# Quick Guide

# *NEAT for RNAseq*

# NGS pipelines made easy

Patrick Schorderet

Patrick.schorderet@molbio.mgh.harvard.edu

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# 1 Introduction

The ***NE***xt generation ***A***nalysis ***T***oolbox (NEAT) is a perl/R package that supports users during the analysis of next generation sequencing (NGS) data.

NEAT provides an easy, rapid and reproducible exploratory data analysis (EDA) tool that allows users to assess their data in less than 24 hours (based on a 200mio read Highseq run). NEAT was developed in two main sections, ChIPpip (RNApip) and ChIPmE (RNAmE). The first section (RNApip) helps users with jobs that are computationally demanding (mapping, filtering, etc). The second section (RNAmE) consists of jobs that can be run locally (on a desktop computer). All four sections are standalone applications.

This quick guide is intended for users who want to run a quick analysis and are comfortable using the default parameters and algorithms. Please refer to the complete guide for more details on in depth analysis.

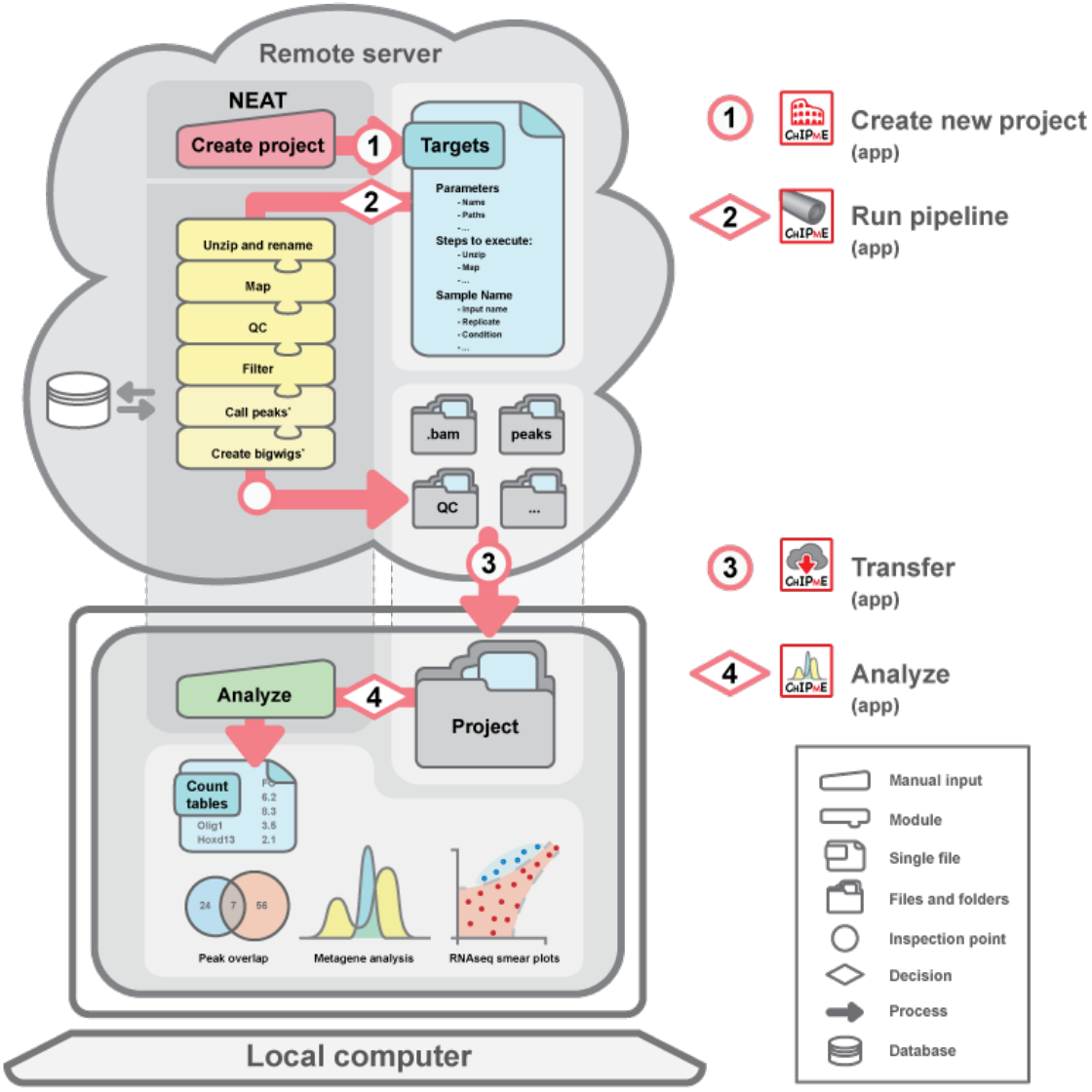



