28,684', 'Analyses 0', 'Remaining days 30', and a 'Copy url' button.

The main workspace area is titled 'Workflow' and contains two dataset cards:

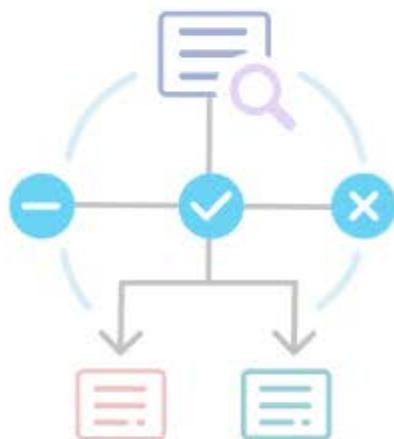
- TranscriptomeMu...** (microarray) #rows: 50 #col.: 27659
- Proteome_Muscle...** (generic) #rows: 50 #col.: 1025

Below these cards, there are two sections:

- All datasets**: A table with columns: Name, Type, # Rows, # Columns, Log, Normalized. It lists the two datasets above. Buttons for 'Remove' and '+ Add' are at the top right.
- All analyses**: A table with columns: Name, Function. It is currently empty, showing a sad face icon and the text "Nothing's here...".

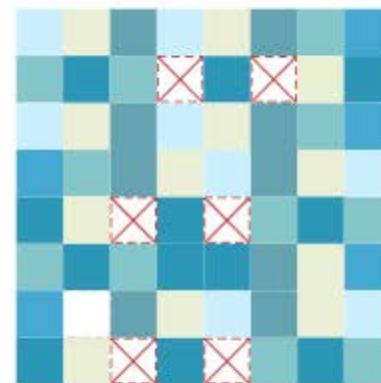


Let's edit data!



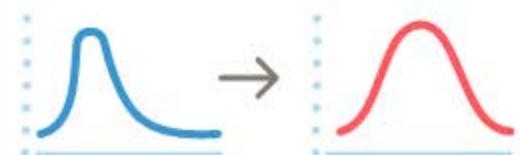
Dataset edition

Provides several ways to modify a dataset.



Missing values

Explore, remove or impute missing values on a dataset.



Normalize dataset

Correct technical biases and prepare your data for further analyses.



Dataset edition

1. Select a dataset and an action ?

Dataset

Infos_50 ▾

Actions

- Transpose
- Change dataset nature
- Change variable (column) types
- Set individual (row) names
- Subset individuals (rows)
- Subset variables (columns)
- Rename categories
- Reorder categories

History

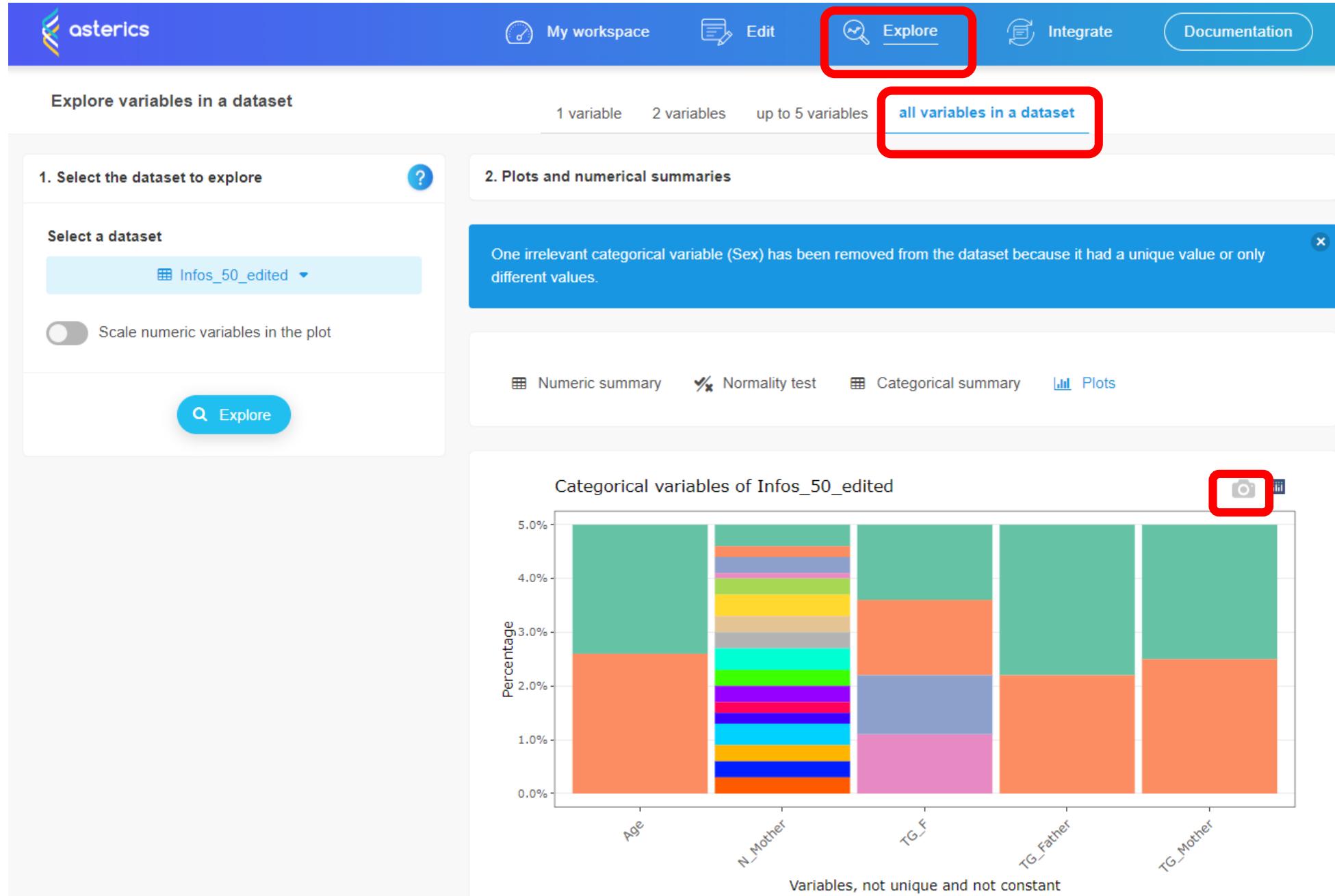
No history to display

Select an action.

Dataset "Infos_50"

# rows	# col.	# missing	% missing	# numeric	# cat.	# logic	# others
50	32	0	0	27	5	0	0

	Sex categorical 0.00%	N_Mother numeric 0.00%	Age categorical 0.00%	TG_Mother categorical 0.00%	TG_Father categorical 0.00%	TG_F categorical 0.00%	Weight numeric 0.00%
F449	M		6 90d	LW	LW	LWLW	733
F458	M		6 90d	LW	LW	LWLW	782
F500	M		8 90d	LW	LW	LWLW	597
F736	M		23 90d	LW	LW	LWLW	597
F744	M		23 90d	LW	LW	LWLW	784
F894	M		33 90d	LW	LW	LWLW	813
F895	M		33 90d	LW	LW	LWLW	389
F455	M		6 90d	LW	MS	MSLW	717
F485	M		8 90d	LW	MS	MSLW	754
F498	M		8 90d	LW	MS	MSLW	704
F738	M		23 90d	LW	MS	MSLW	522
F899	M		33 90d	LW	MS	MSLW	599





My workspace



Edit



Explore



Integrate

Documentation

1 variable

2 variables

up to 5 variables

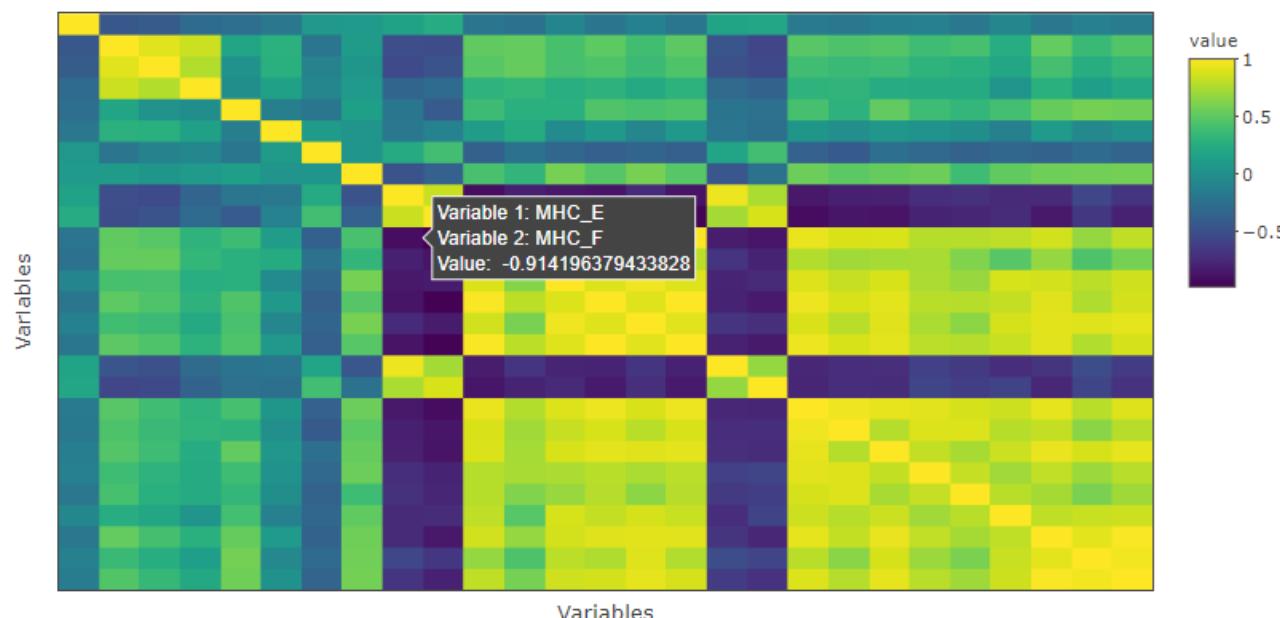
all variables in a dataset

2. Plots and numerical summaries

One irrelevant categorical variable (Sex) has been removed from the dataset because it had a unique value or only different values.

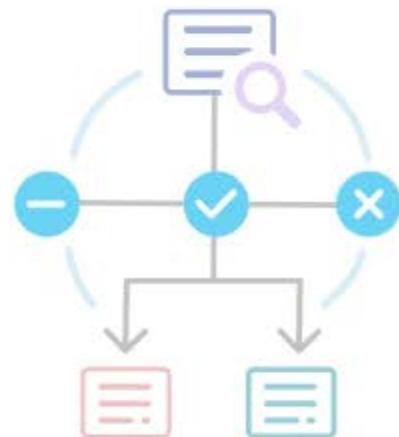
 Numeric summary Normality test Categorical summary Plots

Correlation plot of Infos_50



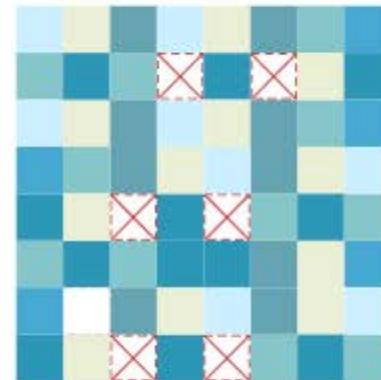


Let's edit data!



Dataset edition

Provides several ways to modify a dataset.



Missing values

Explore, remove or impute missing values on a dataset.



Normalize dataset

Correct technical biases and prepare your data for further analyses.



Which decisions
should I take to handle
missing values?

qPCR data...

# rows	# col.	# missing	% missing	# numeric	# cat.	# logic	# others
257	45	2760	23.87	45	0	0	0
	ATP2B1 numeric 1.17%	BCAS2 numeric 38.52%	BIRC5_F.R numeric 56.03%	CD14 numeric 39.30%	CD3E numeric 0.39%	CIR1 numeric 34.63%	CSNK2A1 numeric 79.38%
P1	0.0559	0.0623	NA	0.1167	0.0191	0.0132	NA
P2	0.0521	0.1207	NA	0.042	0.0828	0.0399	NA
P3	0.153	0.0737	NA	0.0761	0.0773	NA	0.0826
P4	0.0644	0.1384	NA	0.2347	0.0495	0.0555	NA
P5	0.2821	0.246	NA	0.3472	0.2379	0.0441	NA
P6	0.1404	0.1411	0.1522	0.0928	0.0943	NA	NA
P7	0.5945	NA	0.1526	NA	0.2019	0.3797	NA
P8	0.1211	0.1025	0.088	0.1648	0.141	NA	NA



My workspace



Edit



Explore



Integrate

Documentation

Missing values

Explore missing values

Remove missing values

Impute missing values

1. Select the dataset to analyze



Select a dataset

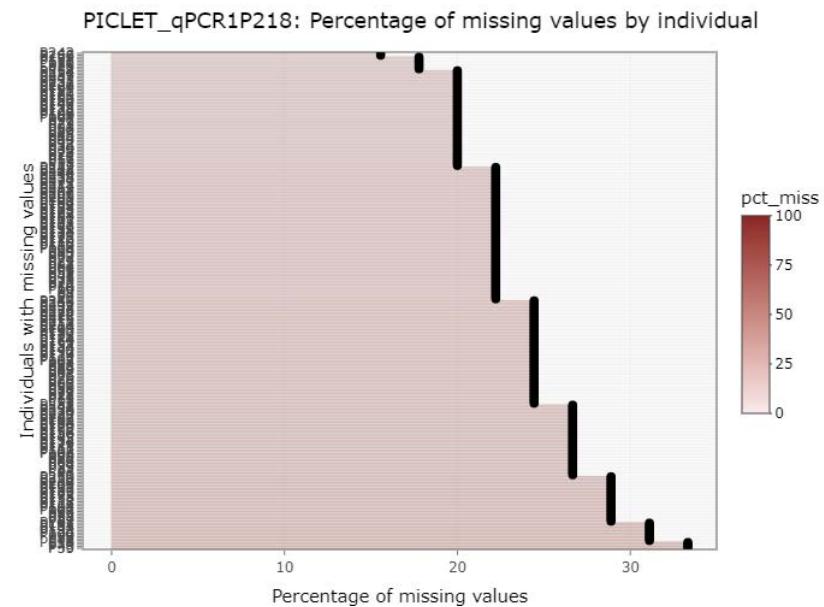
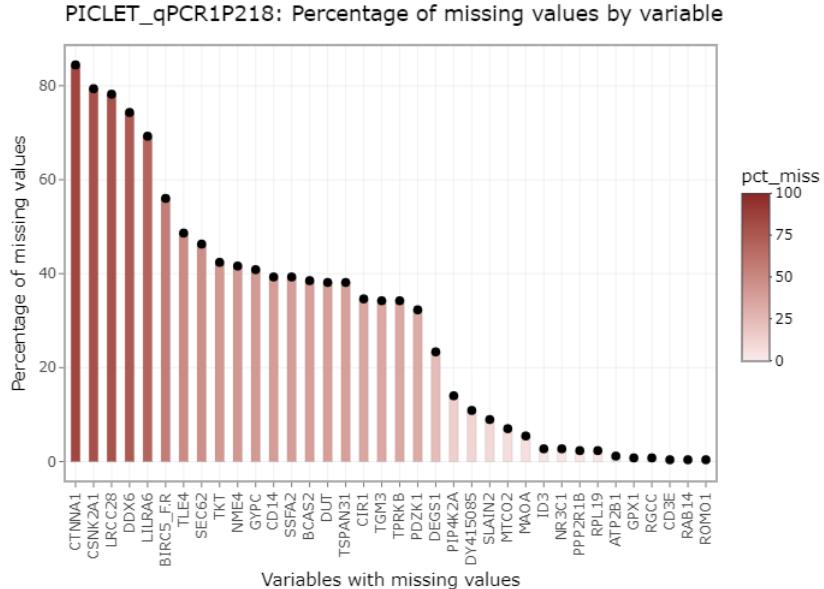
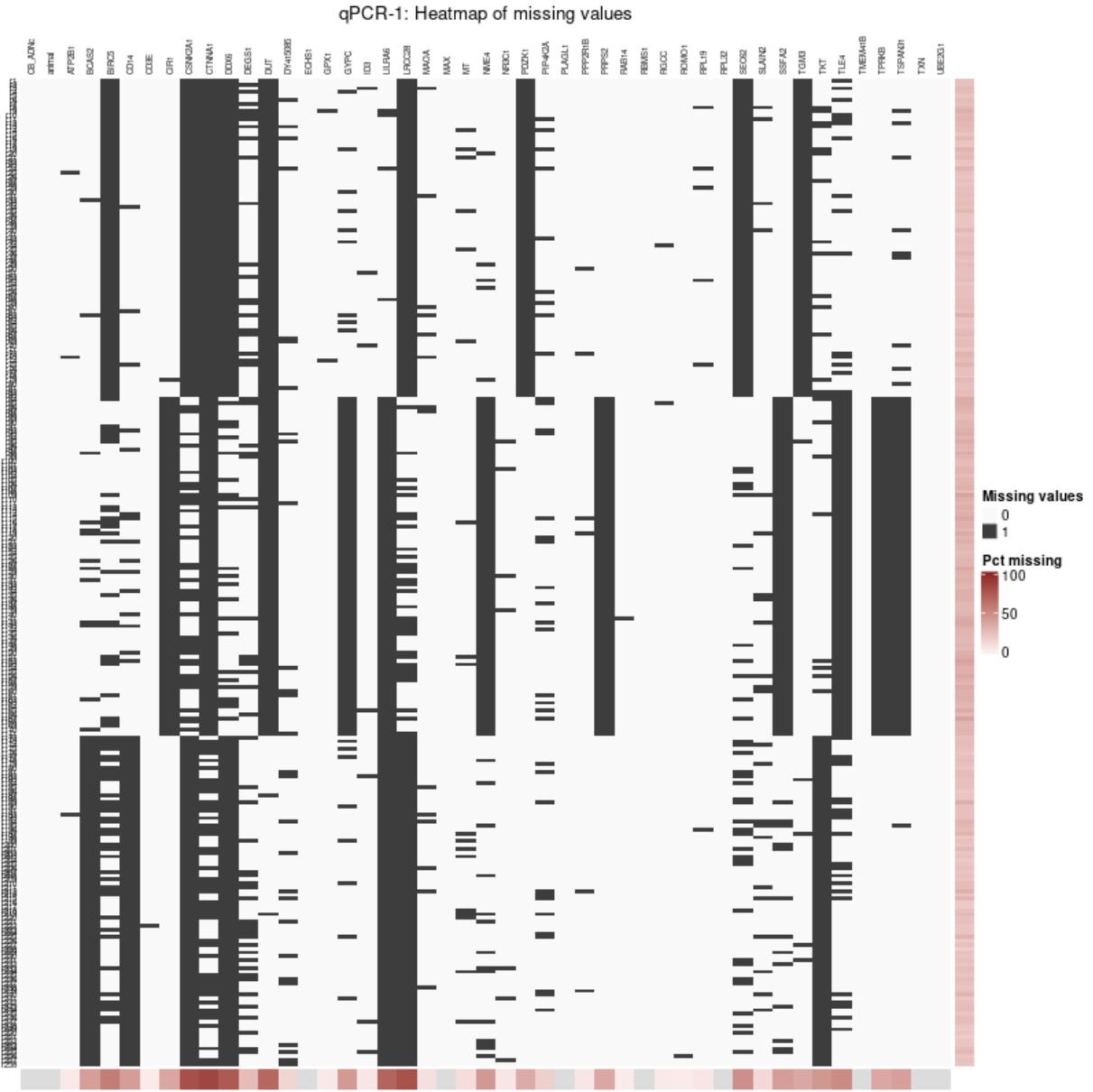
PICLET_qPCR1P218 ▾

2. Plots and numerical summaries

Heatmaps

Plots

Summary





Missing values

Explore missing values

Remove missing values

Impute missing values

1. Select the dataset to analyze 

Select a dataset

PICLET_qPCR1P218 ▾

Direction

* Individuals (rows) ▾

Maximum % of acceptable missingness in individuals/variables



Run

2. Plots and numerical summaries

Purpose of Missing values:

Handle missing values in your datasets

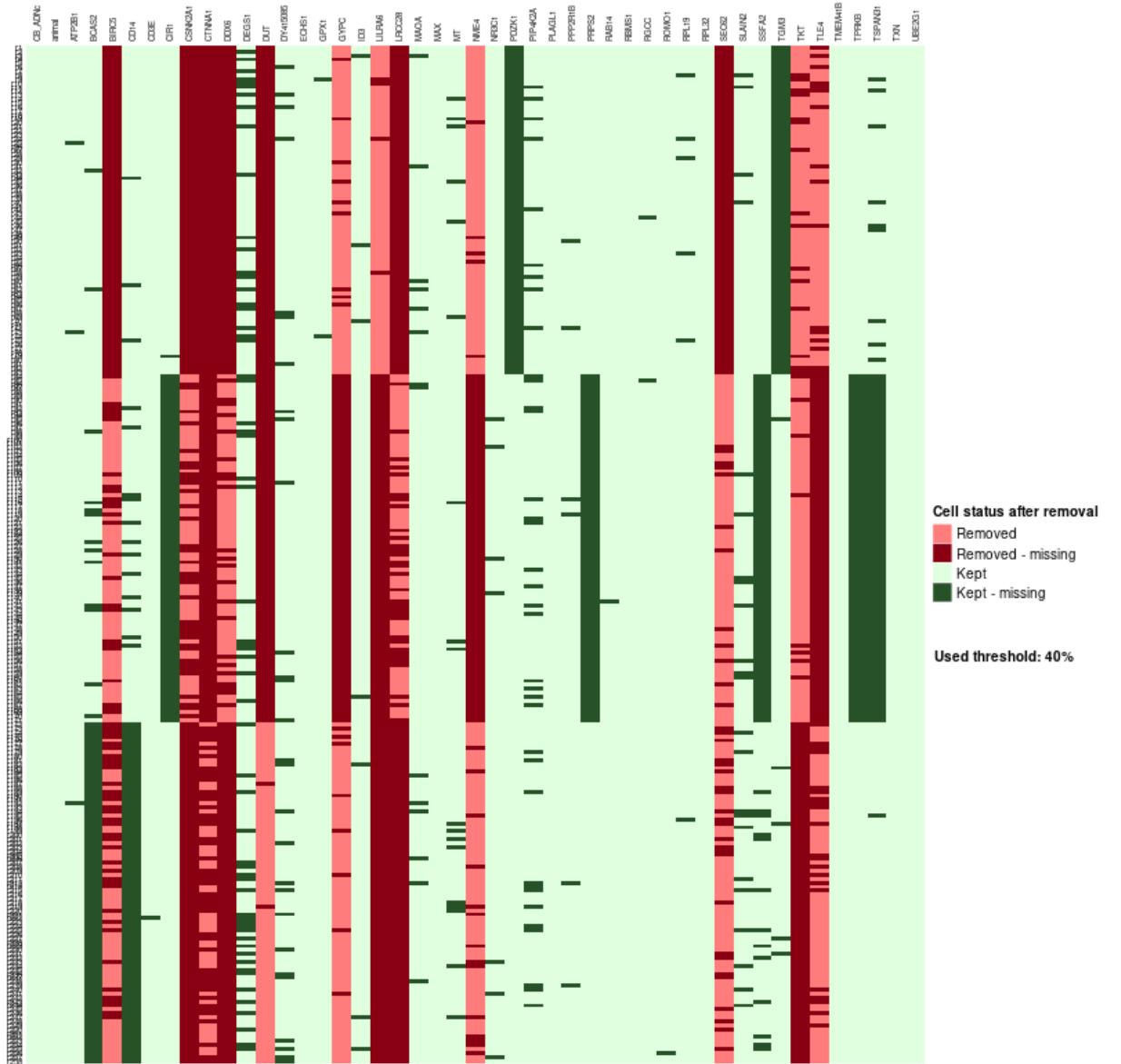
- Explore the distribution of missing values in your dataset
- **Impute** missing values with PCA, k-means or by zero.
- OR **remove** individuals / variables with the largest proportions of missing values.

