

# Thalamic control of sensory selection in divided attention

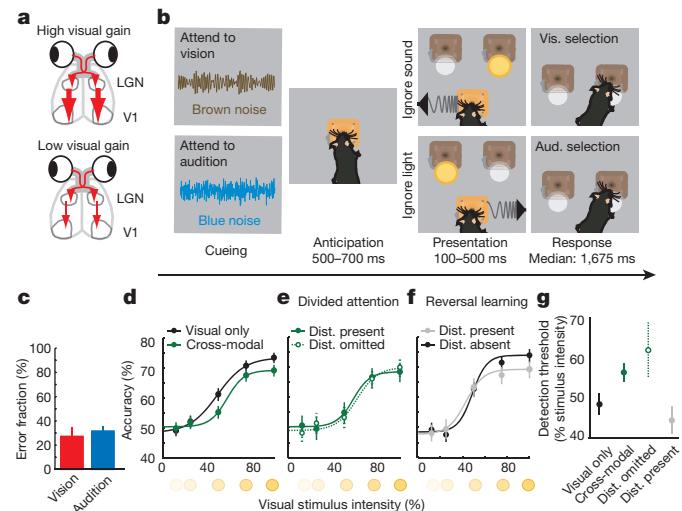
Ralf D. Wimmer<sup>1\*</sup>, L. Ian Schmitt<sup>1\*</sup>, Thomas J. Davidson<sup>2</sup>, Miho Nakajima<sup>1</sup>, Karl Deisseroth<sup>2,3,4</sup> & Michael M. Halassa<sup>1,5,6</sup>

**How the brain selects appropriate sensory inputs and suppresses distractors is unknown.** Given the well-established role of the prefrontal cortex (PFC) in executive function<sup>1</sup>, its interactions with sensory cortical areas during attention have been hypothesized to control sensory selection<sup>2–5</sup>. To test this idea and, more generally, dissect the circuits underlying sensory selection, we developed a cross-modal divided-attention task in mice that allowed genetic access to this cognitive process. By optogenetically perturbing PFC function in a temporally precise window, the ability of mice to select appropriately between conflicting visual and auditory stimuli was diminished. Equivalent sensory thalamocortical manipulations showed that behaviour was causally dependent on PFC interactions with the sensory thalamus, not sensory cortex. Consistent with this notion, we found neurons of the visual thalamic reticular nucleus (visTRN) to exhibit PFC-dependent changes in firing rate predictive of the modality selected. visTRN activity was causal to performance as confirmed by bidirectional optogenetic manipulations of this subnetwork. Using a combination of electrophysiology and intracellular chloride photometry, we demonstrated that visTRN dynamically controls visual thalamic gain through feedforward inhibition. Our experiments introduce a new subcortical model of sensory selection, in which the PFC biases thalamic reticular subnetworks to control thalamic sensory gain, selecting appropriate inputs for further processing.

To dissect the circuit basis of sensory selection, we sought a behaviour capable of dividing attention across modalities in the freely behaving mouse. Building on the rich history of visual neuroscience<sup>6–8</sup>, we focused our investigations on visual processing under conditions in which vision was behaviourally selected or suppressed (Fig. 1a). As such, we developed and validated a two-alternative forced-choice task in which mice selected between conflicting visual and auditory stimuli on a trial-by-trial basis. Stimuli indicated the location where a mouse had to nose poke to obtain a reward. Trial availability and target modality were signalled through binaurally emitted noise. For some mice, brown noise (10-kHz low-pass-filtered white noise) signalled a visual target (Fig. 1b, top) whereas blue noise (11-kHz high-pass-filtered white noise) signalled an auditory target (Fig. 1b, bottom). Cueing was counterbalanced across mice, with no effect on performance noted. By design, the task was asymmetric, with a visual detection component (flash from a light-emitting diode (LED) appearing to the right or left) and an auditory discrimination component (upsweep, 10–14 kHz; downsweep, 16–12 kHz). Multiple quality control metrics ensured that mice performed this task using the biasing cues (brown and blue noise) rather than low-level alternating strategies (Extended Data Fig. 1). Performance on the two modalities was balanced ( $n = 15$  mice, Fig. 1c).

