## 431 Class 08

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### Today's Agenda

- Working again with the dm1000 data from Class 07
- Confidence Intervals around a Mean
- Building Visualizations to Compare Distributions
- Confidence Intervals for a Difference between Means

### Today's Packages

```
1 library(broom) ## tidying model output
2 library(ggridges) ## help building ridgeline plots
3 library(Hmisc) ## smean.cl.boot() and smean.cl.norm()
4 library(janitor)
5 library(kableExtra) ## for table neatening
6 library(naniar)
7 library(patchwork)
8 library(readxl) ## new today, read in .xls or .xlsx files
9 library(tidyverse)
10
11 theme_set(theme_bw())
```

### Data Ingest

Today, we'll use an Excel file (.xls, rather than .csv) to import the dm1000 data.

```
1 dm1000 <- read_excel("c08/data/dm_1000.xls") |>
2   clean_names() |>
3   mutate(across(where(is.character), as_factor)) |>
4   mutate(subject = as.character(subject))
```

- The readxl package is a non-core part of the tidyverse, and also includes functions called read\_xls() and read\_xlsx().
- Visit https://readxl.tidyverse.org/.

#### The dm1000 tibble

dm1000# A tibble: 1,000 × 17 age insurance n income subject ht sbp dbp ldl wt a1c tobacco  $\langle chr \rangle$ <dbl> <fct> <dbl> <dbl> <dbl> <dbl> <dbl> <dbl> <fct> 1.63 103. 1 M - 0.00155 Medicaid 29853 145 70 6.4 221 Current 2 M - 000252 Commercial 31248 1.75 112. 151 77 8.5 116 Never 3 M - 000369 Medicare 1.65 74.9 127 8.9 52 23362 73 Former 4 M - 000457 Medicaid 26033 1.63 81.4 125 6.8 122 Never 74 1.69 10.3 5 M-0005 68 Medicare 85374 92.6 120 73 94 Never 6 M-0006 56 Medicaid 31273 1.71 54.6 127 75 12.3 NA Current 7 M - 000754 Commercial 25445 1.68 81.6 114 81 6.5 100 Current  $\bigcirc$   $\mathbf{x}_{\mathbf{f}}$   $\bigcirc$   $\bigcirc$   $\bigcirc$   $\bigcirc$ 

# Estimating a Population Mean from a Sample

Suppose our sample in dm1000 is a random sample from the population of all Cuyahoga County residents between the ages of 31-75 receiving care for diabetes.

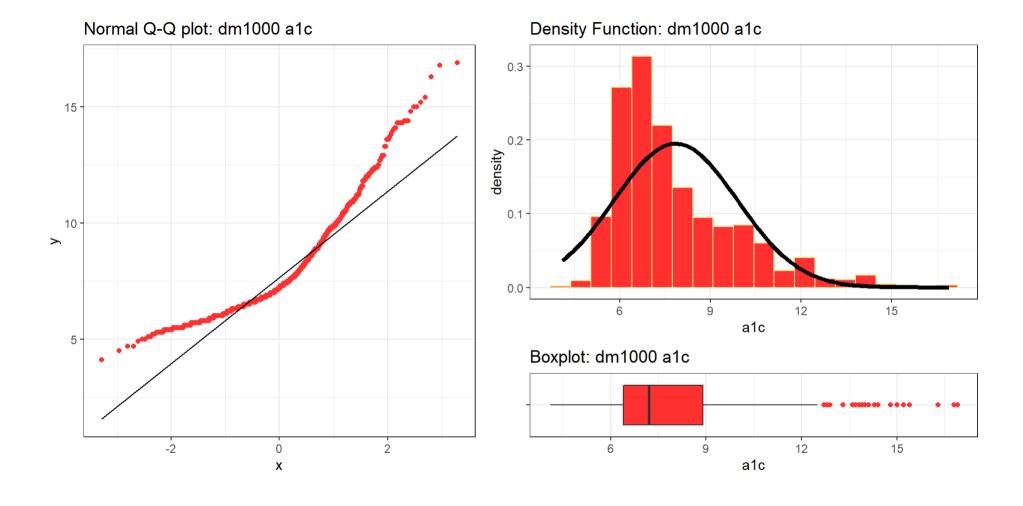
What's a good estimate for the mean Hemoglobin A1c of the people in that population?

