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Spaceflight alters expression of microRNA during T-cell activation

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Abstract

Altered immune function has been demonstrated in astronauts during spaceflights dating back to Apollo and Skylab; this could be a major barrier to long-term space exploration. We tested the hypothesis that spaceflight causes changes in microRNA (miRNA) expression. Human leukocytes were stimulated with mitogens on board the International Space Station using an onboard normal gravity control. Bioinformatics showed that miR-21 was significantly up-regulated 2-fold during early T-cell activation in normal gravity, and gene expression was suppressed under microgravity. This was confirmed using quantitative real-time PCR ($n = 4$). This is the first report that spaceflight regulates miRNA expression. Global microarray analysis showed significant ($P < 0.05$) suppression of 85 genes under microgravity conditions compared to normal gravity samples. *EGR3*, *FASLG*, *BTG2*, *SPRY2*, and *TAGAP* are biologically confirmed targets and are co-up-regulated with miR-21. These genes share common promoter regions with pre-mir-21; as the miR-21 matures and accumulates, it most likely will inhibit translation of its target genes and limit the immune response. These data suggest that gravity regulates T-cell activation not only by transcription promotion but also by blocking translation via noncoding RNA mechanisms. Moreover, this study suggests that T-cell activation itself may induce a sequence of gene expressions that is self-limited by miR-21.—Hughes-Fulford, M., Chang, T. T., Martinez, E. M., Li, C.-F. Spaceflight alters expression of microRNA during T-cell activation.

Keywords: gene expression, microgravity, epigenetics, immune response

Over 50% of the Apollo astronauts had bacterial or viral infections during flight or within 1 wk of landing (1). Apollo 7 marked humanity's first experience with spaceflight infection when all 3 crew members contracted head colds during their mission. On Apollo 13, one astronaut contracted *Pseudomonas aeruginosa* and experienced intense chills and fever (1). *P. aeruginosa* is an opportunistic pathogen that rarely causes disease unless the person is immunosuppressed. As a result, the U.S. National Aeronautics and Space Administration (NASA) implemented a preflight quarantine program that subsequently reduced the number of reported infections to a single Apollo astronaut (1). To this day, the preflight quarantine program is still actively used in both the U.S. and Russian programs. Even with the precautions, astronauts working on Skylab (2), Shuttle (3, 4), International Space Station (ISS) (3), and Soyuz (5) showed changes in immune function and depressed lymphocyte activation compared to levels before spaceflight. Experiments from Skylab and Shuttle have confirmed that T cells have a suppressed immune response (*in vivo* and *in vitro*) with lower T-cell proliferation/activation, lower IL-2 synthesis and severely reduced IL-2-R α expression (RNA and protein) (6–9). More recently, our ISS experiments proved that microgravity of spaceflight was a major cause of decrease of T-cell activation and altered gene expression in microgravity compared to onboard normal gravity controls (10). Immunosuppression during spaceflight may increase the risk of opportunistic infections. Shuttle astronauts on short duration (11 d) spaceflights had significant increases in early viral gene transcription of the Epstein-Barr virus compared to healthy controls, while astronauts on board the ISS for long-duration spaceflight (180 d) had latent and lytic viral gene expression that resembled activation patterns observed during infectious mononucleosis (3, 11).

MicroRNAs (miRNAs) were first discovered in 1993 in *Caenorhabditis elegans* when miRNA *lin-4* was seen to down-regulate expression of the gene *lin-14*; however, there were no homologs to *lin-4* in other species. The later discovery of miRNAs in other species, including humans, showed that miRNAs are common in eukaryotes. Previous studies have shown that key miRNAs are up-regulated after activation of human T cells (12). Here we present our discovery of dysregulation of miRNA gene expression by gravity during early T-cell activation in spaceflight. We recently proved that the lack of immune response in microgravity occurs at the cellular level (10) and identified promoter regions, transcription factors, and signal transduction pathways regulating human T-cell gene activation under normal and altered gravity conditions (10, 13, 14). To our knowledge, this is the first time that miRNAs have been analyzed using an onboard normal gravity control centrifuge eliminating possible confounders such as launch effects, cosmic radiation, and vibration, leaving only gravity conditions as a variable. Our data suggest that gravity regulates T-cell gene expression by 2 separate mechanisms during immune activation. Additionally, our finding that several miR-21 targets are co-up-regulated with miR-21 during early activation suggests that miR-21 may be involved in a previously unrecognized autoregulatory loop that limits the duration of normal T-cell activation.

MATERIALS AND METHODS

Flight mission profile

Whole peripheral blood was obtained from 4 human donors between the ages of 21 and 55 years. All were in good health, and none was taking medication at the time of donation; no personal health identifiers were recorded. This study was reviewed and approved by the University of California, San Francisco, Committee on Human Research. Red blood cells were lysed and peripheral blood leukocytes isolated using Ficoll gradients. Human T cells were further purified using enrichment columns per manufacturer's instructions (R&D Systems, Minneapolis, MN, USA), which isolate CD3⁺ T cells to 90% or greater purity. Human T cells were resuspended into RPMI 1640 medium with 10% fetal calf serum and loaded at 6×10^6 cells per chamber into the spaceflight hardware, Lycis. The spaceflight hardware consisted of 8 cassettes, each with 4 chambers. Some units had smart-button temperature data storage capability, and temperatures of the containers were tracked for the duration of the mission. Purified T cells from each donor were kept separate and loaded into 4 individual chambers in separate cassettes for the following treatments: microgravity nonactivated, microgravity activated, 0.5 g activated, and normal gravity activated. Units were loaded and handed over to the Russian team 14 h before launch and were stored in insulated pouches. The experimental units were launched into space on board the Soyuz 13S rocket from Baikonur, Kazakhstan, on September 18, 2006. Before the in-flight experimental procedures, cassettes were kept in ambient temperatures, which ranged from 16 to 22°C. Cassettes were transferred to the ISS immediately after docking of the Soyuz vehicle on September 20 (flight day 3) and placed into the Kubik incubator centrifuge at 36.5°C. All samples were preincubated at 36.5°C for 30 min. The normal gravity and 0.5 g units were placed in the central centrifuge positions and centrifuged with an applied normal gravity or 0.5 g force, respectively. The microgravity units were placed in static positions for continued microgravity exposure. After 30 min of preincubation, microgravity-nonactivated units were fixed by addition of RNALater (Qiagen, Valencia, CA, USA), removed from the incubator, and stored at 4°C. The microgravity-, 0.5 g-, and normal gravity-activated units were activated with a final concentration of 10 µg/ml Con A and 4 µg/ml anti-CD28. These cassettes were replaced into Kubik on either the centrifuge or static positions and activated for 1.5 h. Activation was stopped with the addition of RNALater, and the units were then moved to 4°C storage. All units were returned to earth on board the Soyuz 12S vehicle on September 28. After landing on September 29, samples were kept at 4°C and handed over to the PI team at the Moscow airport on September 30. Frozen samples were transported to the investigators' laboratory in San Francisco for analysis.

RNA isolation

RNA was isolated by the RNeasy Mini kit (Qiagen) according to the manufacturer's protocol. The samples were then stored at -80°C until further analysis. Isolated RNA from spaceflight samples were all of high quality. Although only a limited amount of small RNA was recovered with this method, there was an abundant amount of miR-21 that could be detected by gene array and quantitative real-time PCR (qRT-PCR).