How to set options?

- "Direction" is used to choose to remove either entire individuals (rows) or entire variables (columns).
- The chosen percentage corresponds to the maximum acceptable missingness. For instance, if 30% is chosen, it means that all individuals or variables with more than 30% of missing values will be removed.

Check help  for further advice.

qPCR-1: original dataset showing variables that will be removed





Missing values

Explore missing values

Remove missing values

Impute missing values

1. Select the dataset to analyze



Select a dataset

PICLET_qPCR1P218 ▾

Imputation method

PCA

Type of variables to impute

Auto

Run

2. Plots and numerical summaries

Purpose of Missing values:

Handle missing values in your datasets

- Explore the distribution of missing values in your dataset
- **Impute** missing values with PCA, k-means or by zero.
- OR **remove** individuals / variables with the largest proportions of missing values.

How to set options?

Imputation method can be chosen according to what you know on the missing values and subsequent analyses:

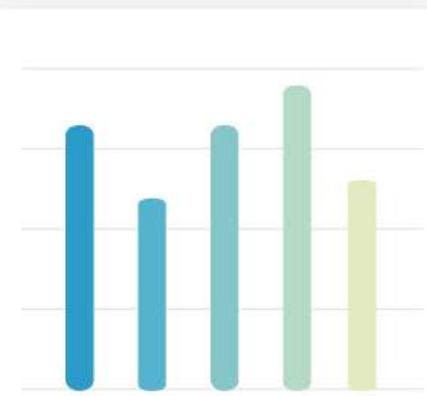
- **imputation by zeros** is dedicated to cases where missing values are due to a measurements below the detection threshold. It is a very basic approach to address this problem but certainly not the best;
- **imputation by PCA** is well designed when you want to use PCA, MFA, or PLS (for instance) afterwards because it best preserves the projection of your individuals on PC axes;
- **imputation by k-nearest neighbors** is based on the idea that two individuals that are similar on observed values also have similar values for unobserved variables. It best preserves the distances between individuals and is well adapted prior clustering.

The last two methods are only valid when data are missing at random.

In addition, for PCA and KNN you can choose to impute **only certain types of variables** (only numerical or only categorical variables). Setting this option to "Auto" imputes only variables of the most present type while "Mixed" imputes both numerical and categorical variables.

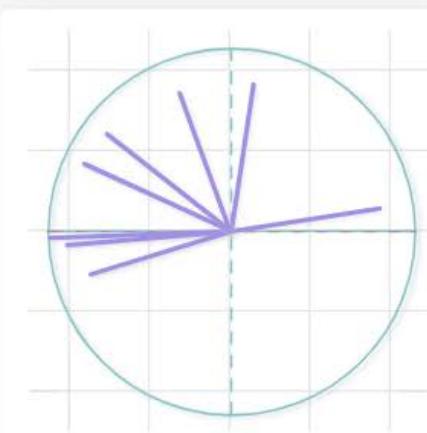


Let's explore a dataset!



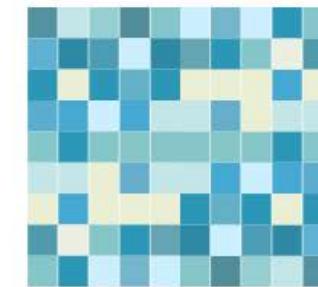
Explore variables in a dataset

Obtain numerical summaries and plots for a few variables.



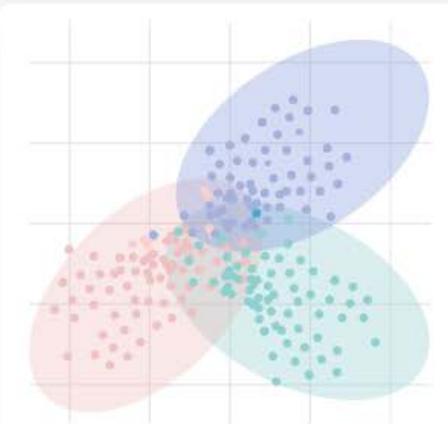
Explore a dataset with PCA

Perform Principal Component Analysis on a dataset.



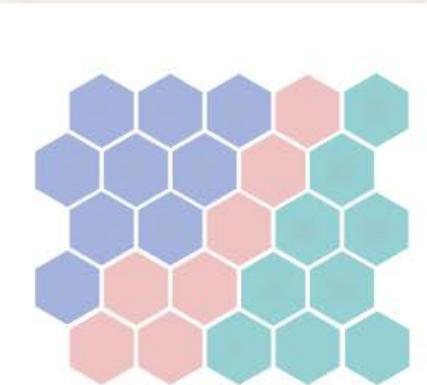
Explore a dataset with a heatmap

Obtain the heatmap of a dataset.



Clustering

Cluster the individuals of a dataset.



Self-Organizing Map

Use SOM as a clustering and visualization method.



Explore a dataset with PCA

1. Display individuals on components



Component to display on X (horizontal)

1



Component to display on Y (vertical)

2



Color



PICLET_qPCR...

Select a variable

Shape



Dataset

Select a variable

Size

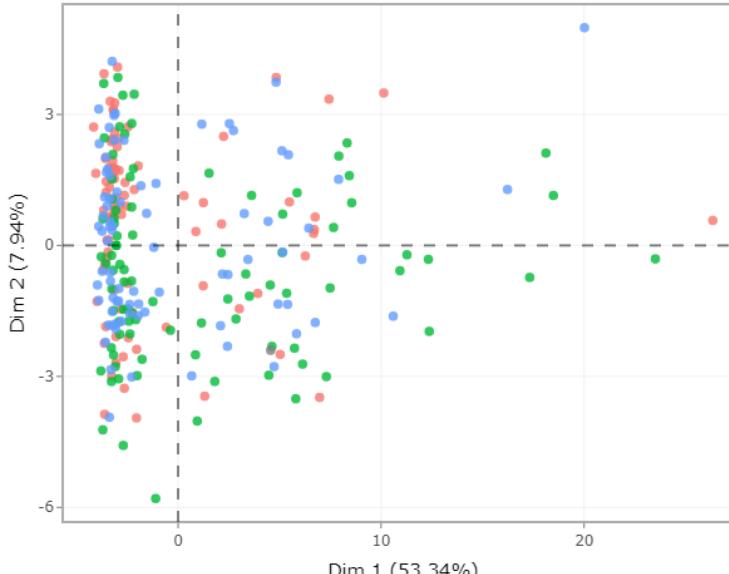


Dataset

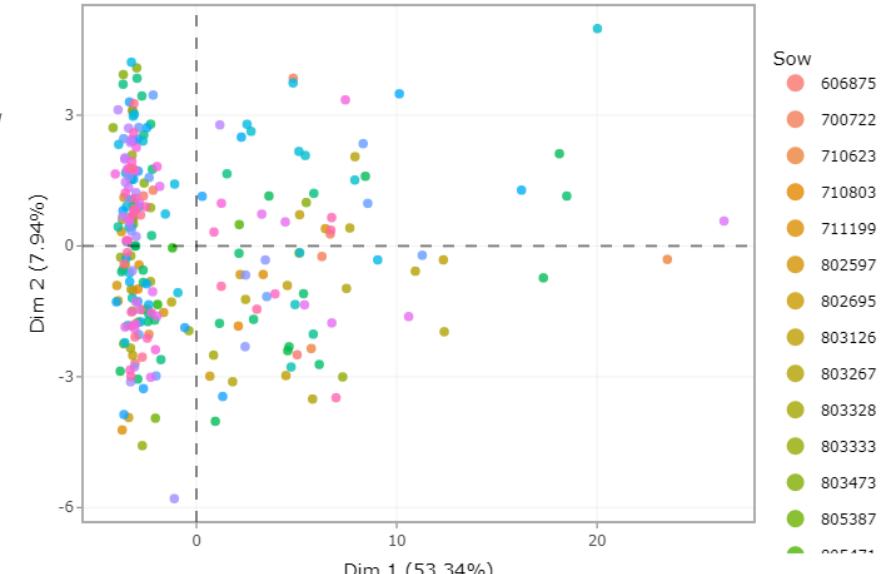
Select a variable

Plot individuals

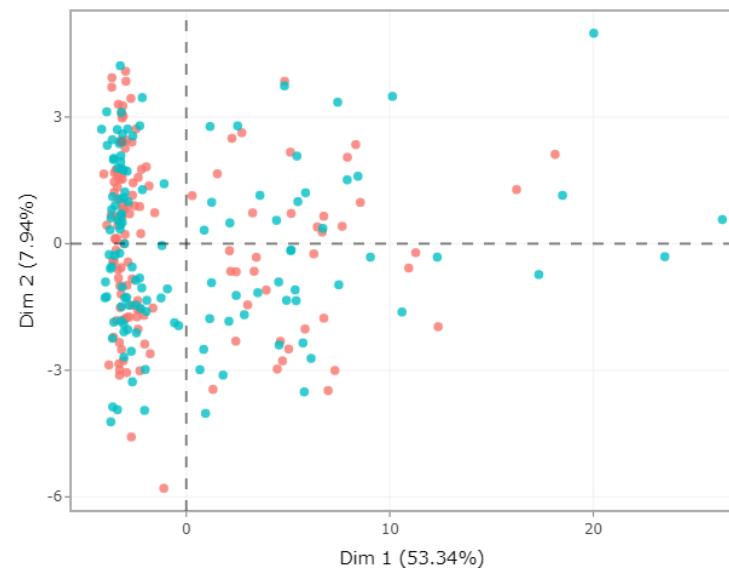
PCA on Data_qPCR_Piclet_e_Imputed_1: projection of individuals on the F



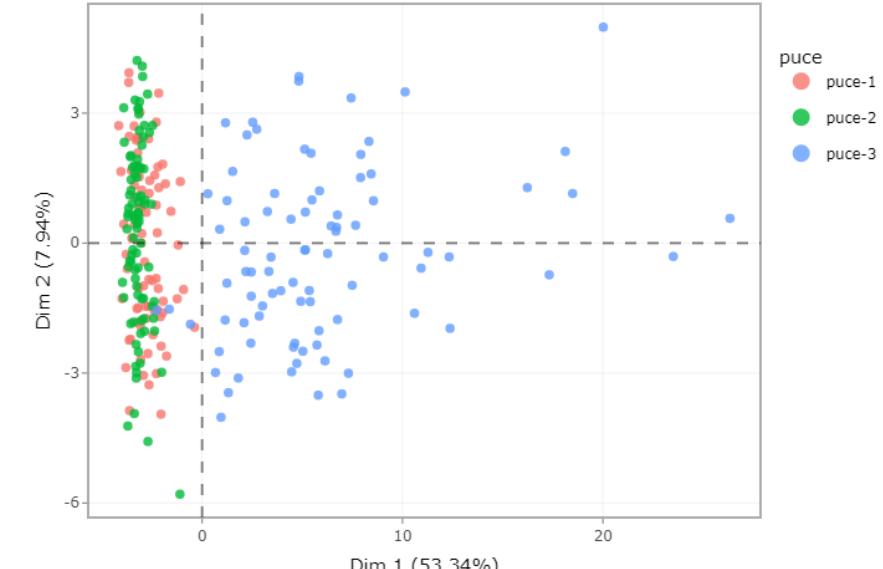
PCA on Data_qPCR_Piclet_e_Imputed_1: projection of individuals on the F



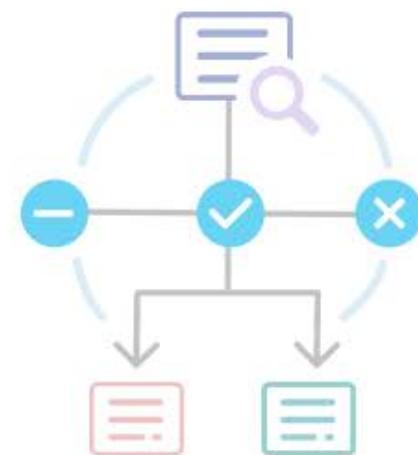
PCA on Data_qPCR_Piclet_e_Imputed_1: projection of individuals on the F



PCA on Data_qPCR_Piclet_e_Imputed_1: projection of individuals on the F

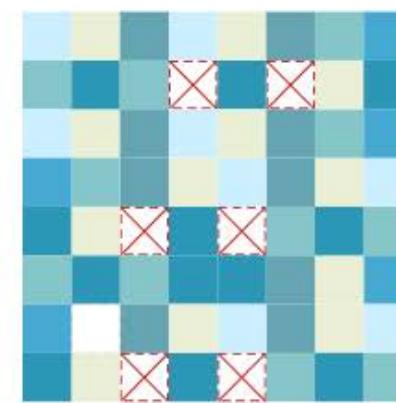


Let's edit data!



Dataset edition

Provides several ways to modify a dataset.



Missing values

Explore, remove or impute missing values on a dataset.



Normalize dataset

Correct technical biases and prepare your data for further analyses.

Normalize dataset

1. Select a dataset, choose a normalization type
and set options

Select a dataset

PICLET_qPCR1P218_Imputed_2 ▾

Choose normalization type

Correct batch effect

Log-transformation

Centering and scaling to unit variance

Correct batch effect

Quantile normalization

Run

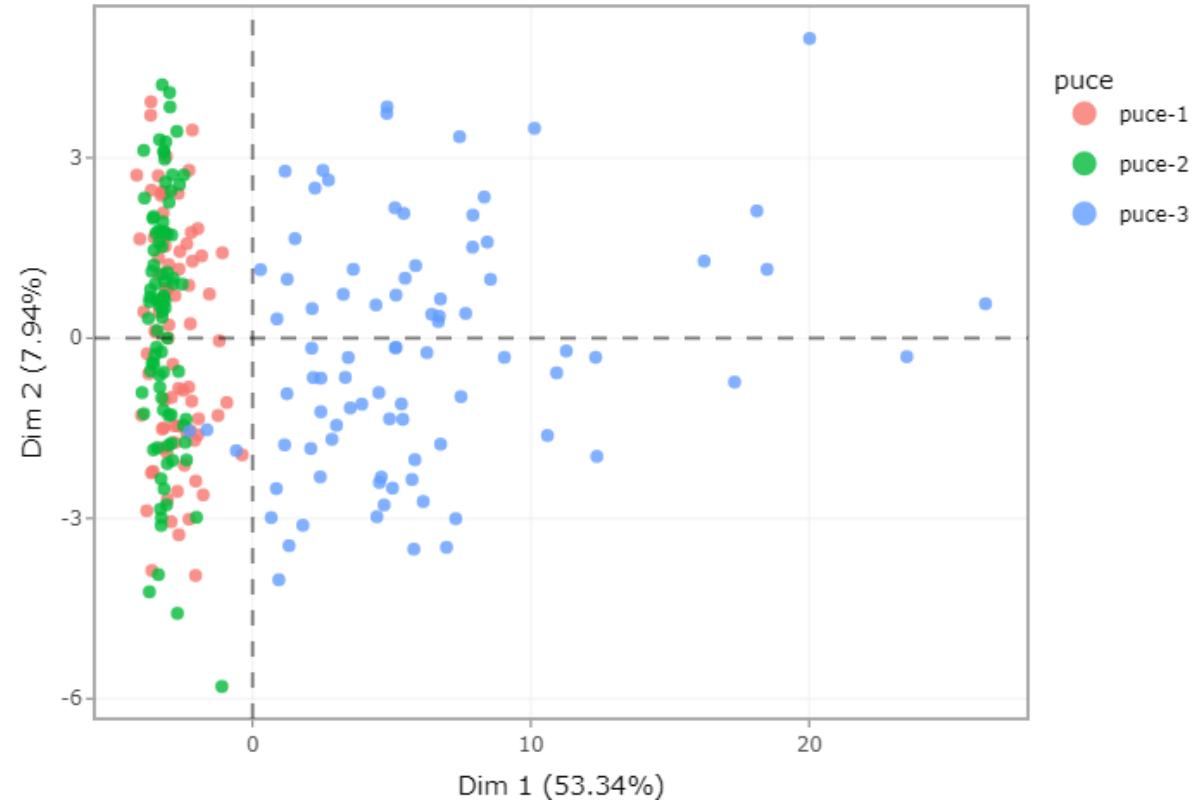
2. Dataset

Purpose of Normalize dataset:

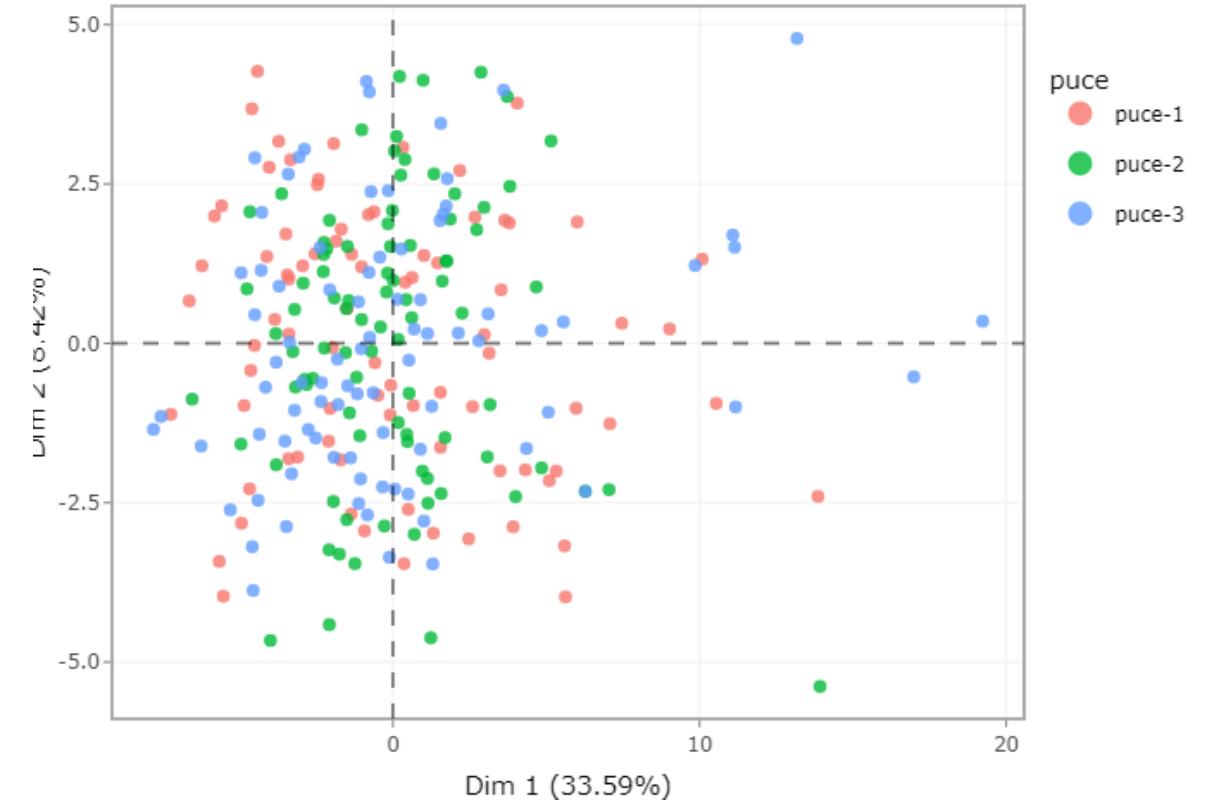
Correct technical biases and prepare your datasets for further analyses

- Basic transformations (log, ...).
- Normalizations specific of certain data types (RNA-seq, ...).
- Normalization for the correction of an explicit batch effect.
- All normalization methods come with diagnostic plots.

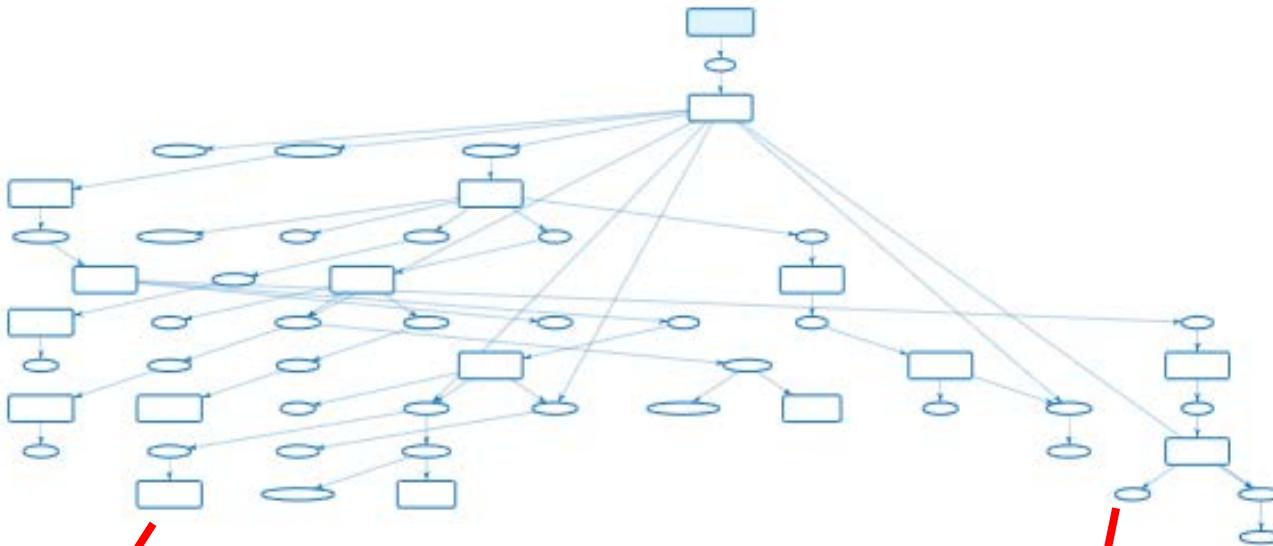
PCA on Data_qPCR_Piclet_e_Imputed_1: projection of individuals on the F



PCA on Data_qPCR_Piclet_e_Imputed_1_normalized_1: projection of ind



Workflow



All datasets

<input type="checkbox"/>	Name	Type	# Rows	# Columns	Log	Normalized	
<input type="checkbox"/>	PICLET_qPCR1P21...	generic	257	45	no	no	
<input type="checkbox"/>	PICLET_qPCR1P21...	generic	257	25	nc	nc	
<input type="checkbox"/>	PICLET_qPCR1P21...	generic	257	45	nc	nc	
<input type="checkbox"/>	PICLET_qPCR1P21...	generic	257	25	nc	nc	

Remove

Add

All analyses

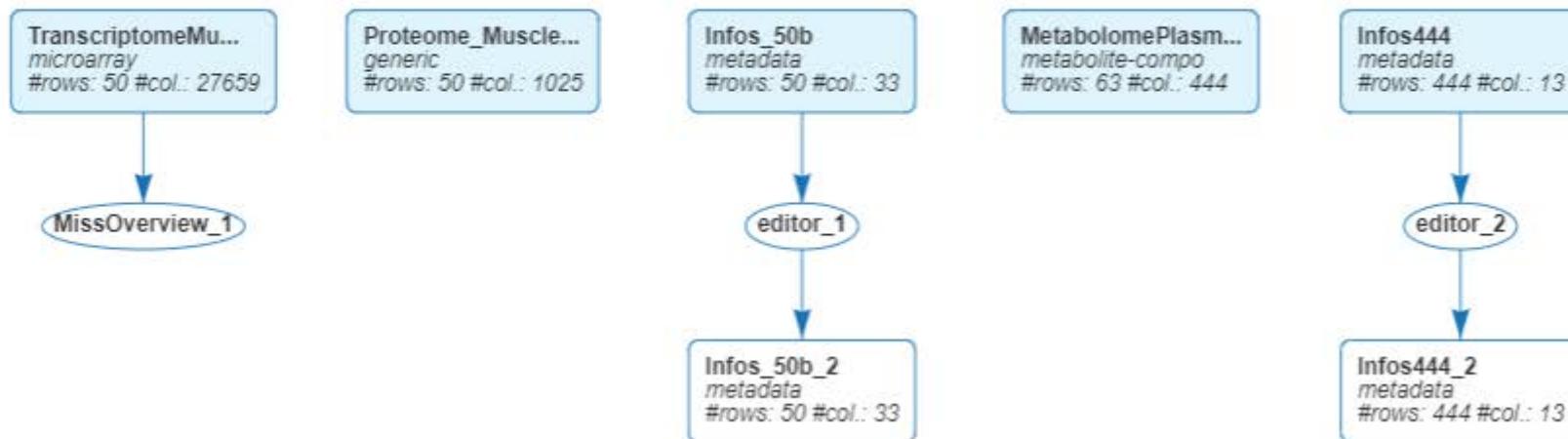
<input type="checkbox"/>	Name	Function
<input type="checkbox"/>	editor_1	r_edit_dataset
<input type="checkbox"/>	UnivariateDatas...	r_univariate_dataset
<input type="checkbox"/>	MissOverview_1	r_missing_overview
<input type="checkbox"/>	MissShowRemove_...	r_missing_showremove
<input type="checkbox"/>	MissShowRemove_...	r_missing_showremove
<input type="checkbox"/>	PCAobj_1	r_famfd
<input type="checkbox"/>	imputeMissing_1	r_missing_impute
<input type="checkbox"/>	imputeMissing_2	r_missing_impute

Remove

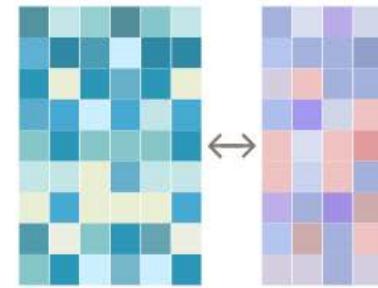
Export report

?