- How would we make this estimate using our data?
- What would we want to know about the data?
- DTDP

### Hemoglobin A1c in the dm1000 sample

```
p1 \leftarrow ggplot(dm1000, aes(sample = a1c)) +
    geom qq(col = "firebrick1") +
    geom gg line(col = "black") +
    theme(aspect.ratio = 1) +
 5
     labs(title = "Normal Q-Q plot: dm1000 a1c")
   p2 \leftarrow ggplot(dm1000, aes(x = a1c)) +
     geom histogram (aes (y = stat(density)),
 9
                     bins = 20, fill = "firebrick1", col = "khaki") +
10
     stat function (fun = dnorm,
11
                    args = list(mean = mean(dm1000$a1c, na.rm = TRUE),
12
                                 sd = sd(dm1000\$a1c, na.rm = TRUE)),
13
                    col = "black", lwd = 1.5) +
14
     labs(title = "Density Function: dm1000 a1c")
15
   p3 < - ggplot(dm1000, aes(x = a1c, y = "")) +
17
    geom boxplot(fill = "firebrick1", outlier.color = "firebrick1") +
18
    labs(title = "Boxplot: dm1000 a1c", v = "")
```

### Hemoglobin A1c in the dm1000 sample



### Numerical Summaries of A1c in dm1000

 Should we assume the A1c values are drawn from a Normal distribution?

```
1 mosaic::favstats(~ a1c, data = dm1000) |>
2   kbl(digits = 2) |>
3   kable_styling(font_size = 32, full_width = FALSE)
```

 min	Q1	median	Q3	max	mean	sd	n	missing
4.1	6.4	7.2	8.9	16.9	7.85	2.04	985	15

- The kbl() and kable\_styling() functions are from the kableExtra package.
  - Read more about kableExtra in its vignette.

## What if we assume the A1cs are Normally distributed?

Then we could use a linear regression model to obtain a 95% confidence interval for the population mean.

#### What is the model we've fit here?

```
1 m1

Call:
lm(formula = alc ~ 1, data = dm1000)

Coefficients:
(Intercept)
7.853
```

 This "intercept only" model simply predicts the mean value of our outcome, a1c.

### Interpreting this interval? (1/3)

• Our 95% confidence interval for the population mean A1c is (7.73, 7.98).

We are estimating the mean of the **population** based on a mean from our *sample* of 1000 observations taken (at random, we assume) from that population.

• Sadly, this **doesn't** mean we're 95% confident that the actual population mean is in that range, even though lots and lots of people (incorrectly) assume it does.

### Interpreting this interval? (2/3)

• Our 95% confidence interval for the population mean A1c is (7.73, 7.98).

Essentially, we have 95% confidence in the **process** of fitting confidence intervals this way. If we fit 100 such confidence intervals to a variety of data sets, we have some reason to anticipate that 95 of them will contain the actual unknown value of the population mean.

### Interpreting this interval? (3/3)

• Our 95% confidence interval for the population mean A1c is (7.73, 7.98).

That's oversimplifying a little, and a particular concern in this case is that the data are somewhat skewed (at any rate, not very close to Normally distributed) and this may impact our ability to generate accurate and efficient confidence intervals via a linear model like this.

### An Equivalent Approach

We could use a t test to obtain a 95% confidence interval for the population mean.

• This is exactly the same result as we obtain from the linear model m1.

### **Another Equivalent Approach**

We could use a function called smean.cl.normal() from the Hmisc package to obtain the same 95% confidence interval for the population mean.

```
1 smean.cl.normal(dm1000$a1c, conf.int = 0.95)
Mean Lower Upper
7.852893 7.725167 7.980619
```

Again, this is the same result as we have seen previously.

# What if we weren't willing to assume that the A1c values came from a Normal distribution?

 Use the bootstrap to estimate the population mean with 95% confidence.

```
1 set.seed(2022431) # why do we set a seed here?
2 smean.cl.boot(dm1000$a1c, conf.int = 0.95)
Mean Lower Upper
```

```
7.852893 7.726066 7.979513
```

### Bootstrap CI with a different seed

```
1 set.seed(1234567) # what happens if we change the seed
2 smean.cl.boot(dm1000$a1c, conf.int = 0.95)
```

```
Mean Lower Upper 7.852893 7.725378 7.978716
```

• The smean.cl.boot() function comes from the Hmisc package.