**Fig.1 NEAT architecture.** NGS data can be analyzed using NEAT in less than a day. Users follow a logical 4-step process, including the creation of a new project, running the pipeline on a remote server or in the cloud, transferring the data to a local computer and proceeding to the analysis.

# 2 NEAT

## 2.1 Introduction to NEAT

A central feature of NEAT is its ability to perform repetitive tasks on complex sample setups while managing batch submissions and cluster queuing. NEAT can easily be implemented in any institution with limited to no programming experience. The workflow has been designed to efficiently run on a computer cluster using a distributed resource manager such as TORQUE. NEAT has been developed by and for wet-lab scientists as well as bioinformaticiens to ensure user-friendliness, management of complicated experimental setups and reproducibility in the big data era.

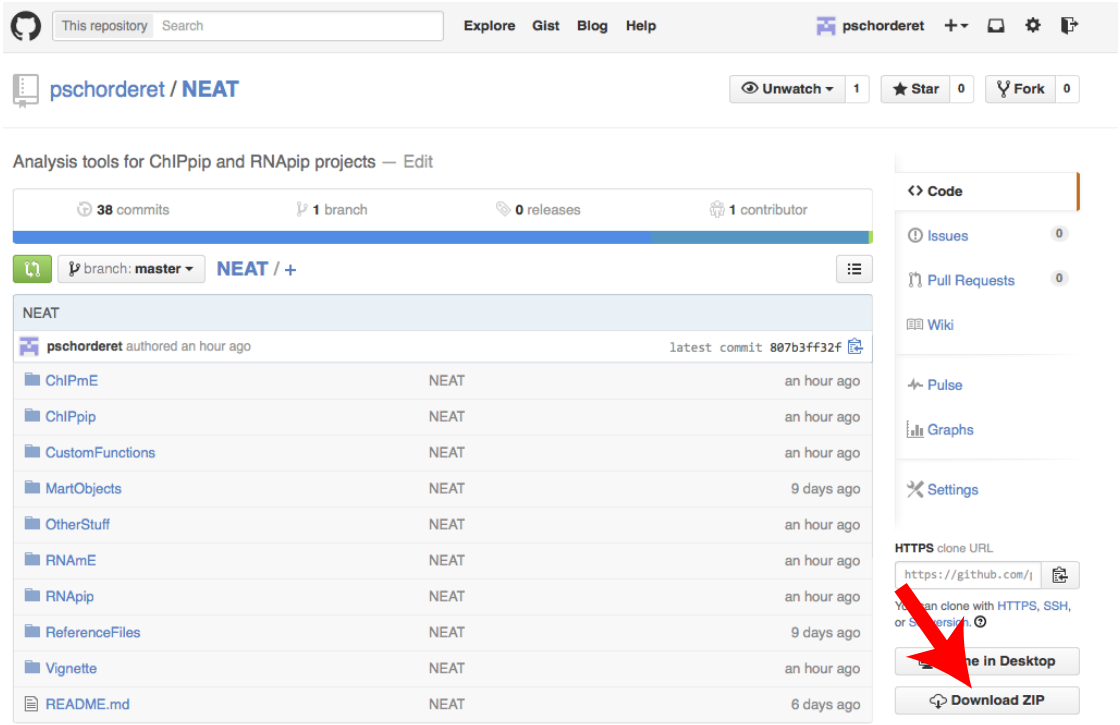
To start using NEAT for RNAseq data analysis, please follow the tutorial. This will walk you through the analysis of a small test dataset (provided as part of RNApip) using your own computer cluster. This will also ensure NEAT and its dependencies are correctly installed before submitting large, memory-savvy analysis.

