

# Differential expression (F2-73, Elite and PI127826 2020 plants)

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# 1 Data import

## 1.1 Import scaled counts

```
scaled_counts <- read.delim("../Supplemental_data_RNA-seq/scaled_counts.tsv",  
                             check.names = F,  
                             stringsAsFactors = F)
```

```
scaled_counts <- read.delim("../Supplemental_data_RNA-seq/scaled_counts.tsv",  
                             check.names = F,  
                             stringsAsFactors = F) %>%  
  mutate(gene = gsub(pattern = "mRNA:", replacement = "", x = gene)) %>%  
  dplyr::select("gene", "F2.73", "Elite_2020", "PI127826_2020")  
head(scaled_counts)
```

##	gene	F2.73	Elite_2020	PI127826_2020
## 1	Solyc00g005000.2.1	3.494954	0.0000	0.000000
## 2	Solyc00g005020.1.1	0.000000	0.0000	2.650106
## 3	Solyc00g005040.2.1	55.919260	49.2653	13.250530
## 4	Solyc00g005050.2.1	1382.254220	1383.7123	1810.022376
## 5	Solyc00g005060.1.1	0.000000	0.0000	0.000000
## 6	Solyc00g005070.1.1	0.000000	0.0000	0.000000

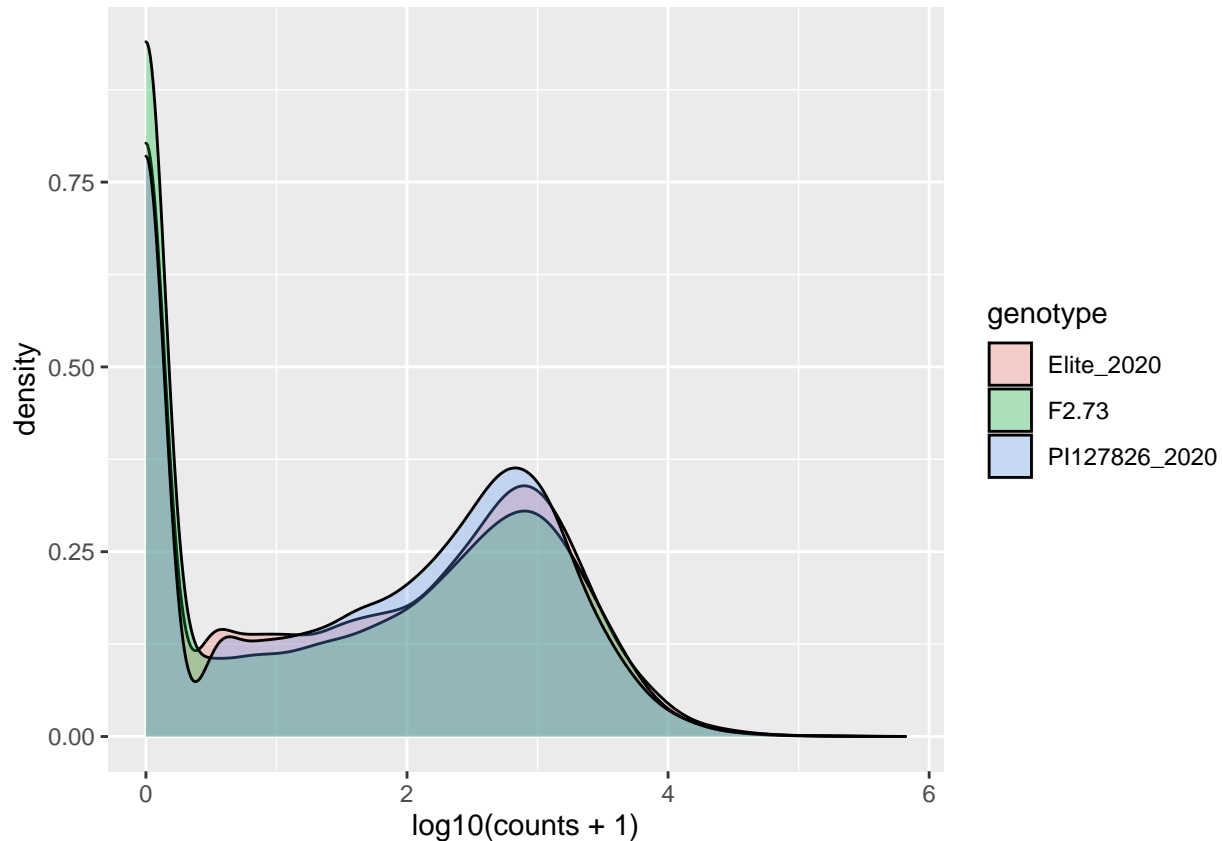
## 1.2 Mapping summary

```
# read.csv("../Supplemental_data_RNA-seq/mapping_summary.csv",  
#           stringsAsFactors = F,  
#           check.names = F) %>%  
#   knitr::kable()
```

