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# Identification of Metagenome-Assembled Genomes Containing Antimicrobial Resistance Genes, Isolated from an Advanced Water Treatment Facility

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Here, we present 95 metagenome-assembled genomes (MAGs) that harbor antimicrobial resistance genes, isolated from samples obtained in a large advanced wastewater reclamation facility prior to microfiltration. The MAGs were not in abundance after filtration at the facility and represent a useful resource to the water treatment community at large.

#### **ABSTRACT**

Here, we present 95 metagenome-assembled genomes (MAGs) that harbor antimicrobial resistance genes, isolated from samples obtained in a large advanced wastewater reclamation facility prior to microfiltration. The MAGs were not in abundance after filtration at the facility and represent a useful resource to the water treatment community at large.

#### **ANNOUNCEMENT**

Human society faces an acceleration of water scarcity due to increasing population, pollution, and land use. Such water scarcity also brings a decrease in water quality, as demonstrated by increased eutrophication, among other concerns (1). Treatment and reuse of water are critical tools in combating water stress globally. Reuse of wastewater carries the potential risk of transmission of small-molecule metabolites and antibiotic resistance genes (ARGs) (2), while its use relieves stress on natural sources of water. Previously, biofilms were sampled from the Orange County Water District (OCWD) Advanced Water Purification Facility (AWPF) in Southern California, which showed differences in the microbial communities in both influent and biofilms on microfiltration and reverse osmosis membranes (3). In a recent study that more fully characterized both biofilms and large volumes of water throughout the treatment process, we described both microbial diversity and load decreasing across this well-engineered system (4). Here, we have expanded and enhanced this previous work (4) by identifying metagenome-assembled genomes (MAGs) that contain ARGs to expand our knowledge of all microbial lineages harboring ARGs that are present within an ultrapurified water facility.

Water and biofilm were sampled at the OCWD AWPF as previously described (4). Briefly, all water samples were filtered using a large-volume concentrator (LVC) dialysis filter cartridge system (Innova Prep LLC, Drexel, MO). Approximately 60 to 100 liters of water were filtered per sample, prior to concentration onto replicate 25-mm (diameter), 0.22-µm (pore size) nitrocellulose filters. Filters were placed in BashingBead lysis tubes (Zymo, Inc., Irvine, CA) containing 750 µl of DNA/RNA Shield (Zymo, Inc.) to preserve samples onsite. Biofilm samples were similarly placed in lysis tubes with DNA/RNA Shield. DNA was extracted using the Zymo Microbiomics DNA/RNA coextraction kit (Zymo, Inc.). Libraries were prepared as previously described in full (4) using the Nextera XT library prep kit (Illumina, San Diego, CA), modified to use 13 cycles of PCR. All samples were sequenced on an Illumina HiSeq 3000 instrument using PE150 chemistry. Two biofilm samples from filtration units at the AWPF obtained from Leddy et al. (3) were processed as described previously and sequenced using SE100 chemistry. Sequence reads were error corrected using BayesHammer v3.12.0 (5) and assembled using MEGAHIT v1.1.3 (6) to produce a single coassembly. Reads were mapped to the assembly with Bowtie 2 v2.3.4.1 (7) and binned into MAGs using Anvi'o v v5 and CONCOCT (8, 9). MAG completion and redundancy estimates were also computed within Anvi'o. MAGs were queried for ARGs using DeepARG (10) and identified phylogenetically by GTDB-Tk v0.3.0 (11). Default parameters were used for all software unless otherwise noted.

A total of 95 MAGs were selected after manual curation within Anvi'o and by DeepARG that contained 185 open reading frames putatively identified as antimicrobial resistance genes. Of note, two MAGs were identified by GTDB-Tk as belonging to the "Candidatus Gracilibacteria" and "Candidatus Patescibacteria" lineages (12). Others identified included an unclassified Dongiaceae MAG that was previously associated with a wastewater treatment facility (13). A more detailed list of assembly statistics and taxonomy of all MAGs can be found in Table 1. As reported previously, no metagenomic sequence and therefore no MAGs were identified after barrier filtration within the system due to a lack of sufficient extractable or amplifiable DNA (4). Future work will further describe the ARGs and will identify non-ARG-containing MAGs and determine how they impact the operation of the OCWD AWPF.

TABLE 1.