Microarray sample preparation and analysis

Three of the 4 human volunteer samples of high RNA quality, as determined by the 260 nm/280 nm values (range, 1.8–2.0, with the majority of samples at 2.0), were chosen for microarray hybridization. RNA was amplified and biotinylated using the MessageAmp II-Biotin enhanced kit per the manufacturer's instructions (Ambion, Austin, TX, USA). A total of 10 µg of biotinylated aRNA was hybridized on to the Human U133 Plus 2.0 Array (Affymetrix, Santa Clara, CA, USA) through the University of California, San Francisco, Gladstone Institute Genomics Core. MIAME (minimum information about a microarray experiment)-compliant microarray data can be found under the accession number [GSE38836](https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE38836) and are posted online (<http://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE38836>).

Microarray analysis

Microarray data were analyzed by GeneSpring GX 13.0 software (Agilent Technologies, Santa Clara, CA, USA). Data were normalized using the GC-RMA algorithm, and baseline normalization was set to the average level of expression in microgravity-nonactivated T cells. Low signal probes (below 20%) were filtered to remove background noise, and statistical analysis was performed using 1-way ANOVA and the Benjamini-Hochberg multiple testing correction with a *P* value of ≤ 0.05 . *Post hoc* Tukey analysis was performed to identify 85 genes with significant differences in regulation between the normal gravity- and microgravity-activated samples. Genes were filtered for fold change that was 2-fold or greater between the normal gravity- and microgravity-activated samples. MiRNA target gene clustering analysis was performed in GeneSpring GX 13.0 software using Pearson's centered metric with centroid linkage.

Gene targets of miR-21

Predicted gene targets for miR-21 were found using TargetScan 6.2 ([15](#)). Targets with a context+ score of -0.10 or less were considered. Experimentally confirmed miRNA targets for miR-21 were identified using DIANA-TarBase v.7.0, a manually curated database ([16](#)). miRNA targets from these databases were compared with the list of 85 genes differentially regulated in gravity conditions to identify 16 genes that had predicted and/or confirmed miR-21 target sites in the 3'-UTR region. *FASLG*, a predicted gene target of miR-21, passed a significance cutoff at $P \leq 0.06$ but not $P \leq 0.05$ and was included in further analysis because of its well-established role in T-cell signaling and function.

qRT-PCR

Details of qRT-PCR methods have been previously published ([10](#)). At the end of the amplification period, melting curve analysis was performed to confirm the specificity of the amplicon. RNA samples were normalized to cyclophilin A (CPhi), also known as kinase C peptidyl prolyl isomerase A (PPIA), as a internal standard. PPIA expression is stable between normal gravity and microgravity conditions. Relative quantification of gene expression was calculated by the

$2^{-\Delta\Delta C_t}$ equation. All data derived using qRT-PCR were from independent donor biologic samples ($n = 4$).

miRNA reverse transcription

miR-21 miRNA was detected and quantified using the mirVana qRT-PCR miRNA detection kit (Ambion) according to the manufacturer's protocol. Total RNA (100 ng) was added to 10 μ l reverse transcription (RT) reaction buffer containing mirVana RT buffer, mirVana RT primer (mir21 or 5S internal standard), and ArrayScript enzyme mix. The RT reaction was incubated at 37°C for 10 min, then 95°C for 10 min to inactivate. A no-template control was also performed for each primer set. For miRNA qRT-PCR, cDNA from the RT reaction (10 μ l) was added to a total of 25 μ l qRT-PCR mixture containing 5 μ l of 5 \times SYBR Green mirVana PCR buffer, 0.5 μ l of 50 \times ROX, 0.5 μ l mirVana PCR primers (miR21 or 5S internal standard), and 1 U of SuperTaq polymerase (Applied Biosystems, Foster City, CA, USA). PCRs were carried out in a Bio-Rad MyiQ Single-Color Real-Time PCR Detection System (Bio-Rad, Hercules, CA, USA). The thermal profile was 50°C for 2 min, denaturation at 95°C for 3 min, followed by 40 amplification cycles at 95°C for 15 s and at 60°C for 30 s. Fluorescence was measured and used for quantitative purposes. At the end of the amplification period, melting curve analysis was performed to confirm the specificity of the amplicon. RNA samples were normalized to 5S rRNA internal standard, which was stable under all gravity conditions. Relative quantification of gene expression is calculated by using the $2^{-(C_t \text{ gene T} - C_t \text{ 5S T}) - (C_t \text{ gene MA} - C_t \text{ 5S MA})}$ equation, where threshold cycle (C_t) gene T represents the calculated C_t of a time point of each sample other than microgravity activated (MA). All data derived by qRT-PCR were from independent biologic samples ($n = 4-8$).

RESULTS

miR-21 is dysregulated in true ISS microgravity during T-cell activation

Using bioinformatics, we analyzed the gene expression T-cell activation of 3 individual human donors under normal gravity on board, and microgravity conditions that were incubated, activated, and fixed on the ISS. In our initial bioinformatics analysis, we found that there was 1 miRNA that had reduced gene expression in true microgravity; in normal gravity, miR-21 was increased 2-fold, while in microgravity miR-21 was not significantly changed compared to onboard normal gravity controls ($n = 4$). This gene array analysis was confirmed by mirVana qRT-PCR. Expression was corrected to internal standard 5S rRNA. A small amount of mRNA from 0.5 g onboard samples ($n = 4$) was available for analysis. With a 0.5 g fractional gravity, the expression of miR-21 was increased over the microgravity sample.

Analysis of mRNA gene arrays for significantly dysregulated microgravity genes

Global microarray analysis revealed significant suppression of 85 genes under microgravity conditions compared to gene up-regulation in onboard normal gravity-activated samples. We analyzed the altered gene expression of activated T

cells in microgravity using software showing either proven miR-21 interaction (TarBase), predicted seed sequences (TargetScan), or both. [Table 1](#) provides an abbreviated list of 13 key target genes that significantly changed gene expression within 1.5 h after activation in spaceflight.

TABLE 1.

Potential and proven miR-21 targets are differentially regulated in normal gravity and microgravity

Gene array fold change, normal gravity vs. microgravity	Gene symbol	Gene title	Source
23.24	<i>EGR1</i>	Early growth response 1	TarBase
11.83	<i>EGR3</i>	Early growth response 3	TargetScan
5.86	<i>FASLG</i>	Fas ligand (TNF superfamily, member 6)	TarBase + TargetScan
5.33	<i>SPRY1</i>	Sprouty homolog 1,	TargetScan
5.19	<i>BTG2</i>	BTG family, member 2	TarBase + TargetScan
4.63	<i>MYC</i>	v-Myc myelocytomatosis viral oncogene homolog	TarBase
4.32	<i>IRF4</i>	Interferon regulatory factor 4	TarBase
4.03	<i>SPRY2</i>	Sprouty homolog 2	TarBase + TargetScan
3.78	<i>TAGAP</i>	T-cell Rho GTPase activating protein	TargetScan
44.10	<i>CD40LG</i>	T-cell activation GTPase TNF superfamily	No miR-21 seed sequence
2.91	<i>CD83</i>	CD83 molecule	TarBase
2.84	<i>REL</i>	v-Rel	TarBase
2.55	<i>NT5E (CD73)</i>	5' Nucleotidase, ecto (CD73)	TarBase + TargetScan
2.32	<i>CD69</i>	CD69 molecule	TargetScan

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Relative fold increases of gene expression from activated T cells in onboard normal gravity compared with those in microgravity were computed using gene array bioinformatics. Potential hsa-miR-21-5p target genes shown have differentially regulated between normal gravity and microgravity conditions. T cells from 3 independent donors were flown in space and activated with concanavalin A/anti-CD28 in microgravity or in