## 2 QC plots

### 2.1 Plot density counts

```
scaled_counts %>%  
  pivot_longer(- gene, names_to = "genotype", values_to = "counts") %>%  
  ggplot(aes(x = log10(counts + 1), fill = genotype)) +  
  geom_density(alpha = 0.3)
```



The density of genes with low count values ( $\log_{10} = 0.3$ ) is lower for F2-73 but seems to be comparable for other genes.

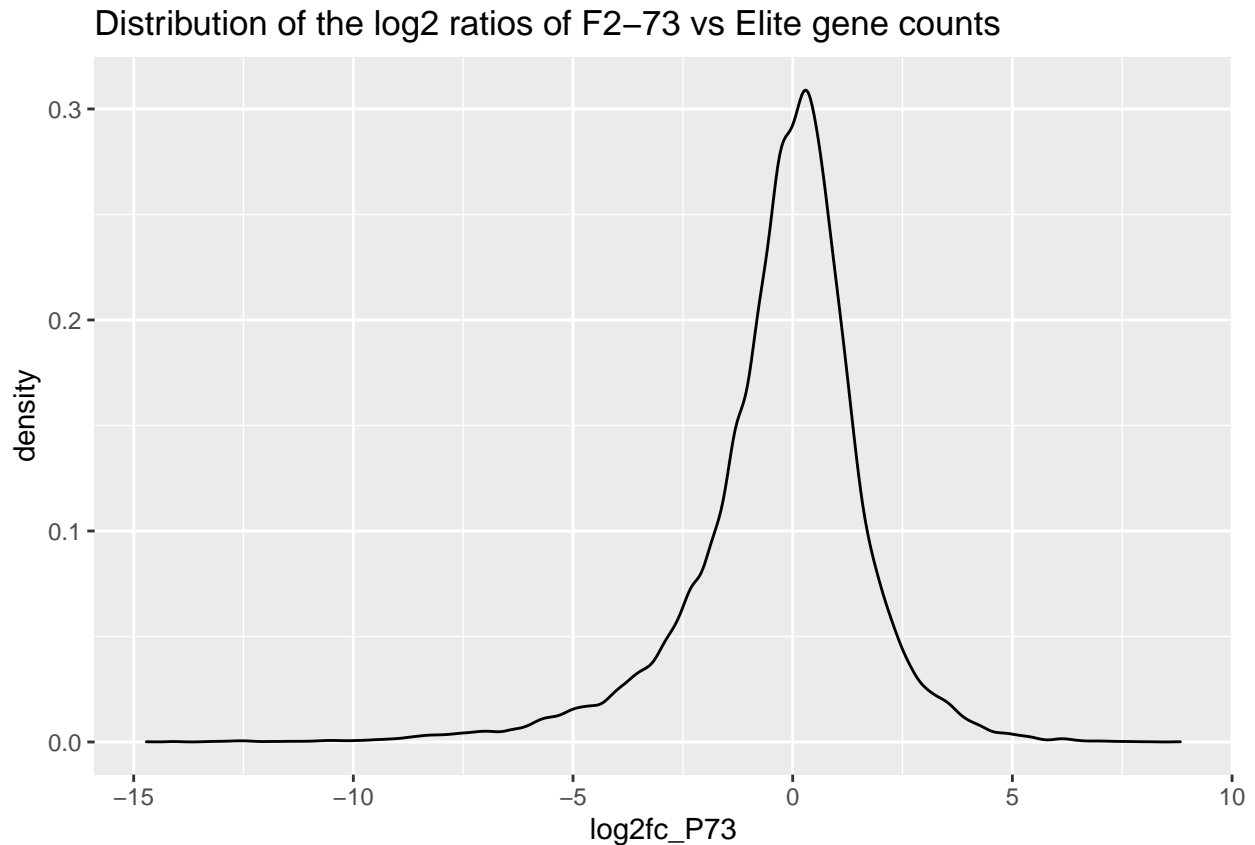
### 2.2 Plot log2ratio F2-73 vs Elite

First, let's extract genes with counts > 0

```
genes_sums <- scaled_counts %>% column_to_rownames("gene") %>% rowSums()  
genes_non_null <- genes_sums[genes_sums > 0]  
genes_non_null <- names(genes_non_null)
```

```
scaled_counts %>%  
  filter(gene %in% genes_non_null) %>%  
  mutate(log2fc_P73 = log2(`F2.73`/Elite_2020)) %>%  
  ggplot(aes(x = log2fc_P73)) +  
  geom_density() +  
  ggtitle("Distribution of the log2 ratios of F2-73 vs Elite gene counts")
```

```
## Warning: Removed 5110 rows containing non-finite values (stat_density).
```

