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## A member of the CONSTANS-Like protein family is a putative regulator of reactive oxygen species homeostasis and spaceflight physiological adaptation

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### Abstract

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A feature of the physiological adaptation to spaceflight in *Arabidopsis thaliana* (*Arabidopsis*) is the induction of reactive oxygen species (ROS)-associated gene expression. The patterns of ROS-associated gene expression vary among *Arabidopsis* ecotypes, and the role of ROS signalling in spaceflight acclimation is unknown. What could differences in ROS gene regulation between ecotypes on orbit reveal about physiological adaptation to novel environments? Analyses of ecotype-dependent responses to spaceflight resulted in the elucidation of a previously uncharacterized gene (*OMG1*) as being ROS-associated. The *OMG1* 5' flanking region is an active promoter in cells where ROS activity is commonly observed, such as in pollen tubes, root hairs, and in other tissues upon wounding. qRT-PCR analyses revealed that upon wounding on Earth, *OMG1* is an apparent transcriptional regulator of *MYB77* and *GRX480*, which are associated with the ROS pathway. Fluorescence-based ROS assays show that *OMG1* affects ROS production. Phylogenetic analysis of *OMG1* and closely related homologs suggests that *OMG1* is a distant, unrecognized member of the CONSTANS-Like protein family, a member that arose via gene duplication early in the angiosperm lineage and subsequently lost its first DNA-binding B-box1 domain. These data illustrate that members of the rapidly evolving COL protein family play a role in regulating ROS pathway functions, and their differential

regulation on orbit suggests a role for ROS signalling in spaceflight physiological adaptation.

**Keywords:** *Arabidopsis thaliana*, B-box domain, CONSTANS-Like, ecotype-specific, microgravity, reactive oxygen species, spaceflight, transcription factor, uncharacterized genes

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Plants adjust to spaceflight by changing patterns of gene expression. The analysis of how different *Arabidopsis* plants respond to space resulted in the discovery of a previously uncharacterized gene (*OMG1*), which was consistently induced in space. This research revealed that OMG1 belongs to the CONSTANS-Like protein family and is involved in maintaining the reactive oxygen species pathway, which appears to be an important feature of the physiological adaptation of plants to the spaceflight environment. It also shows that spaceflight experiments can be used as a platform to discover novel gene function and provide further insights into terrestrial plant biology.

## Introduction

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The complex networks and signalling pathways that equip plants to cope with environmental challenge developed over evolutionary time in terrestrial environments. Exposing plants to novel environments presents opportunities to understand how plants adjust to conditions outside their evolutionary experience. Spaceflight is one such novel environment, and understanding how plants function outside of Earth's boundaries is also integral to space exploration. Plants respond to spaceflight in ways that are dependent on species, ecotype, genetics and even organs ([Cowles et al. 1984](#); [Hoson et al. 2002](#); [Soga et al. 2002](#); [Paul et al. 2013a](#); [Ferl et al. 2014](#)). Indeed, even a single gene difference can influence the spaceflight response ([Paul et al. 2017](#)), thus investigating ecotype-dependent spaceflight transcriptomes can reveal strategies for physiological adaptation to spaceflight while expanding understanding of gene function in terrestrial environments.

The physiological impact of spaceflight is reflected in the patterns of gene expression. Irrespective of plant growth hardware, duration in space or age of plants, one consensus is that spaceflight elicits responses from pathways associated with cell wall remodelling, cell polarity, phytohormone-mediated processes, as well as general stress responses, such as wounding, pathogen defence, heat shock and reactive oxygen species (ROS) ([Joo et al. 2001](#); [Paul et al. 2005](#), [2012b](#), [2013a](#); [Correll et al. 2013](#); [Zupanska et al. 2013](#); [Ferl et al. 2014](#); [Mazars et al. 2014](#); [Sugimoto et al. 2014](#); [Zhang et al. 2014](#); [Kwon et al. 2015](#); [Schüler et al. 2015](#)). In addition, a large proportion of genes differentially expressed in spaceflight encode unknown proteins.

Spaceflight-grown *Arabidopsis* roots tend to be smaller, have fewer lateral roots and shorter root hair development than their terrestrial controls ([Paul et al. 2012a](#); [Kwon et al. 2015](#)). Aspects of these morphologies in ecotype Columbia (Col-0) suggest a relationship between the down-regulation of several peroxidase genes in spaceflight ([Kwon et al.](#)

[2015](#)).

In this study, a spaceflight-induced gene of unknown function identified from roots was investigated for its potential involvement in ROS-related functions. AT1G05290, which we have dubbed *Orbitally Manifested Gene 1 (OMG1)*, was one of the highest significantly ( $P = 0.0005$ ) induced genes in the Advanced Plant Experiment 01 (APEX01) spaceflight data set (fold change = 12) ([Paul et al. 2013a](#)). Two distinct biochemical assays typically used to measure ROS activity indicated that knocking out OMG1 alters the production of ROS. Co-expression networks around *OMG1* show correlations to genes involved in ROS-associated oxidative phosphorylation pathways [Obayashi et al. 2014](#). *OMG1* was also expressed in organs known to highly utilize ROS during development, such as root hairs and pollen tubes ([Shin and Schachtman 2004](#); [Potocký et al. 2007](#); [Winter et al. 2007](#); [Lassig et al. 2014](#)). In addition, *OMG1* is an unrealized member of the CONSTANS-Like family of transcription factors. These findings, originating with the novel environment of spaceflight, further the understanding of plant growth and development and provide insight into the role of a heretofore unknown gene.

## Materials and Methods

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### ROSMETER analysis

The ROSMETER tool assesses the ROS-related transcriptomic signature in a large-scale *Arabidopsis* gene expression data set ([Rosenwasser et al. 2013](#)). *Arabidopsis* gene expression profiles from two independent spaceflight experiments, APEX01 and Advanced Plant Experiment 03-2 (APEX03-2), along with their corresponding ground controls, were used to correlate differential gene expression in spaceflight with specific ROS species and origins. The APEX01 experiment utilized two ecotypes and both were planted on each plate of the experiment. Since the entire plate was harvested on orbit to a single tube, the subsequent transcriptomic analyses comprised 60 % Wassilewskija (WS) and 40 % Col-0 ([Paul et al. 2013b](#)). In the APEX03-2 data set, WS and Col-0 were kept on separate plates, harvested into separate tubes and analysed individually. A data set from Col-0 compared to WS vertically grown in standard laboratory conditions was included as an additional control ([Schultz et al. 2017](#)). From each data set, Affymetrix Probeset-ID, Gene Locus ID,  $P$ -value and fold change were listed and submitted to ROSMETER. The heatmap was constructed using the GENE-E software with hierarchical clustering using the Matrix value offered in the software.

### Plant materials and growth conditions

Seeds from both Col-0 and WS were prepared for APEX01 and APEX03-2 as previously described ([Paul et al. 2012b, 2013b](#); [Sng et al. 2014](#); [Fert and Paul 2016](#)). Briefly, plants were cultured on Phytigel™ 10 cm Petri plates and grown either in the Advanced Biological Research System (ABRS) hardware (APEX01), or in the Veggie hardware (APEX03-2) under continuous lighting of  $\sim 80\text{--}135 \mu\text{mol m}^{-2} \text{s}^{-1}$  PAR. In this study, the effects of the spaceflight

environment were evaluated, which includes ambient radiation, microgravity and attending effects on convection. In both experiments, the ground control plants were grown with a 24- to 48-h schedule delay that allowed the comparable ground growth chambers to be programmed with the precise environmental conditions that the International Space Station (ISS)-grown plants received on orbit.

Plants grown for additional laboratory analyses were grown on Phytigel™ 10 cm plates or in soil, and placed in a greenhouse growth chamber on a 24-h light cycle ( $\sim 80 \mu\text{mol m}^{-2} \text{s}^{-1}$  PAR) at 22 °C.

## OMG1 promoter::GUSmgfp5 transgenic plants and analyses

A 0.8-kb 5' flanking region upstream of the OMG1 (AT1G05290) initial Met was amplified from *Arabidopsis* gDNA using primer pair A [see [Supporting Information—Table S1](#)], which contains restriction enzyme sites BamHI in the forward primer and NcoI in the reverse. The sequenced and confirmed BamHI/NcoI fragment containing the OMG1 5' flanking region was cloned into pCAMBIA1303 to create the OMG1p::GUSmgfp5 fusion construct. Although this region may not represent the full complement of motifs that regulate OMG1, the 0.8-kb 5' flanking region does serve to promote OMG1 transcription and will be referred to as the basal promoter. The construct was then transformed into *Agrobacterium* (*Agrobacterium tumefaciens*) strain GV3101 for floral dip transformation into WS plants ([Clough and Bent 1998](#)). Ten independent OMG1p::GUSmgfp5 transgenic lines grown on selection media (250 mg L<sup>-1</sup> Carbenicillin and 50 mg L<sup>-1</sup> Hygromycin) were screened for T3 homozygosity. Plants grown on Phytigel plates were used for histochemical localization of  $\beta$ -glucuronidase (GUS) ([Jefferson et al. 1987](#)).

Pollen and pollen tube analyses were performed as described ([Li et al. 1999](#)). T1 heterozygous populations of pollen grains and tubes were incubated with X-Gluc to reveal the specificity of the histochemical localization in transformed pollen. Basal OMG1 promoter activity in plants requires  $\sim 3$  h of staining before detection of GUS in the various locations, whereas GUS activity induced by wounding was observed within 30 min of staining.

In the GUS qRT-PCR wounding experiments with OMG1p::GUSmgfp5, comparable leaves from three individual plants were crimped with forceps and left intact on the plant for the duration of each time point (30 min, 1, 4 and 16 h). Leaves were harvested to liquid nitrogen and stored at  $-80$  °C until RNA extraction. In the mock and immediate time point, leaves were either unwounded or crimped and immediately frozen in liquid nitrogen. Total RNA was extracted using RNAeasy (Qiagen) according to the manufacturer's instructions. cDNA transcribed from total RNA was used to analyse the transcript abundance of *GUSA*. Primer pairs B and C [see [Supporting Information—Table S1](#)] were used for the analyses. *GUSA* transcript abundance was normalized to UBQ11 (AT4G05050).

## Gene expression analyses of OMG1 and other downstream genes

OMG1 RT-qPCR analyses used TaqMan™ (Applied Biosystems) on spaceflight material from APEX01 and APEX03-2. *OMG1* gene expression was normalized to *UBQ11* primer and probe sequences D and E [see [Supporting Information—Table S1](#)]. The *omg1* knockout (KO) line (SALK\_045742C) was validated for a homozygous T-DNA insertion using primers F. Homozygous lines were selfed until all progeny were homozygous and screened for the absence of full-length *OMG1* transcript [see [Supporting Information—Fig. S1](#)]. RT-qPCR analyses of *OMG1* expression induced by a time course wounding assay were performed on wild-type (WT) Col-0, WS and *omg1* lines. Three individual plants with leaves of similar ages and sizes were wounded by crushing between two glass slides. The wounded leaves were left attached to the plant for the allotted times (mock, immediate, 5, 10, 30 min and 1 h) before being harvested into liquid nitrogen and stored at  $-80^{\circ}\text{C}$ . RT-qPCR analysis was conducted using SYBR Green master mix (Applied Biosystems) along with the primer pairs C, D (without probe), G–H and L–R [see [Supporting Information—Table S1](#)].

## Fluorescent biochemical ROS assays

For the ROS accumulation assay, intact leaves from WT Col-0, *omg1* KO and *rbohD* KO (SALK\_070610) were wounded with a 10  $\mu\text{L}$  pipette tip and immediately immersed into the fluorescent probe 5-(and 6)-carboxy-2',7'-dichlorodihydrofluorescein diacetate (DCF-DA) (Sigma) solution as described in ([Beneloujaephajri et al. 2013](#)). Briefly, wounded and unwounded leaves were vacuum infiltrated in the dark for 20 min in 60  $\mu\text{M}$  of DCF-DA in standard buffer. Leaves were then rinsed in standard buffer and observed under GFP bandpass filters using the Olympus BX51 compound light microscope at  $10\times$  magnification with 55 ms exposure time. Quantification of fluorescence intensity using ImageJ software was previously described ([Jakic et al. 2017](#)).

A luminol-based ROS assay was done by measuring hydrogen peroxide produced by leaf discs when exposed to flg22. Discs ( $0.125\text{ cm}^2$ ) of 6-week-old WT Col-0, *omg1* and *rbohD* plants were incubated in 100  $\mu\text{L}$  ddH<sub>2</sub>O overnight in 96-well luminometer plates. Water was replaced with the reaction solution of 40  $\mu\text{M}$  luminol (Sigma-Aldrich) and 10  $\mu\text{g mL}^{-1}$  horseradish peroxidase (Sigma-Aldrich) supplemented with 50 nM of flg22 or water for the mock controls. ROS measurements expressed as means of RLU (relative light units) were taken at 2-min intervals for a period of 38 min.

## Phylogenetic analysis

Protein sequences were identified by rpsblast search of the NR database ([Marchler-Bauer and Bryant 2004](#); [Marchler-Bauer et al. 2015](#); [NCBI Resource Coordinators 2016](#)); full-length protein sequences containing a single CCT domain (pfam06203) with  $e\text{-value} < 0.01$  were considered potential OMG1 homologs. Protein sequences were aligned using Clustal Omega v1.2.3 ([Sievers et al. 2011](#)), PROBALIGN ([Roshan and Livesay 2006](#)), MUSCLE v3.8.31 ([Edgar 2004](#)), MSAProbs v0.97 ([Liu et al. 2010](#)), PROBCONS ([Do et al. 2005](#)) and mafft-einsi v7.215 ([Kato and Standley 2013](#)), with default parameters. Alignments were left unprocessed or processed by Gblocks v0.91 to remove potentially

ambiguous regions ([Talavera and Castresana 2007](#)). The minimum number of sequences for a flank position (-b2) was 3/5 the total number of sequences. The maximum number of contiguous non-conserved positions (-b3) was 10. The minimum block length (-b4) was 5, and gap positions were allowed (-b5 = a). Other Gblocks parameters were left at default values. Initial maximum likelihood phylogenies were constructed from each alignment using FastTree v2.1.7 with default parameters ([Price et al. 2010](#)). Initial trees were used as starting trees for full maximum likelihood reconstruction using RAxML v8.2.8 ([Stamatakis 2014](#)). Maximum likelihood phylogenies produced from each alignment were converted to a clade presence-absence matrix using the Super Tree Toolkit v0.1.2 ([Hill and Davis 2014](#)), and a supertree was inferred from this matrix using the BINCAT model in RAxML ([Nguyen et al. 2012](#)). Additionally, all individual sequence alignments were concatenated into a single supermatrix, which was used to infer the maximum likelihood protein family phylogeny using RAxML, with the best-fit evolutionary model selected by AIC ([Wheeler et al. 1995](#)). A consensus of ‘supertree’ and ‘supermatrix’ results was plotted using FigTree v1.4.3.