Detailed taxonomy, accession information, and assembly statistics of MAGs

Bin name	Taxonomy <u>a</u>	GenBank assembly accession no.	Total length (bp)	N <sub>50</sub> (bp)	GC content (%)	Completeness (%)	Redund (%
Bin_1_1	Mycobacterium sp. AWPF1	SSGB00000000	2,707,225	10,968	68.44	38.85	0.72
Bin_1_3	Mycobacterium sp. AWPF2	<u>SSGA00000000</u>	4,217,119	13,813	68.81	60.43	3.6
Bin_1_5	Unclassified  Actinobacteria family  UBA10799 bacterium  AWPF1	<u>SSET00000000</u>	3,552,223	35,771	69.31	84.17	8.63
Bin_1_8	Unclassified  Actinobacteria family  UBA10799 bacterium  AWPF2	SSES00000000	1,125,909	10,906	67.03	22.3	9.35
Bin_10_1	Polynucleobacter sp.	<u>SSFP00000000</u>	1,928,064	15,906	42.88	78.42	7.19
Bin_11_1	Tolumonas sp.	SSEX00000000	1,832,732	10,968	47.81	53.24	2.88
Bin_11_4	Unclassified  Cyclobacteriaceae bacterium	SSEN00000000	2,496,402	8,305	45.51	36.69	9.35
Bin_11_5	Cyclobacteriaceae ELB16-189 sp.	VIGN00000000	1,066,491	8,282	43.86	36.69	12.23
Bin_12_3	Unclassified  Hyphomicrobiaceae bacterium	SSEH00000000	2,999,406	8,617	55.82	7.19	12.23
Bin_14_1	Novosphingobium sp.	SSFU000000000	1,517,683	8,392	64.2	29.5	0.72

Bin	Taxonomy <u>a</u>	GenBank	Total	N <sub>50</sub> (bp)	GC	Completeness	
name		assembly accession no.	length (bp)		content (%)	(%)	(%
Bin_14_3	Unclassified  Gammaproteobacteria bacterium AWPF1	SSEJ00000000	2,472,121	8,119	63.57	51.08	14.39
Bin_14_4	Pseudoxanthomonas sp.	<u>SSFJ00000000</u>	1,848,156	8,064	67.96	34.53	0
Bin_14_5	Unclassified  Gammaproteobacteria bacterium AWPF2	SSEI00000000	1,859,286	10,791	62.7	43.17	41.73
Bin_15_1	Nitrosomonas sp. AWPF2	SSFW00000000	2,557,120	18,443	43.85	83.45	1.44
Bin_15_2	Burkholderiaceae UBA7693 sp.	SSGW00000000	2,650,165	9,223	49.03	22.3	0.72
Bin_15_3	Moranbacterales UBA1568 sp.	<u>SSGE00000000</u>	580,840	8,401	50.03	48.2	2.88
Bin_15_5	Unclassified  Moranbacterales  UBA1568 sp. AWPF1	SSEE00000000	880,789	8,806	45.6	66.91	19.42
Bin_15_6	Unclassified  Moranbacterales  UBA1568 sp. AWPF2	SSED000000000	1,461,468	7,107	48.36	52.52	15.83
Bin_17_3	Unclassified  Rhodocyclaceae bacterium AWPF3	SSDZ00000000	3,614,409	8,707	60.2	60.43	10.79
Bin_19_1	Ottowia sp. AWPF1	<u>SSFT00000000</u>	2,464,482	7,364	68.69	17.27	2.16
Bin_19_2	Ottowia sp. AWPF2	<u>SSFS00000000</u>	2,330,319	8,499	69.66	42.45	6.47
Bin_19_5	Thermomonas sp.	<u>SSFC00000000</u>	1,187,954	11,860	67.14	38.13	0.72
Bin_2_1	Unclassified  Thermomicrobiales family UBA6265 bacterium	SSDT00000000	2,680,321	10,114	60.7	49.64	7.91
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Bin name	Taxonomy <u>a</u>	GenBank assembly accession no.	Total length (bp)	N <sub>50</sub> (bp)	GC content (%)	Completeness (%)	Redund
Bin_20_1	Niabella sp.	<u>SSFZ00000000</u>	1,214,478	6,588	38.77	33.81	7.91
Bin_21_1	Zoogloea sp.	<u>SSDR00000000</u>	2,929,105	10,734	64.98	55.4	1.44
Bin_21_3	Limnohabitans sp. AWPF1	<u>SSGJ00000000</u>	1,670,846	9,983	63.22	19.42	0.72
Bin_23_2	Flavobacterium GCA_002422095.1	<u>SSGM00000000</u>	1,906,623	10,479	32.59	66.19	9.35
Bin_23_3	Flavobacterium sp.	<u>SSGL00000000</u>	2,075,117	12,249	32.14	33.09	2.88
Bin_24_1	Rhodoferax sp.	<u>SSFF00000000</u>	3,654,053	136,395	62.3	99.28	7.19
Bin_24_2	Limnohabitans sp. AWPF2	<u>SSGI00000000</u>	2,967,058	22,882	63.25	73.38	5.04
Bin_24_4	Pseudorhodobacter sp.	<u>SSFK00000000</u>	3,302,298	8,369	63.29	49.64	10.79
Bin_25_1	Unclassified  Rhodocyclaceae bacterium AWPF1	SSDY00000000	2,178,082	10,805	64.25	62.59	2.16
Bin_25_2	Flavobacteriales PHOS-HE28 sp.	SSGN00000000	4,081,487	21,145	62.07	76.98	7.91
Bin_25_4	Dechloromonas sp.	<u>SSGP00000000</u>	3,029,215	19,114	62.22	58.99	5.76
Bin_26_1	Unclassified  Sphingomonadales bacterium	SSDV000000000	3,053,326	60,967	64.22	99.28	0.72
Bin_26_2	Mycobacterium mageritense	<u>SSGC00000000</u>	7,650,754	60,194	66.88	79.14	5.76
Bin_26_3	Unclassified  Dongiaceae  bacterium	SSEL00000000	5,719,975	142,361	61.3	99.28	5.04
Bin_27_1	Thauera aminoaromatica	SSFD000000000	5,236,562	14,873	67.89	73.38	8.63
Bin_27_2	Unclassified  Burkholderiaceae	SSER000000000	3,032,333	8,502	67.32	22.3	0.72