• Workflow

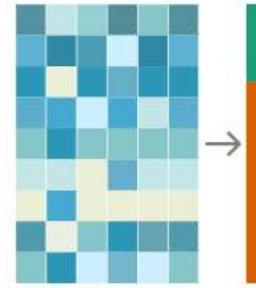


Let's integrate data!



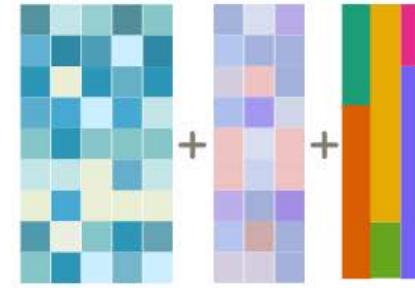
Integrate two datasets with PLS

Performs Partial Least Squares analysis on two datasets.



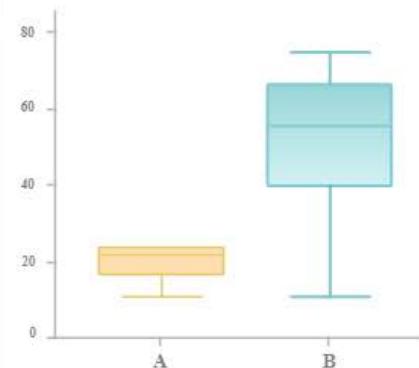
Integrate two datasets with PLS-DA

Performs Partial Least Squares analysis on two datasets.



Integrate datasets with MFA

Perform Multiple Factor Analysis on several datasets.



Differential analysis

Perform differential analysis for all numeric variables of a dataset.



Integrate two datasets with PLS

Preprocessing

Run PLS

Explore individuals

Explore variables

Extract new data

1. Select the datasets to integrate



Datasets

Proteome_Muscle_50 Infos_50_edited

Integrate

2. Plots and summaries

General information

Venn diagram

Upset plot

Description of datasets before filtering common individuals (rows)

Name	# rows	# col.	# missing	% missing	# numeric	# cat.	# logic	# others
Infos_50_edited	50	32	0	0	26	6	0	0
Proteome_Muscle...	50	1025	0	0	1025	0	0	0

Description of datasets after filtering common individuals (rows)

Name	# rows	# col.	# missing	% missing	# numeric	# cat.	# logic	# others
Infos_50_edited	50	32	0	0	26	6	0	0
Proteome_Muscle...	50	1025	0	0	1025	0	0	0

Integrate two datasets with PLS

Preprocessing

Run PLS

Explore individuals

Explore variables

Extract new data

1. Display individuals on components



Component to display on X (horizontal)

1



Component to display on Y (vertical)

2



Color



Info_50b_e

TG_F



Shape



Info_50b_e

Age



Size



Info_50b_e

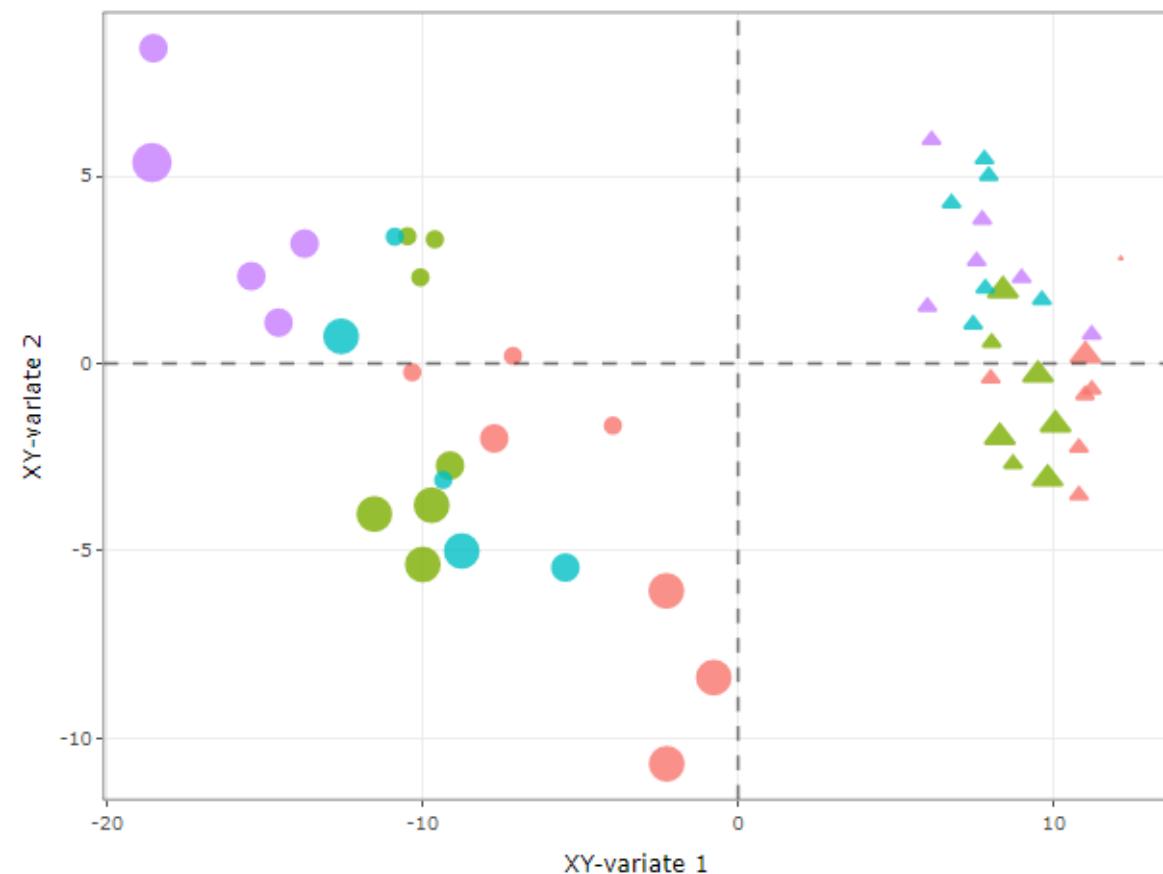
Weight



Plot individuals

2. Plot individuals on components

PLS on Info_50b_e, Proteome_Muscle_50: XY-space



- (TG_F, Age)
- (LWLW, 110d)
 - △ (LWLW, 90d)
 - (LWMS, 110d)
 - ▲ (LWMS, 90d)
 - (MSLW, 110d)
 - △ (MSLW, 90d)
 - (MSMS, 110d)
 - ▲ (MSMS, 90d)

Add to report

Integrate two datasets with PLS

Preprocessing

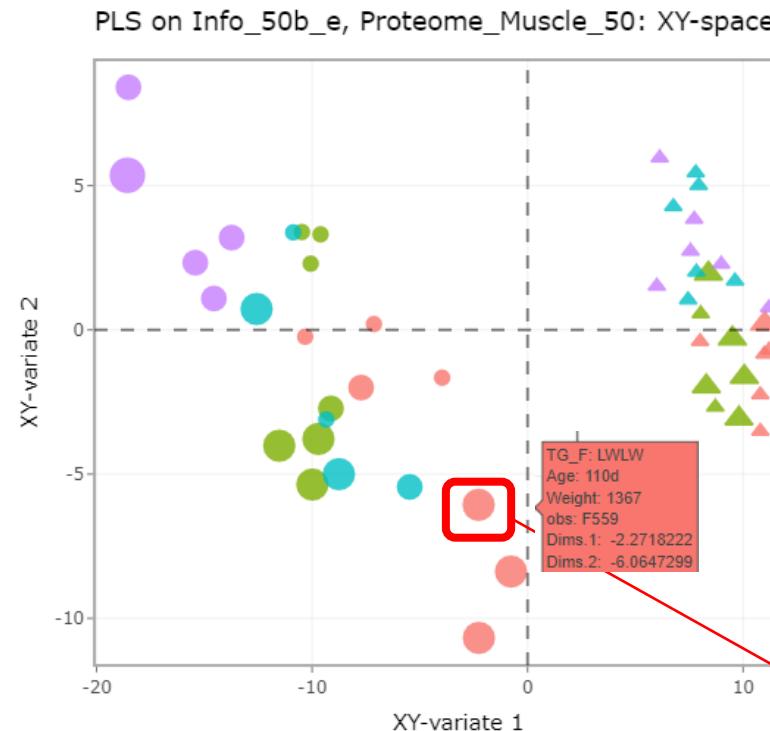
Run PLS

Explore individuals

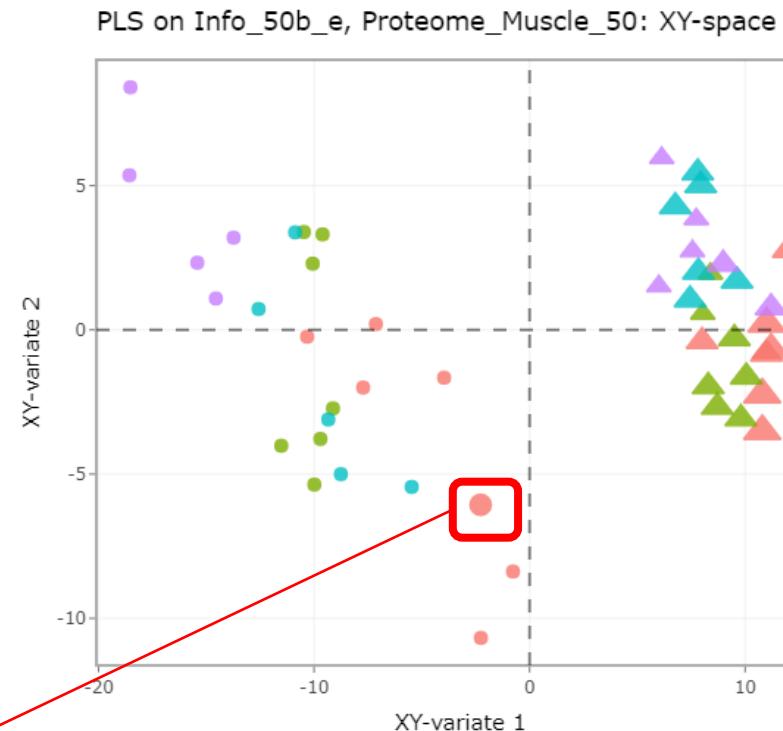
Explore variables

Extract new data

Size = weight



Size = MHC_E



Heavy but developmental delay

Integrate two datasets with PLS

Preprocessing

Run PLS

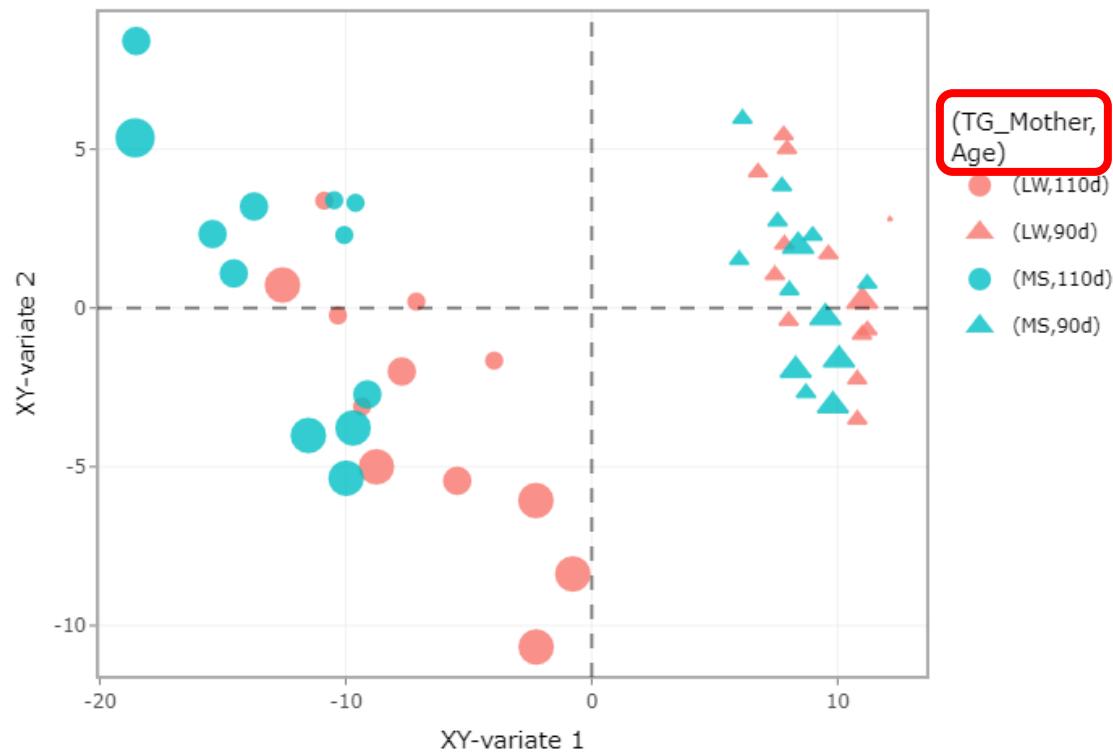
Explore individuals

Explore variables

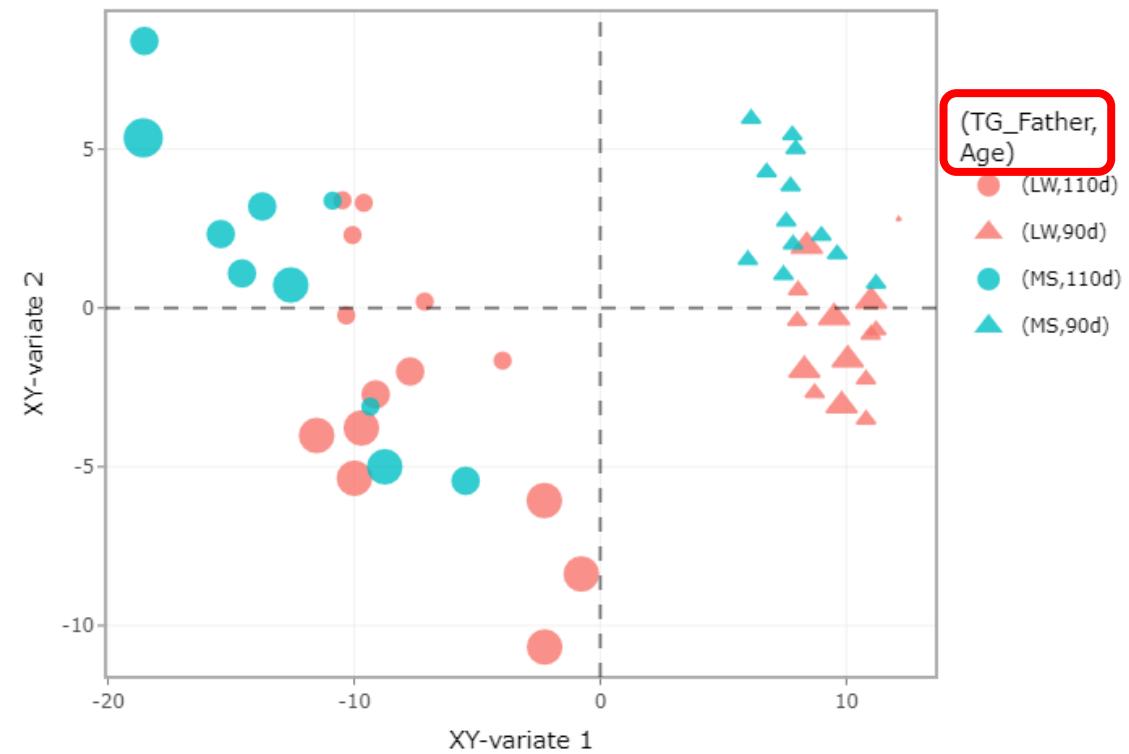
Extract new data

Size = weight

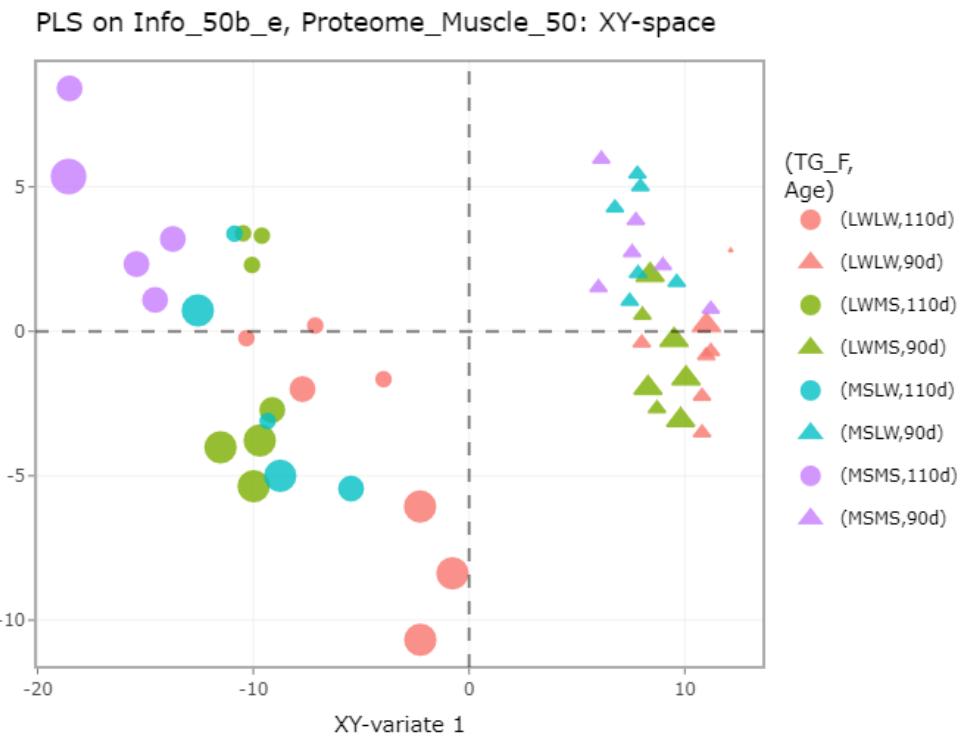
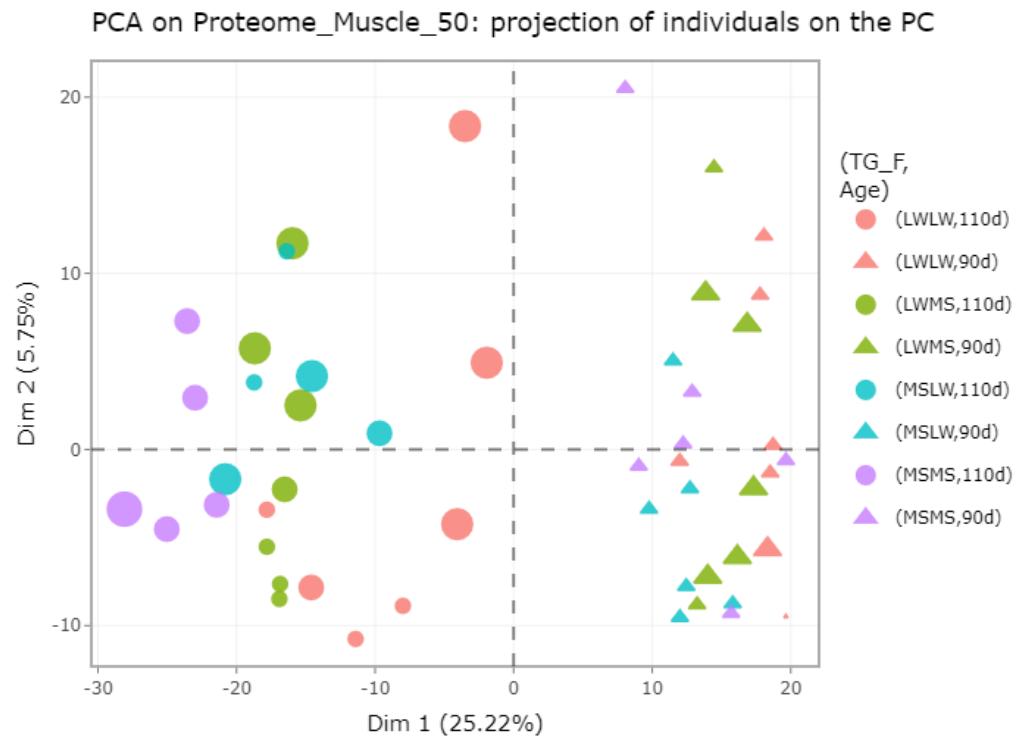
PLS on Info_50b_e, Proteome_Muscle_50: XY-space