Protein structural modelling of the B-box2 domain was done using both the Phyre2 and Chimera ([Pettersen et al. 2004](#)). The closest OMG1 homolog, c2junA of the human Midline-1 (MID1) with a 92 % confidence level was used to predict the overall folding of the B-box domains in OMG1. The predicted structural model of OMG1 was compared to the x-ray crystal structure of c2junA using Chimera and MatchMaker. Side chains positioned near the zinc ions were displayed to predict the potential of OMG1 binding to zinc ions.

## 35S promoter:OMG1cds-sGFPS65T fusion construct for subcellular localization

The coding sequence of OMG1 (RefSeq ID:[NP\\_172021.2](#)) was amplified from cDNA using primer pair I [[see Supporting Information—Table S1](#)]. The PCR fragment was sequenced before being cloned into the pCAMBIA1303 along with sGFP(S65T) ([Chiu et al. 1996](#)) gene via SpeI and BstEII restriction sites using primer pairs J [[see Supporting Information—Table S1](#)]. Primer pair K was then used to amplify the entire OMG1cds-sGFP(S65T), which was validated by sequencing and cloned into pCAMBIA1302 vector via NcoI and BstEII restriction sites. For subcellular localization, *Agrobacterium*-GV3101 containing 100 µM acetosyringone, 0.05 % Silwet-77 and the pCAMBIA1302 vector or pCAMBIA1302-OMG1cds-sGFPS65T fusion vector were incubated in the dark for 20 min at 28 °C. Leaves of 4-week-old tobacco (*Nicotiana benthamiana*) seedlings were injected and GFP signal was observed after 2 days at 25 °C in the dark. DAPI was used to indicate the nucleus.

## Results

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### ROS-producing genes are differentially expressed in response to spaceflight in an ecotype-specific manner

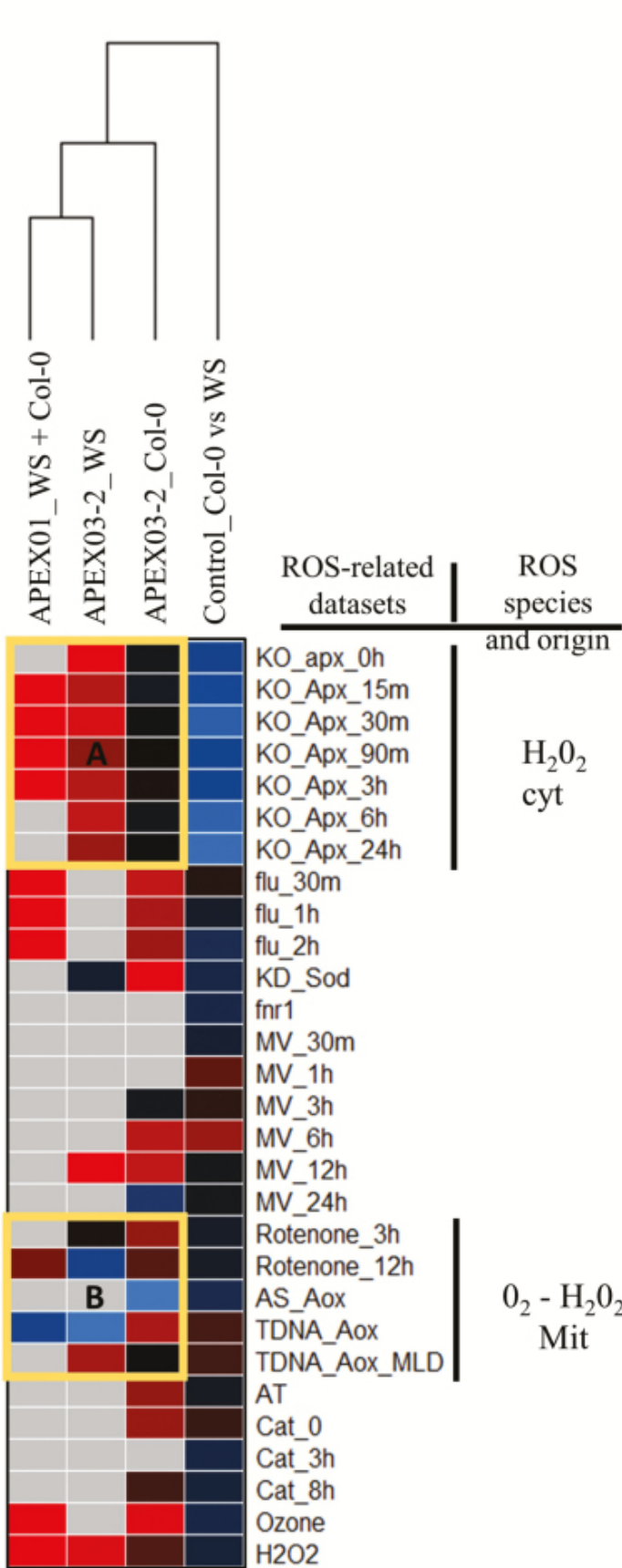
The differential gene expression data sets of the three independent spaceflight experiments and their comparable ground

controls were used in the ROSMETER analyses. The heatmap shows the positive-(red), negative-(blue) and no correlation-(black) between the ROS indices and the queried spaceflight transcriptomes ([Fig. 1A](#)). A general positive correlation of ROS-producing genes was observed in all the spaceflight transcriptomes and hierarchical clustering displayed a higher similarity with the APEX03-2\_WS data set compared to the APEX03-2\_Col-0 data set. Two interesting ROS correlation groups were observed in the heatmap. Group A showed that only the spaceflight transcriptome of WS positively correlated with the cytoplasmic ascorbate peroxidase knockout (KO-*APX1*) experiments, which represent cytoplasmic H<sub>2</sub>O<sub>2</sub> ROS indices, whereas in the Col-0 background cytosolic ROS shows no correlation. Group B showed ecotype-related differences in the correlations associated with mitochondrial stress. Specifically, TDNA-AOX-MLD and Retenone-3h indices showed that if one ecotype had a positive correlation the other had no correlation. In addition, in the Retenone-12h and TDNA-AOX indices, the mixed ecotype transcriptome shows a similar trend to one of the two ecotypes. The control data set, which reflects terrestrial ecotype differences between Col-0 and WS, showed that many of the genes differentially expressed between the two ecotypes are associated with the ROS-scavenging mechanism. A representative transcriptome heatmap from the APEX03-2 spaceflight experiment that compared several significantly ( $P < 0.01$ ) differentially expressed ( $>1$  or  $-1 < \log_2$  fold change) peroxidase genes between 4-day-old Col-0 and WS ecotypes illustrated that Col-0 alters the expression of many more peroxidase genes in spaceflight than does WS ([Fig. 1B](#)). Peroxidases, an integral part of ROS signalling in cell wall remodelling, cell elongation, auxin catabolism, and response to abiotic and biotic stress, ([Shigeto and Tsutsumi 2016](#)) were found to be differentially expressed in spaceflight.

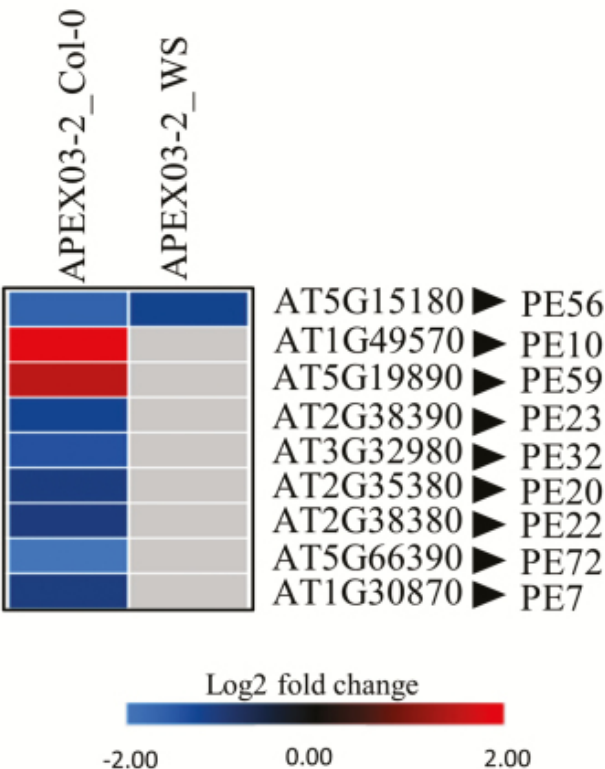
Figure 1.



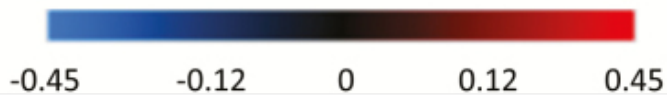
**A** ROS Correlation



**B** Gene Expression



Negative Correlations Positive



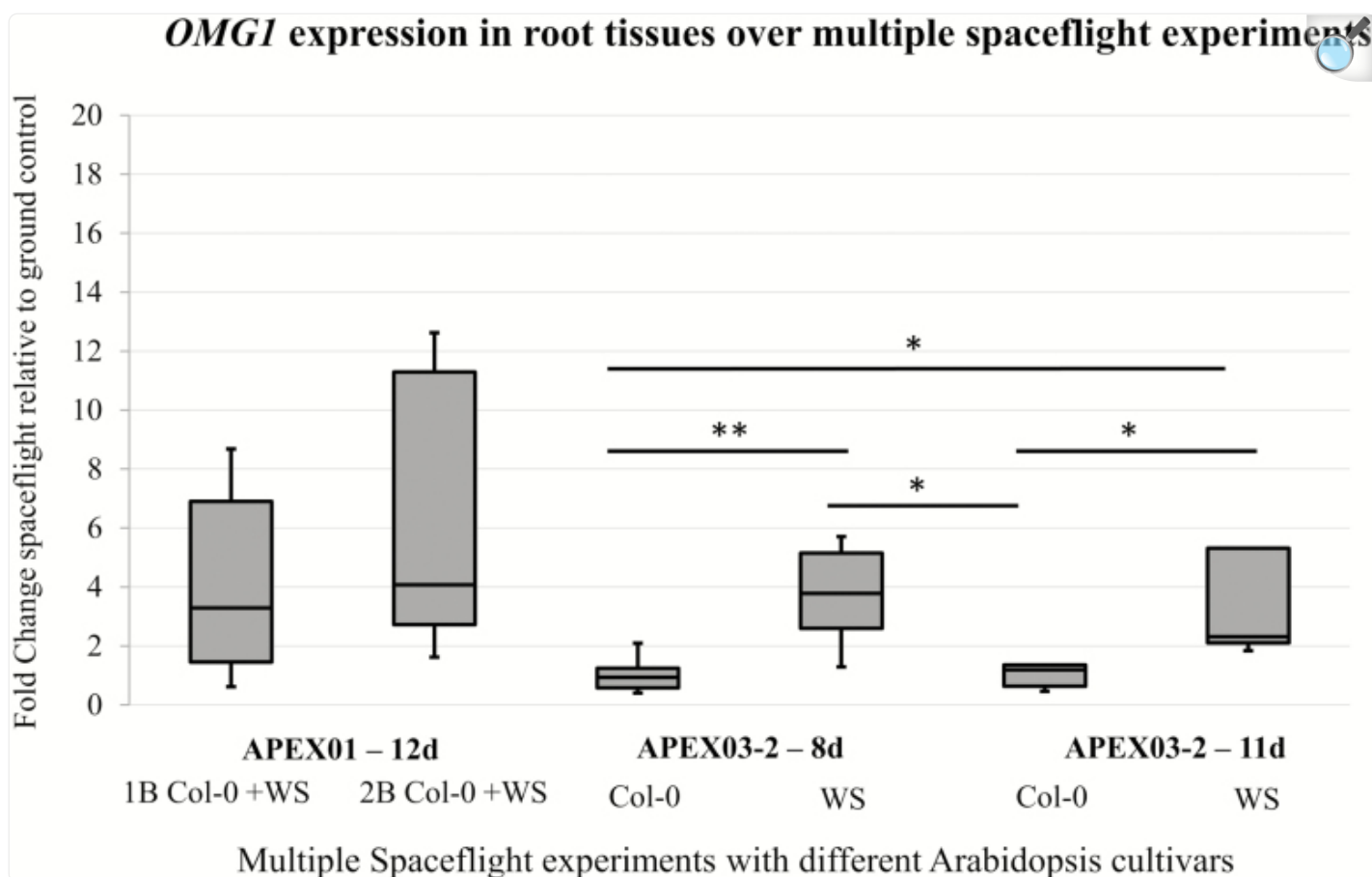
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Correlations between spaceflight transcriptome and ROS-related microarrays revealed that ROS is induced in spaceflight in an ecotype-dependent manner. (A) The microarray (APEX01) and the RNAseq (APEX03-2) analyses of spaceflight transcriptional profile relative to the ground control profile were correlated to known ROS-related data sets producing specific ROS species from specific locations within the cell using the bioinformatics tool called ROSMETER ([Rosenwasser et al. 2013](#)). Positive correlations (red) show ROS production, whereas negative correlations (blue) show ROS scavenging. Black cells suggest no correlations, whereas gray cells indicate that there are insufficient gene numbers for a statistical correlation. Clusters highlighted in yellow boxes denote regions of interest. Group A showed that only the spaceflight transcriptome of WS positively correlated with the cytoplasmic ascorbate peroxidase knockout (KO-*APX1*) experiments, which represent cytoplasmic H<sub>2</sub>O<sub>2</sub> ROS indices. Group B showed ecotype-related differences in the correlations associated with mitochondrial stress. (B) Peroxidase genes having statically ( $P < 0.01$ ) differential expression ( $>1$  or  $-1 < \log_2$  fold) between spaceflight and ground control in both WS and Col-0 ecotypes are depicted. Blue is indicative of down-regulation, red is indicative of up-regulation and gray indicates no statistically significant differential expression.

## OMG1 transcript is up-regulated in WS ecotype across multiple spaceflight experiments

*OMG1* expression was up-regulated in WS roots across multiple spaceflight experiments (APEX01-1B, APEX01-2B and APEX03-2) but remained unchanged in Col-0 ([Fig. 2](#)). The Box-Whisker plot of [Fig. 2](#) illustrates the median fold change of *OMG1* expression between spaceflight and the ground control from a total of nine replicates from each sample. Each spaceflight experiment represents a distinct launch and orbital plant growth event. In both the APEX01 experiments, seeds germinated on orbit grew for 12 days before being harvested. *OMG1* expression shows a 4-fold change in spaceflight compared to ground controls in APEX01-2B and 3.5-fold change in APEX01-1B. In both analyses, the large spread between the maximum and the minimum can be attributed to the mixture of ecotypes. In a subsequent spaceflight experiment, APEX03-2, plants grew for 8 or 11 days on the ISS before being harvested. In the APEX03-2 8-day-old plants, Col-0 showed a median fold change of 1 (no change), whereas WS showed a 3.9-fold increase in spaceflight. In the APEX03-2 11-day-old plants, Col-0 showed a median fold change of 1 and WS showed a 2.1-fold increase in expression.

Figure 2.