Bin	Taxonomy <u>a</u>	GenBank	Total	N <sub>50</sub> (bp)	GC	Completeness	Redunc
name		assembly	length		content	(%)	(%
		accession no.	(bp)		(%)		
	bacterium						
Bin_29_1	Unclassified	<u>SSEC00000000</u>	6,629,733	18,228	66.91	85.61	7.91
	<i>Mycobacterium</i> bacterium						
Din 20 2	Mycobacterium	SSGD00000000	5,009,268	45 207	66.94	99.28	7.19
DIII_29_2	arupense	<u>550D0000000</u>	3,009,208	43,297	00.94	99.20	7.19
Bin_3_3	Chryseobacterium	<u>SSGV00000000</u>	2 503 246	11 245	36.61	22.3	0
	cucumeris		_,,	,			
Bin_3_4	Unclassified	<u>SSEO00000000</u>	1,501,328	17,275	33.51	50.36	0.72
	Chitinophagaceae						
	bacterium						
Bin_3_8	Unclassified	<u>SSEU00000000</u>	2,309,706	6,910	35.54	57.55	19.42
	Bacteroidia bacterium						
Bin_30	Unclassified  Burkholderiaceae	<u>SSEQ00000000</u>	6,171,595	8,294	70.29	47.48	4.32
	bacterium AWPF1						
Bin 31 2	Aeromicrobium sp.	SSHC00000000	1,861,791	18,902	58.23	82.73	1.44
	Unclassified	SSEM00000000			58.83	3.6	5.04
B.II3 .	Desulfurella		0,200,000	10,201	20.02	3.0	2.01
	bacterium						
Bin_35_1	Pseudomonas	<u>SSFN00000000</u>	5,048,709	26,274	63.26	51.8	2.16
	monteilii						
Bin_35_2	Thiobacillus	<u>SSFB00000000</u>	3,564,805	105,954	62.36	86.33	5.76
	GCA_002343685.1						
Bin_35_3	Unclassified	SSEB00000000	3,246,569	241,139	65.99	86.33	2.16
	Nevskiaceae bacterium AWPF1						
Rin 36 2	Crocinitomicaceae	SSGS00000000	1,134,521	8 082	36.56	17.27	0
DIII_30_2	UBA5422 sp. AWPF1	000000000000000000000000000000000000000	1,134,341	0,002	30.30	11.41	U
	1	l	l		I		