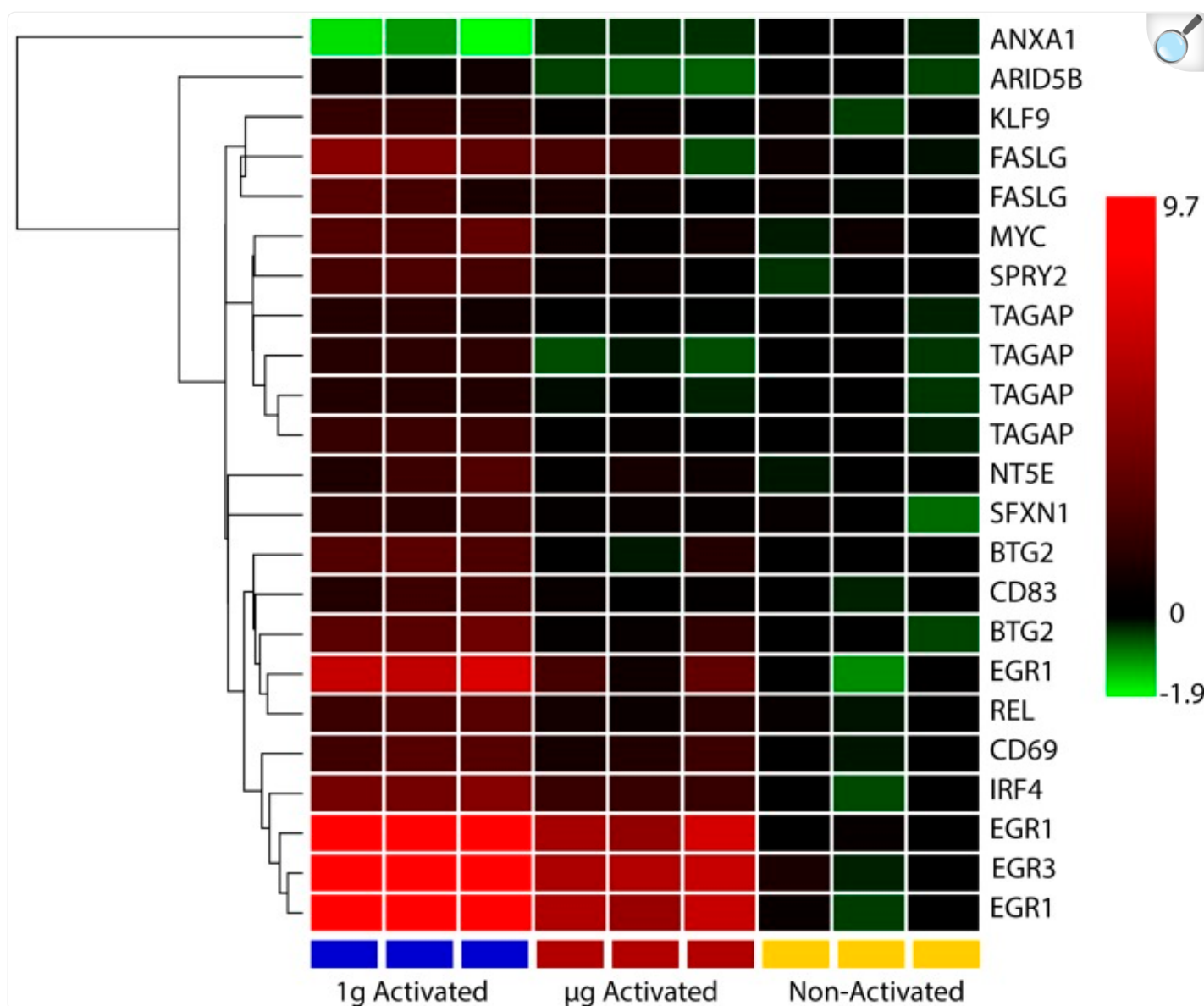
normal gravity. Genes that had altered expression between microgravity and normal gravity were identified by 1-way ANOVA (≤ 0.05) with Benjamini-Hochberg multiple test correction and *post hoc* Tukey test. *FASLG* was not quite significant at $P \leq 0.05$ but was significant at $P \leq 0.06$ and thus is included in the list. Potential hsa-miRNA-21-5p targets were identified using TargetScan (predicted miRNA targets) and TarBase (confirmed miRNA–gene interactions).

Of the 85 gravity-sensitive genes, 17 were defined as miR-21 targets. Of these, 13 were immune-related miR-21 target genes, including *EGR3*, *CD69*, *FASLG*, *SPRY1*, *BTG2*, *SPRY2*, and *TAGAP*. All the immune-related targets were biologically confirmed. We further confirmed *via* gene array analysis using qRT-PCR that *FASLG*, *SPRY2*, *BTG2*, and *TAGAP* gene expression was gravity sensitive.

Heat map of gene expression

Seventeen significant gene targets of miR-21 from flown individual donors ($n = 3$) were differentially regulated after 1.5 h of activation under normal gravity and microgravity conditions on board the ISS ($P \leq 0.05$) ([Fig. 1](#)). Samples were incubated and activated in the same incubator and fixed on board the ISS. In some cases, the gene array contained multiple gene probe sets targeted to different regions of the gene. Here we saw multiple probe sets for *FASLG*, *TAGAP*, and *EGR1* showing the same trends of significant inhibition of gene expression under different gravities. The activation profile in the normal gravity-activated samples is fairly uniform across donors, indicating that common genes and pathways were stimulated in all 3 donors after activation with concanavalin A/anti-CD28. Gene expression in the microgravity flown samples was lower and less uniform than the activated normal gravity samples. In some cases, the microgravity profile was essentially the same as the nonactivated samples, indicating suppression of gene expression in all 3 donors. The 3 conditions had distinct profiles across the 17 predicted miR-21 gene targets, demonstrating clear differences in early activation gene expression between the onboard normal gravity-activated, microgravity-activated, and nontreated controls.

Figure 1.



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Heat map of predicted miR-21 target genes showing differentially regulated 1.5 h after T-cell activation in microgravity (μg) and normal gravity (1g). Genes were grouped based on expression profile using Pearson's centered metric with centroid linkage. Rows represent gene probes; columns represent expression profiles of donors. Heat map scale ranges from green (down-regulation) to red (up-regulation) for log₂ normalized expression values. Blue-labeled columns are normal gravity-activated samples, red-labeled columns are microgravity-activated samples, and yellow-labeled columns are microgravity-nonactivated T cells. A total of 17 unique miR-21 gene targets are shown with differential expression between normal gravity and microgravity condition ($P \leq 0.05$; *FASLG* $P \leq 0.06$). There is considerable variability between donors in

nonactivated samples and some variability in microgravity-activated donors. Gene expression in normal gravity-activated samples becomes considerably more uniform 1.5 h after activation.