PLS on Info_50b_e, Proteome_Muscle_50: XY-space



PCA vs PLS



Size = weight



Integrate two datasets with PLS

Preprocessing

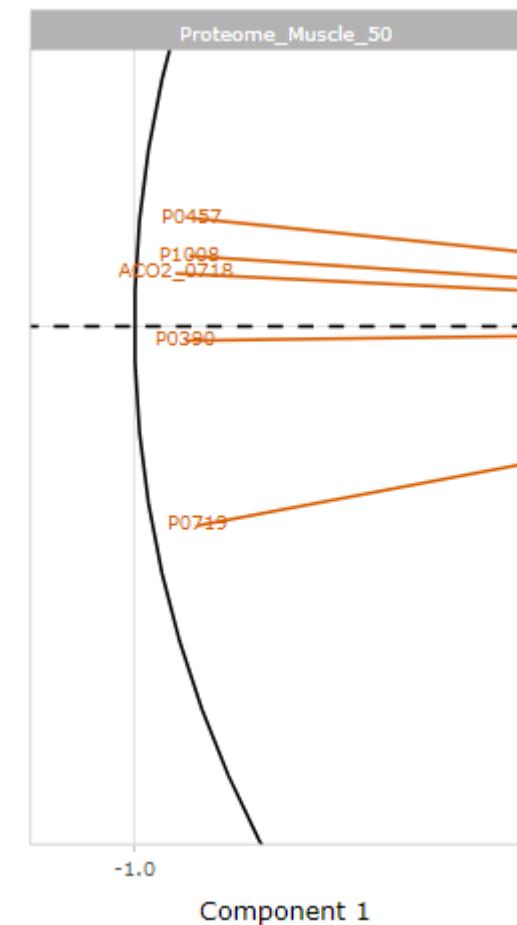
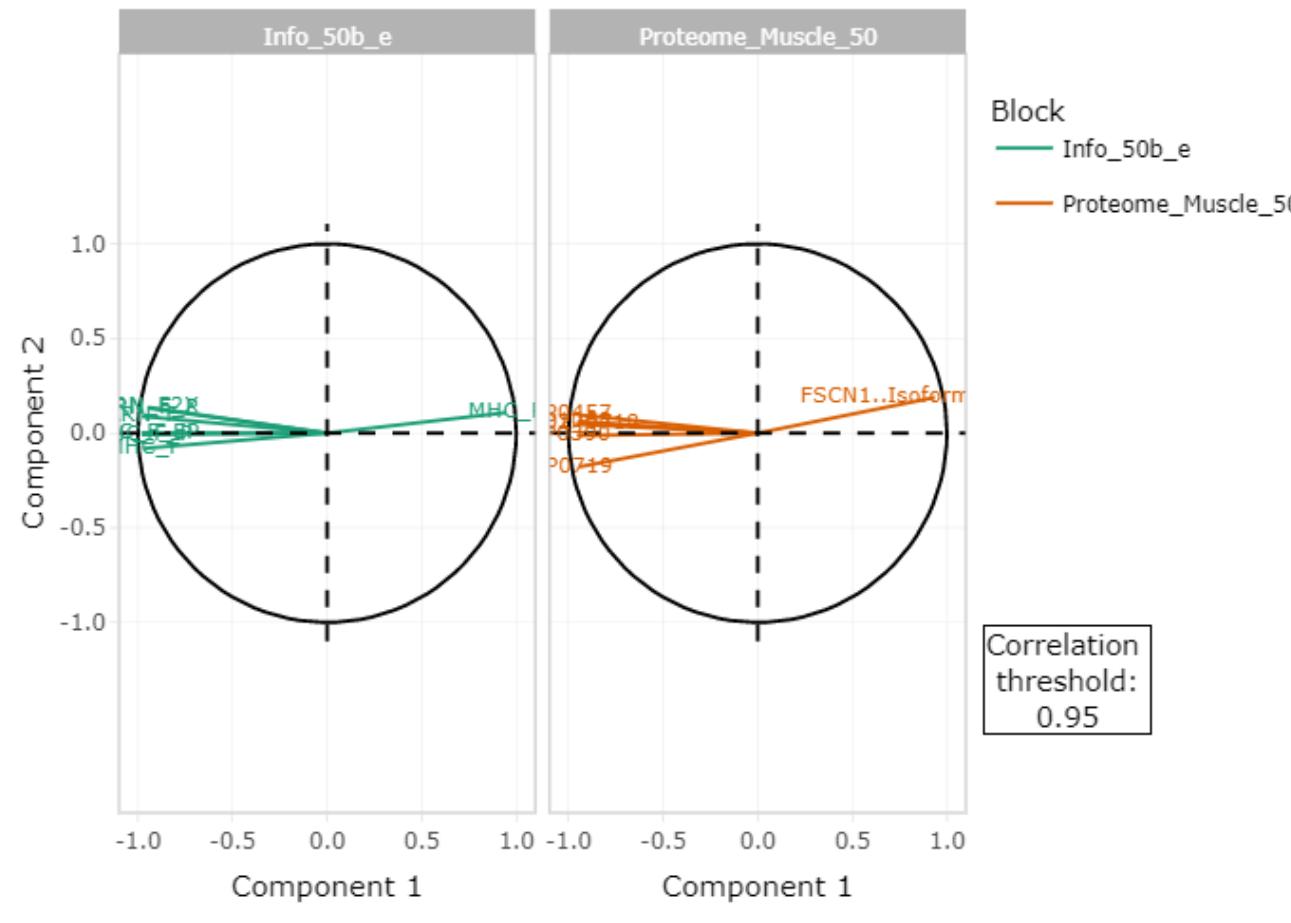
Run PLS

Explore individuals

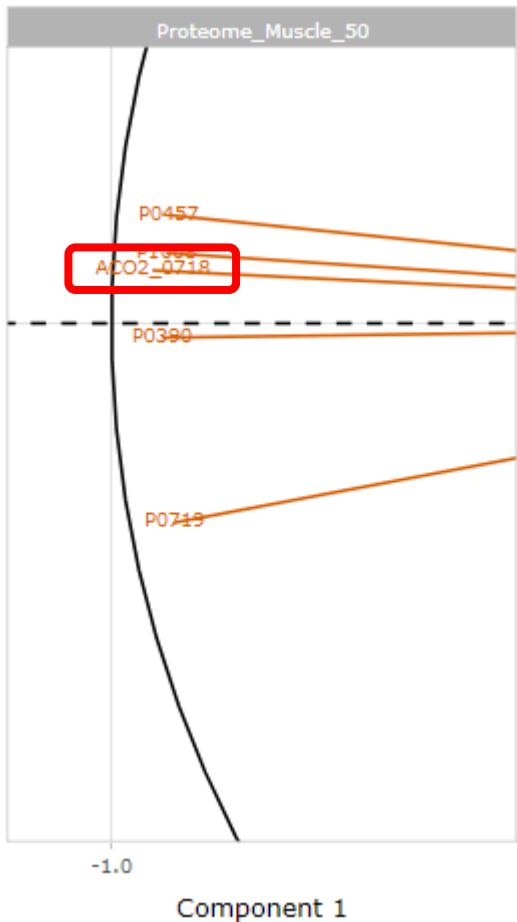
Explore variables

Extract new data

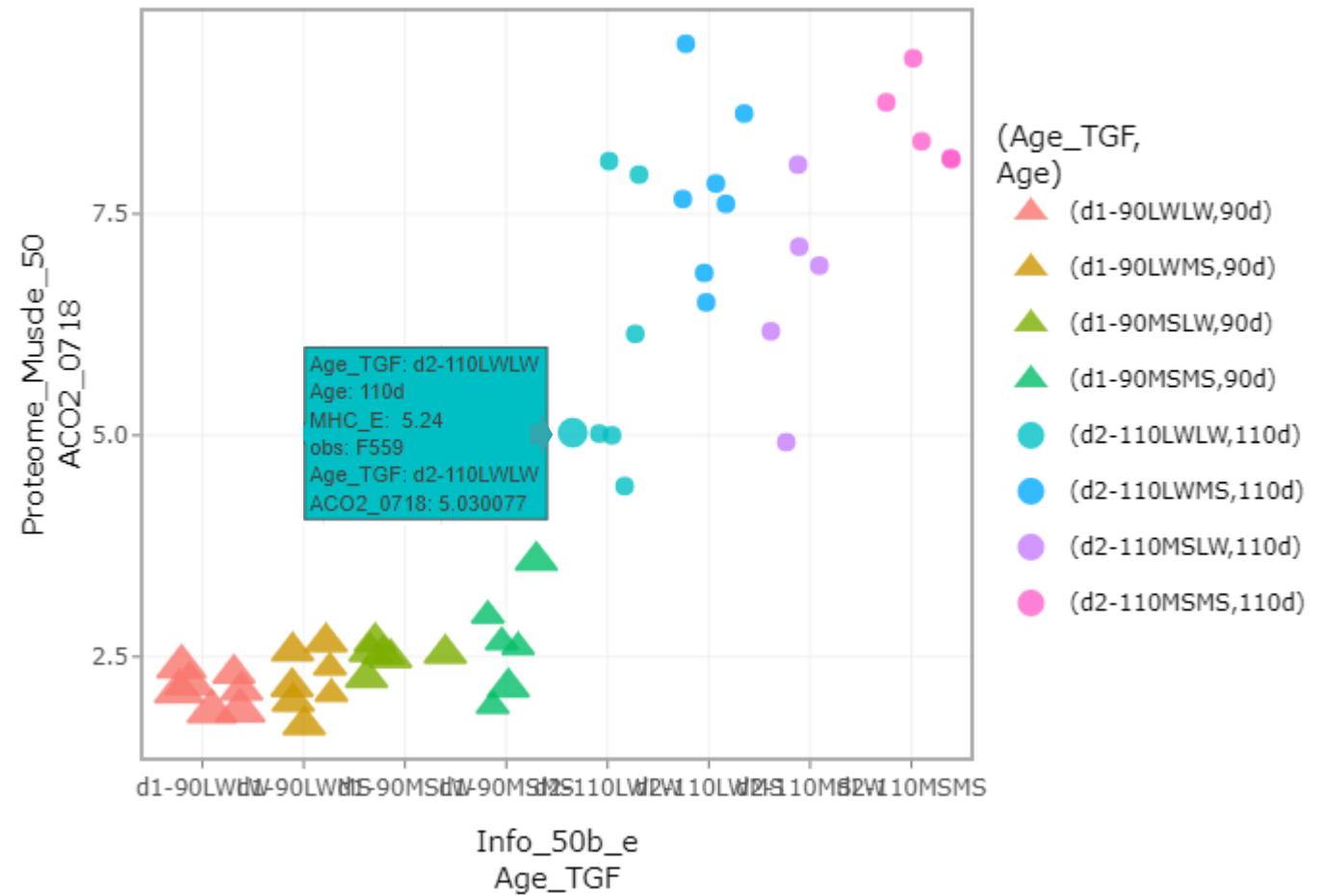
PLS on Info_50b_e, Proteome_Muscle_50: correlations of variables with



Explore variables in a dataset



Size = MHC_E





Integrate two datasets with PLS

Preprocessing

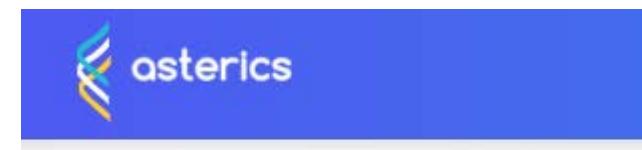
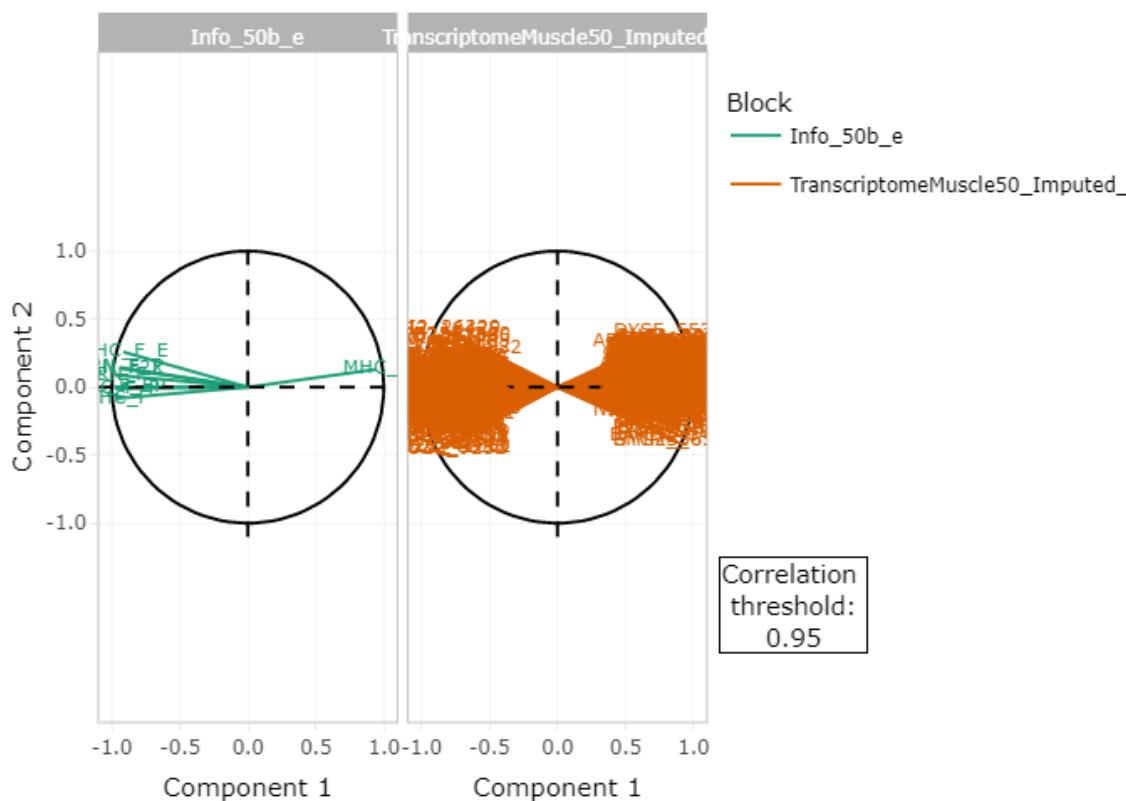
Run PLS

Explore individuals

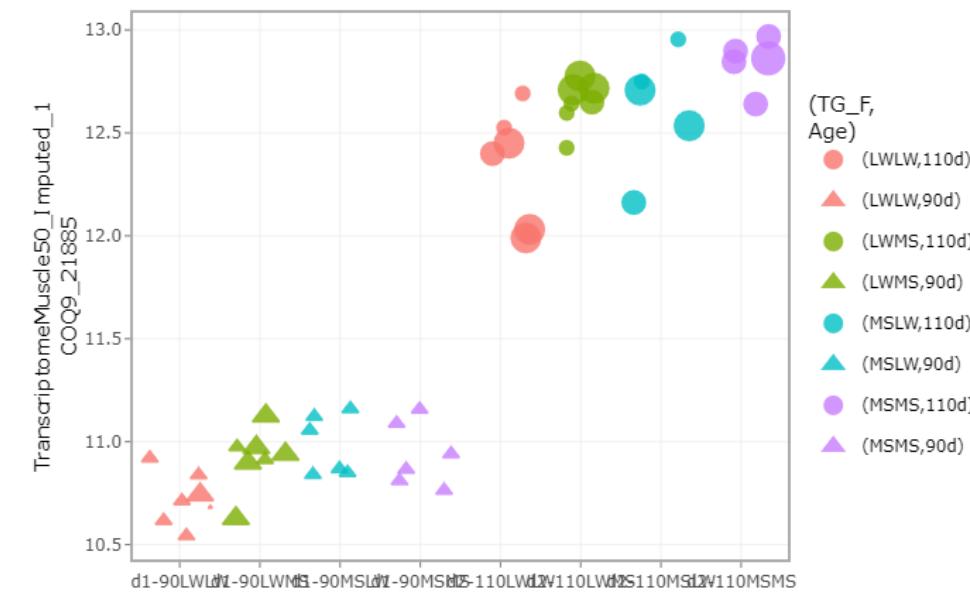
Explore variables

Extract new data

PLS on Info_50b_e, TranscriptomeMuscle50_Imputed_1: correlations of

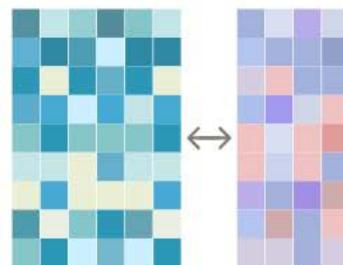


Explore variables in a dataset



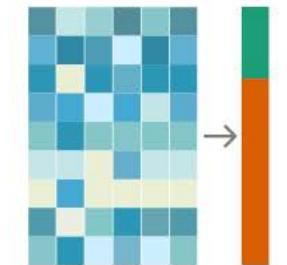
Size = weight

Let's integrate data!



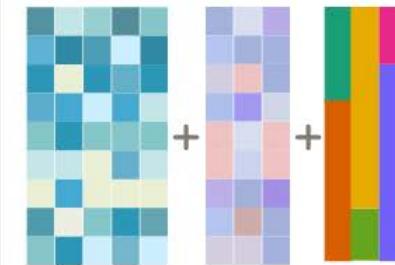
Integrate two datasets with PLS

Performs Partial Least Squares analysis on two datasets.



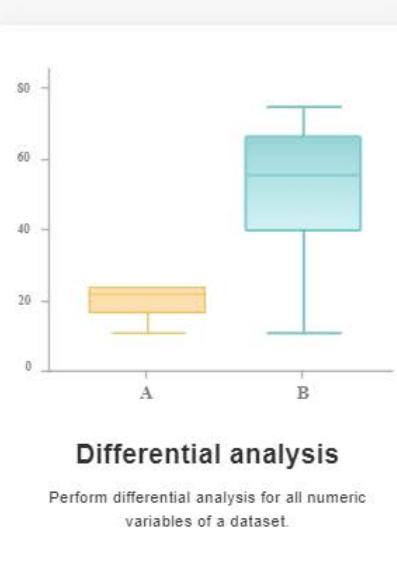
Integrate two datasets with PLS-DA

Performs Partial Least Squares analysis on two datasets.



Integrate datasets with MFA

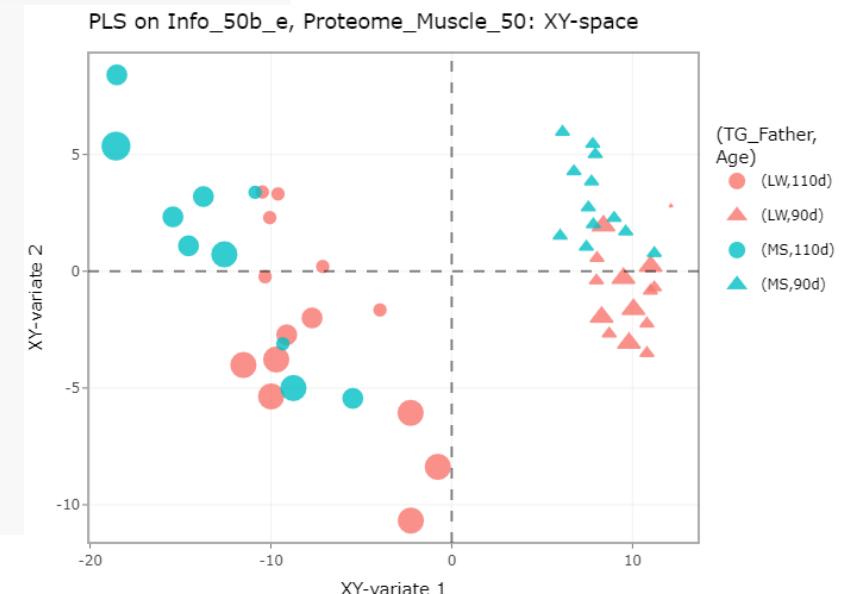
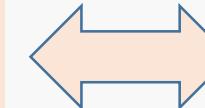
Perform Multiple Factor Analysis on several datasets.



Differential analysis

Perform differential analysis for all numeric variables of a dataset.

It is easy to discriminate the age but what about the genotype?



(TG_Father, Age)
● (LW,110d)
▲ (LW,90d)
● (MS,110d)
▲ (MS,90d)