Bin	Taxonomy <u>a</u>	GenBank	Total	N <sub>50</sub> (bp)	GC	Completeness	Redun
name		assembly	length		content	(%)	(%
Bin_38_1	Nitrosomonas sp. AWPF1	accession no.  SSFV000000000	( <b>bp</b> ) 2,613,687	22,274	(%) 44.56	57.55	0
Bin_39_1	Pelomonas sp.	<u>SSFQ00000000</u>	3,131,775	9,407	69.9	30.22	2.16
Bin_4_1	Cupriavidus sp.	<u>SSGQ00000000</u>	3,464,703	7,277	64.08	16.55	2.88
Bin_4_3	Rhizobium sp. AWPF1	SSFH00000000	4,147,977	10,631	61.05	18.71	1.44
Bin_42_2	Methylophilus methylotrophus	<u>SSGG00000000</u>	1,188,656	14,601	49.11	28.06	2.88
Bin_43_1	Chryseobacterium sp.	<u>SSGU00000000</u>	2,092,908	10,855	38.01	44.6	7.91
Bin_43_5	Crocinitomicaceae UBA5422 sp. AWPF2	<u>SSGR00000000</u>	641,979	9,434	42.11	35.25	17.27
Bin_44_1	Betaproteobacteriales <u>UBA11063</u> sp.  AWPF1	<u>SSGY00000000</u>	2,803,113	14,546	36.96	86.33	2.16
Bin_45_1	Betaproteobacteriales UBA11063 sp. AWPF2	SSGX00000000	2,467,391	70,519	35.22	79.14	1.44
Bin_45_4	"Candidatus Gracilibacteria" UBA5532 sp.	SSGK00000000	668,037	10,255	37.87	58.27	0.72
Bin_48_1	Afipia sp.	<u>SSHB00000000</u>	3,379,245	12,763	61.87	54.68	2.88
Bin_48_2	Rhizobium sp. AWPF2	<u>SSFG00000000</u>	2,353,640	12,778	60.82	38.13	0.72
Bin_5_1	Lysobacter sp.	<u>SSGH00000000</u>	4,387,597	162,300	65.61	94.24	5.76
Bin_5_5	Unclassified  Xanthomonadaceae bacterium	SSEV000000000	3,535,438	16,226	63.31	47.48	0.72
Bin_5_7	Unclassified  Rhodocyclaceae bacterium AWPF2	SSDX00000000	1,708,980	12,103	65.16	33.09	0.72

Bin name	Taxonomy <u>a</u>	GenBank assembly	Total length	N <sub>50</sub> (bp)	GC content	Completeness (%)	Redund
паше		accession no.	(bp)		(%)	(70)	( >0
Bin_51_4	Unclassified Saccharimonadales UBA4665	SSDW00000000	818,540	13,215	50.23	61.87	8.63
Bin_52_1	Pseudomonas alcaligenes	<u>SSFO00000000</u>	3,682,067	12,539	64.72	71.22	3.6
Bin_52_2	Pseudomonas sp. AWPF1	SSFM00000000	1,540,019	11,651	61.81	17.27	4.32
Bin_54_1	Nitrosomonas oligotropha	<u>SSFX00000000</u>	2,754,397	36,184	49.22	67.63	0.72
Bin_54_3	Nitrosomonas GCA_002083595.1	<u>SSFY00000000</u>	2,292,023	8,445	48.32	29.5	6.47
Bin_55_1	46-32 GCA 001898405.1	<u>SSHF00000000</u>	5,043,020	214,644	44.27	94.96	0
Bin_55_2	Methylophilus sp.	<u>SSGF00000000</u>	2,747,675	94,685	50.49	66.91	2.16
Bin_56_1	Thiothrix sp.	SSFA00000000	4,155,808	145,664	44.77	69.78	3.6
Bin_56_2	Pedobacter sp.	SSFR00000000	2,341,987	206,071	38.94	98.56	0.72
Bin_57_1	Rheinheimera sp.	<u>SSFI00000000</u>	3,863,184	45,966	52.26	63.31	2.88
Bin_57_2	UBA7239 sp.	<u>SSEW00000000</u>	1,251,491	7,537	52.91	38.13	9.35
Bin_59_2	Unclassified  Nevskiaceae bacterium AWPF2	SSEA00000000	2,177,696	27,134	59.28	27.34	5.04
Bin_63_2	Unclassified WS6 bacterium	<u>SSDS00000000</u>	1,242,531	11,680	44.02	34.53	9.35
Bin_64_3	Romboutsia sp.	<u>SSFE00000000</u>	381,413	7,806	27.61	45.32	0
Bin_65	Crocinitomicaceae 40-80 sp.	<u>SSGT00000000</u>	2,395,197	12,217	40.24	70.5	2.16
Bin_67	Unclassified  Spirochaetes class  UBA12135 bacterium	SSDU000000000	1,932,890	10,511	31.46	5.04	0