Bootstrap is a resampling method where large numbers of samples of the same size are repeatedly drawn, with replacement, from a single original sample.

### 95% Cls for Population Mean A1c

Approach	<b>Estimate</b>	95% CI	Assume Normality?		
Linear Model	7.853	(7.725, 7.981)	Yes		
Bootstrap	7.853	(7.726, 7.978)	No		

- How does this match up with our understanding of the distribution of the A1c data?
- What's an appropriate number of decimal places to use here?

```
1 dm1000 |> select(a1c) |> head(15) |> as.vector()

$a1c

[1] 6.4 8.5 8.9 6.8 10.3 12.3 6.5 5.4 7.1 8.9 6.6 6.4 11.1 11.8 6.5
```

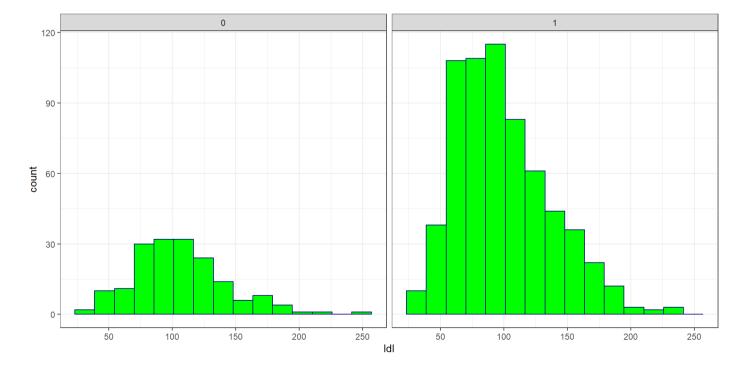
• Might the fairly large sample size (n = 985 non-missing) have something to do with these results?

# Comparing Two Distributions

# Is LDL higher or lower among adults with diabetes who have a statin prescription?

```
1 ggplot(data = dm1000, aes(x = ldl)) +
2  geom_histogram(bins = 15, fill = "green", col = "navy") +
3  facet_wrap(~ statin)
```

Warning: Removed 178 rows containing non-finite values (stat\_bin).

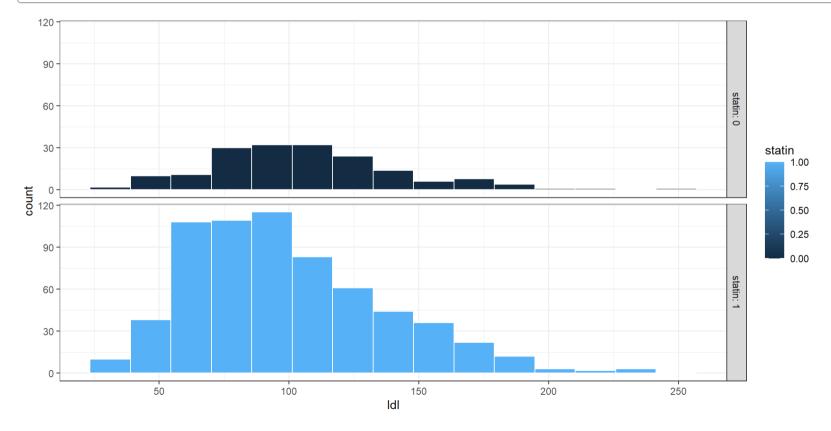


### How might we improve this plot?

- 1. Remove the warning about non-finite (missing) values.
- 2. Place the histograms vertically to ease comparisons.
- 3. Fill the histograms differently for statin and no statin.
- 4. Augment the labels (0 and 1) to show they identify statin use.