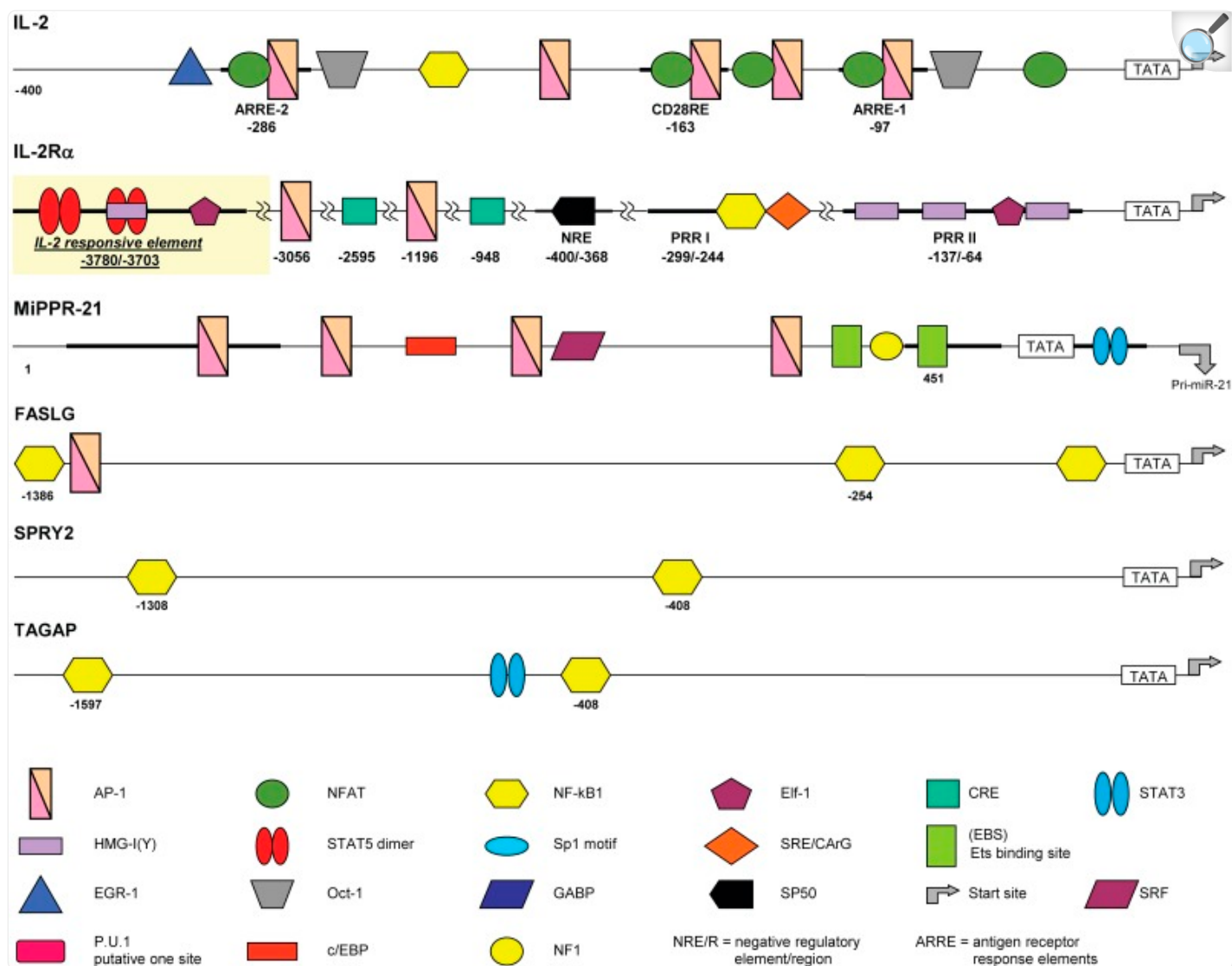
qRT-PCR of immediate early genes in nonactivated, normal gravity onboard activated, and onboard microgravity-activated T cells with 3'-UTR miR-21 seed sequences

We used qRT-PCR to analyze miR-21 targets *BTG2*, *TAGAP*, *SPRY2*, and *FASLG* in these ISS samples that were activated and fixed in orbit. These genes were selected because they contained the seed sequence in the 3'-UTR region. In addition, 4 genes (*BTG2*, *TAGAP*, *SPRY2*, and *FASLG*) had previously been confirmed to have interaction with miR-21. We also analyzed nuclear factor of κ light polypeptide gene enhancer in B-cell inhibitor α (*NFKBIA*) and CD40 ligand (*CD40LG*), 2 genes that are key in early T-cell activation. All 6 genes exhibit gravity-sensitive gene expression. It is notable that many of the miR-21 targets are co-up-regulated with miR-21 during the early hours of T-cell activation.

Predictions of transcription factor locations in promoter regions of gravity-sensitive genes

We used bioinformatics software, the Match program of TRANSFAC Pro, oPOSSUM, target genes to predict transcription factor locations in the promoter regions of gravity-sensitive mRNA. Key immune genes share common transcription factors with miR-21, showing that at the initiation of activation, both the positive immune genes share increased gene expression with the miR-21 precursor pre-miR-21 ([Fig. 2](#)).

Figure 2.



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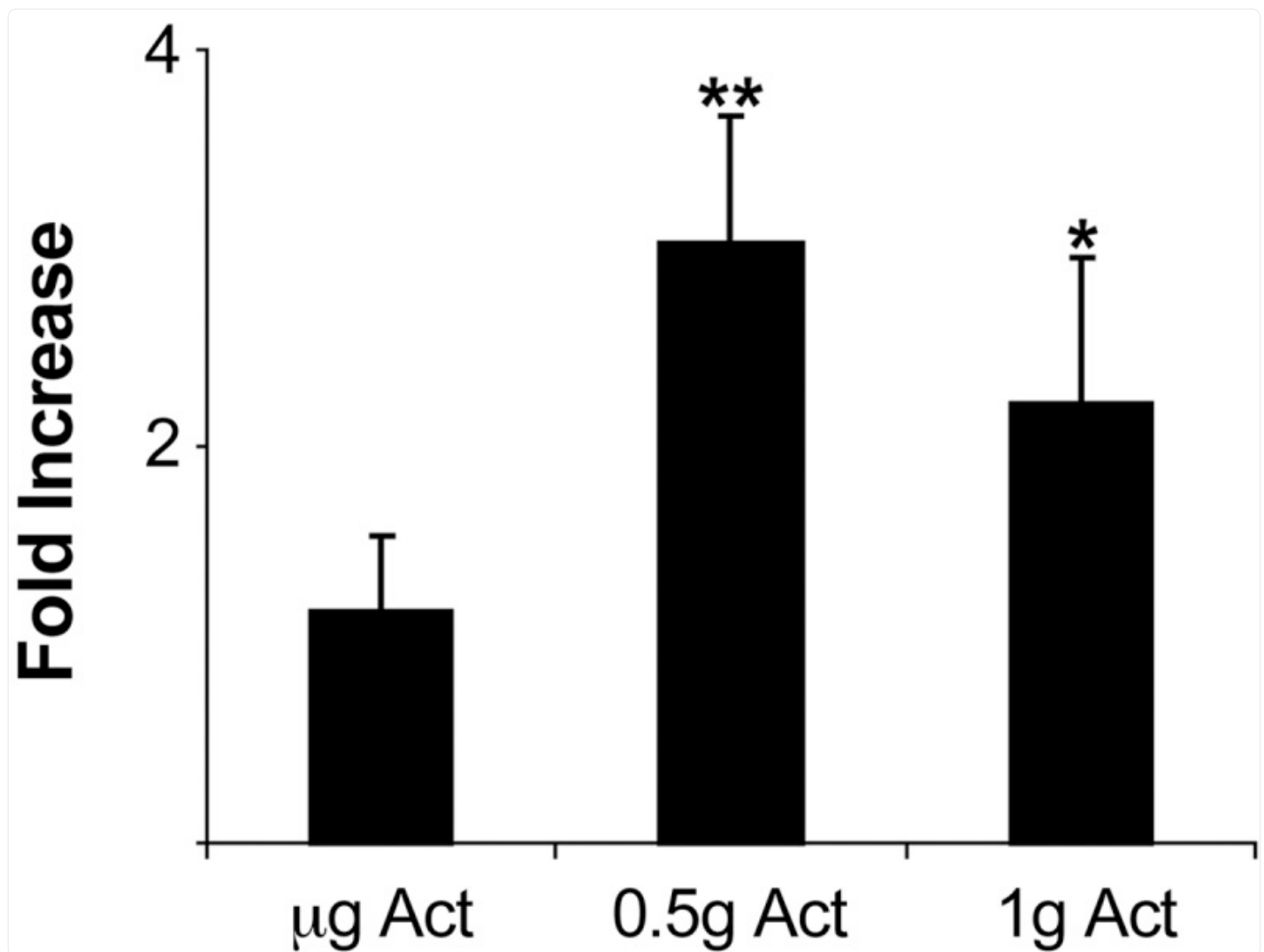
Promoter regions of *IL-1*, *IL-2-R α* , *Premir-21*, *TAGAP*, *SPRY2*, and *FASLG* are induced by AP1 and/or NF- κ B-1 α ; 2 of transcription factors in early gene expression (these 2 transcription factors are found in promoter region of nearly 40% of T-cell early induced genes).