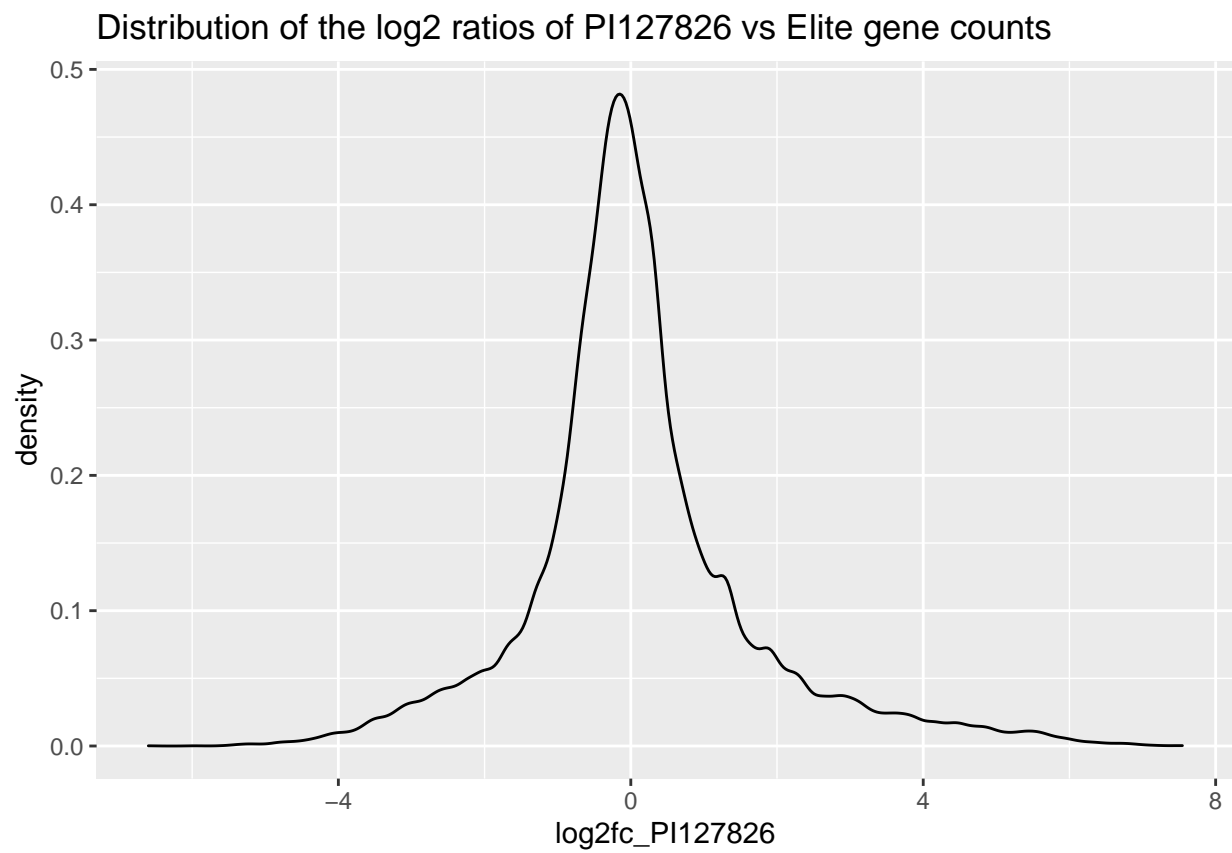


The log2ratio distribution looks OK. Compared to the log2 ratios of F2-28, it is more skewed towards negative log2ratios.

### 2.3 Plot log2ratio PI127826 vs Elite

```
scaled_counts %>%  
  filter(gene %in% genes_non_null) %>%  
  mutate(log2fc_PI127826 = log2(PI127826_2020/Elite_2020)) %>%  
  ggplot(aes(x = log2fc_PI127826)) +  
  geom_density() +  
  ggtitle("Distribution of the log2 ratios of PI127826 vs Elite gene counts")
```

```
## Warning: Removed 3633 rows containing non-finite values (stat_density).
```



### 3 Compute DE genes based on log2ratio Z-score

#### 3.1 Calculate log2ratios

A positive log2ratio for F2-73 and PI127826 means that the gene is more expressed in F2-28 and PI127826 (relative to the Elite line).

Let's calculate the log2ratio and remove the "Infinite" values.

```
log2ratio <-
  scaled_counts %>%
  filter(gene %in% genes_non_null) %>%
  mutate(log2ratio_P73 = log2(`F2.73`/Elite_2020)) %>%
  mutate(log2ratio_PI127826 = log2(PI127826_2020/Elite_2020)) %>%
  select(gene, log2ratio_P73, log2ratio_PI127826) %>%
  filter(!grepl(pattern = "Inf", x = log2ratio_P73)) %>%
  filter(!grepl(pattern = "Inf", x = log2ratio_PI127826))

head(log2ratio)
```

```
##           gene log2ratio_P73 log2ratio_PI127826
## 1 Solyc00g005040.2.1    0.182773531      -1.8945217
## 2 Solyc00g005050.2.1   -0.001520983        0.3874636
## 3 Solyc00g005840.2.1   -0.957797227        0.6485176
## 4 Solyc00g005860.1.1  -2.615592607      -3.0148159
## 5 Solyc00g006470.1.1 -10.568964562        1.7963727
## 6 Solyc00g006490.2.1  -0.683533784        0.1157466
```