All fastq files from the test data have been subsetted to ca. 15'000 reads. This data comes from an unpublished 50bp single end (SE) sequencing experiment although NEAT can deal with paired-end (PE) sequencing as well. For more information on the test data provided in this tutorial, please read below.

Although this quick guide is intended for scientists with no programming experience, users will require access to a remote server. Users are thereby required to have SSH accessibility with a username and a password. Please refer to your system administrator to obtain such credentials.

## 2.2 Install NEAT

To install NEAT, download the NEAT repository from GitHub to any directory on your computer cluster.




In this tutorial, we will suppose the *NEAT/* directory was saved to the user's desktop on a local computer (*~/Desktop*) and that it will be saved to their home directory (*/home/*) on the remote server.

Make sure the folder is named *NEAT* and not *NEAT-master*. Start the transfer to the remote folder by typing the following command line in a terminal window:

```
[~Desktop]$ rsync -avz ~/Desktop/NEAT username@serveradress.edu:/home/
```

Enter your password and allow for the folders to transfer.

# 3 NEAT Part 1 : RNApip

## 3.1 Before running NEAT for RNAseq

Please make sure all R packages required for NEAT (RNApip and RNAmE) are installed on your computer (refer to the R manual) prior to running NEAT. Refer to the *Version information and required packages* section below for more information.

In addition, all four NEAT standalone applications run through *Automator*, a MacOS application standardly installed on most modern Apple computers, please make sure it is installed on your computer.

Finally, make sure your *Terminal* software is closed before launching RNAmE.

## 3.2 Running NEAT part 1.1 (RNApip)

### 3.2.1 Creating a new RNApip project

The first step to run NEAT on ChIPseq data is to create a new RNApip project. Double-click the *1_NewProject.app* found in the NEAT directory *~/Desktop/NEAT/*RNA*mE/* and follow the prompts.


#### 3.2.1.1 Filling in the Targets.txt file

Creating a new project should create the central part of NEAT: the Targets.txt file. The Targets.txt file can be found in the *~/MY_NEW_RNA_PROJECT/DataStructure/* directory and is the backbone of NEAT. It contains all the information specific to your experiment and your computer cluster, including the names of files, the paths to the reference genomes, the steps to execute, the name of your samples, their relationships, etc.

**IMPORTANT**: Sample names including inputs and fastq files cannot start with the letter 'n' (small or capital) as this is the universal perl symbol for carriage return.

The *Targets.txt* file is the most important piece of NEAT and users are expected to invest the time to ensure all paths and parameters exist and are correctly set. However, once set, most of these parameters will not change on a specific computer cluster (users from a same institute will use the same paths). We therefore suggest to more advanced users that they modify the *original* Targets.txt template file (see below).

All parameters of the Targets file should be self-explanatory. Here is a brief summary:

| | | |
|---|---|---|
| My_personal_email | : | If users would like to be notified by emailed when the cluster has finished. This will only work if your computer cluster has activated the emailing feature (please check with system administrator). To ensure servers are not overwhelmed by email |

services, ChIPpip is configured in such a way as to notify users only if the pipeline has terminated properly (with no error). Users may change this parameter by modifying the QSUB_header.sh template file found in *./ChIPpip/scripts/*.

My_personal_ssh : SSH parameters. Refer to your computer core facility

My_project_title : This is the name of the folder of your project on the remote server [automatically generated by ChIPpipCreateNewProject].

Reference_genome : The genome your data will be aligned to. Make sure your core faility has this genome reference installed on your cluster and that the extensions of the files are '.fa'.