Bin	Taxonomy <u>a</u>	GenBank	Total	N <sub>50</sub> (bp)	GC	Completeness	
name		assembly accession no.	length (bp)		content (%)	(%)	(%
Bin_68_2	Unclassified  Moranbacterales class UBA1568 bacterium	SSEF00000000	1,023,203	34,589	53.99	79.14	5.76
Bin_7_2	Unclassified  Burkholderiaceae bacterium AWPF2	SSEP00000000	2,384,147	7,064	63.49	12.95	0.72
Bin_7_3	Alicycliphilus sp.	<u>SSHA00000000</u>	3,015,590	10,496	66.2	43.17	5.76
Bin_7_4	Aquabacterium sp. AWPF1	<u>SSGZ00000000</u>	1,352,035	7,819	65.11	21.58	0
Bin_7_5	Pseudomonas sp. AWPF2	<u>SSFL00000000</u>	1,978,447	9,650	63.83	51.08	10.79
Bin_7_6	Aquabacterium sp. AWPF2	<u>VKOJ00000000</u>	2,228,505	10,843	63.58	33.81	2.16
Bin_71	Unclassified  Leptospira bacterium	SSEG00000000	1,795,065	38,625	54.29	6.17	0
Bin_8_1	Acinetobacter sp. AWPF1	SSHE00000000	2,045,996	23,569	41.75	75.54	2.16
Bin_8_2	Thiothrix sp. AWPF1	<u>SSEZ00000000</u>	1,300,697	7,394	44.99	30.22	1.44
Bin_8_3	Thiothrix sp. AWPF2	<u>SSEY00000000</u>	2,137,450	7,937	45.31	47.48	16.55
Bin_8_5	Acinetobacter sp. AWPF2	SSHD000000000	1,422,583	9,315	42.74	74.1	7.19
Bin_9_3	Unclassified  Elusimicrobia order  F11 bacterium	SSEK00000000	2,256,510	6,750	63.91	51.8	10.79
Bin_9_4	Dokdonella sp.	<u>SSGO00000000</u>	1,632,646	10,049	62.2	31.65	2.16

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<sup>&</sup>lt;sup>a</sup>Taxonomy given was identified using GTDB-tk and is the taxonomy reported within the NCBI accession

record.

## Data availability.

Raw sequence reads are available under BioProject accession number <u>PRJNA428383</u>. Whole-genome sequences are available under the sequential BioSample accession numbers <u>SAMN10026417</u> to <u>SAMN10026511</u>, which include annotations produced with the Prokaryotic Gene Annotation Pipeline (PGAP). <u>Table 1</u> contains individual Web links to each bin assembly and annotation.

#### **ACKNOWLEDGMENTS**

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#### REFERENCES

- 1. Moss B. 2011. Allied attack: climate change and eutrophication. Inland Waters 1:101–105. doi: 10.5268/ IW-1.2.359. [DOI ] [Google Scholar ]
- 2. Garner E, Chen C, Xia K, Bowers J, Engelthaler DM, McLain J, Edwards MA, Pruden A. 2018. Metagenomic characterization of antibiotic resistance genes in full-scale reclaimed water distribution systems and corresponding potable systems. Environ Sci Technol 52:6113–6125. doi: 10.1021/acs.est.7b05419.