DISCUSSION

miRNAs are a class of small noncoding RNAs that act as posttranscriptional regulators of gene expression and play fundamental roles in regulating immune response and autoimmunity (17). In the data presented here, we show for the

first time an altered profile of miRNA expression under true microgravity compared to its onboard normal gravity controls; all samples were isolated from donors on earth, incubated, activated, and fixed in ISS on-orbit operations. The miR-21 differential expression seen in the global gene analysis was confirmed by mirVana qRT-PCR ([Fig. 3](#)). Of interest, samples from onboard 0.5 g samples demonstrated that fractional gravity is capable of restoring gene expression of miR-21.

Figure 3.



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Real-time confirmation of miR-21 down-regulation of gene expression in CD4⁺ T cells flown on ISS. Cells were placed in microgravity (µg), 0.5g, or normal gravity (1g) for 3 h before activation. At 1.5 h after activation with concanavalin A/anti-CD28, RNA was stabilized in RNALater. Total RNA was isolated, and miR-21 expression was quantified using mirVana miRNA detection kit. Expression was corrected to internal standard 5S rRNA. Bars represent mean ± SD of independent biologic replicates ($n = 6$). * $P < 0.05$; ** $P < 0.001$ with 2-tailed Student t test against activated microgravity samples (µg Act).

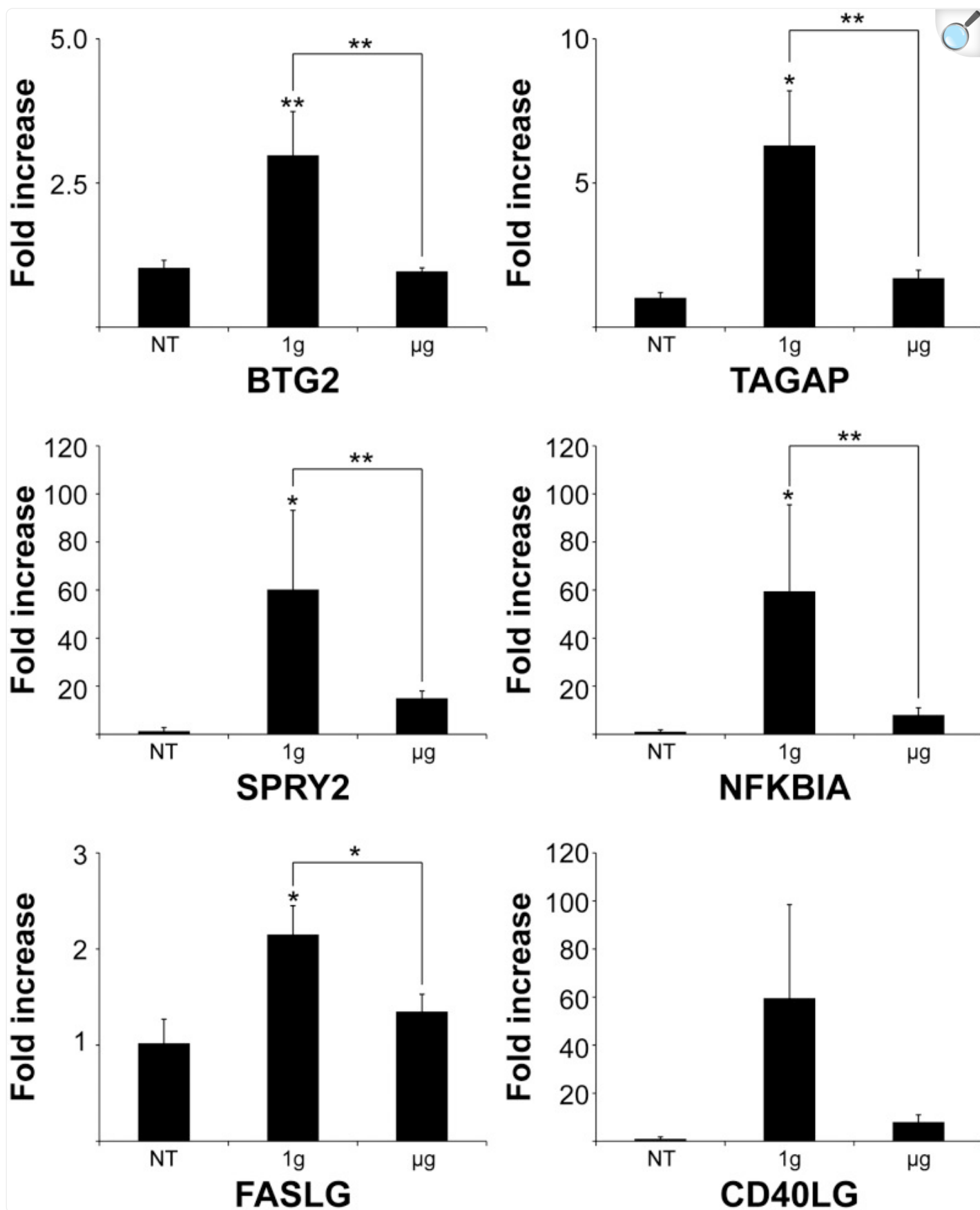
Previous *in vitro* studies by our laboratory using an onboard normal gravity control ([10](#)) demonstrated that T-cell activation was suppressed in microgravity, independent of other systemic factors. Several studies have also

demonstrated that immunosuppression occurs *in vivo* during spaceflight in humans (1–4) and mice (18). The direct effects of microgravity on early T-cell activation described here eliminates confounding systemic effects and shows that the immune system needs gravity to function normally.

Global gene analysis of the ISS samples revealed altered gene expression in spaceflight vs. normal gravity onboard controls 1.5 h after activation, showing changes in 85 genes associated with T-cell activation. Of the 85 differentially expressed genes, 17 were computationally predicted or experimentally verified as miR-21 target genes. Table 1 provides an abbreviated list of 13 T-cell-relevant targets that are predicted miR-21 targets from the miR-21 seed sequence (UAGCUUAU) in the gene by TargetScan. Of the 17 predicted genes, *EGR1*, *FASLG*, *SPRY2*, *BTG2*, *REL*, and *MYC* have been biologically confirmed by others as true targets as described in the TarBase database.