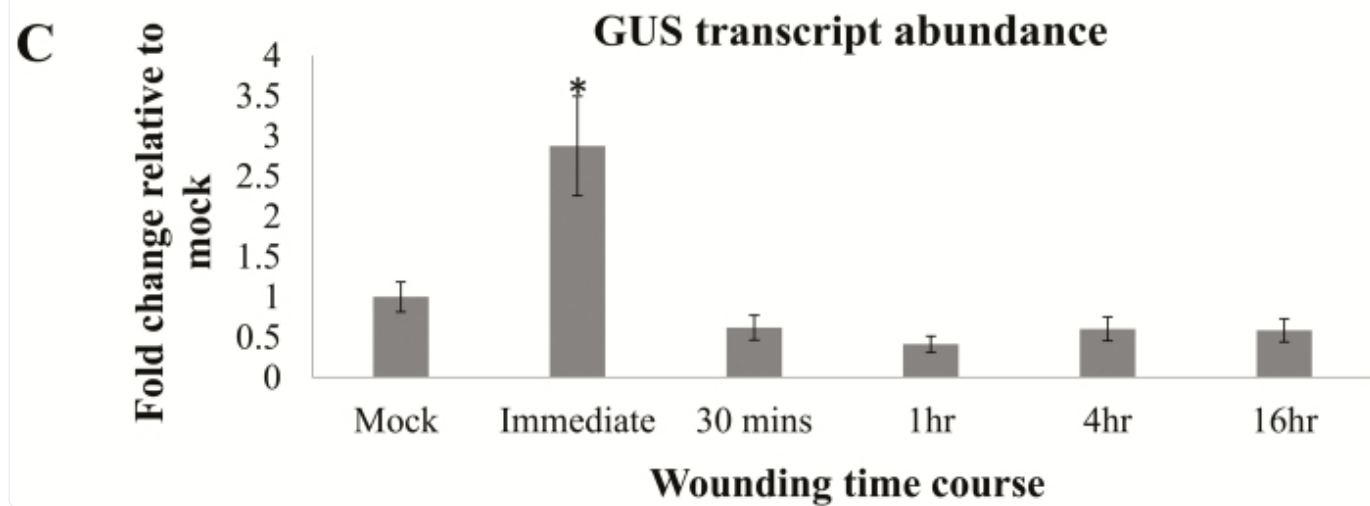
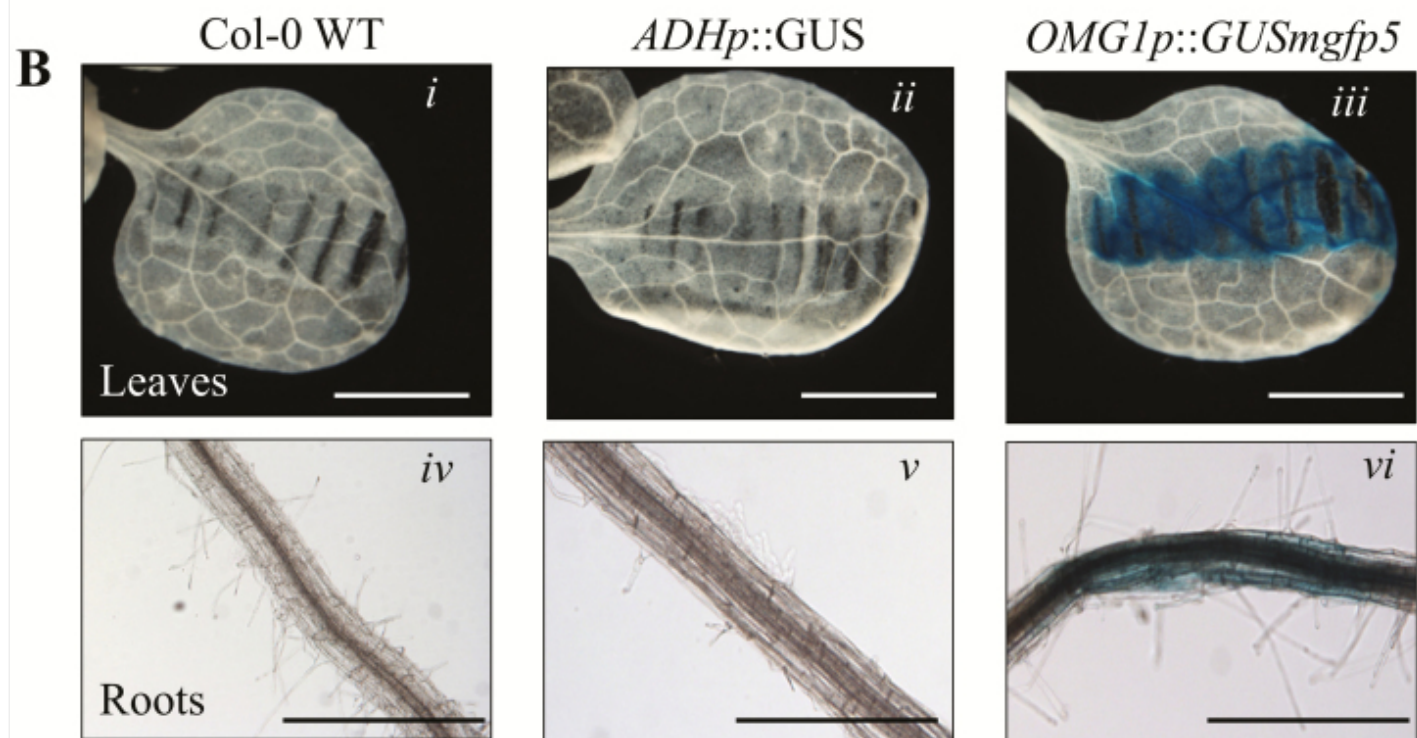
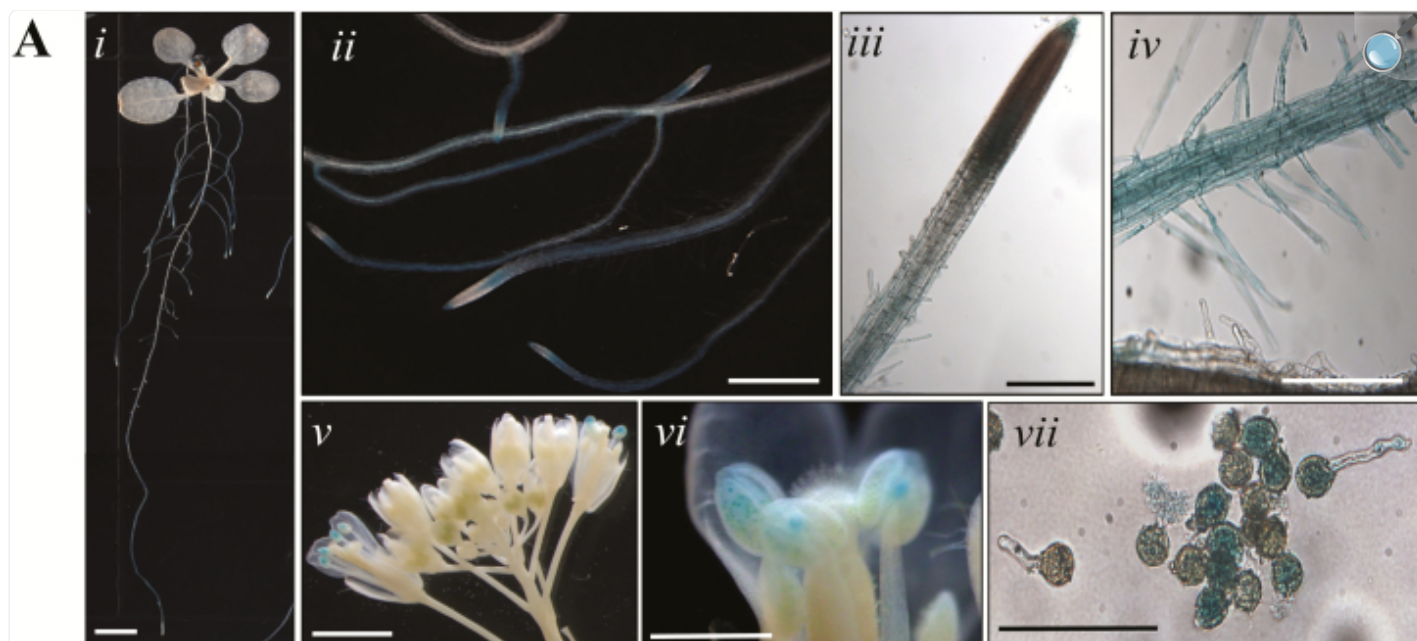
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OMG1 transcripts are up-regulated in WS but unchanged in Col-0 consistently over multiple spaceflight experiments. Ecotypes WS or Col-0 were grown onboard the ISS for the number of days (d) indicated, harvested and compared to comparable ground controls. OMG1 transcripts for flight and ground control samples assayed with qRT-PCR. Spaceflight experiment APEX01-2B launched to the ISS via STS131 and analyses published in [Paul \*et al.\* \(2013b\)](#). The same RNA used in the 12d APEX01 microarrays was used for qRT-PCR shown here. Spaceflight experiment APEX01-1B launched to the ISS via STS129, and spaceflight experiment APEX03-2, 8 day and 11 day was launched to the ISS via SpaceX05. Boxplots were constructed from nine replicates for each sample. Expressions were normalized to UBQ11 and plotted spaceflight relative to ground control using results generated from TaqMan™ qRT-PCR. Spaceflight experiments show that OMG1 transcript is up-regulated in spaceflight relative to ground controls in WS plants, whereas in Col-0 it remains largely unchanged. Significance  $^*(P < 0.01)$ ,  $^{**}(P < 0.001)$  between samples is calculated by doing a single-factor ANOVA followed by *post hoc* Student's *t*-test with Bonferroni correction.

## The OMG1 basal promoter activity was observed in roots, pollen tubes and upon wounding

Histochemical GUS staining revealed some basal *OMG1* promoter activity in roots and pollen but showed heightened activity upon wounding in the roots and leaves. T3 homozygous *OMG1*p::*GUSmgfp5* transgenic plants were used to characterize the activity of the *OMG1* promoter. In roots, basal *OMG1* promoter activity was observed at low levels in the maturation zone of both the primary and lateral roots and root hairs ([Fig. 3A](#), i–iv). In addition, basal promoter activity was also observed in pollen grains and pollen tubes ([Fig. 3A](#), v–vii). A T1 heterozygous population of pollen grains grown in pollen germinating media ([Fig. 3A](#), vii) was used to visualize the specificity of the GUS enzymatic reactions in transformed pollen. Serendipitously, wounded plant tissues were observed to have heightened *OMG1* promoter activity ([Fig. 3B](#)).

Figure 3.





A transgenic reporter line *OMG1* promoter::*GusGFP* in the WS ecotype background shows *OMG1* 5' flanking region activity. (A) Transgenic *Arabidopsis* plants of various ages underwent histochemical GUS staining with X-Gluc. The images show representative distribution patterns of GUS staining in (i) maturation zone of roots, (ii–iii) lateral roots, (iv) root hairs of the maturation zone, (v–vii) pollen and pollen tubes. Scale bars: (i–ii) 1 mm; (iii–vii) 100  $\mu$ m. (B) *OMG1* 5' flanking region activity was also observed upon wounding with a pair of forceps. Wild-type (WT) Col-0, *Adh* promoter::*Gus* transgenic line were used as a control to ensure GUS enzymatic activity was specific for *OMG1* 5' flanking region. Both leaf (i–iii) and root tissue (iv–vi) from the various lines were used. Scale bars: 100  $\mu$ m. (C) Intact leaf tissues were wounded and left for the allotted times before they were harvested for qRT-PCR analysis. The GUS transcript abundance fold change relative to mock was done to access the rapid wounding response of *OMG1* 5' flanking region. All error bars represent the standard error of the mean (SEM) of triplicate representative experiments. \* indicate significant ( $P < 0.05$ ) difference between immediate and the various wounding time points by doing a single-factor ANOVA followed by *post hoc* Student's *t*-test with Bonferroni correction.

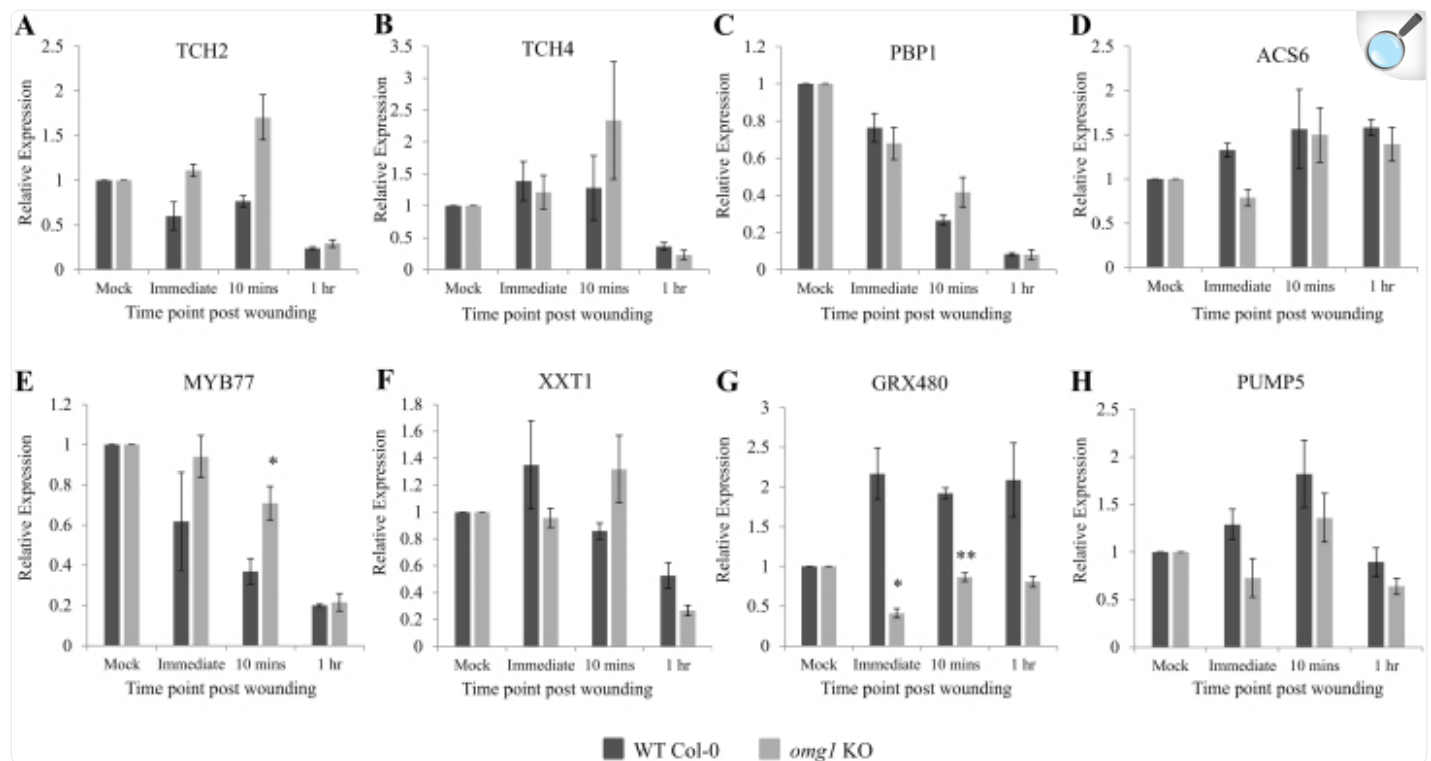
To ensure that *OMG1* promoter activity was specific to wounding, leaf and root tissues from WT Col-0, *ADHp*::*GUS* (Chung and Ferl 1999) and *OMG1p*::*GUSmgfp5* lines were wounded with a pair of forceps. In this wounding experiment, GUS staining was observed only in the *OMG1p*::*GUSmgfp5* line. A qRT-PCR analysis of *GUS* transcripts from leaves wounded at different time points showed that *OMG1* promoter activity heightens immediately after wounding and dissipates after 30 min (Fig. 3C).