A total of **21054** have a finite log2ratio in both F2-73 vs Elite and PI127826 vs Elite.

#### 3.2 Calculate Z-scores and associated p-values

Let's calculate the Z-score of the log2ratio + its associated p-value

```
log2ratio_zscores_pvals <-
  log2ratio %>%
  mutate(zscore_P73 = scale(log2ratio_P73, center = T, scale = T)) %>%
  mutate(zscore_PI127826 = scale(log2ratio_PI127826, center = T, scale = T)) %>%
  mutate(pval_P73 = pnorm(q = abs(zscore_P73), mean = 0, sd=1, log.p = FALSE, lower.tail=FALSE)) %>%
  mutate(pval_PI127826 = pnorm(q = abs(zscore_PI127826), mean = 0, sd=1, log.p = FALSE, lower.tail=FALSE)) %>%
  arrange(desc(log2ratio_P73)) %>%
  as_tibble()

head(log2ratio_zscores_pvals)
```

```
## # A tibble: 6 x 7
##   gene log2ratio_P73 log2ratio_PI127~ zscore_P73[,1] zscore_PI127826~
##   <chr>      <dbl>      <dbl>      <dbl>      <dbl>
## 1 Soly~         8.82         3.63         4.67         2.41
## 2 Soly~         8.10         6.21         4.29         4.17
## 3 Soly~         7.74         1.63         4.11         1.05
## 4 Soly~         7.57         5.70         4.03         3.82
## 5 Soly~         7.36         7.17         3.92         4.82
## 6 Soly~         7.17         3.63         3.82         2.41
## # ... with 2 more variables: pval_P73[,1] <dbl>, pval_PI127826[,1] <dbl>
```