### LDL stratified by statin (Plot 2)

```
1 tempdat <- dm1000 |> filter(complete.cases(ldl, statin))
2
3 ggplot(data = tempdat, aes(x = ldl, fill = statin)) +
4    geom_histogram(bins = 15, col = "white") +
5    facet_grid(statin ~ ., labeller = "label_both")
```

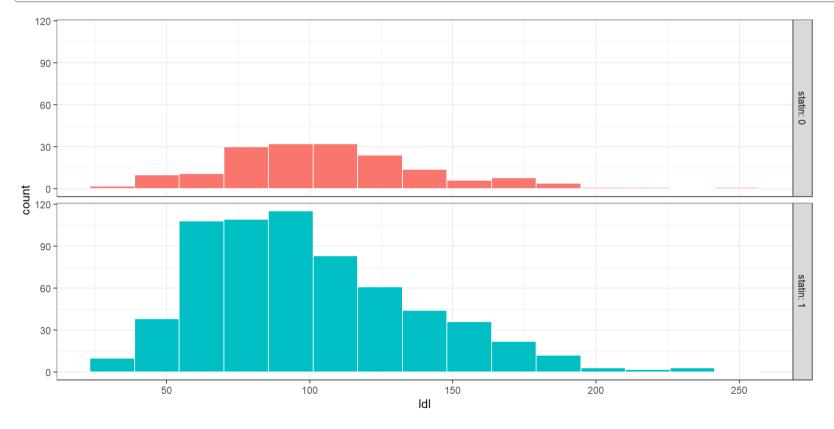


### Problems with the previous plot

- 1. Statin is actually a two-category variable (1 and 0 are just codes) but the legend is treating it as if it was a numeric variable.
- 2. Do we actually need the legend (called a guide in R) or can we remove it?

### But statin is categorical? (Plot 3)

```
1 tempdat <- dm1000 |> filter(complete.cases(ldl, statin))
2 ggplot(data = tempdat, aes(x = ldl, fill = factor(statin))) +
3    geom_histogram(bins = 15, col = "white") +
4    facet_grid(statin ~ ., labeller = "label_both") +
5    guides(fill = "none")
```



### **Faceting**

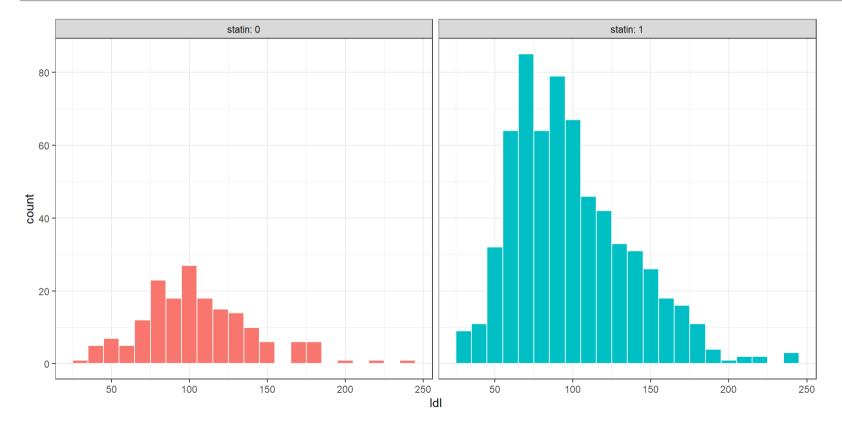
It's very useful to split data into groups and plot each group separately to make comparisons across the groups. We can then draw those subplots side by side.

We have two main tools: facet\_wrap() and facet\_grid()

- facet\_wrap(~ grp1) to obtain plots within each grp1 arranged into horizontal subpanels and wrapping around, like words on a page.
- facet\_grid(grp1 ~ .) to obtain plots within each grp1 arranged vertically (vertical subpanels)
- facet\_grid(grp1 ~ grp2) to obtain plots within each combination of grp1 and grp2 with vertical and horizontal subpanels.