## OMG1 regulates the expression of MYB77 and GRX480 known to be induced in spaceflight and upon wounding

Eight genes [see [Supporting Information—Table S2](#)] were used to elucidate potential functional interactions with *OMG1*. Since *OMG1* promoter activity was shown to rise immediately after wounding (Fig. 3C), a wounding time course of mock, immediate, 10 min and 1 h was used to screen the expression of the genes listed in [Supporting Information—Table S2](#) that could be affected by knocking out *OMG1*. Two genes, *GRX480* (At1g28480) and *MYB77* (At3g50060), out of eight genes tested showed significant differences (Student's *t*-test  $P < 0.05$ ) in gene expression between WT and *omg1* lines (Fig. 4D and E). The expression of *GRX480* was repressed after wounding in the *omg1* line and did not increase as did the WT line. On the other hand, *MYB77* expressed in the *omg1* line showed a significant delay in down-regulating gene expression when compared to the WT at the 10-min post-wounding time point. *MYB77* and *GRX480* were significantly differentially expressed between Col-0 and WS (Fig. 5). Col-0 showed a median fold change of 1.7 and WS showed a 3-fold increase in *MYB77* expression in spaceflight relative to ground controls (Fig.

5A). In the *GRX480* analyses, Col-0 showed a median fold change of 0.3 and WS showed a median fold change of 1.3 in spaceflight relative to ground controls (Fig. 5B).

Figure 4.

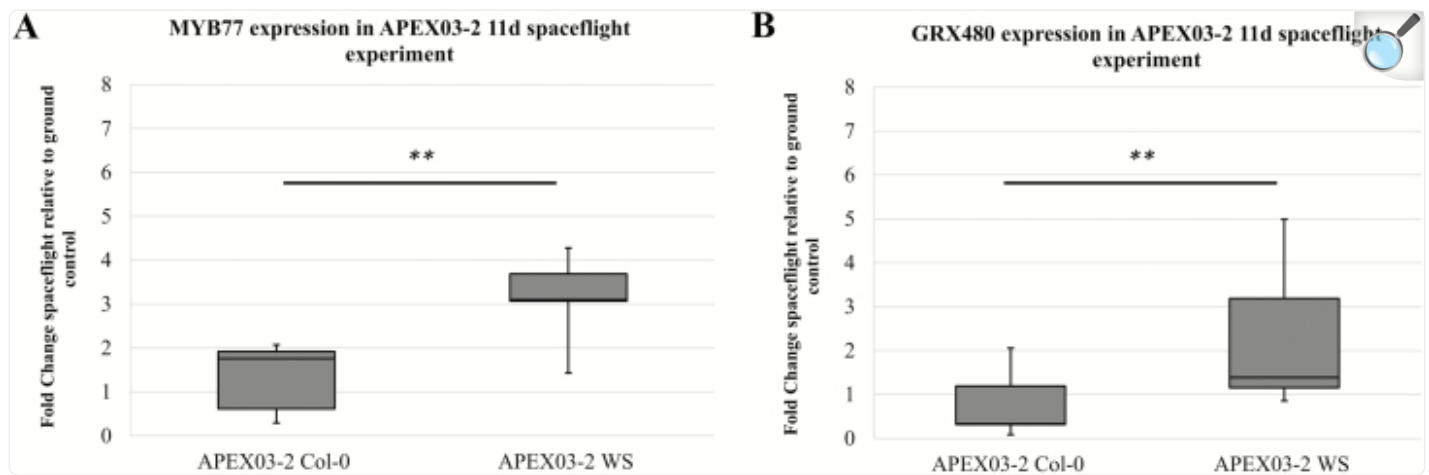


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*OMG1* regulates the rapid wounding response of *MYB77* and *GRX480*. WT Col-0 and *omg1* KO intact leaf tissues were untouched (mock) or wounded with a pair of forceps and left for the indicated time points before harvesting for qRT-PCR analyses. Expression of the indicated genes (A–H) shared between spaceflight, rapid wounding and pollen development [see [Supporting Information—Table S2](#)] in the WT (Col-0) and *omg1* KO leaves was analysed. Untouched leaves (mock), immediately after wounding, 10-min post-wounding and 1-h post-wounding were the indicated time points that the transcripts were measured. The average relative expression was normalized to a housekeeping gene (UBQ11) and presented relative to the mock controls. \* indicate significance ( $P < 0.05$ ), \*\* ( $P < 0.001$ ) between WT Col-0 and *omg1* which was calculated by doing a two-factor ANOVA followed by *post hoc* Student's *t*-test. All error bars represent the standard error of the mean of triplicate representative experiments.



Figure 5.



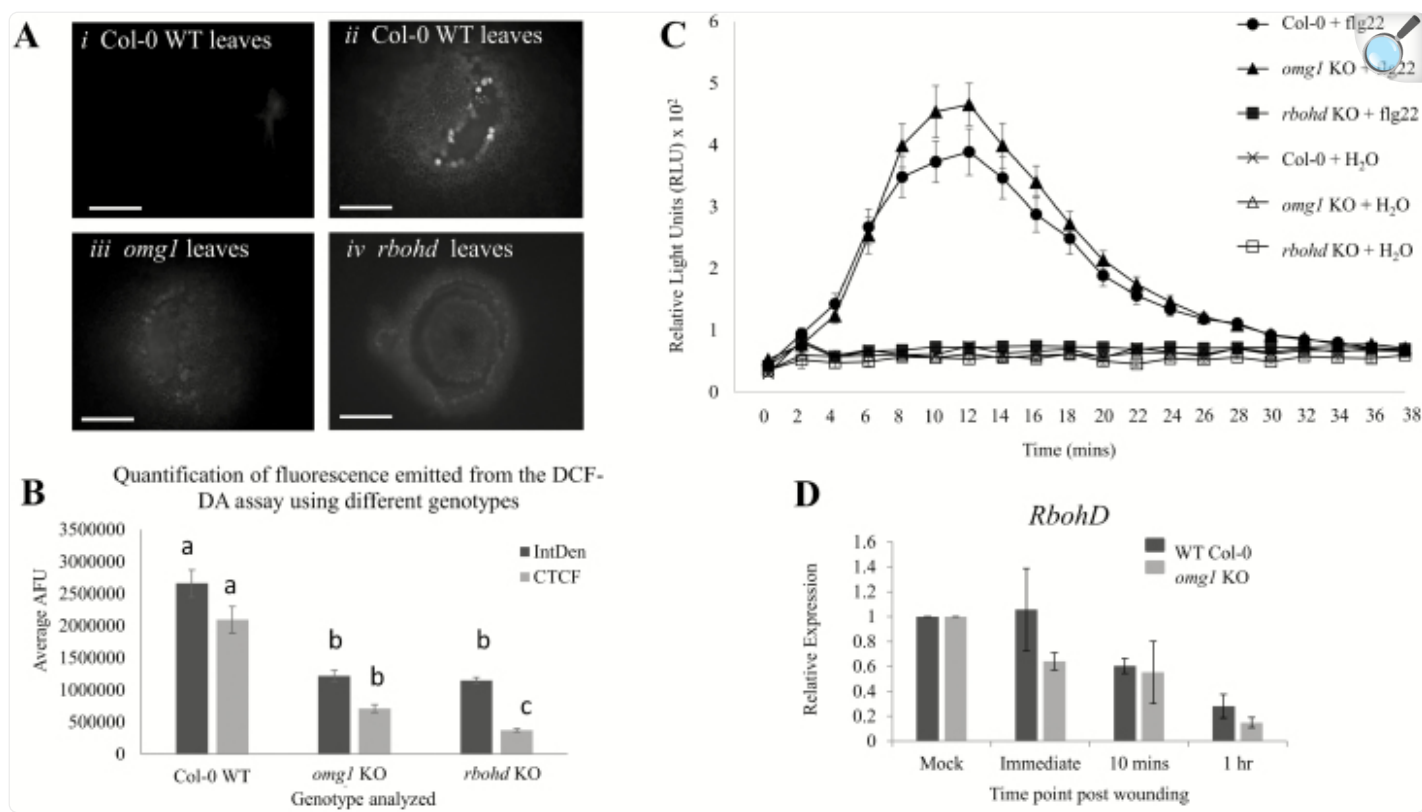
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*MYB77* and *GRX480* expression of APEX03-2 spaceflight experiment showed higher fold change in WS compared to the Col-0 ecotype. (A) qRT-PCR done on APEX03-2 11d spaceflight root tissue show that *MYB77* transcript in spaceflight relative to ground controls is significantly ( $P = 0.001$ ) different between WS and Col-0 plants. The fold change of *MYB77* expression is higher in the WS ecotype compared to the Col-0 ecotype. (B) qRT-PCR showed a significant ( $P = 0.007$ ) difference in the expression fold change of *GRX480* between Col-0 and WS plants. Boxplots were derived from nine replicates normalized to UBQ11 and plotted spaceflight relative to ground control. \*\* indicate ( $P < 0.01$ ) significant difference between Col-0 and WS was calculated by doing a Student's *t*-test.

## ROS accumulation upon wounding and flg22 induction is altered in *omg1* KO leaves compared to Col-0 leaves

Wild-type Col-0, *omg1* and respiratory burst oxidase homology D (*rbohD*) KO plants, which are delayed in ROS accumulation (Miller *et al.* 2009), were subjected to mechanical wounding with a pipette tip. The production of hydroxyl, peroxy and other ROS activity within the cells was measured with DCF-DA assay. Green fluorescence at the site of wounding was captured after 20 min with light microscopy. Wild-type Col-0 leaves produced ROS around the wound site, whereas neither *omg1* nor *rbohD* showed prominent production of ROS (Fig. 6A). Quantification of fluorescence using ImageJ showed significant differences (Student's *t*-test  $P < 0.01$ ) between Col-0 and the mutant lines (Fig. 6B).

Figure 6.



[Open in a new tab](#)

Reactive oxygen species (ROS) accumulation upon wounding and flg22 induction is altered in *omg1* leaves compared to Col-0 leaves. (A and B) DCF-DA ROS assay performed on wounded leaf tissues reveals that OMG1 affects cellular ROS production. (A) DCF-DA is a fluorogenic dye that measures hydroxyl, peroxyl and other ROS activity within the cell. A pipette tip was used to wound the surface of the leaves from the WT line, *omg1* KO line and *rbohD* KO line (a known ROS-deficient mutant). Upon wounding, ROS is usually produced around the site of injury. The DCF-DA reagent is then oxidized by ROS resulting in appearance of green fluorescence. Plants were wounded and immediately placed in the DCF-DA solution for 20 min before visualized on the light microscope. (i) WT Col-0 leaves without wounding, (ii) WT Col-0 leaves, (iii) *omg1* leaves and (iv) *rbohD* leaves were placed in DCF-DA solution immediately after wounding. In the WT plants green fluorescence around the wound site indicates the production of ROS. However, in the *omg1* KO line and the *rbohD* KO line, ROS production was not detected. Images were taken on the Olympus BX51 compound scope at 10 $\times$  magnification, exposure time 55 ms. Scale bars: 100  $\mu$ m. (B) Quantification of fluorescence was performed using integrated density (IntDen) and corrected total cell fluorescence (CTCF) [CTCF = Integrated density – (Area of selected cell  $\times$  Mean fluorescence of background readings)] in ImageJ. Statistical analyses were performed using Student's *t*-test, bar graphs with different letters show significant

difference ( $P < 0.01$ ). (C and D) Higher elicitor-induced oxidative burst in *omg1* KO is not associated with altered expression of *RbohD*. (C) Extracellular ROS production was measured using a luminol-based assay in leaf discs from *omg1* KO, *rbohD* KO and WT Col-0 plants after elicitation with water or flg22. Experiments were repeated at least three times and results are a mean of 24 samples. (D) The relative expression of *RbohD* in the WT (Col-0) and *omg1* KO leaves was analysed in a wounding assay untouched leaves (mock), immediately after wounding, 10-min post-wounding and 1-h post-wounding. The average relative expression was normalized to a housekeeping gene (UBQ11) and presented relative to the mock controls. No significant difference in *RbohD* expression between Col-0 and *omg1* KO plants was observed when the Student's *t*-test analysis was performed. All error bars represent the standard error of the mean of triplicate representative experiments.

Based on luminol ROS assays OMG1 affects the elicitor-induced generation of extracellular ROS. Flg22-induced hydrogen peroxide production by leaf discs of *omg1* mutants was ~20 % higher compared to Col-0. Water-treated controls along with the *rbohD* leaf discs showed no production of flg22-induced H<sub>2</sub>O<sub>2</sub> ([Fig. 6C](#)). Since elicitor-induced extracellular ROS is governed by *RbohD*, the measurement of *RbohD* expression between Col-0 and *omg1* lines in a wounding time course was analysed ([Fig. 6D](#)). The relative expression of *RbohD* showed no significant difference between WT and *omg1* leaves.