Comparing visual detection under cross-modal and visual-only conditions suggested divided attention between vision and audition in the cross-modal task (Fig. 1d). Specifically, the visual detection threshold

was higher in cross-modal trials ( $n = 4$  mice,  $P < 0.05$ , bootstrap comparison; see Extended Data Fig. 2 for single-mouse examples and fixed lapse rate analysis). This difference persisted even when the conflicting auditory stimulus was randomly but systematically removed (Fig. 1e and Extended Data Fig. 3), suggesting that diminished visual perception under cross-modal conditions is a result of expectation (top-down)



**Figure 1 | Cross-modal divided attention in the mouse.** **a**, Hypothesized control of visual gain under cross-modal conditions (LGN, lateral geniculate nucleus; V1, primary visual cortex). **b**, Task design. A mouse is simultaneously informed about trial availability and the nature of the target stimulus through binaurally delivered noise. In this schematic, brown noise denotes ‘attend to vision’ and blue noise denotes ‘attend to audition’. Following a variable anticipation, during which the mouse is required to hold its snout in a centrally located poke, conflicting auditory and visual stimuli are presented. By design, the task is asymmetric, having a visual detection component (presence or absence of light at the reward location) and an auditory discrimination component (upsweep, turn left; downsweep, turn right). **c**, Mice exhibited comparable performance on visual and auditory trials (mean  $\pm$  s.e.m.,  $n = 15$  mice). **d**, Visual detection performance in cross-modal trials compared to visual-only trials ( $n = 4$  mice,  $\geq 421$  trials per condition). Note that both detection threshold and peak performance were lower in the cross-modal condition. **e**, Eliminating the auditory distractor in the cross-modal condition did not affect the visual detection psychometric function ( $n = 4$  mice,  $\geq 211$  trials per condition). **f**, When mice were not differentially cued but instead ignored the auditory stimulus by learning that it was not rewarded over a full session (reversal learning), visual detection threshold did not change ( $n = 6$  mice,  $\geq 242$  trials per condition). **g**, Visual detection threshold (bootstrap computed) of the pertinent psychometric functions in **d–f**. Error bars in **d–g** are 95% confidence intervals and therefore non-overlap denotes significance of  $P < 0.05$ . Aud., auditory; dist., distractor; vis., visual.

<sup>1</sup>New York University Neuroscience Institute, Department of Neuroscience and Physiology, New York University Langone Medical Center, New York, New York 10016, USA. <sup>2</sup>Department of Bioengineering, Stanford University, Stanford, California 94305, USA. <sup>3</sup>Cracking the Neural Code Program, Stanford University, Stanford, California 94305, USA. <sup>4</sup>Department of Psychiatry and Behavioral Sciences, Stanford University, Stanford, California 94305, USA. <sup>5</sup>Department of Psychiatry, New York University Langone Medical Center, New York, New York 10016, USA. <sup>6</sup>Center for Neural Science, New York University, New York, New York 10003, USA.

\*These authors contributed equally to this work.

rather than sensory interference (bottom-up). Conversely, when mice selected targets based on reward history, detection threshold was unaffected (Fig. 1f,  $n = 6$  mice). Selective suppression of the visual detection threshold in cross-modal trials was observed in both the raw (Fig. 1d, e) and the fitted (Fig. 1g) data. Together, these findings suggest that a cued, trial-by-trial task design is required for investigating sensory selection in divided attention.