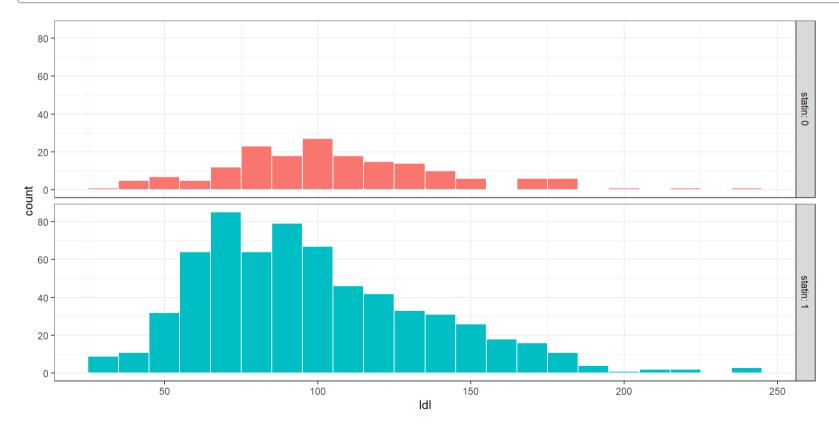
### Using facet\_wrap()

```
1 tempdat <- dm1000 |> filter(complete.cases(ldl, statin))
2 ggplot(data = tempdat, aes(x = ldl, fill = factor(statin))) +
3    geom_histogram(binwidth = 10, col = "white") +
4    facet_wrap(~ statin, labeller = "label_both") +
5    guides(fill = "none")
```



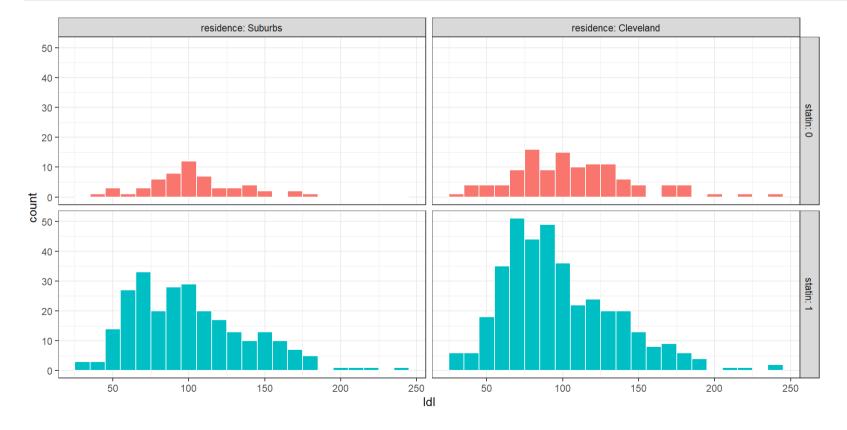
### Using facet\_grid()

```
1 tempdat <- dm1000 |> filter(complete.cases(ldl, statin))
2 ggplot(data = tempdat, aes(x = ldl, fill = factor(statin))) +
3    geom_histogram(binwidth = 10, col = "white") +
4    facet_grid(statin ~ ., labeller = "label_both") +
5    guides(fill = "none")
```



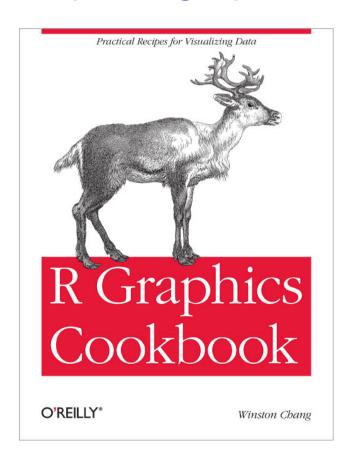
### facet\_grid(): two groupings

```
1 tempdat <- dm1000 |> filter(complete.cases(ldl, statin, residence))
2 ggplot(data = tempdat, aes(x = ldl, fill = factor(statin))) +
3    geom_histogram(binwidth = 10, col = "white") +
4    facet_grid(statin ~ residence, labeller = "label_both") +
5    guides(fill = "none")
```



# My Main Source for ggplot2 Visualization Recipes

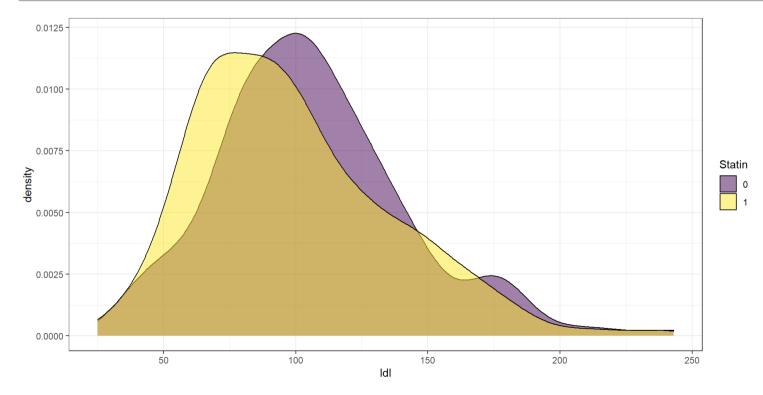
https://r-graphics.org/ (Second Edition)