## OMG1 is a member of the CONSTANS-Like protein family

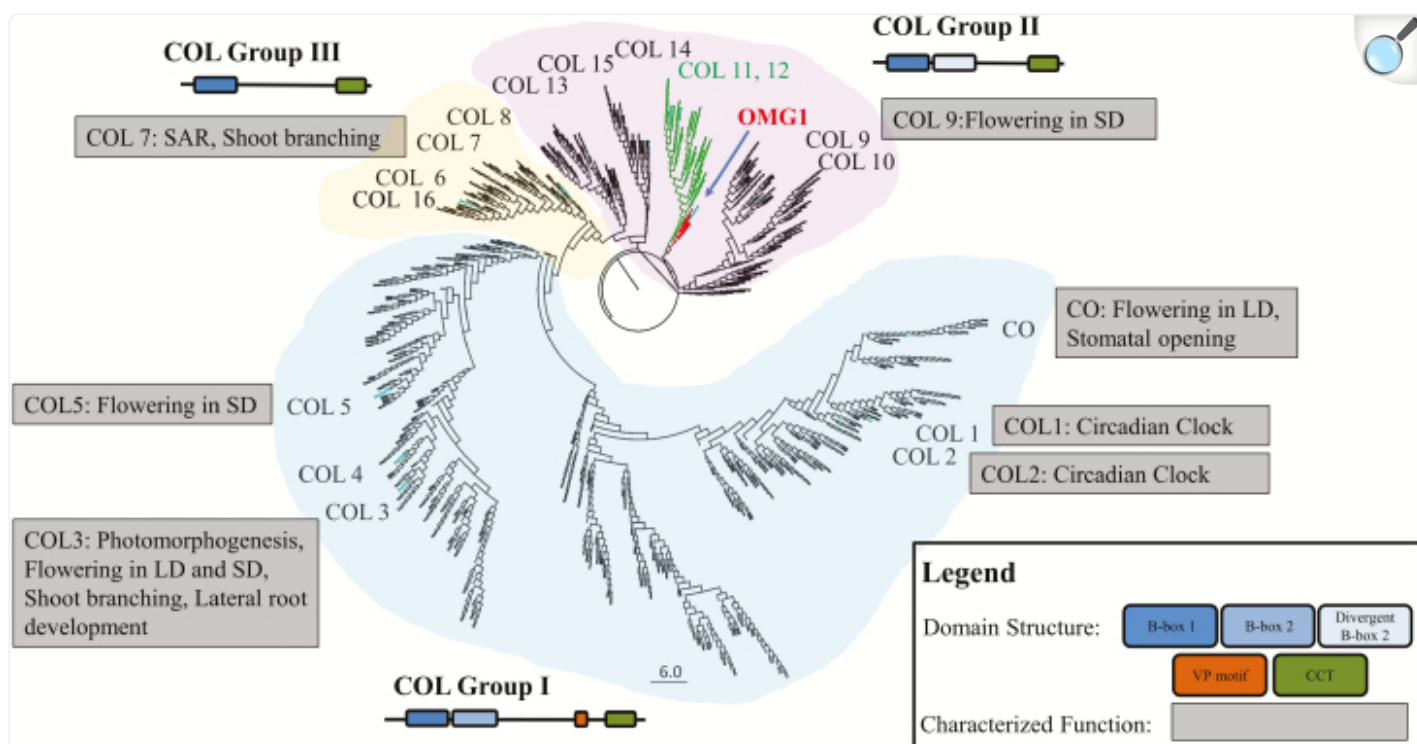
The predicted *OMG1* gene encodes a 369 amino acid protein that contains a C-terminal 43 amino acid sequence that is 84 % sequence similarity to the (CONSTANS (CO), CONSTANS-Like (COL) and TIMING OF CAB EXPRESSION1 (TOC1)) 'CCT' domain ([Berardini et al. 2015](#)). *OMG1* also contains two divergent B-box domains in the N-terminal region [[see Supporting Information—Fig. S2](#)]. The *OMG1* B-box1 domain has 51 % similarity to the B-box1 consensus sequence, and the *OMG1* B-box2 domain has a 94 % similarity to the B-box2 consensus sequence ([Griffiths et al. 2003](#); [Masaki et al. 2005](#); [Gangappa and Botto 2014](#)).

A consensus phylogeny suggests that *OMG1* belongs to group II of the CO and COL family of transcriptional regulators ([Fig. 6](#)). Maximum likelihood phylogenies placed a clade of *OMG1*-related sequences from Brassicas and Monocots (SH-like aLRT > 0.989) close to COL11-12 sequences from across flowering plants [[see Supporting Information—Table S3](#)]. *OMG1* appears closely related to the COL11-12 and COL9-10 lineages, although the precise placement of the *OMG1* group remains unresolved.

Both the *OMG1* and the COL11-12 clades consist of sequences representing both monocots and dicots. In the consensus tree, a single COL11-12-like sequence from the basal flowering plant *Amborella trichopoda* rooted the *OMG1* and

COL11-12 clades (SH-like aLRT > 0.912), suggesting that the OMG1 lineage arose via a gene-duplication event in flowering plants, prior to the divergence of monocots and dicots ([Fig. 7](#)). However, no OMG1-related sequences from Eudicots were observed outside the Brassicaceae family, suggesting that either OMG1 was lost from non-Brassica dicots, or the sequence has diverged so much in these lineages as to be unrecognizable. The different alignment and tree-reconstruction procedures support the early-duplication hypothesis, and make it unlikely that the grouping of monocot and dicot OMG1-related sequences is a phylogenetic artefact.

Figure 7.



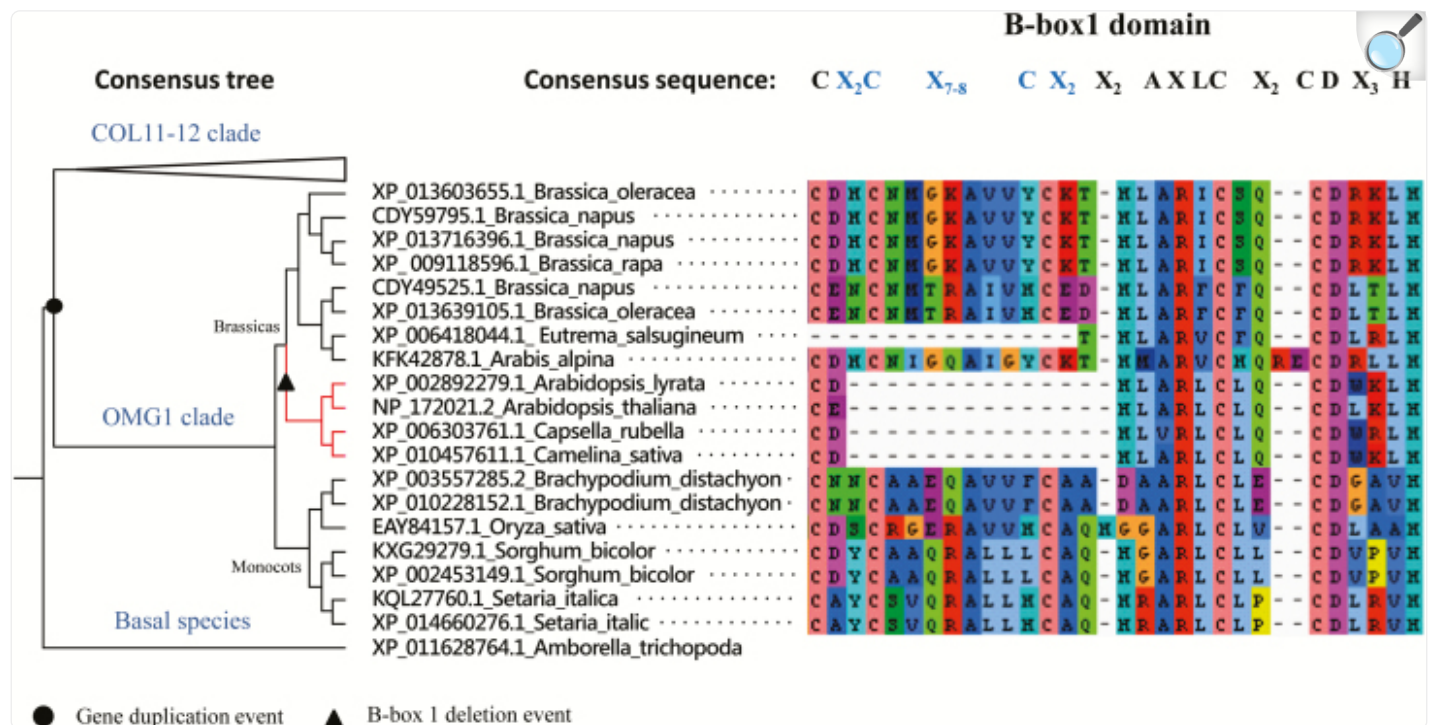
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OMG1 is a member of the CO and COL protein family in *Arabidopsis*. The consensus tree derived from full-length protein sequences containing a single CCT domain of all plants found in the non-redundant (NR) database revealed that OMG1 belongs to the CO and COL family of proteins. In *Arabidopsis* (taxa highlighted in cyan), the COL family can be divided into three groups. Group I (shaded in light blue) comprises of CO, COL1-6 which have domain structures consisting of two B-box domains (type I and type II) at the N-terminal and both a VP motif along with a CCT domain at the C-terminal. Genes in group II (shaded in light purple) comprising of COL9-15 and OMG1 have domain structure consisting of a type I B-box domain and a second diverged B-box2 domain along with a C-terminal CCT domain. The OMG1 clade is highlighted in red, whereas its closest homolog COL11-12 is highlighted in green. Group III (shaded in light orange) comprises of COL6-8 and COL16 have domain structures consisting of a single B-box1 domain and a CCT domain. Listed in the grey boxes are the characterized functions of some of the COL members ([Gangappa and Botto 2014](#)).

The high level of sequence divergence in the OMG1 B-box1 domain is primarily due to a 13-amino acid deletion close to the start of the B-box1 consensus sequence CX-----XAXLCX<sub>2</sub>CDX<sub>3</sub>H ([Fig. 7](#)). This B-box1 deletion was

observed across several *Brassica* sequences nested within the OMG1 clade, but this deletion was not found in monocot OMG1, suggesting it probably occurred late in the Brassicaceae lineage. Although the OMG1 B-box2 domain is largely conserved (Fig. 8), two highly conserved consensus amino acids are different in OMG1 compared to the consensus sequence CX<sub>2</sub>CX<sub>7-10</sub>CX<sub>7</sub>CX<sub>2</sub>CX<sub>5-12</sub>H. The third cysteine in the consensus sequence is serine in OMG1 (C49S), and the histidine at the end of the consensus sequence is leucine (H73L). Examination of B-box2 domains outside the OMG1 clade suggests that these substitutions are unique to the OMG1 clade [see [Supporting Information—Fig. S3](#)].

Figure 8.



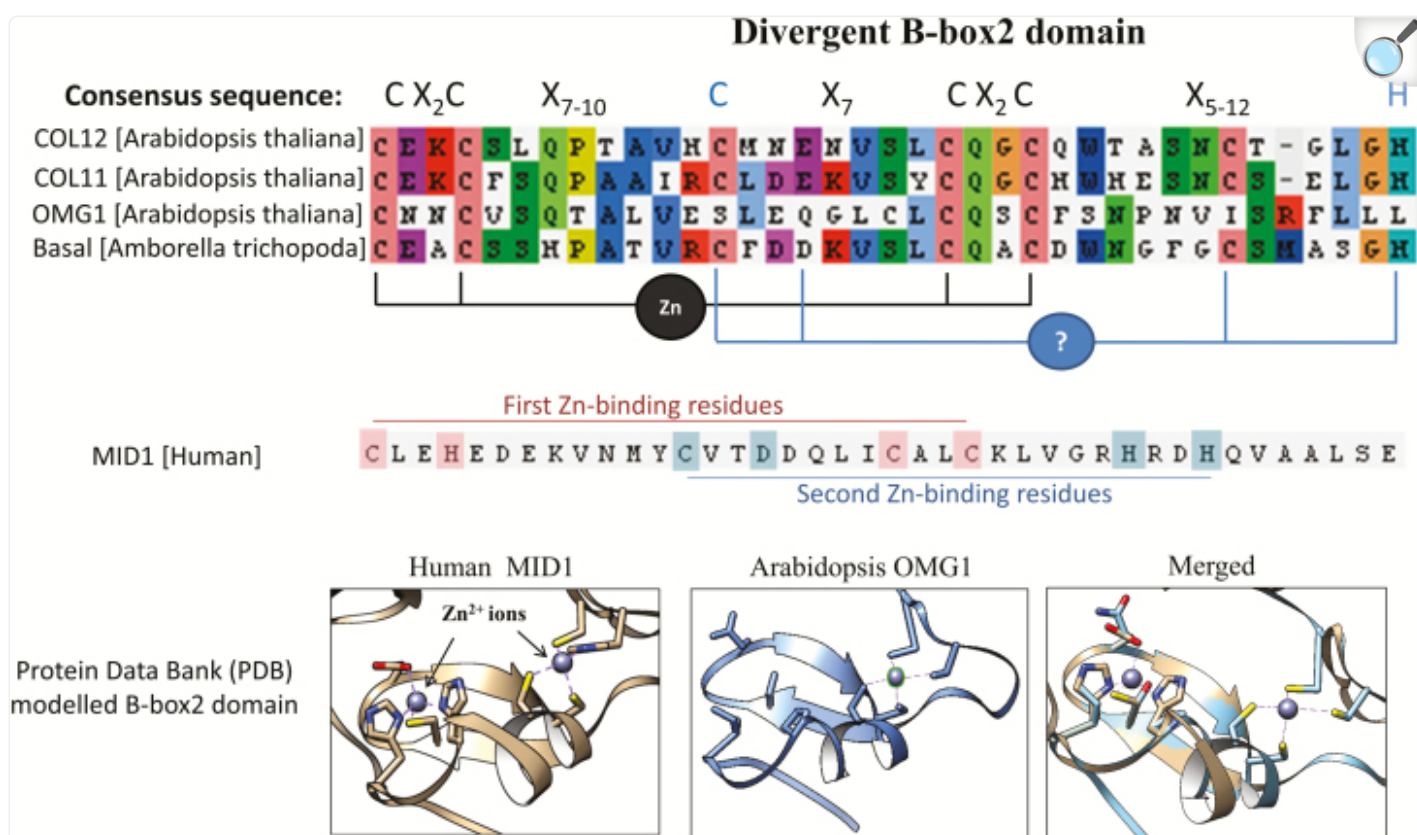
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OMG1 resulted from a gene duplication of COL11 and 12 and underwent a subsequent B-box1 deletion event. OMG1 and CONSTANS-Like (COL) 11-12 are sister clades based on the consensus tree described in Fig. 8, suggesting that OMG1 resulted from a gene duplication of COL11-12 (indicated by the circle). Sequences within the OMG1 clade are grouped into *Brassicas* and *monocots*. The *Brassica* sequences highlighted in red, share a similar B-box1 deletion indicated by the triangle and reflected in the adjacent B-box1 domain sequences derived using the AliView software. The B-box1 consensus sequence (Gangappa and Botto 2014) is listed and regions where the deletion occurs are highlighted in blue. The consensus tree depicted here is not drawn to scale.

The B-box domain family belongs to a subgroup of zinc-binding proteins stabilized by canonical cysteine-rich zinc-finger motifs ([Klug and Schwabe 1995](#)). The inferred structure of Zn-bound OMG1 B-box2 suggests that OMG1 is capable of stably binding at least one zinc ion, via a canonical zinc-finger motif ([Fig. 9](#)). In the model, four conserved cysteine residues surrounding the CX<sub>2</sub>C configuration were spatially positioned in a similar fashion as the amino acids surrounding the first Zn-binding site in the human MID1 template, suggesting this first OMG1 Zn-binding site retains a canonical zinc-finger structure.



Figure 9.