Local_path_to_proj : [Automatically generated by RNApipCreateNewProject].

Local_path_to_NEAT : [Automatically generated by RNApipCreateNewProject].

Proj_TaxonDatabase : See NEAT for RNAseq.

Proj_TaxonDatabaseDic : See NEAT for RNAseq.

Remote_path_to_proj : Full path to your project folder (without the project name) [automatically generated].

Remote_path_to_NEAT : Full path to your NEAT folder. Note that in our example, we have created our project within the RNApip folder itself, but users can freely decide to create a dedicated folder for all of their RNApip projects.

Remote_path_to_orifastq.gz Full path to where your .fastq.gz files are. Usually, your sequencing core facility will let you know where they store these files. Note that all .fastq.gz files can be kept in a single location, they do not need to be copied to your folder.

Remote_path_to_chrLens.dat Path to a .dat file containing chromosome information for your reference genome. Refer to your computer core facility.

Remote_path_to_RefGen.fasta Path to folder containing your reference genome files. Refer to your computer core facility

Aligner_algo_short : "BWA" for standard alignment. If other algorithms are used, modify the *AdvancedSettings.txt* file accordingly.

Paired_end_seq_run : "0" for single end sequencing. "1" for paired end sequencing.

Remove_from_bigwig : Many softwares are incapable to load tracks because they countain unrecognized lines. For the mouse mm9 genome, these correspond to lines starting with 'random' and 'chrM'. You can easily find these for your preferred genome by attempting to load them to a genome browser, which will tell you which lines are 'unrecognized'.

Steps_to_execute_pipe : Users can choose from the following tasks: *unzip*, *qc*, *map* and *filter*. If you do not want to run all of these, simply delete them

for the Targets.txt or rename them. Once ran, RNApip will change the value of these from '*unzip*' to '*unzip_DONE*'. Obviously, a certain hierarchy has to be followed, e.g. attempting to filter reads without having previously mapped them (in the same run or in a previous run) will not work. Note that '*qc*' requires Thomas Girke's systemPipeR package; map requires bwa; the default *peacalling* requires the R package SPP; *filter* and *cleanbigwig* require samtools. Refer below for exact requirements.

Please modify the *Targets.txt* file according to your needs. To modify the Targets.txt file, we suggest users get accustomed to using plain text editors such as TextWrangler as it will avoid including spaces and special characters.

The paths to the reference genomes should be obtained from your computer core facility (system administrator), as they are the ones usually maintaining these up to date. Note here that the reference genome files should have an '*.fa*' extension (e.g. mm9.fa). Please make sure that your core has named these files accordingly as any other extension will lead the pipeline to abort prematurely.

Note here that *#Remote_path_to_RefGen.fasta* refers to the path to the folder in which '*.fa*' files are located, not to the file itself. This is different than in ChIPpip. *#Remove_from_bigwig* can usually be disregarded.

To avoid repeating these steps at each new NEAT (RNApip) project creation, we suggest you modify the *original* Targets.txt file that is used as template when creating a new ChIPpip project. This file can be found in *~/NEAT/RNApip/scripts/NewRNApipProject/DataStructure/Targets.txt*.

## 3.3 Running NEAT Part 1.2 (RNApip)

Once the *Targets.txt* file is correctly set up, users can run the *2_Run_NEAT.app*.


This script will execute the tasks specified in the *Targets.txt* file.

This will launch the pipeline and will prompt a summary of the user's parameters. ChIPpip automatically manages all creations and batch submissions of jobs, dependencies, ordering of files, queuing, etc. If the cluster is using TORQUE, the processes can be followed on the terminal using the ***qstat*** command (type ***qstat*** in your terminal). Briefly, *Q* stands for queuing, *R* for running, *E* for exiting and *H* for holding.

Once the pipeline has finished, it will notify users of its status by email (if applicable).

The mock data provided as a test example should take no more than one hour to run, usually a lot less.