  [DOI ] [PubMed] [Google Scholar ]
- 3. Leddy MB, Hasan NA, Subramanian P, Heberling C, Cotruvo J, Colwell RR. 2017. Characterization of microbial signatures from advanced treated wastewater biofilms. J Am Water Works Assoc 109:E503–E512. doi: 10.5942/jawwa.2017.109.0116. [DOI ] [Google Scholar ]
- 4. Stamps BW, Leddy MB, Plumlee MH, Hasan NA, Colwell RR, Spear JR. 2018. Characterization of the microbiome at the world's largest potable water reuse facility. Front Microbiol 9:2435. doi: 10.3389/fmicb.2018.02435. [DOI ] [PMC free article] [PubMed] [Google Scholar ]

- 5. Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol 19:455–477. doi: 10.1089/cmb.2012.0021. [DOI ] [PMC free article] [PubMed] [Google Scholar ]
- 6. Li D, Liu CM, Luo R, Sadakane K, Lam TW. 2015. MEGAHIT: an ultra-fast single-node solution for large and complex metagenomics assembly via succinct de Bruijn graph. Bioinformatics 31:1674–1676. doi: 10.1093/bioinformatics/btv033. [DOI ] [PubMed] [Google Scholar ]
- 7. Langmead B, Salzberg SL. 2012. Fast gapped-read alignment with Bowtie 2. Nat Methods 9:357–359. doi: 10.1038/nmeth.1923. [DOI ] [PMC free article] [PubMed] [Google Scholar ]
- 8. Alneberg J, Bjarnason BS, de Bruijn I, Schirmer M, Quick J, Ijaz UZ, Lahti L, Loman NJ, Andersson AF, Quince C. 2014. Binning metagenomic contigs by coverage and composition. Nat Methods 11:1144–1146. doi: 10.1038/nmeth.3103. [DOI ] [PubMed] [Google Scholar ]
- 9. Eren AM, Esen ÖC, Quince C, Vineis JH, Morrison HG, Sogin ML, Delmont TO. 2015. Anvi'o: an advanced analysis and visualization platform for 'omics data. PeerJ 3:e1319. doi: 10.7717/peerj.1319.

  [DOI ] [PMC free article] [PubMed] [Google Scholar ]
- 10. Arango-Argoty G, Garner E, Pruden A, Heath LS, Vikesland P, Zhang L. 2018. DeepARG: a deep learning approach for predicting antibiotic resistance genes from metagenomic data. Microbiome 6:23. doi: 10.1186/s40168-018-0401-z. [DOI ] [PMC free article] [PubMed] [Google Scholar ]
- 11. Parks DH, Chuvochina M, Waite DW, Rinke C, Skarshewski A, Chaumeil P-A, Hugenholtz P. 2018. A standardized bacterial taxonomy based on genome phylogeny substantially revises the tree of life. Nat Biotechnol 36:996–1004. doi: 10.1038/nbt.4229. [DOI ] [PubMed] [Google Scholar ]
- 12. Rinke C, Schwientek P, Sczyrba A, Ivanova NN, Anderson IJ, Cheng J-F, Darling A, Malfatti S, Swan BK, Gies EA, Dodsworth JA, Hedlund BP, Tsiamis G, Sievert SM, Liu W-T, Eisen JA, Hallam SJ, Kyrpides NC, Stepanauskas R, Rubin EM, Hugenholtz P, Woyke T. 2013. Insights into the phylogeny and coding potential of microbial dark matter. Nature 499:431–437. doi: 10.1038/nature12352. [DOI ] [PubMed] [Google Scholar ]
- 13. Liu Y, Jin JH, Liu YH, Zhou YG, Liu ZP. 2010. *Dongia mobilis* gen. nov., sp. nov., a new member of the family *Rhodospirillaceae* isolated from a sequencing batch reactor for treatment of malachite green effluent. Int J Syst Evol Microbiol 60:2780–2785. doi: 10.1099/ijs.0.020347-0. [DOI ] [PubMed] [Google Scholar ]

### **Associated Data**

This section collects any data citations, data availability statements, or supplementary materials included in this article.

# Data Availability Statement

Raw sequence reads are available under BioProject accession number <u>PRJNA428383</u>. Whole-genome sequences are available under the sequential BioSample accession numbers <u>SAMN10026417</u> to <u>SAMN10026511</u>, which include annotations produced with the Prokaryotic Gene Annotation Pipeline (PGAP). <u>Table 1</u> contains individual Web links to each bin assembly and annotation.

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