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The B-box2 domain of OMG1 is predicted to bind at least one zinc ion. The sequence of B-box2 domain was compared between *Arabidopsis* COL11-12, OMG1 and the basal species *Amborella trichopoda*. The consensus sequence of the B-box2 domain is listed and OMG1 seems to have a relatively conserved B-box2 domain except for the residues highlighted in blue. Residues within the COL sequences that create a zinc-binding interface are indicated by the black lines, whereas residues that might create a second zinc-binding interface are indicated by the blue lines. The human MID1 sequence is also listed and residues that bind to the first zinc ion are highlighted in pink, whereas residues that bind to the second zinc ion are highlighted in blue. OMG1 B-box2 domain was modelled after a well-characterized B-box2 found in the Human MID1 protein located in the Protein Data Bank (PDB). Modelling of the protein was done using the Chimera protein modelling software. In the *Arabidopsis* OMG1 protein, four cysteines are positioned such that they interact with a zinc ion. However, the substitution of amino acids around the second zinc-binding region shows strong evidence for the loss of the second Zn-binding site as modelled in the human MID1 template.

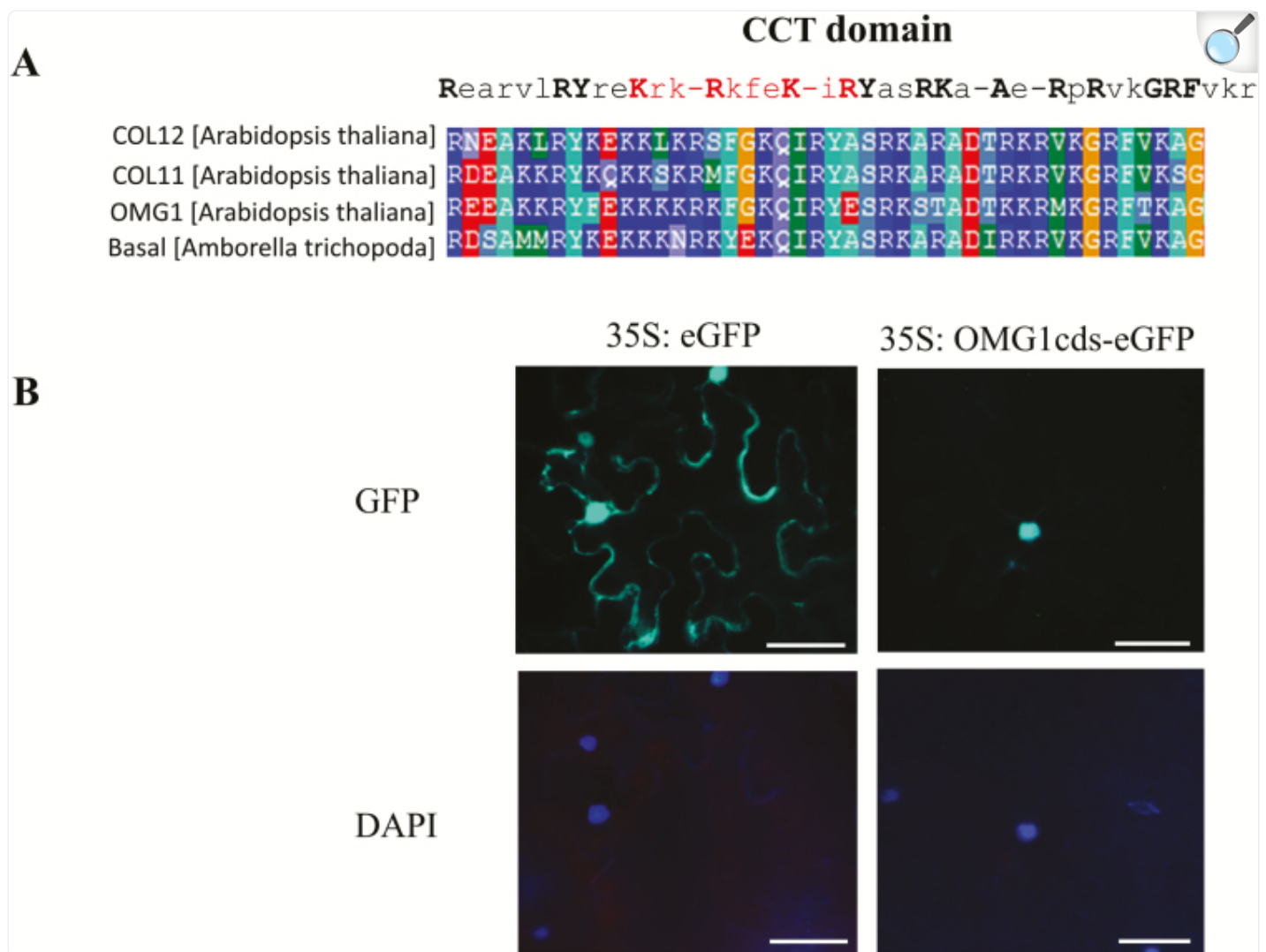


However, radical substitutions at key Zn-binding sites (C49S, E52Q, C67I and H73L) occurring in the second Zn-binding pocket of OMG1's B-box2 motif appear to completely disrupt potential Zn-binding ([Fig. 9](#)). It remains unclear whether the series of indels and substitutions after the OMG1 B-box2 domain could result in a novel structure that facilitates zinc ion binding or a different functional motif. Furthermore, sequences within the OMG1 clade show similar substitutions at most of these conserved Zn-binding residues, implicating the possibility that functional B-box2 in these proteins require only a single Zn ion [see [Supporting Information—Fig. S3](#)].

## OMG1 localizes to the nucleus

The transient expression of a 35S::OMG1-eGFP fusion protein in tobacco leaves indicates that OMG1 localizes in the nucleus. The highly conserved CCT domain in OMG1 has been associated with important functions in transcriptional regulation, DNA binding and nuclear localization ([Robson \*et al.\* 2001](#); [Yan \*et al.\* 2011](#); [Gendron \*et al.\* 2012](#)). Cells transformed with 35S::eGFP alone displayed a uniform expression of eGFP throughout the leaf pavement cells in the vacuoles, nucleus and membrane. However, cells transformed with 35S::OMG1-eGFP fusion were specifically localized to the nucleus, as indicated by the DAPI nuclei-specific counter stain ([Fig. 10](#)).

Figure 10.



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OMG1 has a conserved CCT domain and localizes in the nucleus. (A) The sequence of CCT domain was compared between *Arabidopsis* COL11-12, OMG1 and the basal species *Amborella trichopoda*. OMG1 has a conserved CCT domain (consensus sequence in black) that encodes a nuclear localization signal (highlighted in red). (B) Subcellular localization using leaves of 4-week-old *Nicotiana benthamiana* seedlings that were transiently transformed with *Agrobacterium*-GV3101 carrying the At1g05290cds-eGFP fusion vector or the pCAMBIA1302 (as an overexpressing eGFP control) indicated that OMG1 is localized solely in the nucleus and exhibits no presence elsewhere in the cell. DAPI stains for the nucleus and can be seen to overlap with the eGFP expression. Scale bars: 100  $\mu$ m.

## Discussion

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As sessile organisms, plants have developed sensitive, adaptive strategies to regulate their metabolism in a changing environment ([Noctor et al. 2007](#); [Schwarzländer and Finkemeier 2012](#)). A strategy to avoid oxidative damage relies on the ability to deploy different antioxidants and ROS-scavenging/detoxifying enzymes to efficiently adjust their redox states. When subjected to the novel environment of spaceflight, plants respond by overexpressing more ROS- or redox-associated genes than comparable plants on the ground ([Im et al. 2011](#); [Paul et al. 2012b, 2013b, 2017](#); [Correll et al. 2013](#); [Sugimoto et al. 2014](#)). ROS in its various forms and subcellular production sites can elicit distinct and diverse physiological, biochemical and molecular responses ([Mittler et al. 2004](#); [Gadjev et al. 2006](#); [Swanson and Gilroy 2010](#); [Suzuki et al. 2012](#)). It is possible that plants drew from diverse ROS response strategies to cope with spaceflight, an environment that is outside their evolutionary experience and therefore potentially stressful. The data presented here suggest that *OMG1* is a putative regulator of the ROS signalling pathway associated with wounding and may play a prominent role in regulating the spaceflight ROS response, especially in the WS ecotype where it is strongly induced by spaceflight.

### Spaceflight induces the differential expression of ROS-producing genes in an ecotype-dependent manner

In general, the differentially expressed genes in spaceflight showed a positive correlation to genes producing ROS ([Fig. 1A](#)). Although it is known that ecotype differences can diversify the ROS signatures associated with stress responses on Earth ([Kalbina and Strid 2006](#); [Li et al. 2006, 2008](#)), these results are the first to demonstrate that different ecotypes use different specific subcellular ROS species to respond to the spaceflight environment ([Fig. 1](#)).

Previous morphological differences between Col-0 and WS ecotypes in the spaceflight environment were primarily observed in the roots. Although the overall root length at 11 days on orbit between the two ecotypes showed no significant difference ([Paul et al. 2017](#)), it was noted that the Col-0 ecotype showed a dramatic lag in root development during the first 48-h post-germination in orbit when compared to the WS ecotype, and that there are also ecotype-specific differences in root growth behaviour ([Paul et al. 2012a](#)). One difference between the Col-0 and WS ecotypes is the regulation of class III peroxidase genes ([Passardi et al. 2007](#)). In fact, two genes *PE10* (AT1G49570) and *PE59* (AT5G19890) that were up-regulated in the APEX03-2 Col-0 but remained unchanged in APEX03-2 WS ([Fig. 1B](#)) are involved in controlling the root cell elongation process ([Markakis et al. 2012](#)). The differentially expressed peroxidase genes between the two ecotypes in the APEX03-2 spaceflight transcriptomes ([Fig. 1B](#)) reinforce the notion that different ecotypes utilize ROS differently to respond to their environment and suggest that in WS, these genes were already at a level appropriate for spaceflight acclimation and were therefore unchanged. In addition, the observed down-regulation of peroxidases in Col-0 was consonant with observations previously published ([Kwon et al. 2015](#)). Since peroxidases are necessary for the general oxidoreduction of  $H_2O_2$  in general, the down-regulation of peroxidases in spaceflight

within Col-0 could contribute to Col-0-specific ROS signatures seen in clusters A and B of the ROSMETER analyses ([Fig. 1A](#)).

The current ROSMETER analysis is not intended to define how specific ecotypes utilize peroxidases or regulate specific ROS indices in response to spaceflight. Rather, it is to illustrate that the genetic background of the spaceflight-grown plant had a substantial impact on the specific subcellular ROS species that were used in the physiological response.

## OMG1 transcript is consistently up-regulated in WS ecotype across multiple spaceflight experiments and is induced rapidly upon wounding in both WS and Col-0

*OMG1* is consistently induced in spaceflight in WS but not Col-0. Comparisons of the 5' upstream region, the coding region and the 3' UTR of *OMG1* between the two ecotypes showed no differences in the primary sequence. However, in the intergenic region downstream of the *OMG1* stop codon, a point mutation is observed at positions 2197, 3327, and a two-nucleotide deletion is observed at position 3488 [[see Supporting Information—Fig. S4](#)].

Wassilewskija and Col-0 do regulate *OMG1* differently. *OMG1* showed slight differences in basal expression between Col-0 and WS, and both ecotypes demonstrated immediate up-regulation of *OMG1* after wounding, along with the rapid diminishing of the signal afterwards [[see Supporting Information—Fig. S5A](#)]. However, Col-0 and WS responded very differently in a wounding dosage test, where leaves were subjected to various degrees of damage at the onset of wounding ('immediate' time point) [[see Supporting Information—Fig. S5B](#)]. Irrespective of the extent of the wounded surface area of the leaf, Col-0 expressed *OMG1* at a consistent level, whereas WS expressed *OMG1* in a dose-response manner [[see Supporting Information—Fig. S5C](#)]. These results suggest that *OMG1* is involved in a rapid wounding response in both Col-0 and WS ecotypes but with different modes of regulation.

The WS and Col-0 ecotypes are genotypically distinct, and their transcriptomic responses to environmental and developmental cues can be dissimilar (e.g. [Chen et al. 2005](#); [Kusunoki et al. 2017](#)). The overall spaceflight transcriptomes of WS and Col-0 are dramatically different ([Paul et al. 2017](#)). It is hypothesized that even though *OMG1* is functional in both ecotypes upon wounding, other gene products or pathways present in Col-0 but not WS could compensate for the 'need' for *OMG1* in space, or that these genes were already at a level appropriate for spaceflight acclimation and were therefore unchanged in the spaceflight transcriptome. Although *OMG1*'s role in the physiological adaptation of WS to spaceflight has yet to be discerned, its strong induction suggests that *OMG1* contributes to a successful adjustment to that environment. Irrespective of *OMG1*'s specific role in spaceflight physiology, it is noteworthy that *OMG1*'s overall metabolic function, which was previously unknown, was only discovered because of its role in spaceflight physiology.

## OMG1 is associated with the regulation of ROS

*OMG1* is associated with ROS regulation. The location of *OMG1* promoter region activity coincided with regions of dynamic ROS activity. *OMG1* specifically regulated the expression of two ROS-associated genes (*GRX480* and *MYB77*). And, loss of *OMG1* protein in *omg1* resulted in deviations from WT in two different ROS assays.

The rapid induction of *OMG1* expression immediately after wounding was observed with a promoter-driven GUS reporter qRT-PCR analysis. Rapid induction of transcription (prior to 2-min post-stimulus) has been observed in gravitational and mechanical stimulation of *Arabidopsis* root apical tissues ([Kimbrough et al. 2004](#)). In addition, wounding *Arabidopsis* leaves produces ROS within minutes at the wound site ([Miller et al. 2009](#); [Beneloujaephajri et al. 2013](#)) and this rapid response is necessary for plants to elicit immediate defence/repair mechanisms against herbivore and pathogen attacks. Wounded plants can transmit rapid RBOHD-dependent ROS signals of speeds up to 8.4 cm min<sup>-1</sup> to mediate long-distance cell-to-cell communication ([Miller et al. 2009](#)). However, since no statistical differences were seen in the expression level of *RbohD* in Col-0 WT plants and *omg1* KO plants ([Fig. 6D](#)), this suggests that the changes in extracellular or intracellular ROS are not a result of altered expression of *RbohD* in the *omg1* KO line.

Reactive oxygen species molecules are essential signalling molecules in plant growth and development but can also act as toxic stressors. Therefore, there exists an elaborate and highly redundant network of genes that maintain the delicate ROS homeostasis ([Gechev et al. 2006](#)). Functional *OMG1* is necessary for rapid generation of intracellular ROS at the wound site ([Fig. 6A](#) and [B](#)). However, over time these ROS return to WT levels, presumably facilitated by other genes of redundant function [[see Supporting Information—Fig. S6](#)].

In a recent review, it has been suggested that altered ROS signalling in response to the spaceflight environment could affect plant growth and development indirectly via changes in hormonal influx within the plants ([Schüler et al. 2015](#)). *OMG1* was found to regulate the expression of ROS-associated genes *GRX480* and *MYB77*. *GRX480/GRXC9* is a member of the glutaredoxin (GRX) family of small redox enzymes. *GRX480* is a salicylic acid (SA)-responsive gene induced quickly and transiently by an NPR1-independent pathway, which is associated with the ROS-scavenging/antioxidant network ([Laporte et al. 2012](#); [Zander et al. 2012](#); [Herrera-Vásquez et al. 2015a](#); [Huang et al. 2016](#)). *MYB77* is a member of the R2R3-type transcription factor family that is involved in ROS metabolism through the mitogen-activated protein kinase (MAPK) cascade. The MAPK cascade is also known to play a role in plant development and hormone signalling and includes hormones such as jasmonic acid (JA), SA and ethylene (ET) ([Shin and Schachtman 2004](#); [Tejge et al. 2004](#); [Galvez-Valdivieso et al. 2009](#); [Herrera-Vásquez et al. 2015b](#)).