Integrate two datasets with PLS-DA

Preprocessing

Run PLS-DA

Explore individuals

Explore variables

Extract new data

1. Run PLS-DA

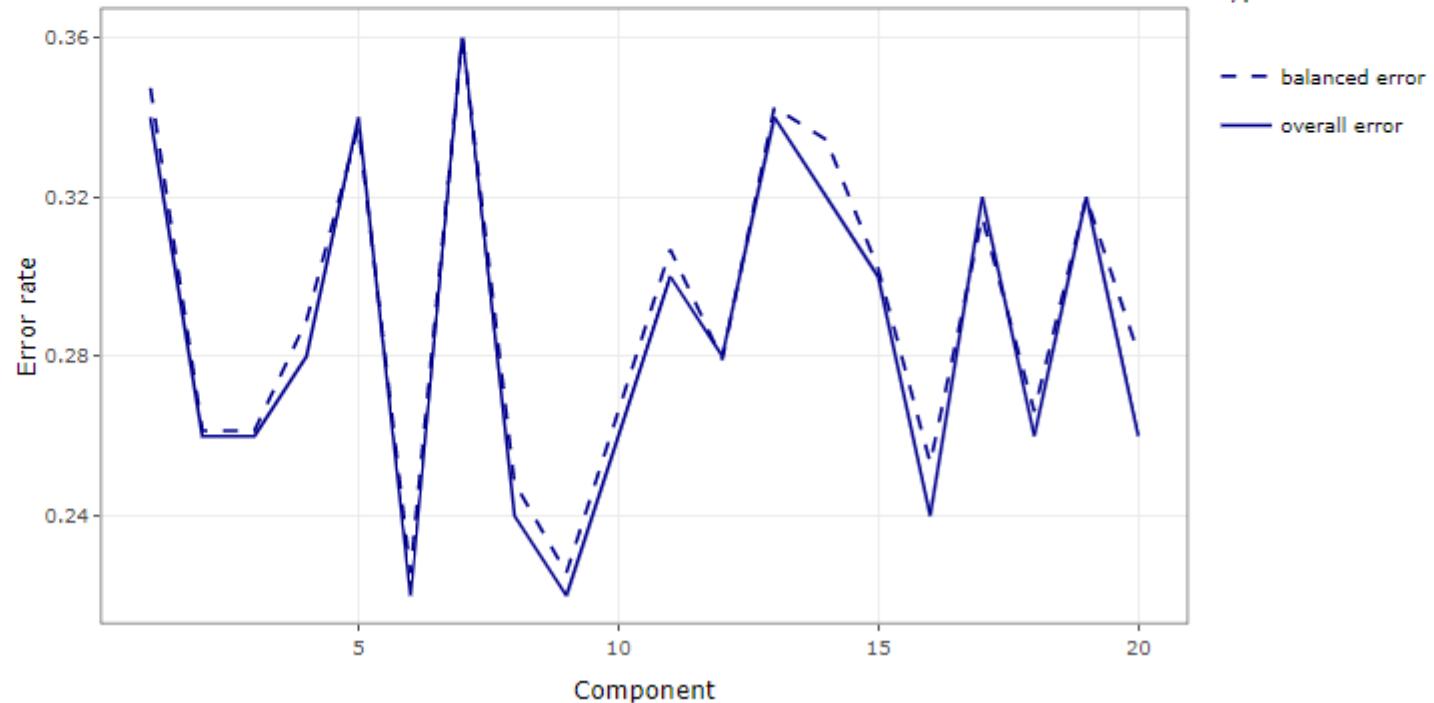


Run

TG_Father
2 levels

2. Plots

Proteome_Muscle_50 predicts Info_50b_e: Classification error rate

Type
 

Integrate two datasets with PLS-DA

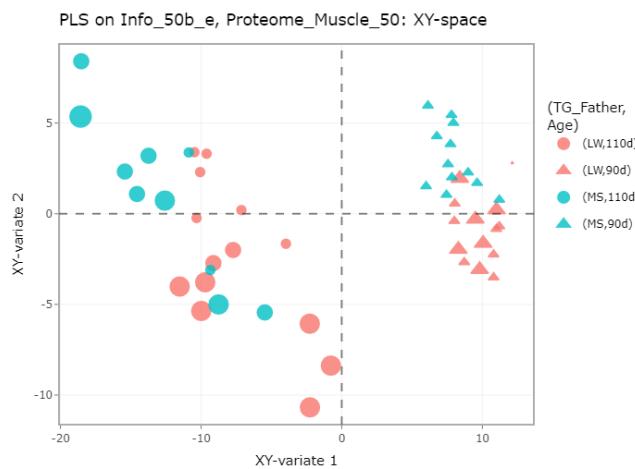
Preprocessing

Run PLS-DA

Explore individuals

Explore variables

Extract new data



Integrate two datasets with PLS-DA

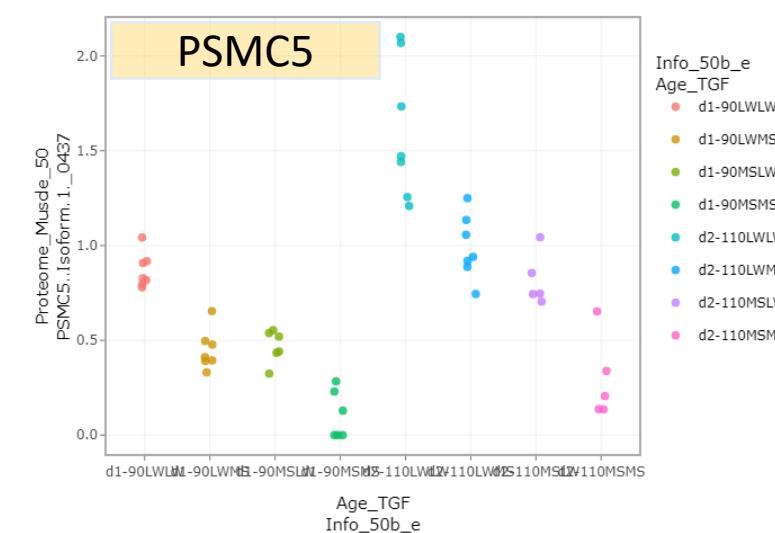
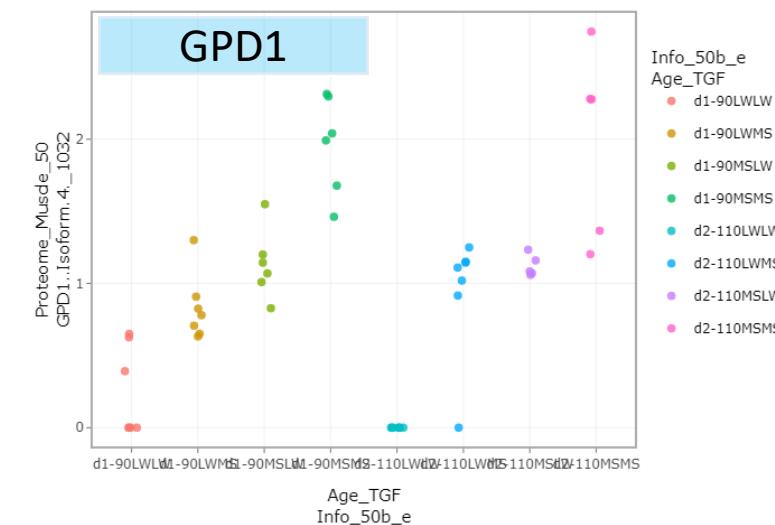
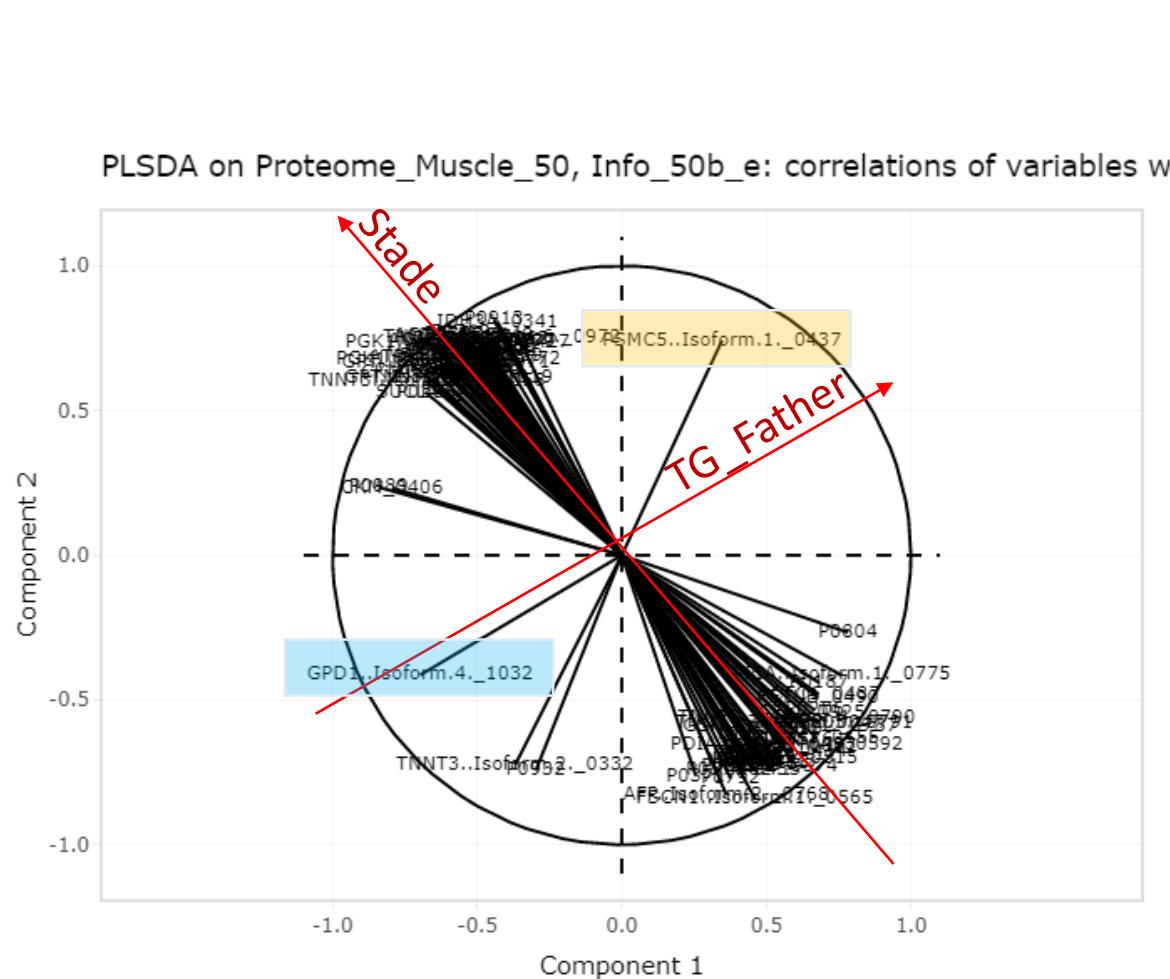
Preprocessing

Run PLS-DA

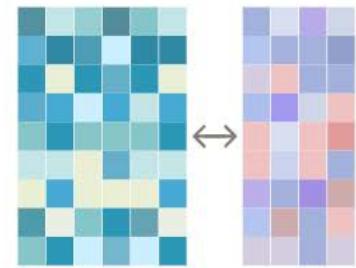
Explore individuals

Explore variables

Extract new data

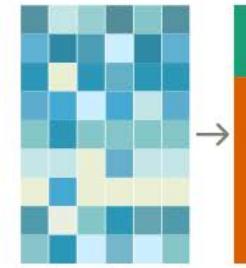


Let's integrate data!



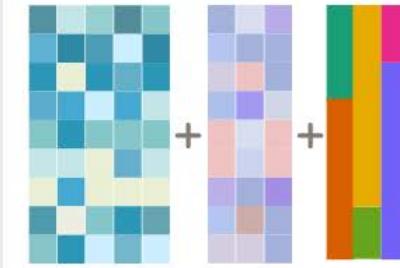
Integrate two datasets with PLS

Performs Partial Least Squares analysis on two datasets.



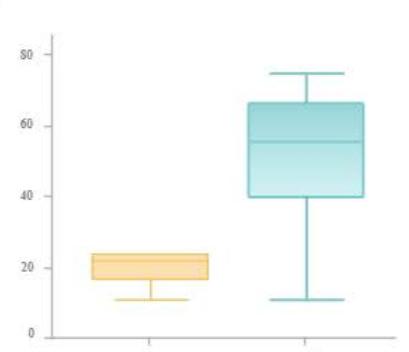
Integrate two datasets with PLS-DA

Performs Partial Least Squares analysis on two datasets.



Integrate datasets with MFA

Perform Multiple Factor Analysis on several datasets.



Differential analysis

Perform differential analysis for all numeric variables of a dataset.



Differential analysis

Preprocessing

Multiple tests

Posthoc tests

Extract dataset

1. Run multiple tests



Type

 parametric non-parametric automatic

Correction

BH

Threshold

0.05

Run

2. Explore results of multiple tests



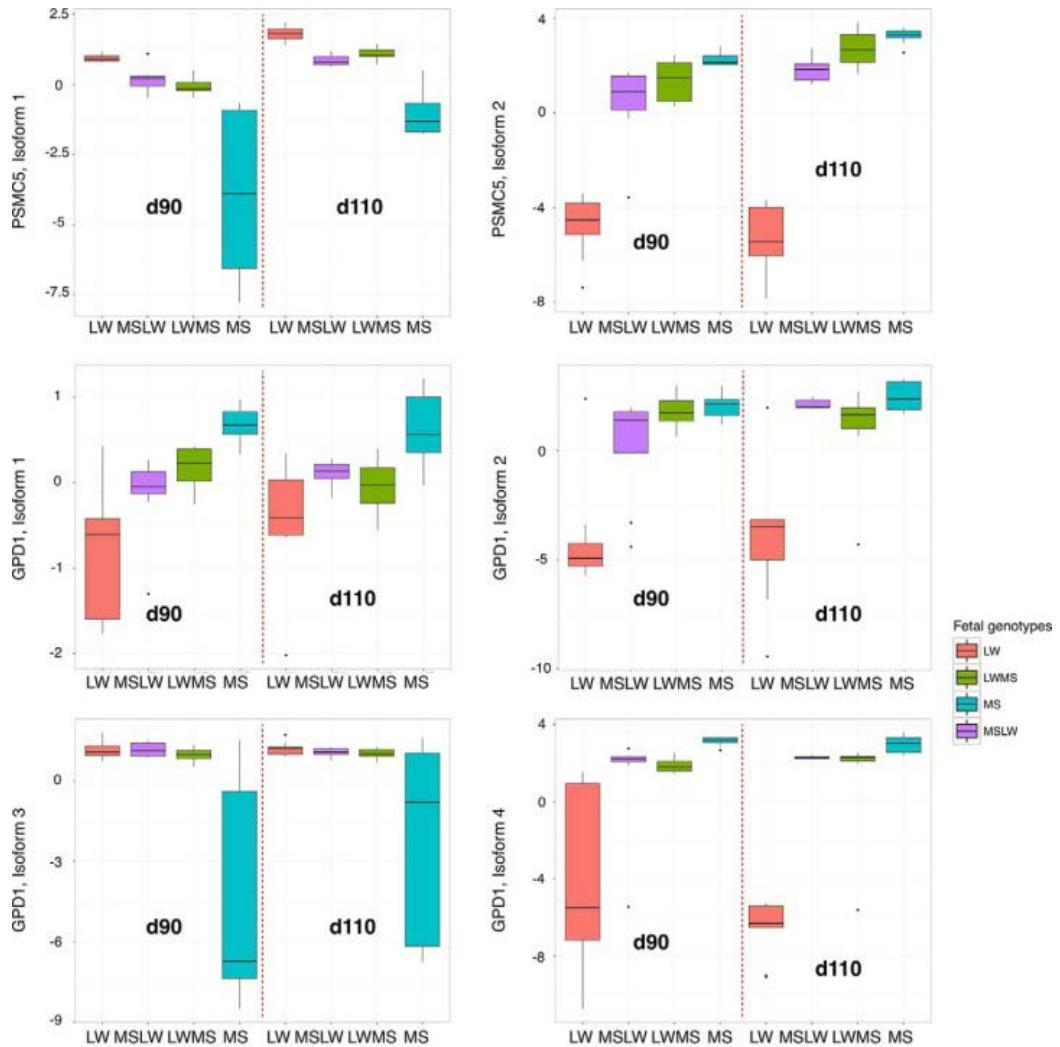
Plots



Summary

Test results (Kruskal-Wallis test, corrected with 'BH') for Proteome_Muscle_50 vs Info_50b_e TG_Father

Variables	Test stats.	p-values	Adj. p-values	Signs
28 proteins BH<0.05				
GPD1..Isoform.4._1032	26.5851	2.5218e-7	0.0003	1
LXN_0198	20.745	0.000005247	0.0021	-1
P0987	20.4733	0.0000060468	0.0021	-1
TNNT3..Isoform.3._0355	18.995	0	0.0022	1
PSMC5..Isoform.1._0437	18.9987	0	0.0022	-1
P0988	19.4156	0	0.0022	1
TF_0852	18.3197	0	0.0027	-1



PSMC5,
Proteasome 26S Regulatory Subunit, ATP-dependent
degradation of ubiquitinated proteins, 5
2 isoforms

PGD1
Glycerol-3-phosphate dehydrogenase 1, cytoplasmic
4 isoforms



Integrate two datasets with PLS-DA

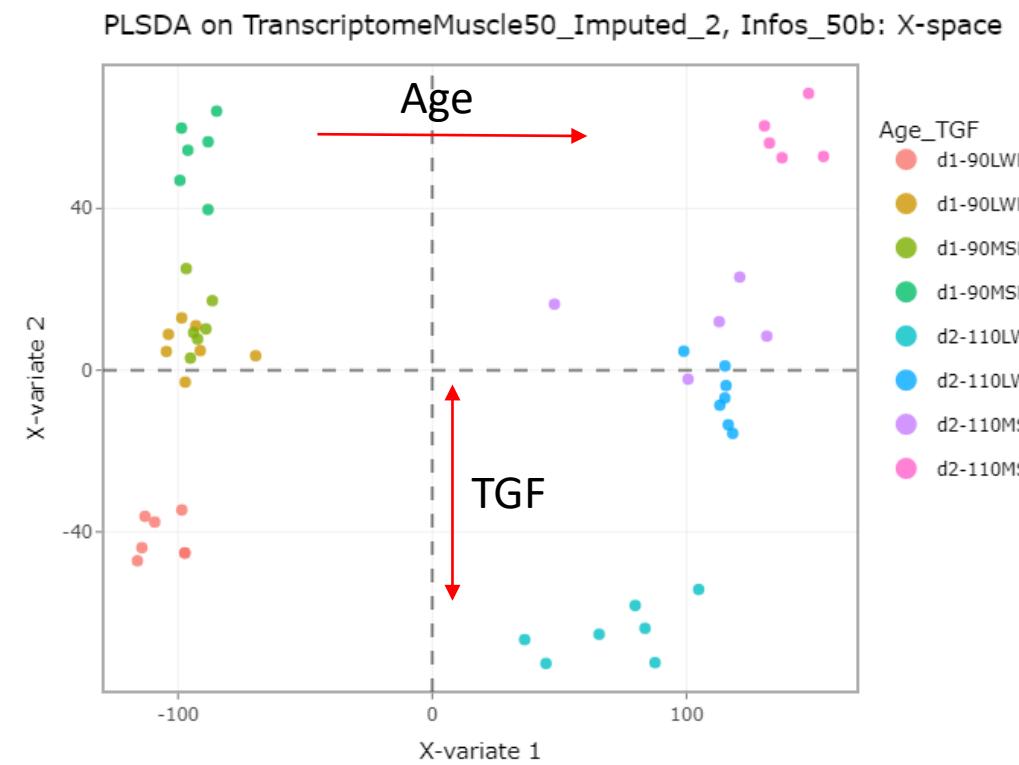
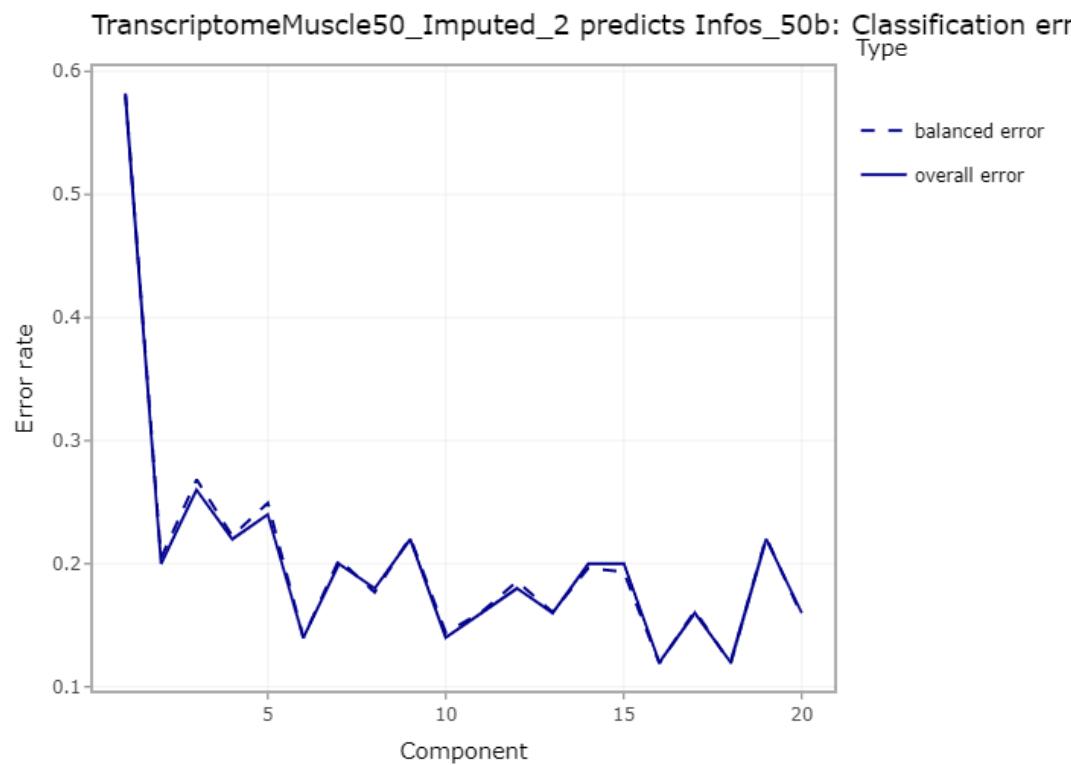
Preprocessing

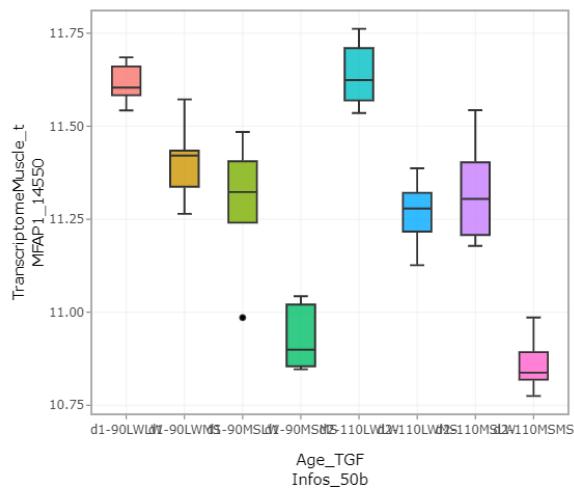
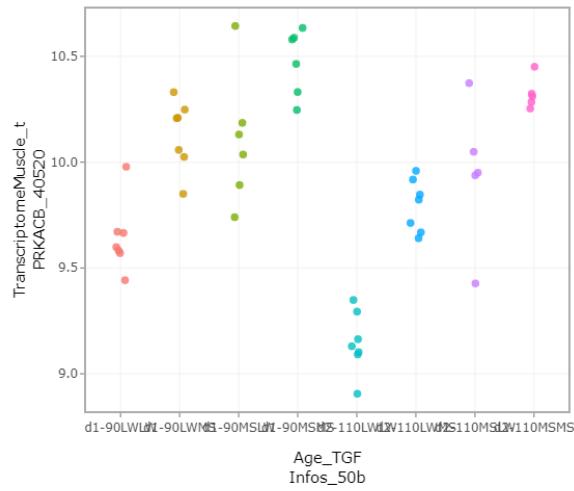
Run PLS-DA

Explore individuals

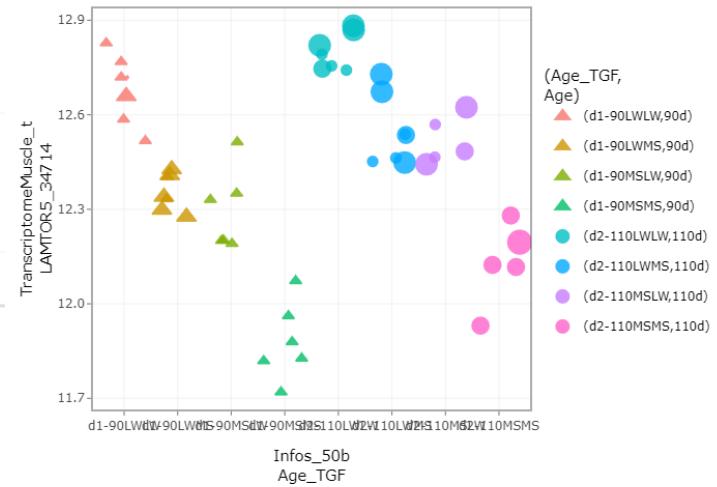
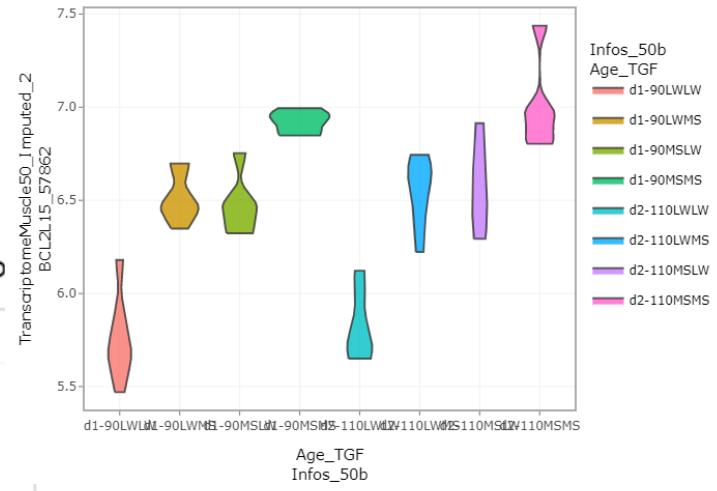
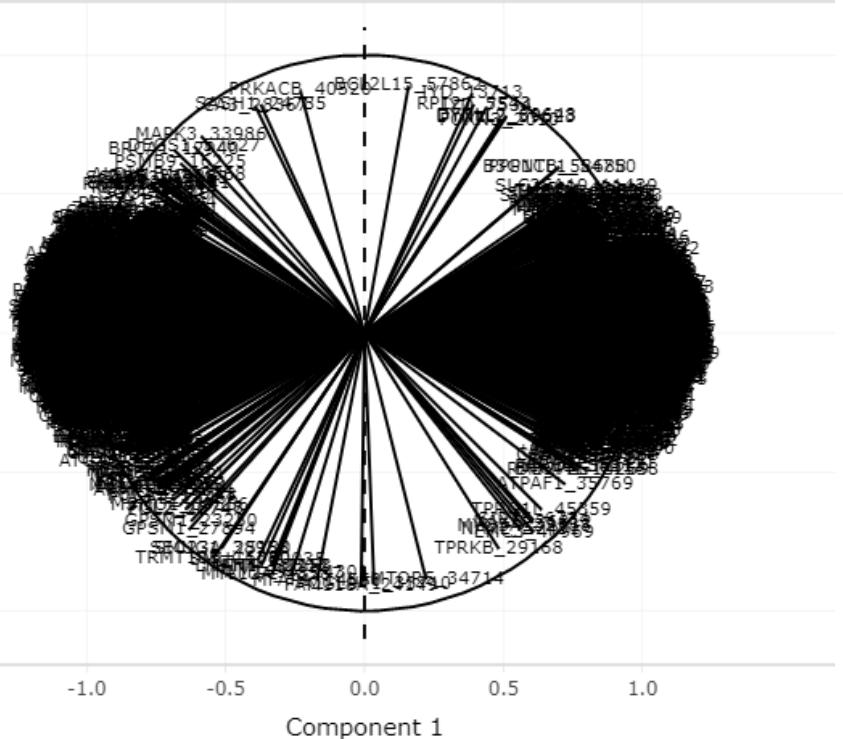
Explore variables

Extract new data

Age_TGF
8 levels



PCA on TranscriptomeMuscle50_Imputed_2, Infos_50b: correlation





Differential analysis

Preprocessing

Multiple tests

Posthoc tests

Extract dataset

1. Run multiple tests



Type

 parametric non-parametric automatic

Correction

BH

Threshold

0.05



2. Explore results of multiple tests



Plots



Heatmap



Summary

Test results (Kruskal-Wallis test, corrected with 'BH') for TranscriptomeMuscle_t vs Infos_50b: Age_TGF

Variables	Test stats.	p-values	Adj. p-values
A4GALT_23662	36.2485	0.000006507	0
AAAS_2894	34.6425	0	0
AAAS_18735	38.7512	0.0000021801	0
AAGAB_36620	41.0106	8.0581e-7	0
AARS2_40325	35.1548	0	0
ABCA3_15222	36.3064	0.0000063452	0
ARCA3_58959	34.623	0	0

Explore a dataset with PCA

Run PCA

Explore individuals

Explore variables

Extract new data

1. Display individuals on components



Component to display on X (horizontal)

1



Component to display on Y (vertical)

2



Color



Infos444_...