We confirmed the global gene array analysis of several of the target genes using qRT-PCR analysis. We have previously confirmed microgravity-mediated suppression of *IL-2*, *IL-2-RA*, *EGR1*, and *INF- γ* gene expression in the immune response (10, 13, 14, 18). Using qRT-PCR, we confirmed that microgravity-induced down-regulation of 4 of the miR-21 target genes, *BTG2*, *TAGAP*, *SPRY2*, and *FASLG*, as well as immune regulators *NFKB1A* and *CD40LG* genes (Fig. 4). We chose *BTG2*, *TAGAP*, and *SPRY2* for qRT-PCR analysis because they had been previously shown to biologically interact with miR-21. *BTG2* is a known factor in T-cell activation and supports proliferation in immune cells (19). Other genes important in immune function, such as *NFKB1A* and *CD40LG*, were selected because of their importance in early T-cell activation. *CD40LG* is a marker of T-cell activation, and here we found that as early as 1.5 h of activation, it is induced in T cells that were normal gravity activated and was inhibited in microgravity-activated cells. NF- κ B-1A binds to REL, RELA, or RELB to form the NF- κ B complex. NF- κ B-1A is associated with cells that express chemokines or cytokines. *TAGAP* is a member of the rho GTPase activator protein superfamily, and *TAGAP* loci have been associated with immune diseases such as rheumatoid arthritis (20–22) and multiple sclerosis (23, 24). *SPRY2* protein down-regulation by miR-21 inhibits proliferation, and when *SPRY2* is overexpressed, it can increase proliferation of HEK293T cells (25, 26). Binding of *FASLG* (TNFSF6) to FAS results in cell apoptosis and cell death. FAS ligand is known to be expressed in activated splenocytes and may be part of an innate mechanism to limit the duration of cell activation.

Figure 4.

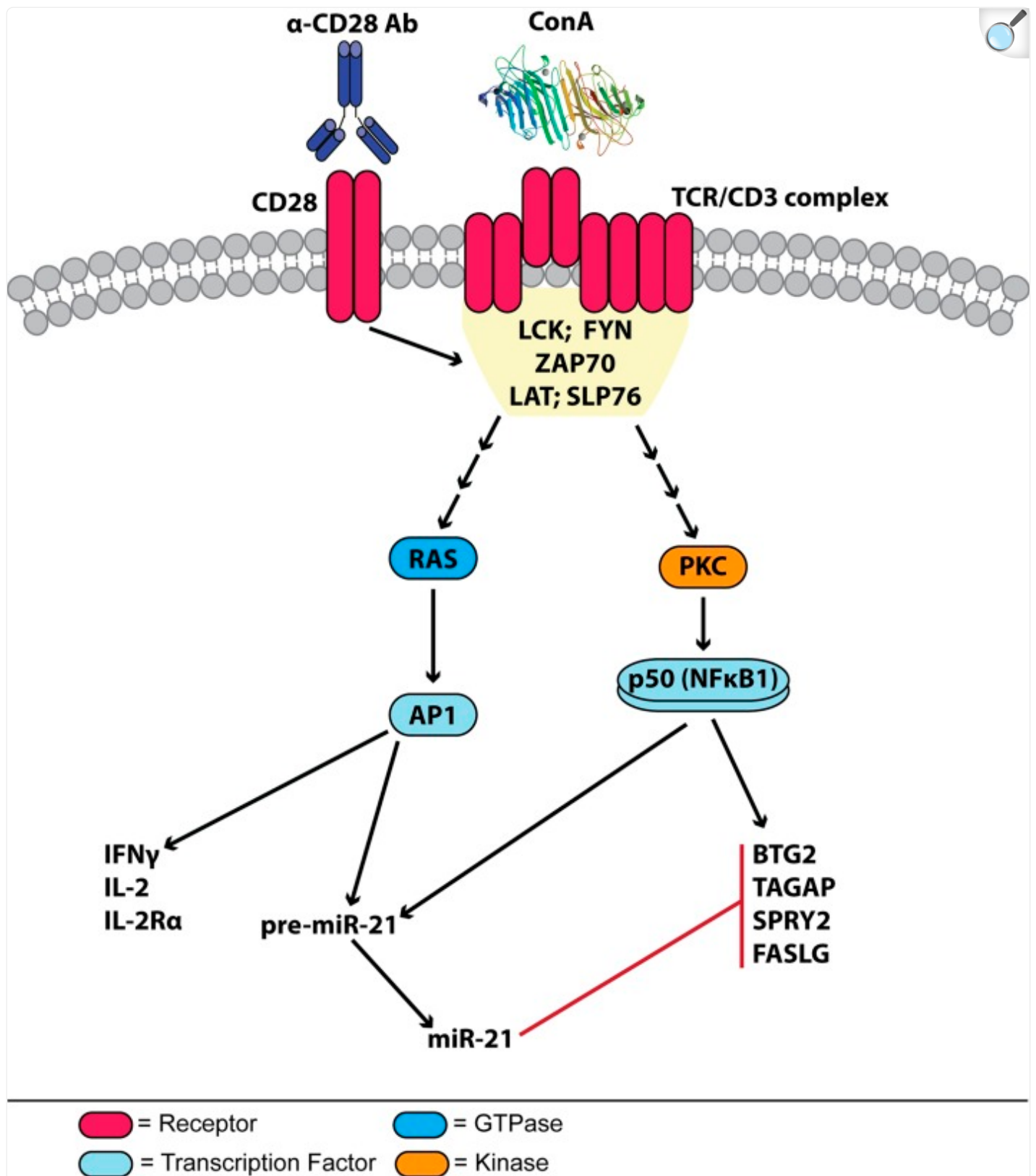


qRT-PCR of genes selected from gene array analysis in nonactivated and concanavalin A/anti-CD28 activated human lymphocytes in flown normal gravity-activated and flown microgravity conditions indicating altered gene expression in ISS spaceflight vs. normal gravity onboard controls (1.5 h). We used qRT-PCR to analyze mir-21 targets *BTG2*, *TAGAP*, *SPRY2*, and *FASLG* in ISS samples that were activated and fixed in orbit. *NFKBIA* and *CD40LG* were also analyzed because they are important in T-cell activation.

These data show that as early as 1.5 h after T-cell activation, key immune genes are induced in an ISS normal gravity environment and are suppressed in microgravity. It is notable that several of the miR-21 targets are initially up-regulated in T-cell activation, and it is possible that as miR-21 levels rise in the cell, this miRNA will cause a reduction in the protein synthesis of these targets and may be another mechanism to limit the time of T-cell activation.

In this laboratory's previous work on activation of T cells, gel shift analysis revealed that the activator protein 1 (AP1) transcriptional factor was activated by protein kinase A and was blocked by a MAPK inhibitor, U016, with AP1 being a key player in T-cell activation (13) (Fig. 5). AP1 is a transcription factor site composed of Fos and Jun family proteins. AP1 is also the key regulator of miR-21 gene induction; Fujita *et al.* (27) demonstrated that miR-21 is regulated by AP1 after phorbol treatment. Moreover, studies by Talotta *et al.* (28) demonstrated that RAS activation of miR-21 is regulated by the AP1 proteins. The promoter region analysis of key immune regulatory genes, including the precursor to miR-21, pre-miR-21, is shown in Fig. 2. The early activation of T cells and the gene induction of miR-21 and 4 of its functional/computational targets share common pathway or pathways of up-regulation by AP1 and NF- κ B.

Figure 5.