# 4 NEAT Part 2 : RNAmE

## 4.1 Running NEAT Part 2.1 (RNAmE)

### 4.1.1 Step 1: Download a RNApip project

To transfer a RNApip project from a remote server to a local computer, double click the RNAmE 3_*Transfer.app*.


Users will be asked to locate the NEAT directory and where they want to save their RNApip project. In this example, the NEAT directory is on our desktop and we will save our RNApip project on the desktop as well.

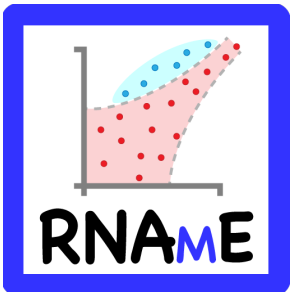
The *3_Transfer.app* will use all the information found in your Targets file to download the RNApip project from the remote server to your local computer. Please be attentive as you will need to enter your ssh passwords several time.

Downloading an entire project should not take more than a few minutes.

## 4.2 Running NEAT Part 2.2 (RNAmE)

### 4.2.1 Step 2: Run a RNAmE analysis

Once the project has been downloaded, users can run the proper RNAmE analysis. To this end, double click the NEAT *4_Analyze.app*.




Users will be asked to locate the NEAT folder (also saved on the desktop in this example). Users will also need to choose a mart object. Finally, users will be asked to locate the RNApip folder (the one just downloaded). In our case, the RNApip project was downloaded to the desktop. Running the analysis using the test data should take less than a minute.

## 4.3 Outputs

### 4.3.1 Logs

Each time ChIPmE is run, a log file is created and named using the date and time. This file is save in the *~/MY_NEW_RNA_PROJECT/logs/* directory. We strongly encourage users to initially look at these files, as any error that might have occurred will be saved there. Usually, if no error is prompted from the terminal, RNAmE has terminated correctly. Also, please note that if there are unrecognized chromosome

names such as random chromosomes, warning messages will appear. In the test data, there are 50 or more warnings. These can usually be disregarded.

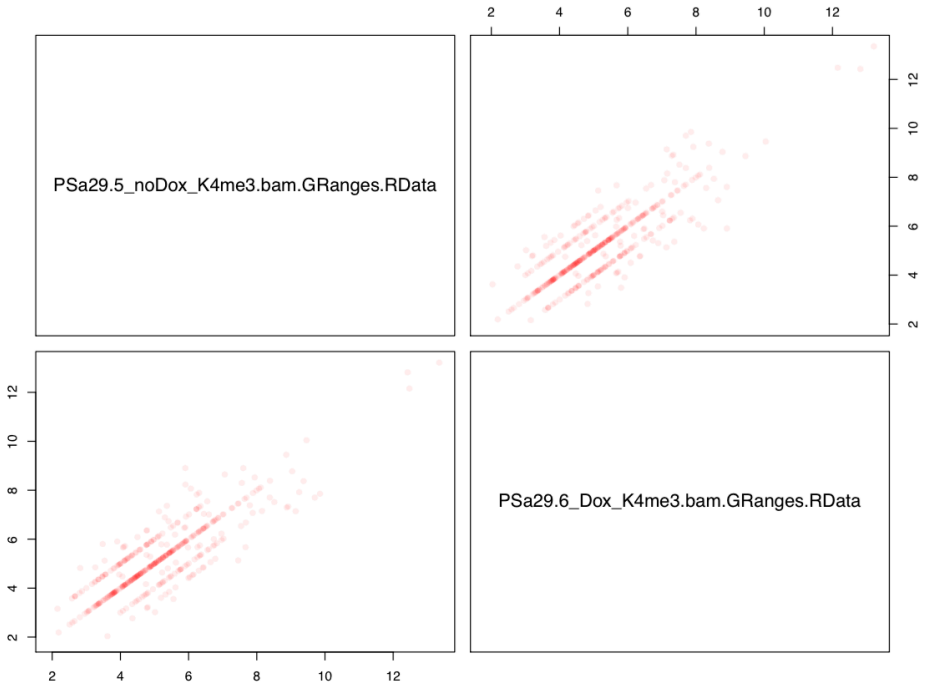
### 4.3.2 Count tables and RPKM tables

RNAmE will generate count tables that should be self-explanatory. In brief, rows correspond to genes and columns correspond to samples.