Age



Shape



Dataset

Select a variable

Size



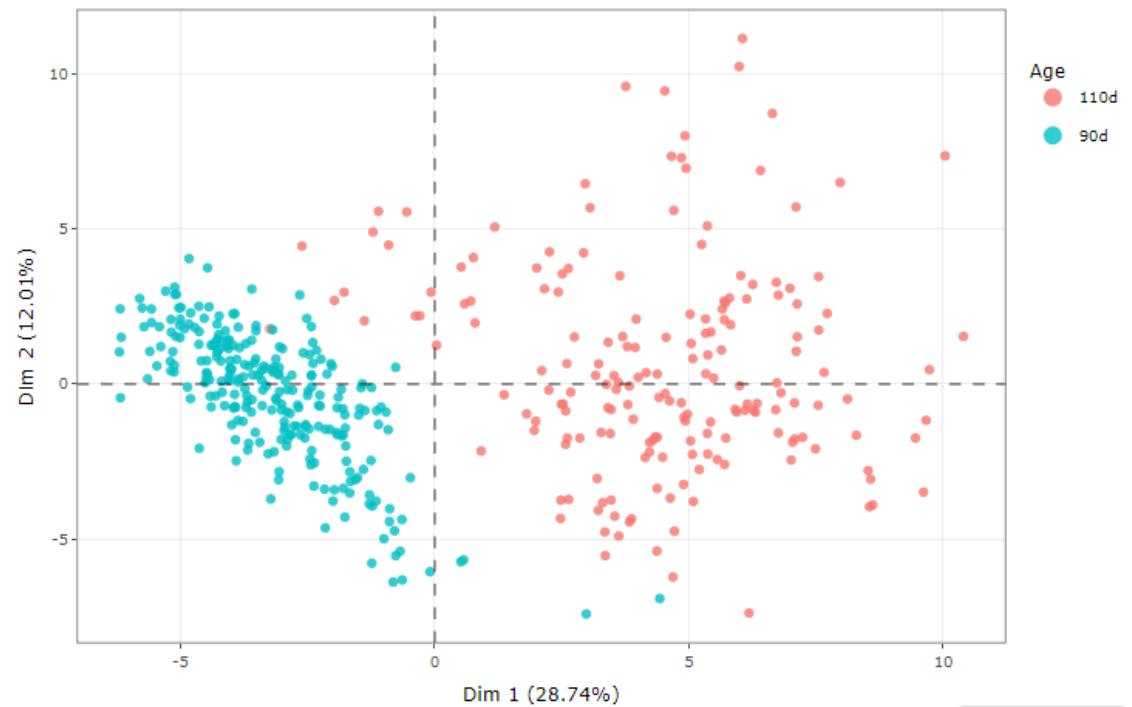
Tr Dataset

Select a variable

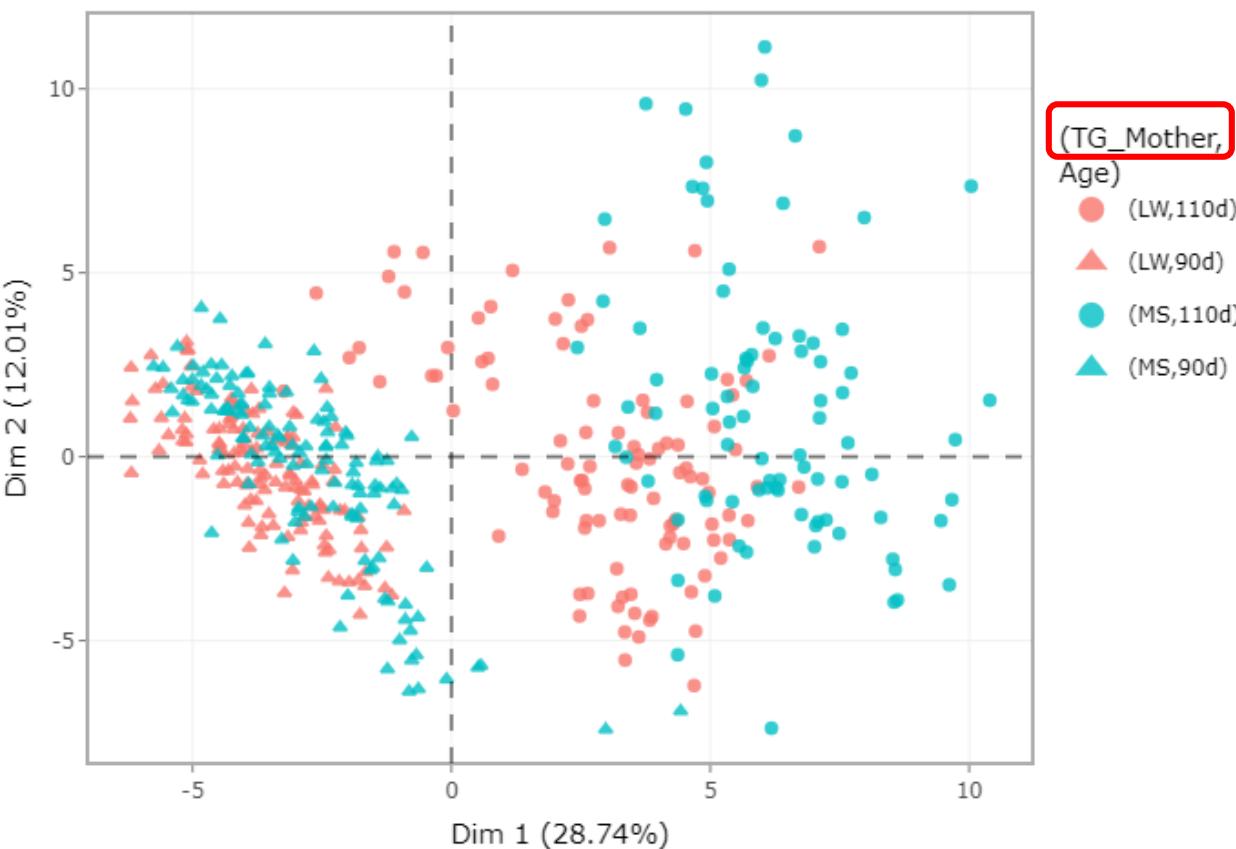
Plot individuals

2. Plot individuals of edited_3 on components

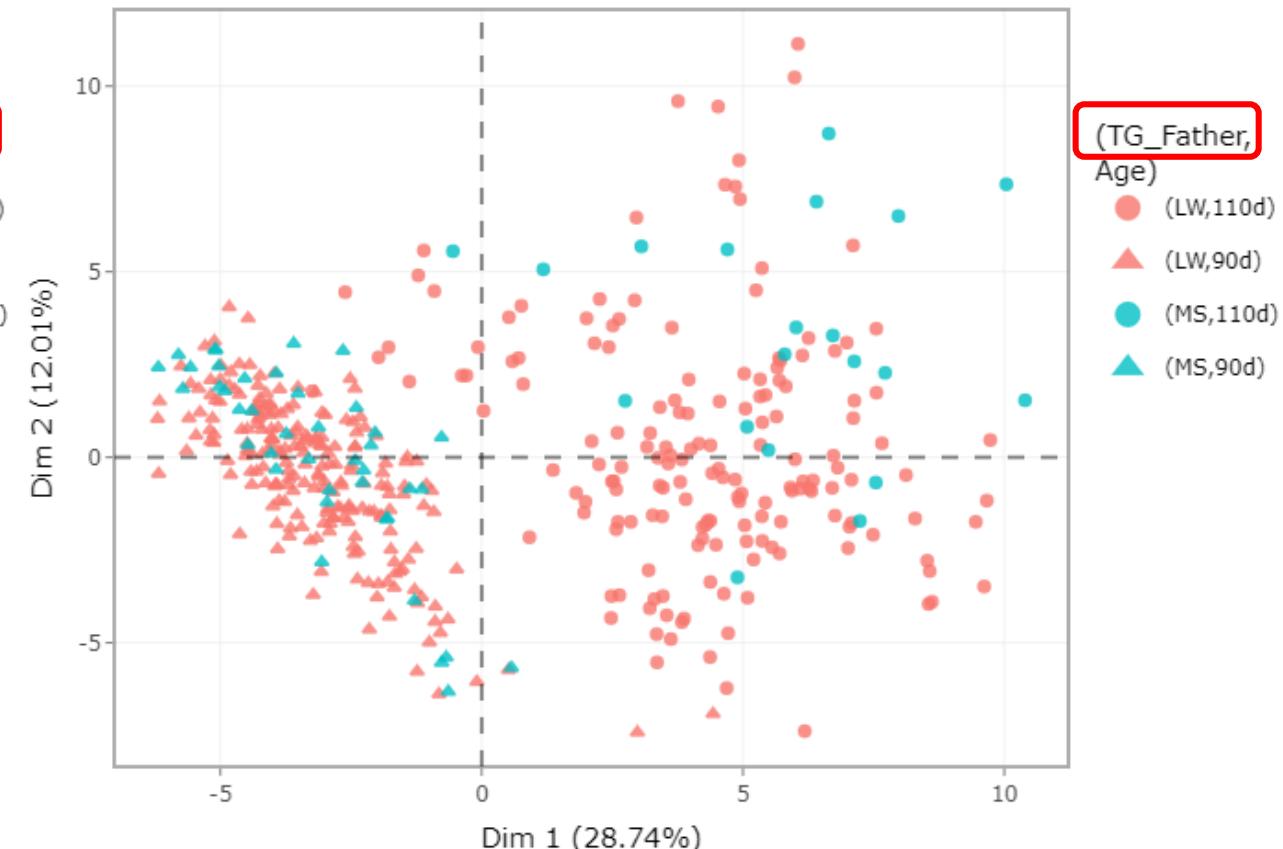
PCA on MetabolomePlasma444_t: projection of individuals on the PC



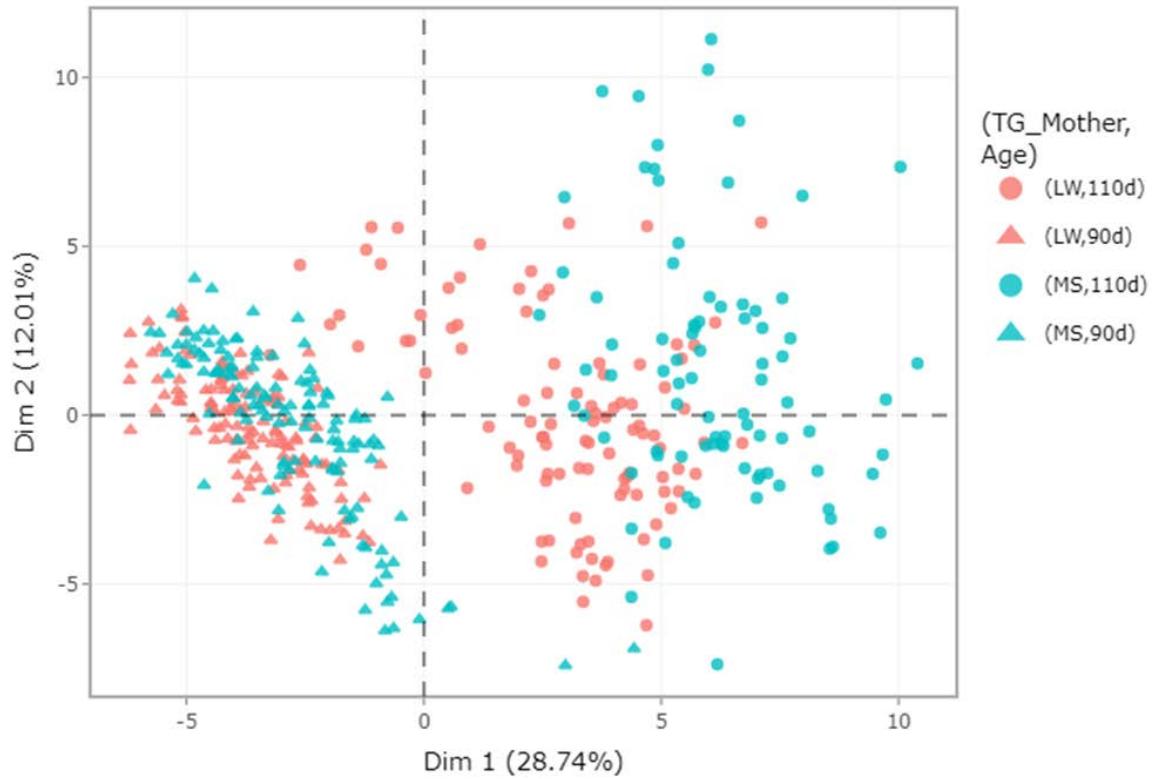
PCA on MetabolomePlasma444_t: projection of individuals on the PC



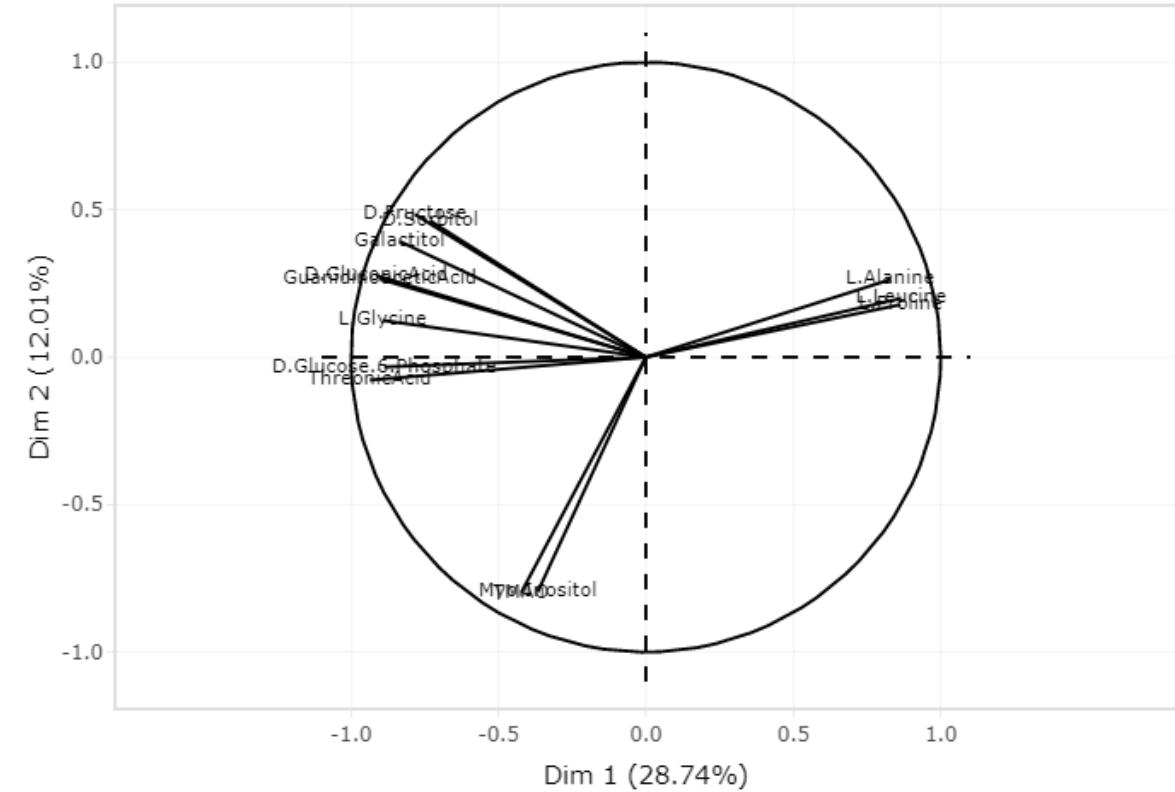
PCA on MetabolomePlasma444_t: projection of individuals on the PC



PCA on MetabolomePlasma444_t: projection of individuals on the PC

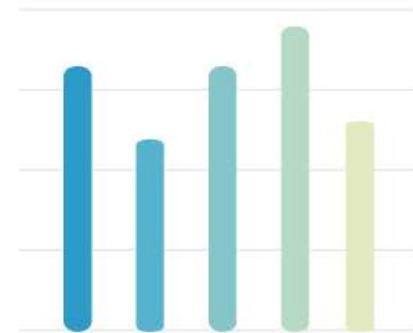


PCA on MetabolomePlasma444_t: correlations of variables with the PCs.
Correlation threshold: 0.85



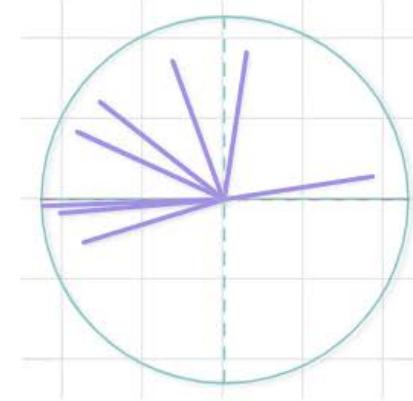


Let's explore a dataset!



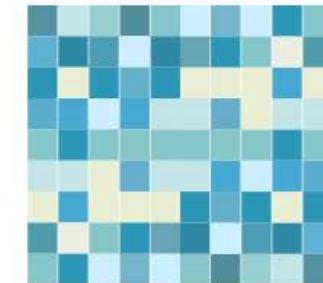
Explore variables in a dataset

Obtain numerical summaries and plots for a few variables.



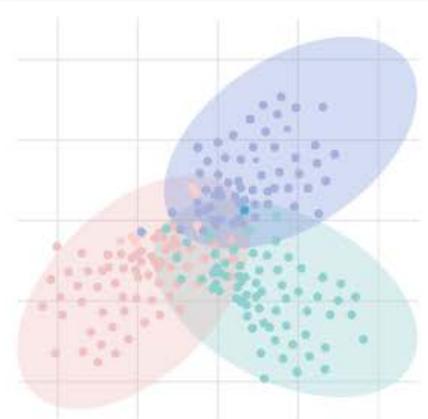
Explore a dataset with PCA

Perform Principal Component Analysis on a dataset.



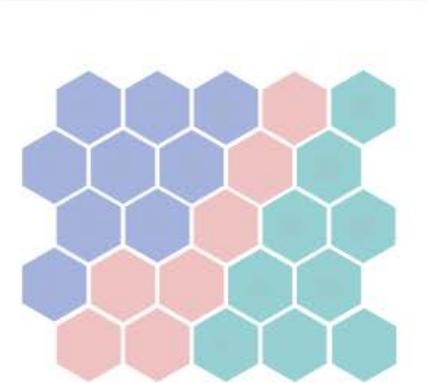
Explore a dataset with a heatmap

Obtain the heatmap of a dataset.



Clustering

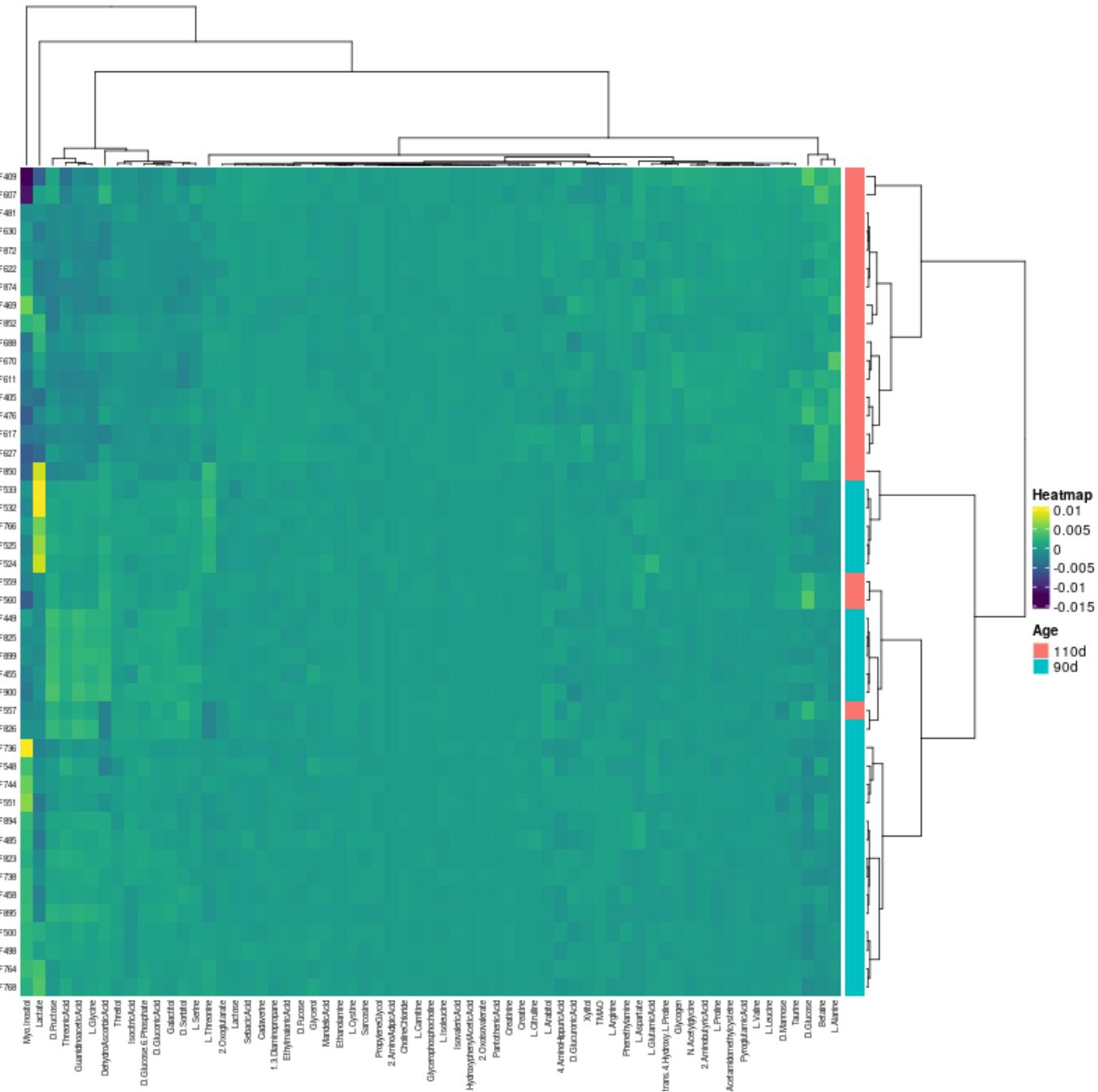
Cluster the individuals of a dataset.