### Comparison of densities

#### This plot ignores the relative frequencies.

```
1 tempdat <- dm1000 |> filter(complete.cases(ldl, statin))
2 ggplot(data = tempdat, aes(x = ldl, fill = factor(statin))) +
3    geom_density(alpha = 0.5) + scale_fill_viridis_d() +
4    labs(fill = "Statin")
```



### **Numerical Summaries for Two Groups**

statin	n	min	med	max	mean	sd
0	176	34	102	243	106.22	36.05
1	646	25	92	241	99.22	37.21

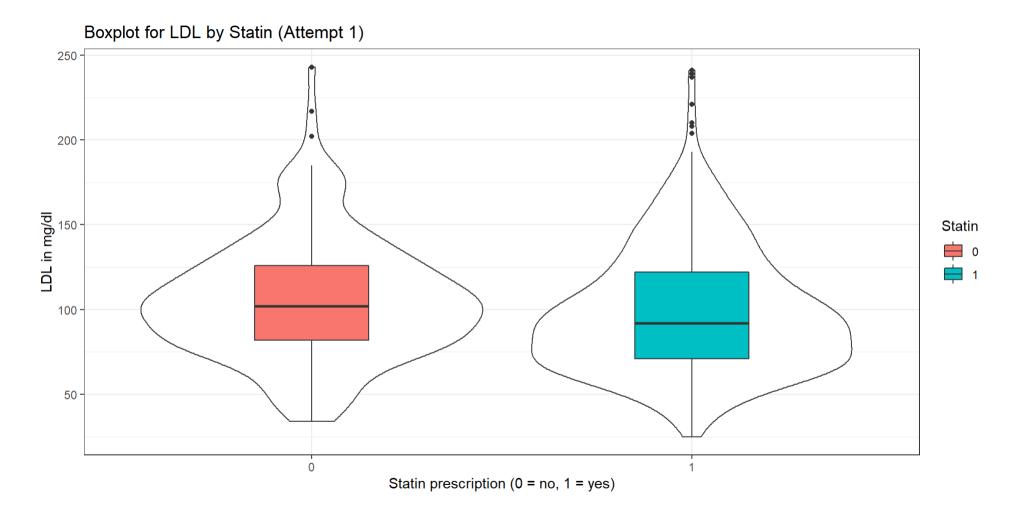
Difference in mean(LDL) between the two samples?