*OMG1*'s role in ROS signalling was established with two frequently used ROS biochemical assays. The DCF-DA assay showed that intracellular ROS production around the wound site was dampened in the *omg1* mutant, whereas flg22-induced extracellular ROS production was 20 % higher than WT in the *omg1* mutant. These conflicting results can be addressed by the different signalling pathways elicited by each stimulus. Responses to mechanical damage can be

localized, systemic or both. In any case, the response involves the generation, translocation, perception and transduction of wound signals to activate the expression of wound-inducible genes. In response to mechanical wounding, the central role for JA signalling is well established. However, many other compounds, such as oligopeptide systemin, oligosaccharides, phytohormones, hydraulic pressure or electrical pulses, have also been proposed to play a role in wound signalling ([León \*et al.\* 2001](#)). Early localized ROS induced by wounding, specifically  $O_2^{\bullet-}$ ,  $H_2O_2$  and singlet oxygen, have been suggested to originate intracellularly from the photosynthetic electron transport chain in chloroplasts ([Morker and Roberts 2011](#)).

On the other hand, flg22, a microbe-associated molecular pattern (MAMP), elicits an early response that includes SA-, JA- and ET-defence signalling pathways. Flg22 also elicits a late response that includes activation of defence-related senescence processes, SA-dependent secretory pathway genes and pathogenesis-related protein 1 (*PR1*) expression ([Denoux \*et al.\* 2008](#)). MAMP-induced ROS have been shown to originate from the apoplast through activation of NADPH oxidases as well as peroxidases ([Jwa and Hwang 2017](#)). Although there are crosstalks between signalling pathways induced by mechanical wounding and flg22, there are also unique pathways induced by each stimulus that could attribute to the different results observed. Details of these mechanisms are still being investigated; however, the results infer that within the Brassicaceae family OMG1 does play a role in the ROS signalling pathway.

## Phylogeny and protein architecture of OMG1

Prior to this study, members of the *Arabidopsis* CO and COL family were thought to only function in photoperiod signalling that involves flowering, circadian rhythms, stomata opening and root-shoot branching ([Gangappa and Botto 2014](#)). Evidence presented here suggests that members of this family play a role in ROS signalling. This, in turn, could aid in the functional characterization of many of the other uncharacterized members such as COL11 and COL12. We suggest that divergent function of OMG1 can be attributed to the evolved B-box1 and B-box2 domains. This is in line with reports on the rapid evolution of COL genes within the Brassicaceae, in which the increased evolutionary rates were observed not only in the middle region of the protein but also in the conserved zinc-finger and basic CCT regions ([Lagercrantz and Axelsson 2000](#)).

## Conclusion

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OMG1 is a CCT domain transcriptional regulator of ROS homeostasis, one that is important to the physiological adaptation of *Arabidopsis* ecotype WS to spaceflight. While investigations are underway to determine the detailed mechanisms of how OMG1 regulates the ROS homeostasis in wounding, and its specific role in spaceflight, the results presented here demonstrate several important concepts for general consideration. First, spaceflight experiments are useful platforms to discover novel gene functions and provide fundamental insights into terrestrial plant biology. Second, the response to spaceflight is genotype-specific and this understanding should be taken into consideration when

defining the overall plant spaceflight response. WS and Col-0 differ in induction of OMG1 in spaceflight. Third, the maintenance of ROS homeostasis plays an important role as plants physiologically adapt to spaceflight. And, last, the CONSTANS-Like family contains members that function well beyond the clock and timing roles typically associated with CCT domains, including ROS signalling.

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## Contributions by the Authors

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N.J.S. designed and conducted the ROS and OMG1-related experiments, analysed the transcriptome data, and wrote the first drafts of the manuscript. B.K. made substantial contributions to the phylogenetic analyses. A.-L.P. and R.J.F. supervised the work, designed and conducted the spaceflight transcriptome experiments, and revised the manuscript.

## Conflict of Interest

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None declared.

## Supporting Information

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The following additional information is available in the online version of this article—

[Table S1](#). List of all the primer pairs used in experiments presented in this paper.

[Table S2](#). List of genes shared between spaceflight, rapid wounding and pollen tube growth transcriptomic data sets.

[Table S3](#). List of all unique plant species represented in the phylogenetic analysis.

[Figure S1](#). Characterization of the *omg1* homozygous knockout (KO) line (SALK\_045742C).

[Figure S2](#). Full-length sequence of *Arabidopsis* OMG1 aligned to *Arabidopsis* COL11-12 and the basal species *Amborella trichopoda*.



[Figure S3](#). B-box2 domain sequence in all species within the OMG1 clade.

[Figure S4](#). Alignment details of OMG1 in Wassilewskija (WS) vs. Columbia (Col-0) cultivar.

[Figure S5](#). Characterization of OMG1 expression in wild-type (WT) Wassilewskija (WS) and WT Columbia (Col-0) with qRT-PCR analysis upon wounding experiments.

[Figure S6](#). Time course of the 5-(and 6)-carboxy-2',7'-dichloro dihydrofluorescein diacetate (DCF-DA) fluorescent reactive oxygen species (ROS) assay described in [Fig. 5](#).

### Supplementary Material

[Click here for additional data file.](#) (2.2MB, pdf)

## Acknowledgements

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## Literature Cited

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1. Beneloujaephajri E, Costa A, L'Haridon F, Métraux JP, Binda M. 2013. Production of reactive oxygen species and wound-induced resistance in *Arabidopsis thaliana* against *Botrytis cinerea* are preceded and depend on a burst of calcium. BMC Plant Biology 13:160. [[DOI](#)] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
2. Berardini TZ, Reiser L, Li D, Mezheritsky Y, Muller R, Strait E, Huala E. 2015. The *Arabidopsis* information resource: making and mining the “gold standard” annotated reference plant genome. Genesis 53:474–485. [[DOI](#)] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
3. Chen WJ, Chang SH, Hudson ME, Kwan WK, Li J, Estes B, Knoll D, Shi L, Zhu T. 2005. Contribution of transcriptional regulation to natural variations in *Arabidopsis*. Genome Biology 6:R32. [[DOI](#)] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]



4. Chiu W, Niwa Y, Zeng W, Hirano T, Kobayashi H, Sheen J. 1996. Engineered GFP as a vital reporter in plants. *Current Biology* 6:325–330. [[DOI](#)] [[PubMed](#)] [[Google Scholar](#)]
5. Chung HJ, Ferl RJ. 1999. *Arabidopsis* alcohol dehydrogenase expression in both shoots and roots is conditioned by root growth environment. *Plant Physiology* 121:429–436. [[DOI](#)] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
6. Clough SJ, Bent AF. 1998. Floral dip: a simplified method for *Agrobacterium*-mediated transformation of *Arabidopsis thaliana*. *The Plant Journal* 16:735–743. [[DOI](#)] [[PubMed](#)] [[Google Scholar](#)]
7. Correll MJ, Pyle TP, Millar KD, Sun Y, Yao J, Edelmann RE, Kiss JZ. 2013. Transcriptome analyses of *Arabidopsis thaliana* seedlings grown in space: implications for gravity-responsive genes. *Planta* 238:519–533. [[DOI](#)] [[PubMed](#)] [[Google Scholar](#)]
8. Cowles JR, Scheld HW, Lemay R, Peterson C. 1984. Growth and lignification in seedlings exposed to eight days of microgravity. *Annals of Botany* 54:33–48. [[DOI](#)] [[PubMed](#)] [[Google Scholar](#)]
9. Denoux C, Galletti R, Mammarella N, Gopalan S, Werck D, De Lorenzo G, Ferrari S, Ausubel FM, Dewdney J. 2008. Activation of defense response pathways by OGs and Flg22 elicitors in *Arabidopsis* seedlings. *Molecular Plant* 1:423–445. [[DOI](#)] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
10. Do CB, Mahabhashyam MS, Brudno M, Batzoglou S. 2005. ProbCons: probabilistic consistency-based multiple sequence alignment. *Genome Research* 15:330–340. [[DOI](#)] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
11. Edgar RC. 2004. MUSCLE: multiple sequence alignment with high accuracy and high throughput. *Nucleic Acids Research* 32:1792–1797. [[DOI](#)] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
12. Ferl RJ, Koh J, Denison F, Paul A-L. 2014. Spaceflight induces specific alterations in the proteomes of *Arabidopsis*. *Astrobiology* 15:32–56. [[DOI](#)] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
13. Ferl RJ, Paul A-L. 2016. The effect of spaceflight on the gravity-sensing auxin gradient of roots: GFP reporter gene microscopy on orbit. *Npj Microgravity* 2:15023. [[DOI](#)] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
14. Gadjev I, Vanderauwera S, Gechev TS, Laloi C, Minkov IN, Shulaev V, Apel K, Inzé D, Mittler R, Van Breusegem F. 2006. Transcriptomic footprints disclose specificity of reactive oxygen species signaling in *Arabidopsis*. *Plant Physiology* 141:436–445. [[DOI](#)] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
15. Galvez-Valdivieso G, Fryer MJ, Lawson T, Slattery K, Truman W, Smirnov N, Asami T, Davies WJ, Jones AM, Baker NR, Mullineaux PM. 2009. The high light response in *Arabidopsis* involves ABA signaling

between vascular and bundle sheath cells. *The Plant Cell* 21:2143–2162. [[DOI](#)] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]

16. Gangappa SN, Botto JF. 2014. The BBX family of plant transcription factors. *Trends in Plant Science* 19:460–470. [[DOI](#)] [[PubMed](#)] [[Google Scholar](#)]

17. Gechev TS, Van Breusegem F, Stone JM, Denev I, Laloi C. 2006. Reactive oxygen species as signals that modulate plant stress responses and programmed cell death. *BioEssays* 28:1091–1101. [[DOI](#)] [[PubMed](#)] [[Google Scholar](#)]

18. Gendron JM, Pruneda-Paz JL, Doherty CJ, Gross AM, Kang SE, Kay SA. 2012. *Arabidopsis* circadian clock protein, TOC1, is a DNA-binding transcription factor. *Proceedings of the National Academy of Sciences* 109:3167–3172. [[DOI](#)] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]

19. Griffiths S, Dunford RP, Coupland G, Laurie DA. 2003. The evolution of CONSTANS-like gene families in barley, rice, and *Arabidopsis*. *Plant Physiology* 131:1855–1867. [[DOI](#)] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]

20. Herrera-Vásquez A, Carvallo L, Blanco F, Tobar M, Villarroel-Candia E, Vicente-Carbajosa J, Salinas P, Holuigue L. 2015a. Transcriptional control of glutaredoxin GRXC9 expression by a salicylic acid-dependent and NPR1-independent pathway in *Arabidopsis*. *Plant Molecular Biology Reporter* 33:624–637. [[DOI](#)] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]

21. Herrera-Vásquez A, Salinas P, Holuigue L. 2015b. Salicylic acid and reactive oxygen species interplay in the transcriptional control of defense genes expression. *Frontiers in Plant Science* 6:171. [[DOI](#)] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]

22. Hill J, Davis KE. 2014. The Supertree Toolkit 2: a new and improved software package with a Graphical User Interface for supertree construction. *Biodiversity Data Journal* e1053. [[DOI](#)] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]

23. Hoson T, Soga K, Mori R, Saiki M, Nakamura Y, Wakabayashi K, Kamisaka S. 2002. Stimulation of elongation growth and cell wall loosening in rice coleoptiles under microgravity conditions in space. *Plant Cell Physiology* 43:1067–1071. [[DOI](#)] [[PubMed](#)] [[Google Scholar](#)]

24. Huang LJ, Li N, Thurow C, Wirtz M, Hell R, Gatz C. 2016. Ectopically expressed glutaredoxin ROXY19 negatively regulates the detoxification pathway in *Arabidopsis thaliana*. *BMC Plant Biology* 16:200. [[DOI](#)] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]

25. Im YJ, Killens R, Lee AY, Ji M, Lowder C, Grunden AM, Boss WF. 2011. Redesigning plants for spaceflight and beyond: transfer of genes from *Pyrococcus*. *Gravitational Space Research* 23:39. [[Google](#)]

26. Jakic B, Buszko M, Cappellano G, Wick G. 2017. Elevated sodium leads to the increased expression of HSP60 and induces apoptosis in HUVECs. PLoS One 12:e0179383. [[DOI](#)] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
27. Jefferson RA, Kavanagh TA, Bevan MW. 1987. GUS fusions: beta-glucuronidase as a sensitive and versatile gene fusion marker in higher plants. The EMBO Journal 6:3901–3907. [[DOI](#)] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
28. Joo JH, Bae YS, Lee JS. 2001. Role of auxin-induced reactive oxygen species in root gravitropism. Plant Physiology 126:1055–1060. [[DOI](#)] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
29. Jwa NS, Hwang BK. 2017. Convergent evolution of pathogen effectors toward reactive oxygen species signaling networks in plants. Frontiers in Plant Science 8:1687. [[DOI](#)] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
30. Kalbina I, Strid A. 2006. Supplementary ultraviolet-B irradiation reveals differences in stress responses between *Arabidopsis thaliana* ecotypes. Plant, Cell & Environment 29:754–763. [[DOI](#)] [[PubMed](#)] [[Google Scholar](#)]
31. Katoh K, Standley DM. 2013. MAFFT multiple sequence alignment software version 7: improvements in performance and usability. Molecular Biology and Evolution 30:772–780. [[DOI](#)] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
32. Kimbrough JM, Salinas-Mondragon R, Boss WF, Brown CS, Sederoff HW. 2004. The fast and transient transcriptional network of gravity and mechanical stimulation in the *Arabidopsis* root apex. Plant Physiology 136:2790–2805. [[DOI](#)] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
33. Klug A, Schwabe JW. 1995. Protein motifs 5. Zinc fingers. FASEB Journal 9:597–604. [[PubMed](#)] [[Google Scholar](#)]
34. Kusunoki K, Nakano Y, Tanaka K, Sakata Y, Koyama H, Kobayashi Y. 2017. Transcriptomic variation among six *Arabidopsis thaliana* accessions identified several novel genes controlling aluminium tolerance. Plant, Cell & Environment 40:249–263. [[DOI](#)] [[PubMed](#)] [[Google Scholar](#)]
35. Kwon T, Sparks JA, Nakashima J, Allen SN, Tang Y, Blancaflor EB. 2015. Transcriptional response of *Arabidopsis* seedlings during spaceflight reveals peroxidase and cell wall remodeling genes associated with root hair development. American Journal of Botany 102:21–35. [[DOI](#)] [[PubMed](#)] [[Google Scholar](#)]
36. Lagercrantz U, Axelsson T. 2000. Rapid evolution of the family of CONSTANS LIKE genes in plants.