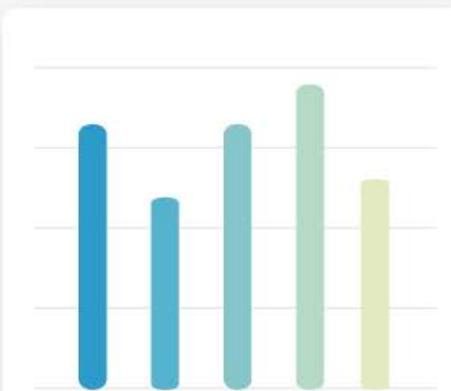
Self-Organizing Map

Use SOM as a clustering and visualization method.

Heatmap of dataset

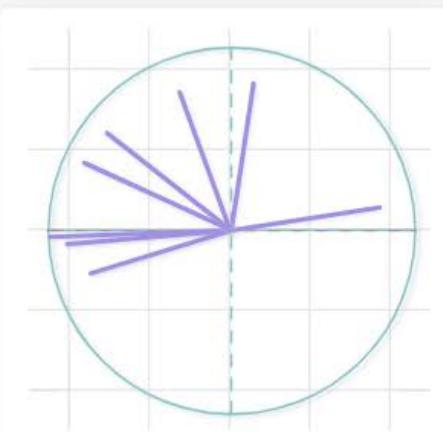


Let's explore a dataset!



Explore variables in a dataset

Obtain numerical summaries and plots for a few variables.



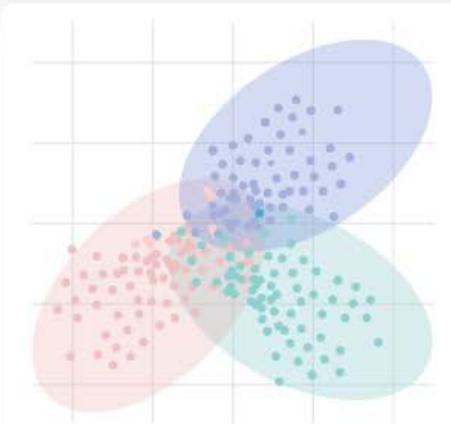
Explore a dataset with PCA

Perform Principal Component Analysis on a dataset.



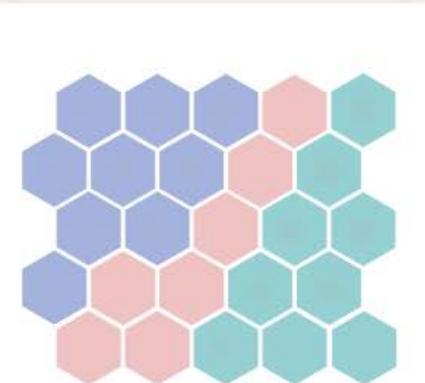
Explore a dataset with a heatmap

Obtain the heatmap of a dataset.



Clustering

Cluster the individuals of a dataset.



Self-Organizing Map

Use SOM as a clustering and visualization method.

Self-Organizing Map

Self-Organizing Map

Explore individuals

Explore prototypes

Superclustering

1. Run a SOM analysis



Select a dataset

Proteome_Muscle_50

Topology

hexagonal

Map length

5

Map width

5

Seed

21171

You can fix the random seed used to ensure reproducibility of the analysis.

Run SOM

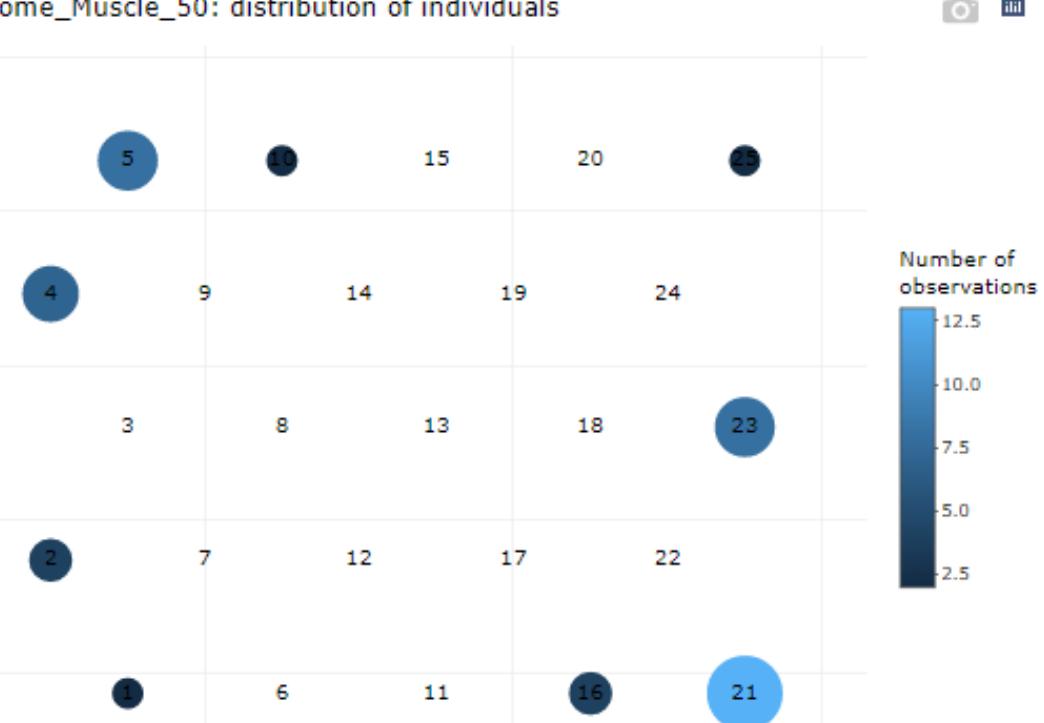
2. Explore SOM clustering

Plots

Summary

Cluster view

Proteome_Muscle_50: distribution of individuals



Self-Organizing Map

Self-Organizing Map

Explore individuals

Explore prototypes

Superclustering

Quality criteria of the SOM map

Topographic error

0.04

Quantization error

781.6191

Correlation ratio

individual

Ratio

P0457	0.9942
P0390	0.9939
ACO2_0718	0.9933
P1008	0.9932
P1057	0.9921
P0240	0.9919
TNNIT3_Isoform_7_1058	0.9914



Quality criteria:

- **topographic error:** is a measure of how well the map is organized (i.e., if two clusters are close, can you trust the fact that corresponding individuals are similar?). This number is always between 0 (best possible organization) and 1 (worst possible organization), with 0 being the value expected for small maps;
- **quantization error:** is a measure of the clustering quality (very similar to within-dispersion in clustering). This number is always non negative (the smaller its value, the better the clustering).



Correlation ratio: This table displays the 20 first variables with the largest percentage of inertia reproduced by the clusters of the map. The largest the percentage of inertia, the more relevant the corresponding variable is to explain differences between clusters.

Self-Organizing Map

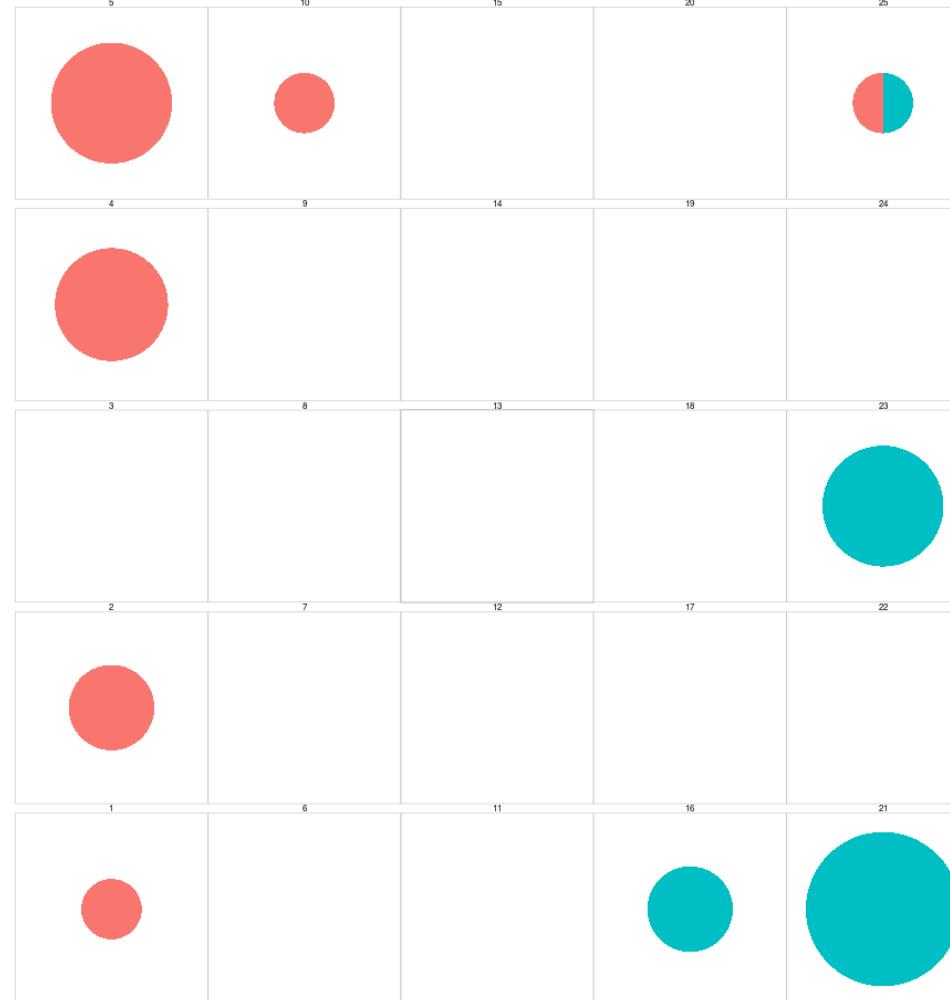
Self-Organizing Map

Explore individuals

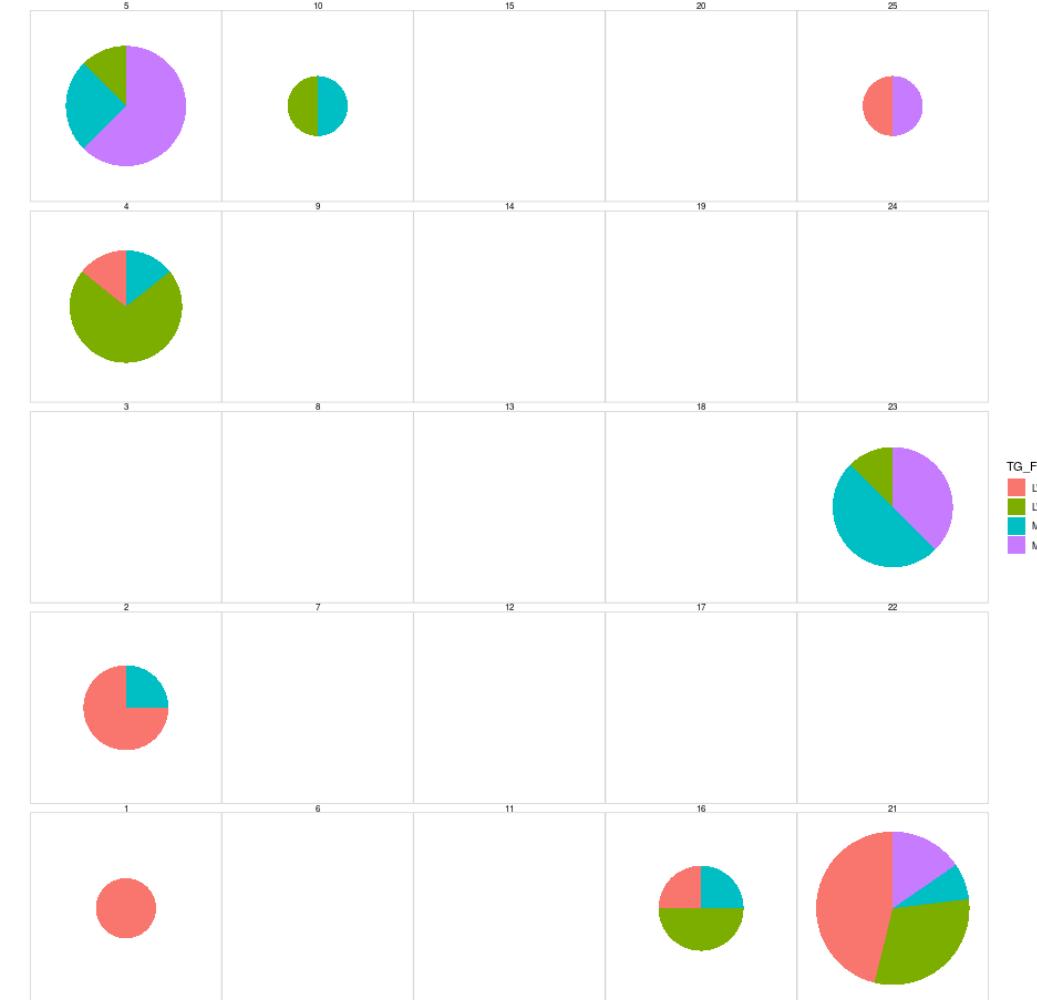
Explore prototypes

Superclustering

SOM on Proteome_Muscle_50: additional variable overview



SOM on Proteome_Muscle_50: additional variable overview



Self-Organizing Map

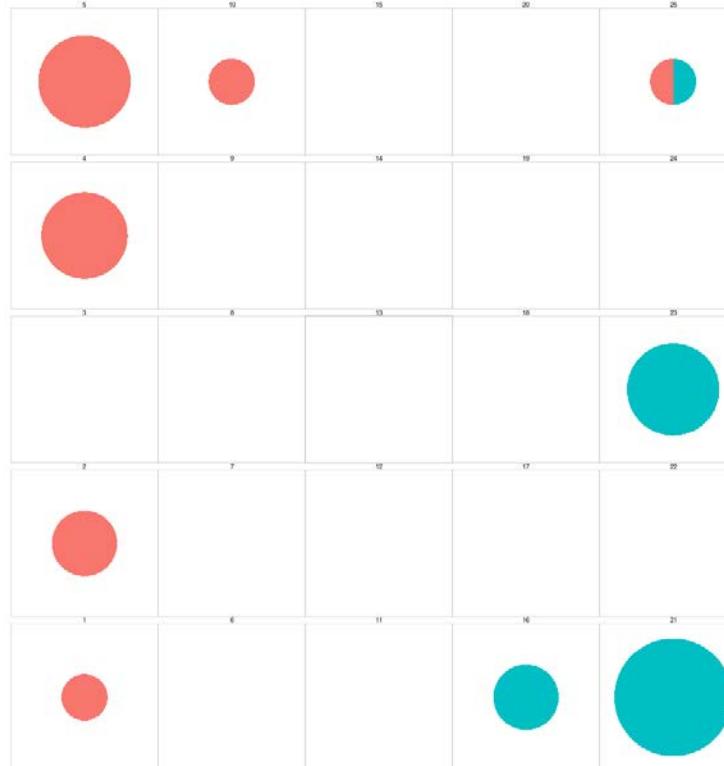
Self-Organizing Map

Explore individuals

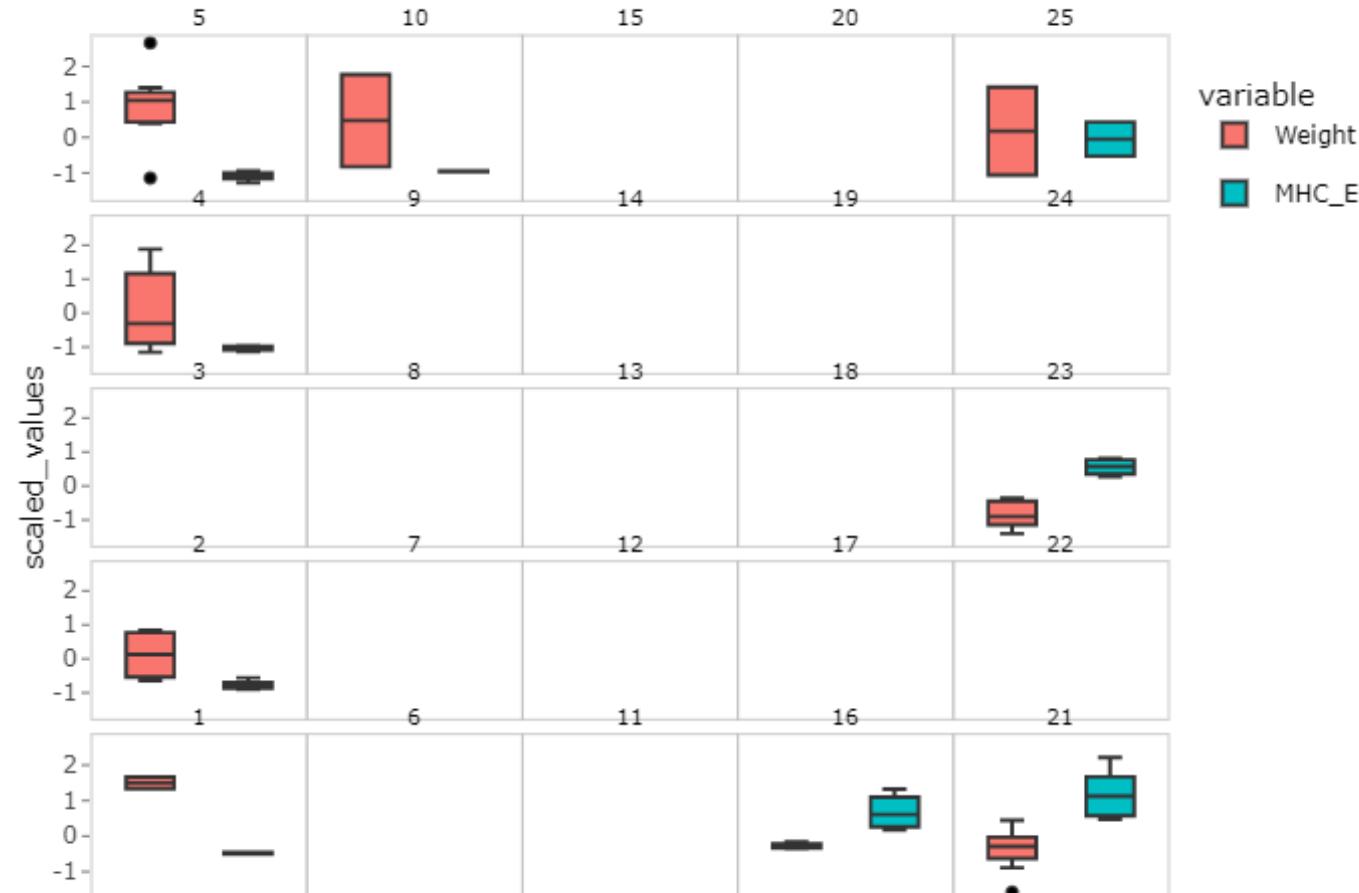
Explore prototypes

Superclustering

SOM on Proteome_Muscle_50: additional variable overview



SOM on Proteome_Muscle_50: additional variable overview



Self-Organizing Map

Self-Organizing Map

Explore individuals

Explore prototypes

Superclustering

1. Display prototypes



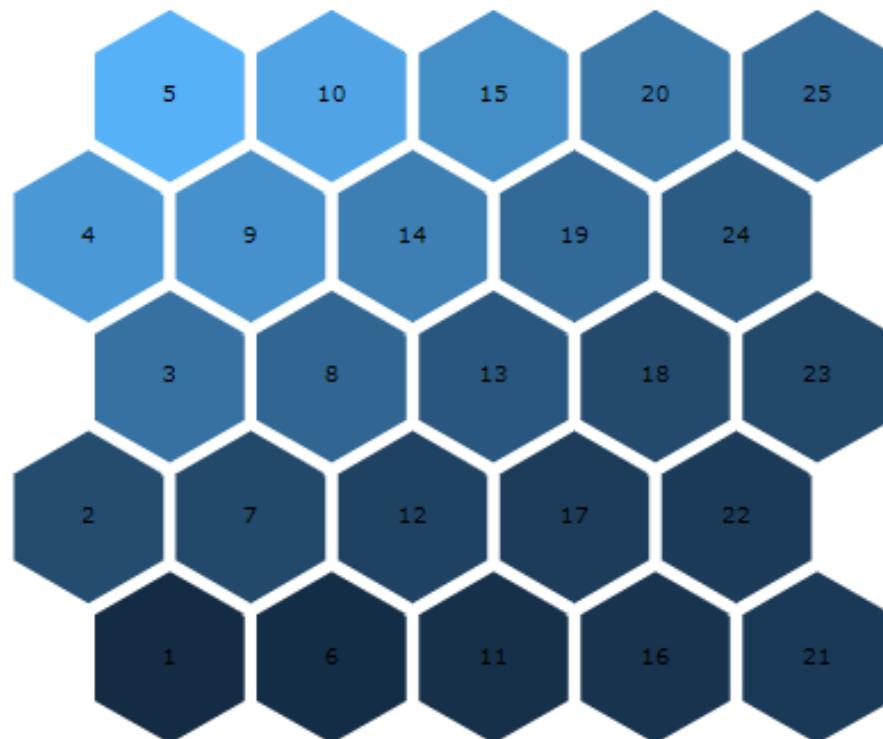
Plot type

Select one variable

Explore individuals

2. Explore prototypes

SOM on Proteome_Muscle_50: prototypes overview



value of
GPD1..Isoform.4._1032
for each prototype



Add to report

Self-Organizing Map

Self-Organizing Map

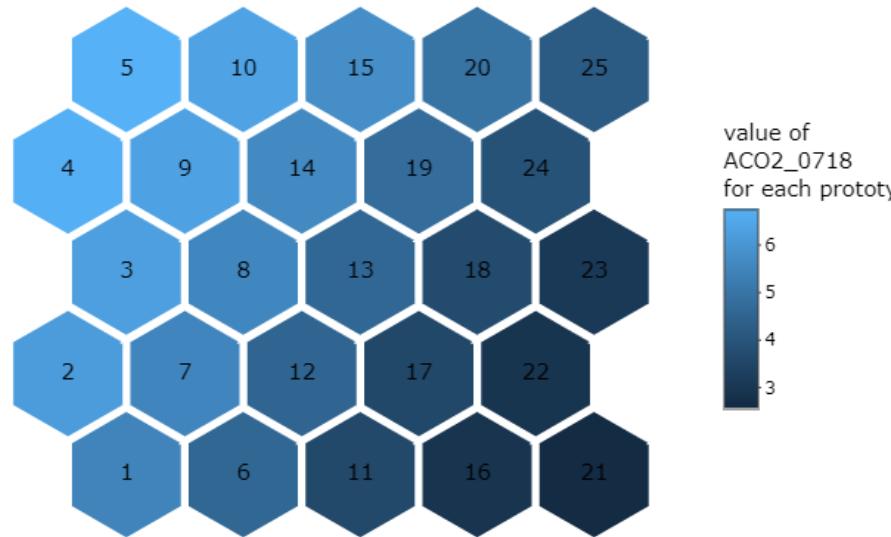
Explore individuals

Explore prototypes

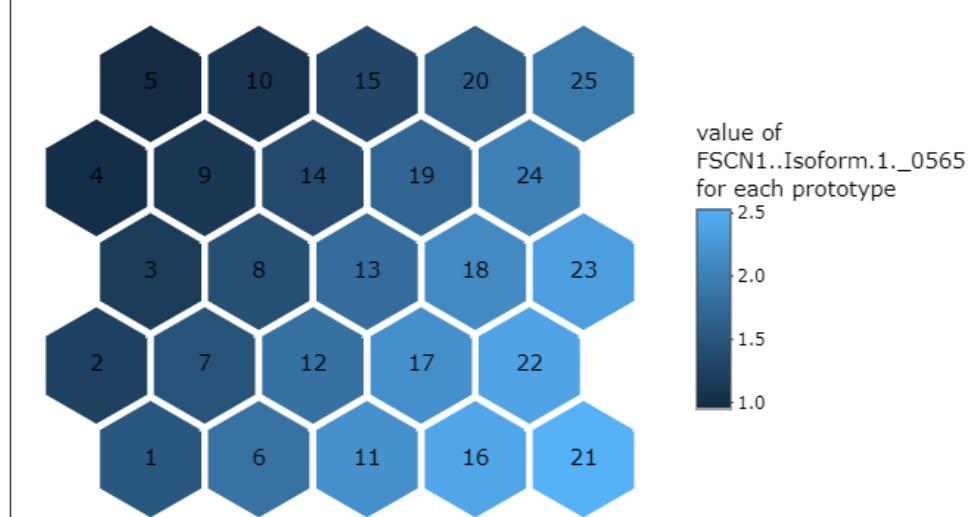
Superclustering

	Corr.Ratio
P0457	0.9942
P0390	0.9939
ACO2_0718	0.9933
P1008	0.9932
P1057	0.9921
P0240	0.9919
TNNT3..Isoform.7._1058	0.9914
FH_0451	0.9911
P0913	0.9911
P0773	0.991
CKMT2_0429	0.9908
P0719	0.9907
NDUFV1_0496	0.9906
ATP5A1_0520	0.9904
P0269	0.9892
PGK1..Isoform.1._0427	0.9891
MDH2_0275	0.988
FSCN1..Isoform.1._0565	0.9875
ACADVL_1051	0.9867
P0276	0.9867