### Using favstats for LDL by Statin

### Comparison Boxplot (LDL by statin)

#### Attempt 1

### Comparison Boxplot (LDL by statin)

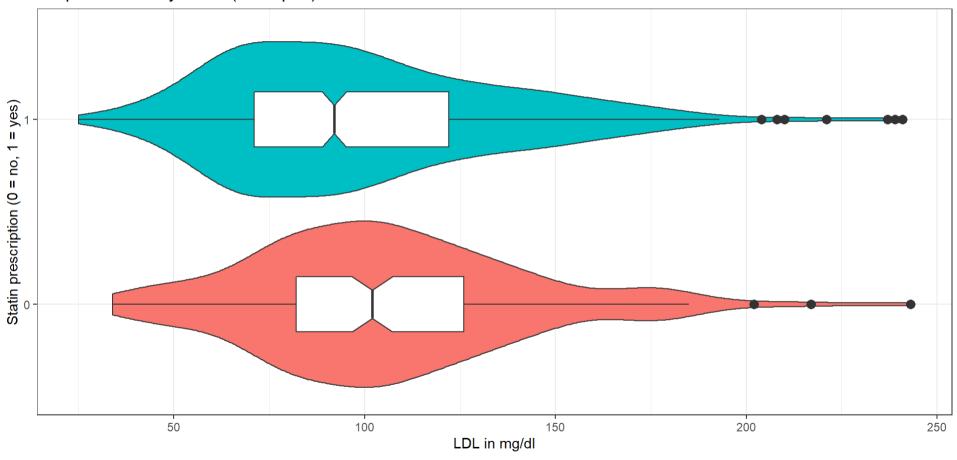


### Try 2: Boxplot for LDL and statin

```
1 tempdat <- dm1000 |> filter(complete.cases(ldl, statin))
2
3 ggplot(data = tempdat, aes(x = factor(statin), y = ldl)) +
4    geom_violin(aes(fill = factor(statin))) +
5    geom_boxplot(width = 0.3, outlier.size = 3, notch = TRUE) +
6    coord_flip() +
7    guides(fill = "none") +
8    labs(x = "Statin prescription (0 = no, 1 = yes)",
9    y = "LDL in mg/dl",
10    title = "Boxplot for LDL by Statin (Attempt 2)")
```

### Try 2: Boxplot for LDL and statin

Boxplot for LDL by Statin (Attempt 2)



## Setting Up Third Try

# Third Try on Boxplot for LDL by Statin Use

# Third Try on Boxplot for LDL by Statin Use

Boxplot for LDL by Statin (Attempt 3) Statin Statin prescription No Statin 200 50 100 150 250 LDL in mg/dl

### 95% CI for difference in means

We want to estimate the difference between the population mean LDL WITH statin and population mean LDL WITHOUT statin.

#### The sample means, you'll remember, are:

```
1 mosaic::favstats(ldl ~ statin, data = dm1000) |>
2  select(statin, n, mean, sd, missing) |>
3  kbl(digits = 2) |> kable_styling(font_size = 24)
```

statin	n	mean	sd	missing
0	176	106.22	36.05	66
1	646	99.22	37.21	112

### 95% CI for difference in means

 If we are willing to assume that LDL follows a Normal distribution in each statin group, then we can use a linear model with one predictor.

```
1 m2 <-lm(ldl \sim statin, data = dm1000)
```

#### Coefficients of Model m2

```
1 m2
Call:
lm(formula = ldl ~ statin, data = dm1000)
Coefficients:
(Intercept) statin
    106.2 -7.0
 1 tidy(m2, conf.int = TRUE, conf.level = 0.95) |>
    select (term, estimate, conf.low, conf.high)
# A tibble: 2 \times 4
 term estimate conf.low conf.high
 1 (Intercept) 106. 101. 112.
2 statin -7.00 -13.2 -0.831
```

## Using t.test to get same result

```
1 mosaic::favstats(ldl ~ statin, data = dm1000) |>
2  select(statin, n, mean, sd, missing) |>
3  kbl(digits = 2) |> kable_styling(font_size = 24)
```

statin	n	mean	sd	missing
0	176	106.22	36.05	66
1	646	99.22	37.21	112

estimate	conf.low	conf.high	
7	0.831	13.17	

# 95% CI for difference between population means via the bootstrap

If we are not willing to assume a Normal distribution for LDL in either the statin or the "no statin" group, then we could use a bootstrap approach.

#### The bootdif function

#### from Love-boost.R

```
`bootdif` <-
     function (y, q, conf.level=0.95, B.reps = 2000) {
        lowq = (1 - conf.level)/2
       q <- as.factor(q)
 4
 5
        a <- attr(Hmisc::smean.cl.boot(y[q==levels(q)[1]],
                               B=B.reps, reps=TRUE), 'reps')
       b <- attr(Hmisc::smean.cl.boot(y[q==levels(q)[2]],
                               B=B.reps, reps=TRUE), 'reps')
       meandif <- diff(tapply(y, q, mean, na.rm=TRUE))</pre>
 9
       a.b <- quantile(b-a, c(lowg, 1-lowg))
10
11
       res <- c(meandif, a.b)
12
       names(res) <- c('Mean Difference', lowg, 1-lowg)</pre>
13
       res
14
```

# Assumptions behind our intervals Assumptions these intervals share:

- random samples from the populations of interest
- independent samples (samples aren't paired or matched)

### Additional assumptions for linear model:

- Normal distribution in each group (statin and "no statin")
- variance in each group (statin and "no statin") is equal