### 4.3.3 Differentially expressed gene plots

RNAmE generates pdf plots that are saved in */EXAMPLE/plots/*. General smear plots are created as a quick EDA between samples and replicates. In additional to smear plots, RNAmE generates all possible combinations of QQplots, with highlighted up and downregulated genes. More information on these genes can be found in the corresponding count tables. Finally, RNAmE generates gene lists meant to be fed to Gorilla, a well-established GO term EDA software. Venn diagrams are produced if selected. Users should note that computational time increases exponentially with the number of datasets as all combinations of Venn plots are created.

The smear plots below are an example of the results obtained using the test data. Obviously, the limited number of reads in the test data makes this plot relatively uninformative, but analyzing your own data will give much better resolution.




## 4.4 Advanced settings

### 4.4.1 Custom mart objects

Custom mart objects can easily be created with your favorites genes/regions. The files are simple .bed files that can either be manually or automatically created using various online tools or can be directly downloaded from genome browsers such as USCS or Ensembl. ChIPmE has been developed to ensure consistency between and within labs. In addition the relative small size of these bed files makes it easy to email/share them. We therefore suggest keeping an up-to-date, centralized folder containing all recurrent mart object files.

### 4.4.2 Bam files and GRanges

Once Granges objects have been created, bam files are no longer required locally. Users are thus free to delete these files as they are often two orders of magnitude larger than GRanges objects (respectively several Gb vs tens of Mb). We do suggest that users backup their .bam files on the remote server.

### 4.4.3 Consolidating projects

Consolidating projects is very easy. Users who intend to do so will simply need to change the *Targets.txt* file and copy-paste the Granges objects (or bam files) from one folder to the other. Other files and folder can be left as is.

# 5 Version information and required packages

Version: 0.5.9-r16

Program: samtools (Tools for alignments in the SAM format)
Version: 0.1.18 (r982:295)

R version 3.1.0 (2014-04-10)
Platform: x86_64-redhat-linux-gnu (64-bit)

R packages (with dependencies) required to run RNApip:

- systemPipeR (systemPipeR_0.99.0)
- …

R packages (with dependencies) required to run RNAmE:

- Rsamtools
- GenomicRanges
- GenomicAlignments
- caTools
- VennDiagram

sessionInfo()

R version 3.1.2 (2014-10-31)

Platform: x86_64-apple-darwin10.8.0 (64-bit)

locale:

[1] en_US.UTF-8/en_US.UTF-8/en_US.UTF-8/C/en_US.UTF-8/en_US.UTF-8

attached base packages:

[1] grid parallel stats4 stats graphics grDevices utils

[8] datasets methods base

other attached packages:

[1] VennDiagram_1.6.9 caTools_1.17.1 GenomicAlignments_1.2.1

[4] Rsamtools_1.18.2 Biostrings_2.34.1 XVector_0.6.0

[7] GenomicRanges_1.18.3 GenomeInfoDb_1.2.3 IRanges_2.0.1
[10] S4Vectors_0.4.0 BiocGenerics_0.12.1

loaded via a namespace (and not attached):
[1] base64enc_0.1-2 BatchJobs_1.5 BBmisc_1.8 BiocParallel_1.0.0
[5] bitops_1.0-6 brew_1.0-6 checkmate_1.5.0 codetools_0.2-9
[9] DBI_0.3.1 digest_0.6.4 fail_1.2 foreach_1.4.2
[13] iterators_1.0.7 RSQLite_1.0.0 sendmailR_1.2-1 stringr_0.6.2
[17] tools_3.1.2 zlibbioc_1.12.0

# 6 Reported bugs

Refer to the complete guide for reported bugs

# 7 Funding

This pipeline was developed with funding from the Swiss National Science Foundation.

# 8 References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