



Birmingham River Champion's Urban Riverfly guidance

Overview

Urban Riverfly one of four citizen science techniques that volunteers are undertaking within the Birmingham River Champions. This technique is led by the Freshwater Biological Association and enables volunteers to monitor river ecosystem health by identifying **macroinvertebrates**. These little critters are great 'bioindicators', meaning that the presence of different species can indicate different pressures (e.g., the absence of sensitive species may indicate pollution). They are therefore known as the 'canaries' of our waterways as they can provide early warning indications of extreme pressures.

Urban Riverfly should take under 1 hour (less than 30-minutes in time) and sampling should ideally be performed **every month** when volunteers are willing and available and weather / flow conditions permit. The sampling should be repeated at the same site (the location of which should be sent to the Birmingham River Champions) and ideally by the same surveyor. For **biosecurity** purposes, groups sampling multiple locations (and particularly multiple rivers) on the same day should change surveyors.

Methods

Site identification

Before undertaking Urban Riverfly, you should identify a section of river that is **easy to access** and **shallow** (below welly height). Ideally, you should sample in habitats called 'riffles' - shallow waters containing fast flows and larger sediments with 'wave' like features on the water surface.



Riffle habitats in rivers, characterised by shallow depths, fast flow velocities, coarse sediments and 'wave-like' disturbances on the water surface

Kick sampling and hand search

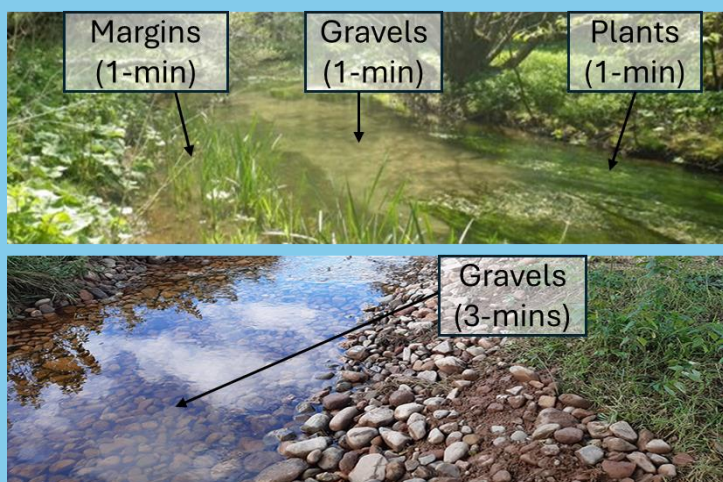
Volunteers should take the D-Frame net into the river and perform a **3-minute kick sample**. This involves disturbing riverbed habitats (e.g., plants, algae, sediments) using the base of your foot in a 'kicking' motion, with the net positioned just below (facing upstream) to collect the disturbed macroinvertebrates. This is easiest when you're stood 'side on' and the riverbed is disturbed in front of the net (you do not need to 'score goals')



and kick into the net as this will clog up your tray). Some key things to remember:

- You should spend **approximately 15-seconds** kicking in each spot.
- You should **only time** when **actively kicking**.
- You should **divide** your time up **proportionally** based on the habitats present (e.g., if your river has equal gravels and plant coverage, you'd spend 1.5-minutes kicking in each).

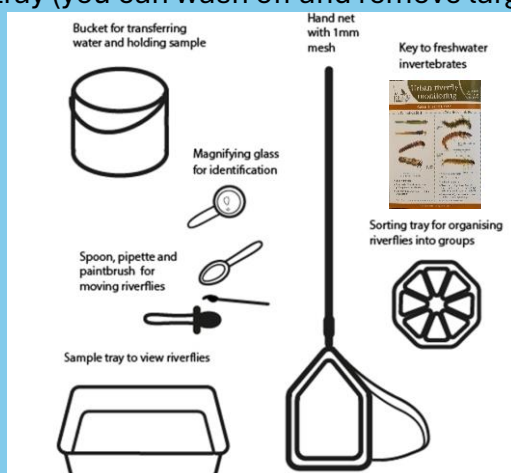
After the kick sample, you should perform a **1-minute hand search**. For this, you should inspect large objects (e.g., wood or rocks) that cannot be disturbed by kicking, and the river margins for 'surface-dwelling' macroinvertebrates (e.g., "pond skaters") and add these to your net (it is okay if you cannot see anything during this period).



A typical stance for a kick sample and different habitats sampled for different time periods based on their proportional occurrence across the channel.

Macroinvertebrate sampling

You should empty your net into a tray containing river water. But if you have a large amount of material it might be easiest to place your sample into a bucket with river water and empty parts in at a time – you should ideally be able to see at least 30% of the white tray (you can wash off and remove large items like rocks and leaves as necessary).



The Urban Riverfly kit and a macroinvertebrate sample with roughly the maximum amount of material that should be put into a tray when identifying specimens.



You should record the number of each identified specimen into one of the following abundance categories: **1-9; 10-99; 100-999; ≥ 1000** . If splitting one sample across multiple trays, you should roughly tally the number of specimens in each tray before coming up with a final abundance category value based on all the trays combined. Urban Riverfly records can be uploaded *via* our [online form](#), where we will generate summary statistics based that will be displayed on our [website](#).

‘Cheats’ for identifying key species

Besides the resources already provided, see this ‘cheat sheet’ for IDing key species:

Species	Identification information
Cased caddisflies	Look out for small stones arranged in a ‘tube’ structure, only record if you can see the caddisfly inside . Other cases can be made out of plants and even snail shells.
Caseless caddisflies	No tails, but posterior hooks . Look out for a hard head and soft body . Two main types: <ul style="list-style-type: none"> • ‘Green sedge’ caddisflies - green, spikey gills on the side of body; • ‘Net-spinning’ caddisflies – dark brown head / light brown body, ‘broom’ like hooks, gills under body
Stoneflies	Stoneflies = 2 tails; Mayflies = 3 tails – both have 6 legs – but mayfly tails are lost easily, not to be mistaken for stoneflies. Also don’t confuse with damselflies (see decision tree below).
Green drake (burrowing) mayfly	
Flat-bodied (stone clinger) mayfly	
Olive mayfly	Most common mayfly. Moves rapidly (torpedo), slimmer body, ‘leaf-like’ gills on side. Don’t confuse with blue-winged olives!
Blue-winged olive mayfly	Darker, wider body than olives with stripes . Tail can stick up like a scorpion . Moves slowly , gills not sticking out.
Freshwater shrimp	Lots of legs , swims on side . Generally peach or cream colour. Contact BRC project team if you see ‘striped’ pattern or spikes near the appendage as this may be the invasive ‘demon’ shrimp.
Freshwater hoglouse	Lots of legs , crawls on front . Dark brown.
Adult beetles	Most common will be jet black all over and small . If confident , see the decision tree below on beetle larvae.
Freshwater worms	Generally pink colour. Unlike non-biting midges, worms have no visible head and don’t swim , also much longer / wriggly .
Freshwater snails	Do not record empty shells. May come in different forms besides more ‘typical’ looking snails, such as ‘ramshorn’ snails (coiled and flat) and ‘limpets’ (wizards hat).
Freshwater leeches	2 key types . One much larger, darker that moves by expanding/contracting (elastic band). The other smaller , curl up in c-shape and have ‘train track’ pattern on back.