Postulated actions of miR-21 and gene expression of early T-cell activation. Hypothesized pathways of expression and down-regulation. miR-21 is known to down-regulate AP1 (28), and from presence of its seed sequence in 3'-UTR of target genes miR-21 is predicted to down-regulate 4 targets approximately 24 h after activation; *BTG2*, *TAGAP*, *SPRY2*, and *FASLG* as miR-21 expression increases. It is possible that miR-21 provides a mechanism of self-limited induction regulating T-cell activation.

Taken together, the early activation of T cells and the gene induction of miR-21 precursor and 4 of its computational targets share common pathways of up-regulation. Therefore, we hypothesize that in early regulation, T-cell activation-induced gene expression is regulated by AP1 and later by protein kinase C. As miR-21 accumulates in the cell in the days after initial activation, it most likely will inhibit translation of miR-21 targets such as *BTG2*, *TAGAP*, *SPRY2*, and *FASLG*. Our previous experiments have demonstrated that AP1 and NF- κ B account for 37% of all significant induction of gene expression 4 h after T-cell activation (13). Figure 5 shows our hypothetical pathway of miR-21 regulation of T-cell activation, illustrating our theory that the pathways that are responsible for early T-cell activation are NF- κ B and AP1. This illustration was deduced from prior work (13, 14) and from Figs. 1, 3, 4.

We have previously shown that *REL*, *IL-2*, *IL-2-RA*, and *INF- γ* are increased during normal gravity activation, and expression is repressed in microgravity (10, 13, 14, 18). Here we present evidence that early activation genes and miR-21 gene expression have increased gene expression during the early hours of T-cell activation. Extensive analysis of the promoter regions seen in Fig. 2 shows that the genes in question share common transcription elements in promoter regions of *IFN- γ* , *IL-2*, *IL-2-RA*, *BTG2*, *TAGAP*, *SPRY2*, *FASLG*, and miR-21 precursor.

We were left with the question of why the miR-21 target genes and miR-21 itself are co-up-regulated the start of T-cell activation. These opposing actions are analogous to pressing the gas pedal while stepping on the brake; however, it takes days for the repressive miR-21 to build up in a cell before blocking translation (29–31). This provides a plausible mechanism for suppression of T-cell activation to limit the immune response. Our results and hypothesis are in line with the findings of Fujita *et al.* (27) showing increased expression of miR-21 by AP1. Moreover, Carissimi *et al.* (32) showed that miR-21 is up-regulated by T-cell receptor activation and that miR-21 reaches peak production at 3 d. In addition, they showed that overexpression of miR-21 in Jurkat cells causes a suppression of T-cell receptor signaling, and they surmised that miR-21 caused suppression of activation downstream of T-cell receptor signal (32), although they were unable to find the mechanism. In mice, miR-21 is a known limiting factor of immune response (33). Here we present evidence that the downstream miR-21 targets *BTG2*, *SPRY2*, *TAGAP*, and *FASLG* may be among the targets regulating suppression. Together, these microgravity experiments support our hypothesized mechanism for miR-21 translational repression of key immune genes. This previously unknown mechanism controlling the immune response has been termed self-limiting induction. This mechanism may apply to other growth induction that is normally self-limiting, such as wound healing. It is possible that in some tissues, dysfunction of this type of mechanism could lead to uncontrolled growth and result in cancer.

Acknowledgments

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Glossary

AP1

activator protein 1

CD40LG

CD40 ligand

CPHI

cyclophilin A aka peptidyl prolyl isomerase A (PPIA)

C_t

threshold cycle

ISS

International Space Station

miRNA

microRNA

NASA

National Aeronautics and Space Administration

NFKB1A

nuclear factor of κ light polypeptide gene enhancer in B-cell inhibitor α

PPIA

peptidyl prolyl isomerase A

qRT-PCR

quantitative real-time PCR

RT

reverse transcription

REFERENCES

1. Hawkins W., Zieglschmid J. (1975) Clinical aspects of crew health. In Biomedical Results of Apollo (Johnston R., Dietlein L., Berry C., eds.), pp. 43–81, National Aeronautics and Space Administration, Washington, DC [[Google Scholar](#)]

2. Kimsey S. (1977) Hematology and Immunology Studies. In Biomedical Results from Skylab (Dietlein R. J. L., ed.), pp. 249–283, National Aeronautics and Space Administration, Washington, DC [[Google Scholar](#)]
3. Stowe R. P., Kozlova E. V., Sams C. F., Pierson D. L., Walling D. M. (2011) Latent and lytic Epstein-Barr virus gene expression in the peripheral blood of astronauts. *J. Med. Virol.* 83, 1071–1077 [[DOI](#)] [[PubMed](#)] [[Google Scholar](#)]
4. Stowe R. P., Mehta S. K., Ferrando A. A., Feedback D. L., Pierson D. L. (2001) Immune responses and latent herpesvirus reactivation in spaceflight. *Aviat. Space Environ. Med.* 72, 884–891 [[PubMed](#)] [[Google Scholar](#)]
5. Konstantinova I. V., Antropova E. N., Legen'kov V. I., Zazhirei V. D. (1973) [Reactivity of lymphoid blood cells in the crew of “Soyuz-6,” “Soyuz-7” and “Soyuz-8” spacecraft before and after flight]. *Kosm. Biol. Med.* 7, 35–40 [[PubMed](#)] [[Google Scholar](#)]
6. Cogoli A. (1997) Signal transduction in T lymphocytes in microgravity. *Gravit. Space Biol. Bull.* 10, 5–16 [[PubMed](#)] [[Google Scholar](#)]
7. Cogoli A., Bechler B., Cogoli-Greuter M., Criswell S. B., Joller H., Joller P., Hunzinger E., Müller O. (1993) Mitogenic signal transduction in T lymphocytes in microgravity. *J. Leukoc. Biol.* 53, 569–575 [[DOI](#)] [[PubMed](#)] [[Google Scholar](#)]
8. Crucian B. E., Cabbage M. L., Sams C. F. (2000) Altered cytokine production by specific human peripheral blood cell subsets immediately following space flight. *J. Interferon Cytokine Res.* 20, 547–556 [[DOI](#)] [[PubMed](#)] [[Google Scholar](#)]
9. Pippia P., Sciola L., Cogoli-Greuter M., Meloni M. A., Spano A., Cogoli A. (1996) Activation signals of T lymphocytes in microgravity. *J. Biotechnol.* 47, 215–222 [[DOI](#)] [[PubMed](#)] [[Google Scholar](#)]
10. Chang T. T., Walther I., Li C. F., Boonyaratanakornkit J., Galleri G., Meloni M. A., Pippia P., Cogoli A., Hughes-Fulford M. (2012) The Rel/NF- κ B pathway and transcription of immediate early genes in T cell activation are inhibited by microgravity. *J. Leukoc. Biol.* 92, 1133–1145 [[DOI](#)] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
11. Payne D. A., Mehta S. K., Tying S. K., Stowe R. P., Pierson D. L. (1999) Incidence of Epstein-Barr virus in astronaut saliva during spaceflight. *Aviat. Space Environ. Med.* 70, 1211–1213 [[PubMed](#)] [[Google Scholar](#)]
12. Contreras J., Rao D. S. (2012) MicroRNAs in inflammation and immune responses. *Leukemia* 26, 404–413 [[DOI](#)] [[PubMed](#)] [[Google Scholar](#)]