SOM on Proteome_Muscle_50: prototypes overview



SOM on Proteome_Muscle_50: prototypes overview



Self-Organizing Map

Self-Organizing Map

Explore individuals

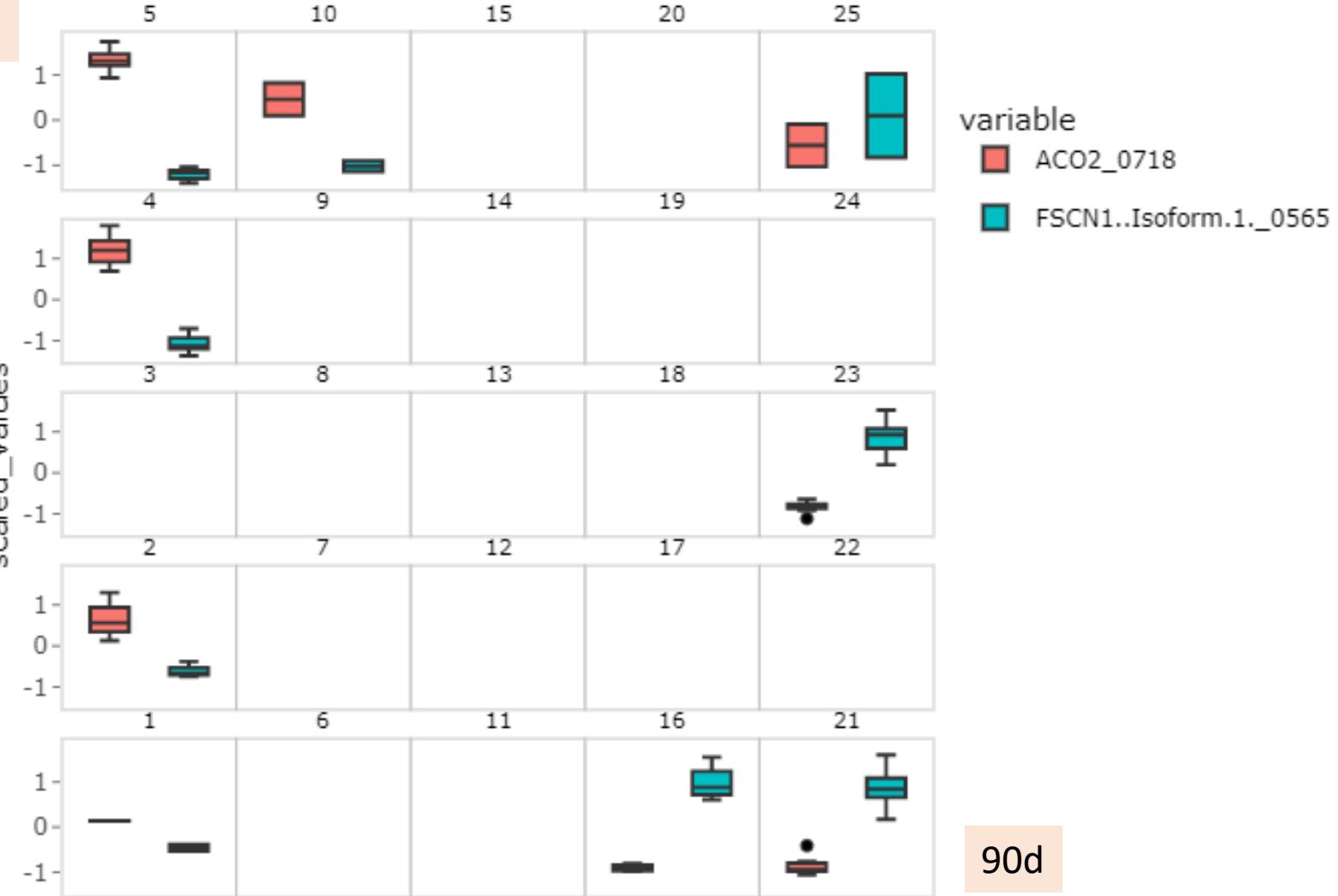
Explore prototypes

Superclustering

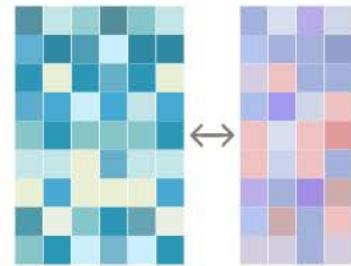
	Corr.Ratio
P0457	0.9942
P0390	0.9939
ACO2_0718	0.9933
P1008	0.9932
P1057	0.9921
P0240	0.9919
TNNT3..Isoform.7._1058	0.9914
FH_0451	0.9911
P0913	0.9911
P0773	0.991
CKMT2_0429	0.9908
P0719	0.9907
NDUFV1_0496	0.9906
ATP5A1_0520	0.9904
P0269	0.9892
PGK1..Isoform.1._0427	0.9891
MDH2_0275	0.988
FSCN1..Isoform.1._0565	0.9875
ACADVL_1051	0.9867
P0276	0.9867

SOM on Proteome_Muscle_50: overview of individuals

110d

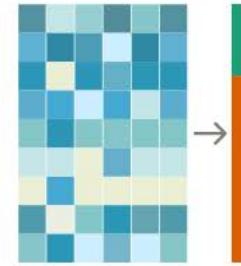


Let's integrate data!



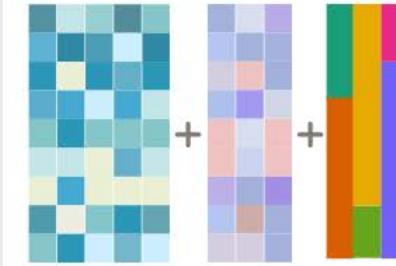
Integrate two datasets with PLS

Performs Partial Least Squares analysis on two datasets.



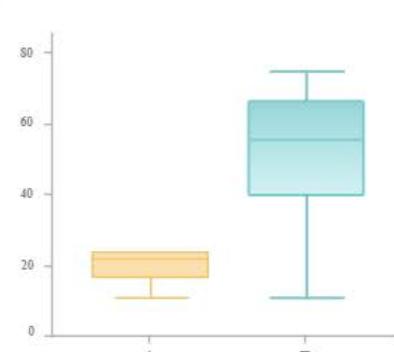
Integrate two datasets with PLS-DA

Performs Partial Least Squares analysis on two datasets.



Integrate datasets with MFA

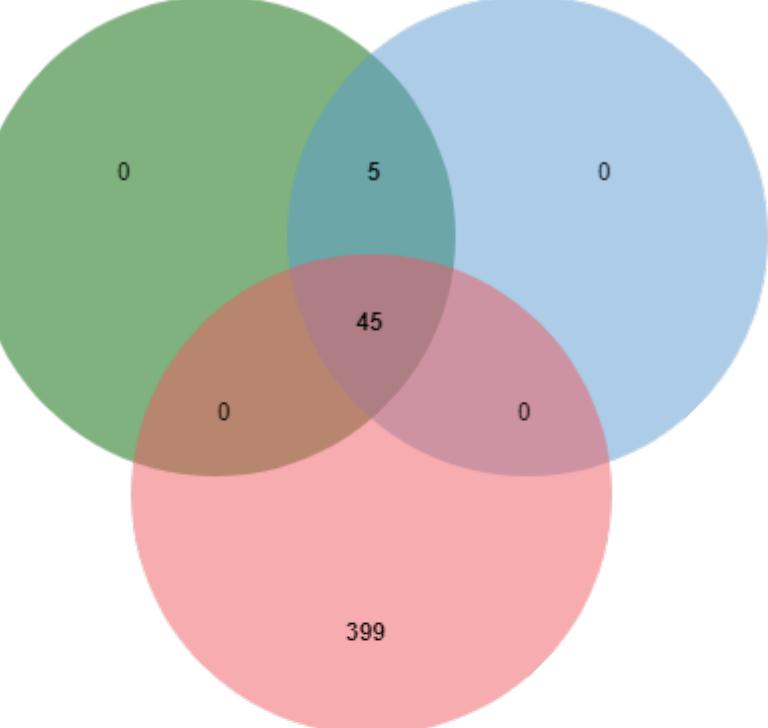
Perform Multiple Factor Analysis on several datasets.



Differential analysis

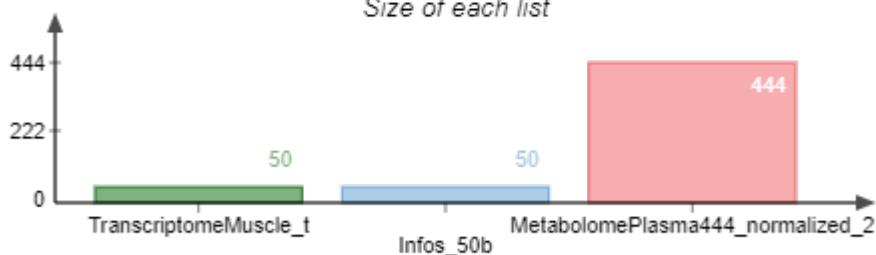
Perform differential analysis for all numeric variables of a dataset.

TranscriptomeMuscle_t Infos_50b

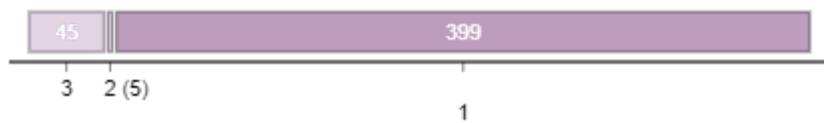


MetabolomePlasma444_normalized_2

Size of each list



Number of elements: specific (1) or shared by 2, 3, ... lists



Integrate datasets with MFA

Preprocessing

Run MFA

Explore individuals

Explore variables

Explore groups

Extract new data

1. Display individuals on components



Component to display on X (horizontal)

1



Component to display on Y (vertical)

2



Color



Info_50b_e

TG_F



Shape



Info_50b_e

Age



Size



Info_50b_e

Weight



Partial individual group



Partial individual group



Partial individuals

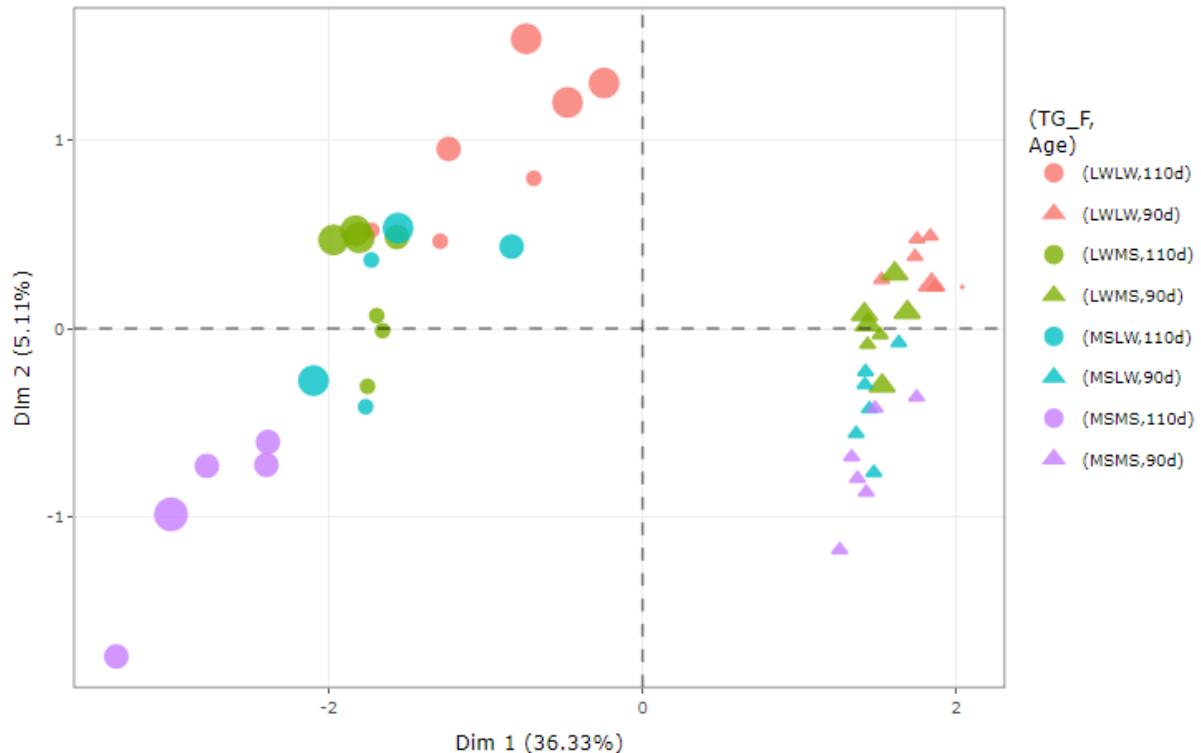


Add an individual

Plot individuals

2. Plot individuals on components

MFA on Info_50b_e, TranscriptomeMuscle50_Imputed_1, Proteome_Muscle_50: projection on Dim 1



Add to report

Integrate datasets with MFA

Preprocessing

Run MFA

Explore individuals

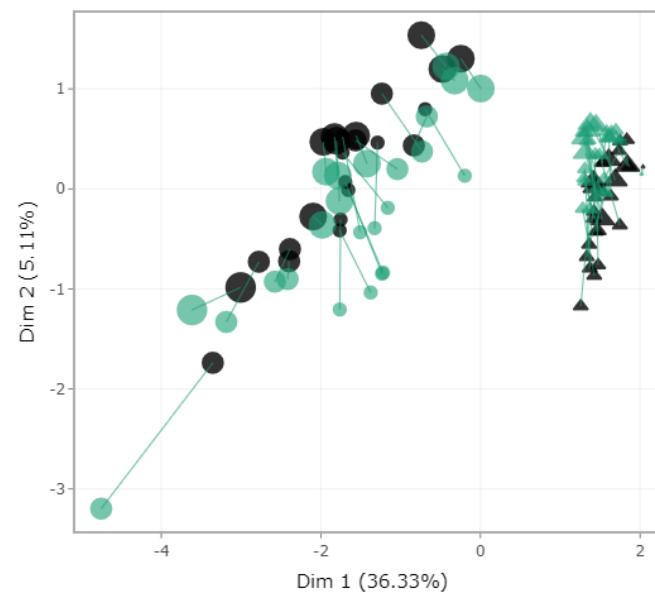
Explore variables

Explore groups

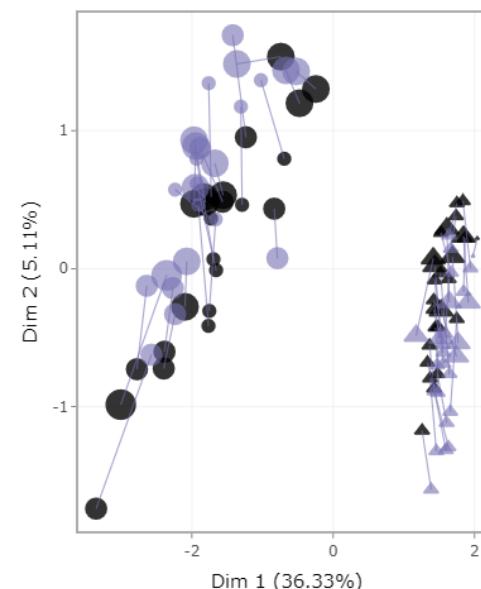
Extract new data

Next steps...

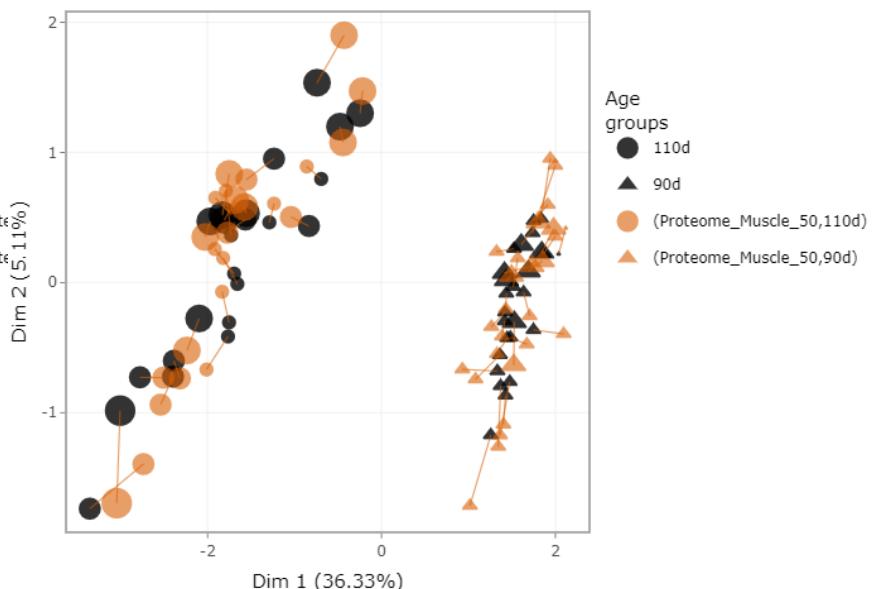
MFA on Info_50b_e, TranscriptomeMuscle50_Imputed_1, Proteome_Muscl



MFA on Info_50b_e, TranscriptomeMuscle50_Imputed_1, Proteome_Muscl



MFA on Info_50b_e, TranscriptomeMuscle50_Imputed_1, Proteome_Muscl



Datasets 8 Variables 29,283 Analyses 6 Remaining days Copy url

Workflow

```

graph TD
    A[TranscriptomeMu...  
microarray  
Rows: 60 Col: 27569] --> B((MissOverview_1))
    B --> C[Proteome_Muscle...  
generic  
Rows: 60 Col: 1026]
    C --> D[Info_600  
metadata  
Rows: 60 Col: 33]
    D --> E(editor_1)
    E --> F[Info_600_2  
metadata  
Rows: 60 Col: 33]
    F --> G[MetabolomePlasm...  
metabolite-compo  
Rows: 63 Col: 444]
    G --> H(editor_2)
    H --> I[Info444  
metadata  
Rows: 444 Col: 13]
    I --> J[MetabolomePlasm...  
metabolite-compo  
Rows: 444 Col: 63]
    J --> K(PCAobj_1)
    K --> L(PCAobj_2)
    L --> M[PCAobj_11d91a2b5246.html]
    L --> N[PCAobj_21d95ea9658d.html]
  
```

WinZip (Non enregistré) - asterics_reports1d94db1c2ee.zip

Fichier Actions Options Aide

Nouveau Ouvrir Favoris Ajouter Extraire Voir Contrôle Assistant

Nom	Type	Modifié	Taille	Taux	Compressé
astericsreport editor_11d91a2b5246.html	Chrome HTML Document	17/08/2022 09:13	1 964 748	73%	528 764
astericsreport editor_21d95b0edb02.html	Chrome HTML Document	17/08/2022 09:13	1 964 745	73%	528 769
astericsreport editor_31d94bd4355e.html	Chrome HTML Document	17/08/2022 09:13	1 964 636	73%	528 729
astericsreport MissOverview_11d923bfd802.html	Chrome HTML Document	17/08/2022 09:13	6 232 916	71%	1 819 891
astericsreport PCAobj_11d97665a528.html	Chrome HTML Document	17/08/2022 09:13	5 418 126	71%	1 547 641
astericsreport PCAobj_21d95ea9658d.html	Chrome HTML Document	17/08/2022 09:13	5 522 510	72%	1 568 596

Sélectionnés 0 fichiers, 0 octets Total 6 fichiers, 22 528Ko

All analyses

Remove Export report ?

<input checked="" type="checkbox"/> Name	Function
> <input checked="" type="checkbox"/> editor_1	r_edit_dataset
> <input checked="" type="checkbox"/> editor_2	r_edit_dataset
> <input checked="" type="checkbox"/> MissOverview_1	r_missing_overview
> <input checked="" type="checkbox"/> PCAobj_1	r_fAMD
> <input checked="" type="checkbox"/> editor_3	r_edit_dataset
> <input checked="" type="checkbox"/> PCAobj_2	r_fAMD

Merci à toute l'équipe d'Asterics
Merci pour votre attention



To be continued...