Given the known role of PFC in top-down control of sensory processing and that our psychophysical measurements revealed top-down engagement in the cross-modal task, we asked whether cross-modal performance was PFC dependent. We targeted the prelimbic cortex because of its known homology to primate dorsolateral PFC<sup>9</sup>. We used the VGAT-ChR2 mouse to perturb PFC function in a temporally precise manner. In this mouse, the light-activated ion channel channelrhodopsin-2 (ChR2) is expressed under the vesicular GABA ( $\gamma$ -aminobutyric acid) transporter promoter (VGAT)<sup>10</sup>. Optogenetic drive in this mouse is known to exert intensity- and duration-dependent inhibition of excitatory neural activity<sup>11,12</sup>. Using this approach, we observed behavioural disruption only when the PFC activity was perturbed during stimulus anticipation (Fig. 2a,  $n = 4$

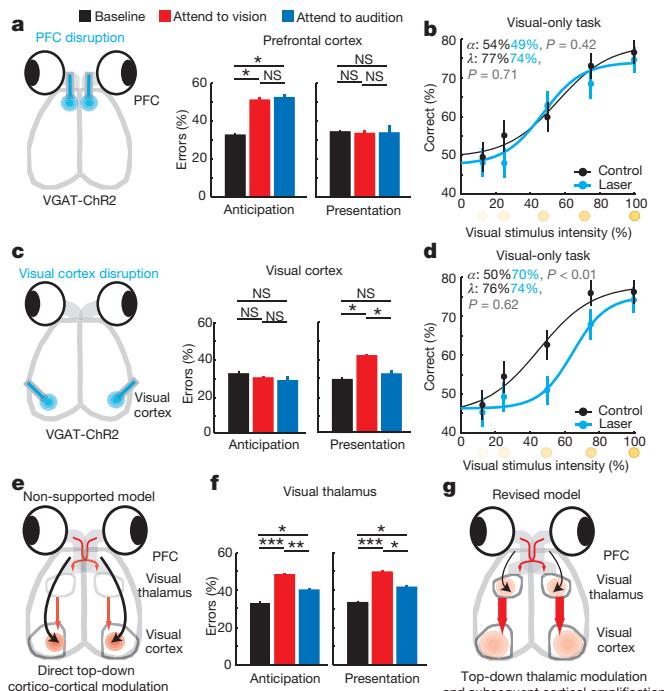
mice). This effect was dependent on the cross-modal nature of the task, as perturbing PFC activity in a visual-only task had no effect on performance regardless of task difficulty (Fig. 2b,  $n = 3$  mice,  $\geq 246$  trials per condition).

We reasoned that the PFC might be exerting its effect on performance by biasing sensory circuits towards target stimulus processing and distractor suppression. Several studies have suggested that this top-down bias is exerted at the level of the sensory cortex<sup>4,13</sup>. We did not find this to be the case in our task; perturbing visual cortical activity diminished visual performance only during stimulus presentation (Fig. 2c). This effect, unlike that of the PFC, was not cross-modal task specific, as it increased the detection threshold in a visual-only task (Fig. 2d,  $n = 3$  mice,  $\geq 239$  trials per condition,  $P < 0.01$ , bootstrap comparison). An analogous manipulation of auditory cortex resulted in a qualitatively similar effect on performance, but with a larger effect size (Extended Data Fig. 4a,  $n = 4$  mice,  $**P < 0.01$ , Wilcoxon rank-sum test), probably owing to the auditory cortical requirement for stimulus discrimination<sup>14</sup>. Together, these findings support the role of sensory cortical areas in stimulus amplification and discrimination, but are inconsistent with their being a locus for top-down bias of sensory processing. Moreover, optogenetic perturbations of frontal regions that project directly to visual and auditory cortices such as the anterior cingulate cortex (ACC) and the lateral orbitofrontal cortex (OFC) did not affect cross-modal performance (Extended Data Fig. 4b, c,  $n = 8$  sessions, 2 mice). In contrast, localized viral injection of AAV-hSyn-DIO-ChR2 into the prelimbic cortex of mice in which Cre recombinase is expressed in inhibitory neurons (VGAT-Cre), followed by optical manipulation, disrupted performance (Extended Data Fig. 4d–h).