### 95% confidence intervals for mean LDL

95% CIs for LDL  $\mu_{NoStatin} - \mu_{Statin}$ 

Approach	<b>Estimate</b>	95% CI
linear model	7.00	(0.83, 13.17)
bootstrap	7.00	(0.82, 13.31)

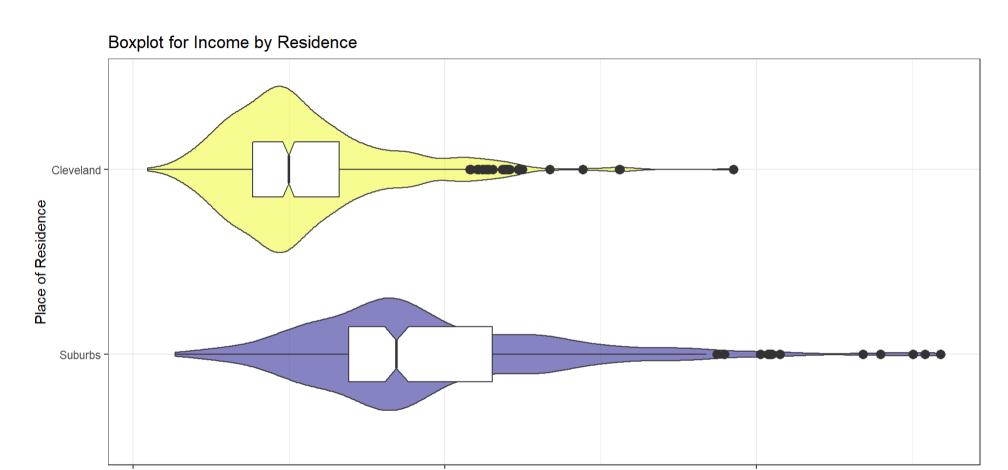
Are our conclusions meaningfully different if we do (or do not) assume Normal population distributions of LDL within each group (statin and no statin)?

## Comparing Income by Residence

```
1 tempdat <- dm1000 |> filter(complete.cases(n_income, residence))
2
3 ggplot(data = tempdat, aes(x = residence, y = n_income)) +
4    geom_violin(aes(fill = residence)) +
5    geom_boxplot(width = 0.3, outlier.size = 3, notch = TRUE) +
6    coord_flip() + guides(fill = "none") +
7    scale_fill_viridis_d(alpha = 0.5, option = "C") +
8    labs(x = "Place of Residence",
9    y = "Median Neighborhood Income ($)",
10    title = "Boxplot for Income by Residence")
```

# Comparing Income by Residence

0e+00



5e+04

Median Neighborhood Income (\$)

1e+05

### Sample Means, Medians

```
1 mosaic::favstats(n_income ~ residence, data = dm1000) |>
2  select(residence, n, mean, median, sd, missing) |>
3  kbl(digits = 0) |> kable_styling(font_size = 32)
```

residence	n	mean	median	sd	missing
Suburbs	371	47251	42223	20333	0
Cleveland	601	27725	24907	13031	0

# Estimating 90% CI for Difference in Means Linear Model for Income by Residence

#### **Bootstrap Approach**

### 90% CI for difference in mean Income

90% CIs for Income  $\mu_{Suburbs} - \mu_{Cleveland}$ 

Approach	<b>Estimate</b>	90% CI
linear model	19527	(17765, 21289)
bootstrap	19527	(17545, 21469)

Are our conclusions meaningfully different if we do (or do not) assume Normal population distributions of neighborhood income within each residence group?

### Save the dm1000 tibble as an R data set

Preserves all changes in your R work (factors, etc.)

We'll load this in when we work with these data in Class 09.

### **Session Information**

```
1 sessionInfo()
R version 4.2.1 (2022-06-23 ucrt)
Platform: x86 64-w64-mingw32/x64 (64-bit)
Running under: Windows 10 x64 (build 22000)
Matrix products: default
locale:
[1] LC COLLATE=English United States.utf8
[2] LC_CTYPE=English United States.utf8
[3] LC MONETARY=English United States.utf8
[4] LC NUMERIC=C
[5] LC TIME=English United States.utf8
attached base packages:
[1] stats graphics grDevices utils datasets methods
                                                                base
```