### 3.3 Add original counts and annotations

Add back the scaled counts.

```
log2ratio_zscores_pvals_with_counts <- inner_join(scaled_counts, log2ratio_zscores_pvals, by = "gene")
```

Add descriptions

```
annots <- read.csv("info/ITAG2.4_loci_gene_descriptions.csv", stringsAsFactors = F)
```

```
final <-  
  log2ratio_zscores_pvals_with_counts %>%  
  mutate(locus = substr(gene, start = 1, stop = 14)) %>%  
  inner_join(x = ., y = annots, by = "locus") %>%  
  as_tibble()
```

```
dim(final)
```

```
## [1] 21054    12
```

### 3.4 Write to CSV file

```
dir.create(path = "./tables_F2-73/", showWarnings = F, recursive = T)  
write.csv(final,  
  file = "tables_F2-73/diff_res_F2-73_or_PI127826_vs_Elite.csv",  
  row.names = F,  
  quote = F)
```

```
final %>% filter(pval_P73 < 0.01) %>% dim()
```

```
## [1] 791    12
```



## 4 MEP and MVA pathway gene analysis

### 4.1 Import MEP and MVA gene identifiers

```
mep_mva_gene_ids <- read.csv("info/mep_mva_terpene_gene_ids.csv",  
                             stringsAsFactors = F)
```

### 4.2 Filter for significant DE genes

Should be significant ( $p < 0.05$ ) in **either** PI127826 vs Elite **AND** F2-28 vs Elite.

```
signif_genes <- filter(final, pval_P73 < 0.05 | pval_PI127826 < 0.05) %>% pull(gene)
```

### 4.3 Keep only MEP and MVA genes significant

```
mep_mva_genes <- inner_join(final, mep_mva_gene_ids)
```

```
## Joining, by = "locus"
```

```
mep_mva_gene_signif <-  
  mep_mva_genes %>%  
  filter(gene %in% signif_genes)
```

```
# show table
```

```
mep_mva_gene_signif %>%  
  select(name, gene, pathway, log2ratio_P73, log2ratio_PI127826, pval_P73, pval_PI127826) %>%  
  knitr::kable()
```

name	gene	pathway	log2ratio_P73	log2ratio_PI127826	pval_P73	pval_PI127826
HMGR	Solyc02g038740.2.1	MVA	5.535523	3.671742	0.001424666	0.007318178
HMGR	Solyc03g032010.2.1	MVA	3.914613	2.923354	0.015629531	0.026619455
HMGR	Solyc03g032020.2.1	MVA	3.315479	2.032462	0.032370075	0.092102361
pMVK	Solyc06g066310.2.1	MVA	2.927697	3.474898	0.049621651	0.010511663
AACT	Solyc07g045350.2.1	MVA	3.651830	2.152733	0.021731436	0.079328502

```
write.csv(mep_mva_gene_signif,  
          file = "tables_F2-73/mep_mva_gene_signif.csv",  
          row.names = F,  
          quote = F)
```

#### 4.4 Plot all MEP and MVA genes

```
for (i in seq_along(mep_mva_genes$gene)){
  tmp_df <- mep_mva_genes[i,]
  tmp_df$title4plot <- paste(tmp_df$name, tmp_df$gene, sep = "_")

  p <-
    tmp_df %>%
    # mutate(plot_title = paste(name, gene, sep = "_")) %>%
    select(title4plot, `F2.73`, Elite_2020, PI127826_2020) %>%
    pivot_longer(- title4plot, names_to = "genotype", values_to = "counts") %>%
    ggplot(. , aes(x = genotype, y = counts, fill = genotype)) +
    geom_bar(stat = "identity") +
    ggtitle(tmp_df$title4plot)

  print(p)
}
```