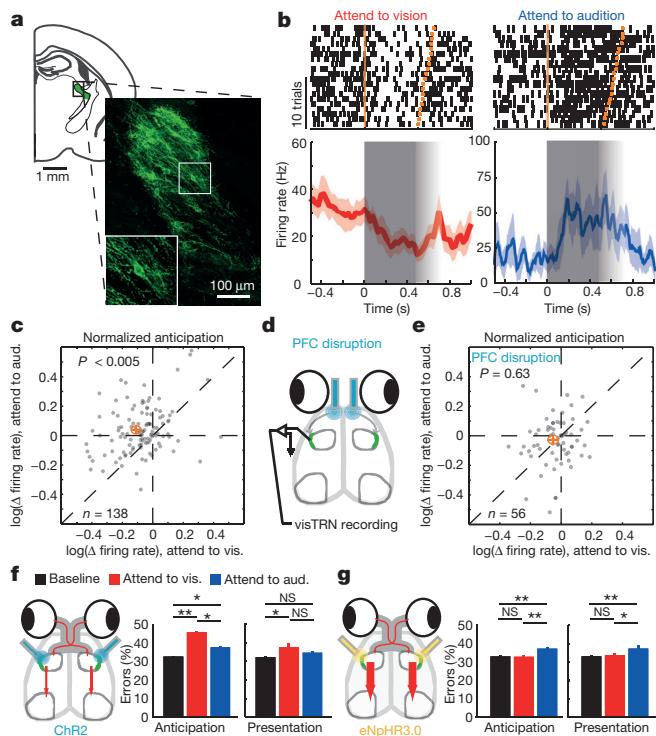
Having shown that direct prefrontal–sensory cortical interaction is unlikely to account for top-down control of visual processing (Fig. 2e), we sought to find the locus of attentional modulation observed in the cross-modal task (Fig. 1c). The sensory thalamus has been implicated in attentional modulation in primates<sup>15–17</sup>, raising the possibility that it could be a locus of top-down sensory bias. Using a similar VGAT-ChR2 strategy, we found that optogenetic perturbation of the visual thalamus (lateral geniculate nucleus; LGN) during either stimulus anticipation or presentation impaired cross-modal performance (Fig. 2f,  $n = 12$  sessions, 3 mice). This suggested that top-down bias of visual detection may be based in the thalamus (Fig. 2g). Visual thalamic manipulation resulted in worsening performance on both auditory and visual trials, implicating intrathalamic interactions in this behavioural effect. Intrathalamic interactions are mediated by a shell of GABAergic neurons surrounding thalamic nuclei known as the thalamic reticular nucleus (TRN)<sup>18</sup>. TRN neurons have been hypothesized to control the gain of thalamic output in a behaviourally relevant manner<sup>19,20</sup>. As such, we asked whether the TRN could be a locus of top-down modulation of the sensory thalamus.

To investigate the role of the TRN in visual gain control in divided attention, we used an intersectional genetic/connectivity strategy to label inhibitory (VGAT-positive) neurons that project to the LGN with retrograde lentiviruses<sup>20</sup> (Fig. 3a and Extended Data Fig. 5a, b) and optogenetically tagged them during extracellular recordings in freely behaving mice (Extended Data Fig. 5c–f).

During cross-modal performance, we observed bidirectional modulation of visTRN neurons consistent with their hypothesized role in behaviourally relevant sensory gain control. Specifically, during ‘attend to vision’ trials, firing rates of these neurons were reduced. In contrast, their rates were elevated during ‘attend to audition’ trials (Fig. 3b (example), Fig. 3c (population;  $n = 138$  cells) and Extended Data Fig. 6). Attentional modulation by the visTRN was eliminated by optogenetic PFC disruption (Fig. 3d, e,  $n = 56$  cells). Although PFC disruption diminished performance, the effects on visTRN firing rates were not simply a covariant of inaccurate performance (Extended Data Fig. 7). In contrast to naturally occurring error trials, during which