13. Boonyaratanakornkit, J. B., Cogoli, A., Li, C. F., Schopper, T., Pippia, P., Galleri, G., Meloni, M. A., and Hughes-Fulford, M. (2005) Key gravity-sensitive signaling pathways drive T cell activation. *FASEB J.* **19**, 2020–2022 [[DOI](#)] [[PubMed](#)]
14. Hughes-Fulford M., Sugano E., Schopper T., Li C. F., Boonyaratanakornkit J. B., Cogoli A. (2005) Early immune response and regulation of IL-2 receptor subunits. *Cell. Signal.* **17**, 1111–1124 [[DOI](#)] [[PubMed](#)] [[Google Scholar](#)]
15. Grimson A., Farh K. K., Johnston W. K., Garrett-Engele P., Lim L. P., Bartel D. P. (2007) MicroRNA targeting specificity in mammals: determinants beyond seed pairing. *Mol. Cell* **27**, 91–105 [[DOI](#)] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
16. Vlachos I. S., Paraskevopoulou M. D., Karagkouni D., Georgakilas G., Vergoulis T., Kanellos I., Anastasopoulos I. L., Maniou S., Karathanou K., Kalfakakou D., Fevgas A., Dalamagas T., Hatzigeorgiou A. G. (2015) DIANA-TarBase v7.0: indexing more than half a million experimentally supported miRNA:mRNA interactions. *Nucleic Acids Res.* **43**, D153–D159 [[DOI](#)] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
17. Baumjohann D., Ansel K. M. (2013) MicroRNA-mediated regulation of T helper cell differentiation and plasticity. *Nat. Rev. Immunol.* **13**, 666–678 [[DOI](#)] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
18. Martinez E. M., Yoshida M. C., Candelario T. L., Hughes-Fulford M. (2015) Spaceflight and simulated microgravity cause a significant reduction of key gene expression in early T-cell activation. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* **308**, R480–R488 [[DOI](#)] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
19. Terra R., Luo H., Qiao X., Wu J. (2008) Tissue-specific expression of B-cell translocation gene 2 (*BTG2*) and its function in T-cell immune responses in a transgenic mouse model. *Int. Immunol.* **20**, 317–326 [[DOI](#)] [[PubMed](#)] [[Google Scholar](#)]
20. Chen R., Stahl E. A., Kurreeman F. A., Gregersen P. K., Siminovitch K. A., Worthington J., Padyukov L., Raychaudhuri S., Plenge R. M. (2011) Fine mapping the TAGAP risk locus in rheumatoid arthritis. *Genes Immun.* **12**, 314–318 [[DOI](#)] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
21. Prasad P., Kumar A., Gupta R., Juyal R. C., Thelma B. K. (2012) Caucasian and Asian specific rheumatoid arthritis risk loci reveal limited replication and apparent allelic heterogeneity in north Indians. *PLoS One* **7**, e31584 [[DOI](#)] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
22. Viatte S., Plant D., Lunt M., Fu B., Flynn E., Parker B. J., Galloway J., Solymossy C., Worthington J., Symmons D. P., Dixey J. J., Young A., Barton A. (2013) Investigation of rheumatoid arthritis genetic susceptibility markers in the early rheumatoid arthritis study further replicates the *TRAF1* association with radiological damage. *J. Rheumatol.* **40**, 144–156 [[DOI](#)] [[PubMed](#)] [[Google Scholar](#)]

23. Akkad D. A., Bellenberg B., Esser S., Weiler F., Epplen J. T., Gold R., Lukas C., Haghighia A. (2015) Multiple sclerosis risk loci correlate with cervical cord atrophy and may explain the course of disability. *Neurogenetics* 16, 161–168 [[DOI](#)] [[PubMed](#)] [[Google Scholar](#)]
24. Ottoboni L., Frohlich I. Y., Lee M., Healy B. C., Keenan B. T., Xia Z., Chitnis T., Guttman C. R., Khoury S. J., Weiner H. L., Hafler D. A., De Jager P. L. (2013) Clinical relevance and functional consequences of the TNFRSF1A multiple sclerosis locus. *Neurology* 81, 1891–1899 [[DOI](#)] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
25. Sayed D., Rane S., Lypowy J., He M., Chen I. Y., Vashistha H., Yan L., Malhotra A., Vatner D., Abdellatif M. (2008) MicroRNA-21 targets Sprouty2 and promotes cellular outgrowths. *Mol. Biol. Cell* 19, 3272–3282 [[DOI](#)] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
26. Wang J. H., Zhou W. W., Cheng S. T., Liu B. X., Liu F. R., Song J. Q. (2015) Downregulation of Sprouty homolog 2 by microRNA-21 inhibits proliferation, metastasis and invasion, however promotes the apoptosis of multiple myeloma cells. *Mol. Med. Rep.* 12, 1810–1816 [[DOI](#)] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
27. Fujita S., Ito T., Mizutani T., Minoguchi S., Yamamichi N., Sakurai K., Iba H. (2008) miR-21 Gene expression triggered by AP-1 is sustained through a double-negative feedback mechanism. *J. Mol. Biol.* 378, 492–504 [[DOI](#)] [[PubMed](#)] [[Google Scholar](#)]
28. Talotta F., Cimmino A., Matarazzo M. R., Casalino L., De Vita G., D'Esposito M., Di Lauro R., Verde P. (2009) An autoregulatory loop mediated by miR-21 and PDCD4 controls the AP-1 activity in RAS transformation. *Oncogene* 28, 73–84 [[DOI](#)] [[PubMed](#)] [[Google Scholar](#)]
29. Ando Y., Yang G. X., Kenny T. P., Kawata K., Zhang W., Huang W., Leung P. S., Lian Z. X., Okazaki K., Ansari A. A., He X. S., Invernizzi P., Ridgway W. M., Lu Q., Gershwin M. E. (2013) Overexpression of microRNA-21 is associated with elevated pro-inflammatory cytokines in dominant-negative TGF- β receptor type II mouse. *J. Autoimmun.* 41, 111–119 [[DOI](#)] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
30. Chusorn, P., Namwat, N., Loilome, W., Techasen, A., Pairojkul, C., Khuntikeo, N., Dechakhamphu, A., Talabnin, C., Chan-On, W., Ong, C. K., Teh, B. T., and Yongvanit, P. (2013) Overexpression of microRNA-21 regulating PDCD4 during tumorigenesis of liver fluke-associated cholangiocarcinoma contributes to tumor growth and metastasis. *Tumour Biol.* 34, 1579–1588 [[DOI](#)] [[PubMed](#)]
31. Qin X., Yan L., Zhao X., Li C., Fu Y. (2012) microRNA-21 overexpression contributes to cell proliferation by targeting PTEN in endometrioid endometrial cancer. *Oncol. Lett.* 4, 1290–1296 [[DOI](#)] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
32. Carissimi, C., Carucci, N., Colombo, T., Piconese, S., Azzalin, G., Cipolletta, E., Citarella, F., Barnaba, V.,

Macino, G., and Fulci, V. (2014) miR-21 is a negative modulator of T-cell activation. *Biochimie* **107**(Pt B), 319–326 [[DOI](#)] [[PubMed](#)]

33. Lu, T. X., Hartner, J., Lim, E. J., Fabry, V., Mingler, M. K., Cole, E. T., Orkin, S. H., Aronow, B. J., and Rothenberg, M. E. (2011) MicroRNA-21 limits *in vivo* immune response–mediated activation of the IL-12/IFN-gamma pathway, Th1 polarization, and the severity of delayed-type hypersensitivity. *J. Immunol.* **187**, 3362–3373 [[DOI](#)] [[PMC free article](#)] [[PubMed](#)]

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