Molecular Biology and Evolution 17:1499–1507. [[DOI](#)] [[PubMed](#)] [[Google Scholar](#)]

37. Laporte D, Olate E, Salinas P, Salazar M, Jordana X, Holuigue L. 2012. Glutaredoxin GRXS13 plays a key role in protection against photooxidative stress in *Arabidopsis*. Journal of Experimental Botany 63:503–515. [[DOI](#)] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]

38. Lassig R, Gutermuth T, Bey TD, Konrad KR, Romeis T. 2014. Pollen tube NAD(P)H oxidases act as a speed control to dampen growth rate oscillations during polarized cell growth. The Plant Journal 78:94–106. [[DOI](#)] [[PubMed](#)] [[Google Scholar](#)]

39. León J, Rojo E, Sánchez-Serrano JJ. 2001. Wound signalling in plants. Journal of Experimental Botany 52:1–9. [[DOI](#)] [[PubMed](#)] [[Google Scholar](#)]

40. Li P, Ainsworth EA, Leahey AD, Ulanov A, Lozovaya V, Ort DR, Bohnert HJ. 2008. *Arabidopsis* transcript and metabolite profiles: ecotype-specific responses to open-air elevated [CO<sub>2</sub>]. Plant, Cell & Environment 31:1673–1687. [[DOI](#)] [[PubMed](#)] [[Google Scholar](#)]

41. Li H, Lin Y, Heath RM, Zhu MX, Yang Z. 1999. Control of pollen tube tip growth by a rop GTPase-dependent pathway that leads to tip-localized calcium influx. The Plant Cell 11:1731–1742. [[DOI](#)] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]

42. Li P, Mane SP, Sioson AA, Robinet CV, Heath LS, Bohnert HJ, Grene R. 2006. Effects of chronic ozone exposure on gene expression in *Arabidopsis thaliana* ecotypes and in *Thellungiella halophila*. Plant, Cell & Environment 29:854–868. [[DOI](#)] [[PubMed](#)] [[Google Scholar](#)]

43. Liu Y, Schmidt B, Maskell DL. 2010. MSAProbs: multiple sequence alignment based on pair hidden Markov models and partition function posterior probabilities. Bioinformatics 26:1958–1964. [[DOI](#)] [[PubMed](#)] [[Google Scholar](#)]

44. Marchler-Bauer A, Bryant H. 2004. CD-Search: protein domain annotations on the fly. Nucleic Acids Research 32:W327–W331. [[DOI](#)] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]

45. Marchler-Bauer A, Derbyshire MK, Gonzales NR, Lu S, Chitsaz F, Geer LY, Geer RC, He J, Gwadz M, Hurwitz DI, Lanczycki CJ, Lu F, Marchler GH, Song JS, Thanki N, Wang Z, Yamashita RA, Zhang D, Zheng C, Bryant SH. 2015. CDD: NCBI's conserved domain database. Nucleic Acids Research 43:D222–D226. [[DOI](#)] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]

46. Markakis MN, De Cnodder T, Lewandowski M, Simon D, Boron A, Balcerowicz D, Doubbo T, Taconnat L, Renou JP, Höfte H, Verbelen JP, Vissenberg K. 2012. Identification of genes involved in the ACC-mediated control of root cell elongation in *Arabidopsis thaliana*. BMC Plant Biology 12:208. [[DOI](#)] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]

47. Masaki T, Tsukagoshi H, Mitsui N, Nishii T, Hattori T, Morikami A, Nakamura K. 2005. Activation tagging of a gene for a protein with novel class of CCT-domain activates expression of a subset of sugar-inducible genes in *Arabidopsis thaliana*. The Plant Journal 43:142–152. [[DOI](#)] [[PubMed](#)] [[Google Scholar](#)]
48. Mazars C, Brière C, Grat S, Pichereaux C, Rossignol M, Pereda-Loth V, Eche B, Boucheron-Dubuisson E, Le Disquet I, Medina FJ, Graziana A, Carnero-Diaz E. 2014. Microgravity induces changes in microsome-associated proteins of *Arabidopsis* seedlings grown on board the international space station. PLoS One 9:e91814. [[DOI](#)] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
49. Miller G, Schlauch K, Tam R, Cortes D, Torres MA, Shulaev V, Dangel JL, Mittler R. 2009. The plant NADPH oxidase RBOHD mediates rapid systemic signaling in response to diverse stimuli. Science Signaling 2:ra45. [[DOI](#)] [[PubMed](#)] [[Google Scholar](#)]
50. Mittler R, Vanderauwera S, Gollery M, Van Breusegem F. 2004. Reactive oxygen gene network of plants. Trends in Plant Science 9:490–498. [[DOI](#)] [[PubMed](#)] [[Google Scholar](#)]
51. Morker KH, Roberts MR. 2011. Light as both an input and an output of wound-induced reactive oxygen formation in *Arabidopsis* leaves. Plant Signaling & Behavior 6:1087–1089. [[DOI](#)] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
52. NCBI Resource Coordinators 2016. Database resources of the National Center for Biotechnology Information. Nucleic Acids Research 44:D7–D19. [[DOI](#)] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
53. Nguyen N, Mirarab S, Warnow T. 2012. MRL and SuperFine+MRL: new supertree methods. Algorithms for Molecular Biology 7:3. [[DOI](#)] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
54. Noctor G, De Paepe R, Foyer CH. 2007. Mitochondrial redox biology and homeostasis in plants. Trends in Plant Science 12:125–134. [[DOI](#)] [[PubMed](#)] [[Google Scholar](#)]
55. Obayashi T, Okamura Y, Ito S, Tadaka S, Aoki Y, Shirota M, Kinoshita K. 2014. ATTED-II in 2014: evaluation of gene coexpression in agriculturally important plants. Plant & Cell Physiology 55:e6. [[DOI](#)] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
56. Passardi F, Dobias J, Valério L, Guimil S, Penel C, Dunand C. 2007. Morphological and physiological traits of three major *Arabidopsis thaliana* accessions. Journal of Plant Physiology 164:980–992. [[DOI](#)] [[PubMed](#)] [[Google Scholar](#)]
57. Paul AL, Amalfitano CE, Ferl RJ. 2012a. Plant growth strategies are remodeled by spaceflight. BMC Plant Biology 12:232. [[DOI](#)] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]

58. Paul A-L, Popp MP, Gurley WB, Guy C, Norwood KL, Ferl RJ. 2005. *Arabidopsis* gene expression patterns are altered during spaceflight. *Advances in Space Research* 36: 1175–1181. [[Google Scholar](#) ]
59. Paul A-L, Sng NJ, Zupanska AK, Krishnamurthy A, Schultz ER, Ferl RJ. 2017. Genetic dissection of the *Arabidopsis* spaceflight transcriptome: are some responses dispensable for the physiological adaptation of plants to spaceflight? *PLoS One* 12:e0180186. [[DOI](#) ] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#) ]
60. Paul A-L, Wheeler RM, Levine HG, Ferl RJ. 2013a. Fundamental plant biology enabled by the space shuttle. *American Journal of Botany* 100:226–234. [[DOI](#) ] [[PubMed](#)] [[Google Scholar](#) ]
61. Paul A-L, Zupanska AK, Ostrow DT, Zhang Y, Sun Y, Li JL, Shanker S, Farmerie WG, Amalfitano CE, Ferl RJ. 2012b. Spaceflight transcriptomes: unique responses to a novel environment. *Astrobiology* 12:40–56. [[DOI](#) ] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#) ]
62. Paul A-L, Zupanska AK, Schultz ER, Ferl RJ. 2013b. Organ-specific remodeling of the *Arabidopsis* transcriptome in response to spaceflight. *BMC Plant Biology* 13:112. [[DOI](#) ] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#) ]
63. Pettersen EF, Goddard TD, Huang CC, Couch GS, Greenblatt DM, Meng EC, Ferrin TE. 2004. UCSF Chimera—a visualization system for exploratory research and analysis. *Journal of Computational Chemistry* 25:1605–1612. [[DOI](#) ] [[PubMed](#)] [[Google Scholar](#) ]
64. Potocký M, Jones MA, Bezvoda R, Smirnoff N, Zárský V. 2007. Reactive oxygen species produced by NADPH oxidase are involved in pollen tube growth. *The New Phytologist* 174:742–751. [[DOI](#) ] [[PubMed](#)] [[Google Scholar](#) ]
65. Price MN, Dehal PS, Arkin AP. 2010. FastTree 2-approximately maximum-likelihood trees for large alignments. *PLoS One* 5:e9490. [[DOI](#) ] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#) ]
66. Robson F, Costa MM, Hepworth SR, Vizir I, Piñeiro M, Reeves PH, Putterill J, Coupland G. 2001. Functional importance of conserved domains in the flowering-time gene *CONSTANS* demonstrated by analysis of mutant alleles and transgenic plants. *The Plant Journal* 28:619–631. [[DOI](#) ] [[PubMed](#)] [[Google Scholar](#) ]
67. Rosenwasser S, Fluhr R, Joshi JR, Leviatan N, Sela N, Hetzroni A, Friedman H. 2013. ROSMETER: a bioinformatic tool for the identification of transcriptomic imprints related to reactive oxygen species type and origin provides new insights into stress responses. *Plant Physiology* 163:1071–1083. [[DOI](#) ] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#) ]
68. Roshan U, Livesay DR. 2006. Probalign: multiple sequence alignment using partition function posterior probabilities. *Bioinformatics* 22:2715–2721. [[DOI](#) ] [[PubMed](#)] [[Google Scholar](#) ]



69. Schüler O, Hemmersbach R, Böhmer M. 2015. A bird's-eye view of molecular changes in plant gravitropism using omics techniques. *Frontiers in Plant Science* 6:1176. [[DOI](#)] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
70. Schultz ER, Zupanska AK, Sng NJ, Paul A-L, Ferl RJ. 2017. Skewing in *Arabidopsis* roots involves disparate environmental signaling pathways. *BMC Plant Biology* 17:31. [[DOI](#)] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
71. Schwarzländer M, Finkemeier I. 2012. Mitochondrial energy and redox signaling in plants. *Antioxidants & Redox Signaling* 18:2122–2144. [[DOI](#)] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
72. Shigeto J, Tsutsumi Y. 2016. Diverse functions and reactions of class III peroxidases. *The New Phytologist* 209:1395–1402. [[DOI](#)] [[PubMed](#)] [[Google Scholar](#)]
73. Shin R, Schachtman DP. 2004. Hydrogen peroxide mediates plant root cell response to nutrient deprivation. *Proceedings of the National Academy of Sciences* 101:8827–8832. [[DOI](#)] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
74. Sievers F, Wilm A, Dineen D, Gibson TJ, Karplus K, Li W, Lopez R, McWilliam H, Remmert M, Soding J, Thompson JD, Higgins DG. 2011. Fast, scalable generation of high-quality protein multiple sequence alignments using Clustal Omega. *Molecular Systems Biology* 7:539. [[DOI](#)] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
75. Sng N, Callahan J, Ferl RJ, Paul A-L. 2014. *Arabidopsis thaliana* for spaceflight applications – preparing dormant biology for passive stowage and on orbit activation. *Gravitational Space Research* 2:2 [[Google Scholar](#)]
76. Soga K, Wakabayashi K, Kamisaka S, Hoson T. 2002. Stimulation of elongation growth and xyloglucan breakdown in *Arabidopsis* hypocotyls under microgravity conditions in space. *Planta* 215:1040–1046. [[DOI](#)] [[PubMed](#)] [[Google Scholar](#)]
77. Stamatakis A. 2014. RAxML version 8: a tool for phylogenetic analysis and post-analysis of large phylogenies. *Bioinformatics* 30:1312–1313. [[DOI](#)] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
78. Sugimoto M, Oono Y, Gusev O, Matsumoto T, Yazawa T, Levinskikh MA, Sychev VN, Bingham GE, Wheeler R, Hummerick M. 2014. Genome-wide expression analysis of reactive oxygen species gene network in Mizuna plants grown in long-term spaceflight. *BMC Plant Biology* 14:4. [[DOI](#)] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
79. Suzuki N, Koussevitzky S, Mittler R, Miller G. 2012. ROS and redox signalling in the response of plants to abiotic stress. *Plant, Cell & Environment* 35:259–270. [[DOI](#)] [[PubMed](#)] [[Google Scholar](#)]

80. Swanson S, Gilroy S. 2010. ROS in plant development. *Physiologia Plantarum* 138:384–392. [[DOI](#)] [[PubMed](#)] [[Google Scholar](#)]
81. Talavera G, Castresana J. 2007. Improvement of phylogenies after removing divergent and ambiguously aligned blocks from protein sequence alignments. *Systems Biology* 56:564–577. [[DOI](#)] [[PubMed](#)] [[Google Scholar](#)]
82. Teige M, Scheikl E, Eulgem T, Dóczi R, Ichimura K, Shinozaki K, Dangl JL, Hirt H. 2004. The MKK2 pathway mediates cold and salt stress signaling in *Arabidopsis*. *Molecular Cell* 15:141–152. [[DOI](#)] [[PubMed](#)] [[Google Scholar](#)]
83. Wheeler WC, Gatesy J, DeSalle R. 1995. Elision: a method for accommodating multiple molecular sequence alignments with alignment-ambiguous sites. *Molecular Phylogenetics and Evolution* 4:1–9. [[DOI](#)] [[PubMed](#)] [[Google Scholar](#)]
84. Winter D, Vinegar B, Nahal H, Ammar R, Wilson GV, Provart NJ. 2007. An “electronic fluorescent pictograph” browser for exploring and analyzing large-scale biological data sets. *PLoS One* 2:e718. [[DOI](#)] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
85. Yan H, Marquardt K, Indorf M, Jutt D, Kircher S, Neuhaus G, Rodríguez-Franco M. 2011. Nuclear localization and interaction with COP1 are required for STO/BBX24 function during photomorphogenesis. *Plant Physiology* 156:1772–1782. [[DOI](#)] [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
86. Zander M, Chen S, Imkampe J, Thurow C, Gatz C. 2012. Repression of the *Arabidopsis thaliana* jasmonic acid/ethylene-induced defense pathway by TGA-interacting glutaredoxins depends on their C-terminal ALWL motif. *Molecular Plant* 5:831–840. [[DOI](#)] [[PubMed](#)] [[Google Scholar](#)]
87. Zhang Y, Wang L, Xie J, Zheng H. 2014. Differential protein expression profiling of *Arabidopsis thaliana* callus under microgravity on board the Chinese SZ-8 spacecraft. *Planta* 241:475–488. [[DOI](#)] [[PubMed](#)] [[Google Scholar](#)]
88. Zupanska AK, Denison FC, Ferl RJ, Paul A-L. 2013. Spaceflight engages heat shock protein and other molecular chaperone genes in tissue culture cells of *Arabidopsis thaliana*. *American Journal of Botany* 100:235–248. [[DOI](#)] [[PubMed](#)] [[Google Scholar](#)]

## Associated Data

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*This section collects any data citations, data availability statements, or supplementary materials included in this article.*



## Supplementary Materials

### Supplementary Material

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