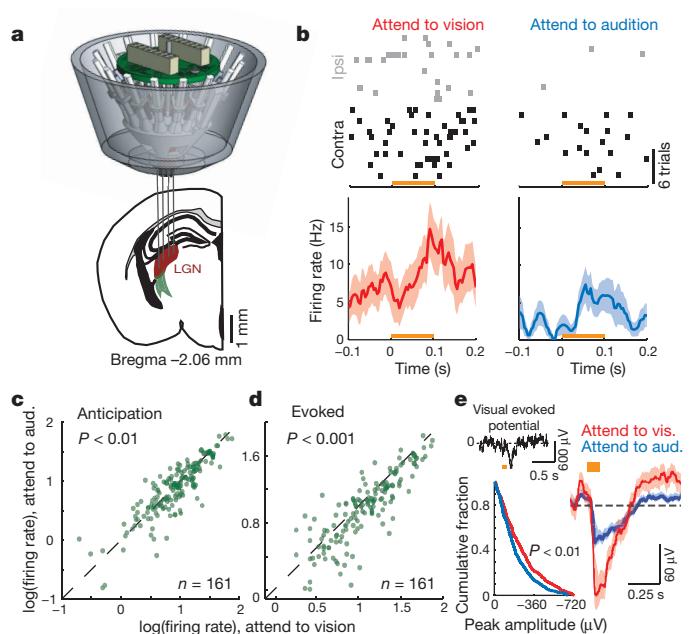
**Figure 2 | Evidence for top-down thalamic modulation in divided attention.** **a**, Disrupting PFC activity by delivering blue laser pulses (50 Hz, 18 ms, 90% duty cycle) impaired task performance at 100% stimulus intensity equally on both modalities only when manipulation was performed during stimulus anticipation ( $n = 4$  mice,  $*P < 0.05$ , Wilcoxon rank-sum test). **b**, The effect was related to the cross-modal nature of the task, not its difficulty, as PFC inhibition did not affect performance on a visual-only task. **c**, Disruption of primary visual cortex during stimulus presentation impaired performance on visual trials ( $n = 4$  mice). **d**, The effect in **c** was related to task difficulty, as the visual detection threshold increased in a visual-only task. **e**, The data in **a** and **c** do not support a causal role for PFC interactions with primary visual cortex in performance. **f**, Perturbing visual thalamic function in a manner similar to cortical perturbations in VGAT-ChR2 mice preferentially diminished performance on visual trials during both anticipation and presentation of target stimuli ( $n = 12$  sessions from 3 mice,  $*P < 0.05$ ,  $**P < 0.01$ ,  $***P < 0.001$ , Wilcoxon rank-sum test). **g**, The finding in **f** supports a model in which PFC activity influences thalamic sensory processing. Bar graphs represent mean  $\pm$  s.e.m. Error bars for psychometric curves are 95% confidence intervals. NS, not significant;  $\alpha$ , detection threshold;  $\lambda$ , lapse rate.



**Figure 3 | PFC-dependent visTRN modulation suggests PFC-TRN functional coupling is required for visual gain control.** **a**, Intersectional tagging of visTRN neurons based on connectivity and genetic identity. Inset, maximum projection of ten 1- $\mu\text{m}$  confocal images. Cells were labelled with ChR2-eYFP and stained with anti-GFP. eYFP, enhanced yellow fluorescent protein; GFP, green fluorescent protein. **b**, Raster of two visTRN neurons triggered on task initiation. Note the reduction of firing rate between the first trigger (trial initiation) and the second one (stimulus presentation) during the ‘attend to vision’ condition, but the opposite during the ‘attend to audition’ condition. Red dotted lines indicate triggers. Fading grey boxes denote the jitter of the anticipatory period. **c**, Group analysis of **b**, showing a scatter plot of responses from visTRN neurons ( $n = 138$  cells from 4 animals,  $P < 0.005$ , Wilcoxon rank-sum test performed over all cells). The orange crosshair indicates mean  $\pm$  95% confidence interval. **d**, PFC activity was disrupted during stimulus anticipation to examine the effect on visTRN activity. **e**, PFC disruption diminished visTRN modulation of attention. **f, g**, Behavioural performance is causally dependent on visTRN modulation of attention. **f**, Optogenetic activation of retrograde-tagged visTRN neurons resulted in performance diminishing on visual trials (mean  $\pm$  s.e.m.,  $n = 12$  sessions from 3 mice,  $*P < 0.05$ ,  $**P < 0.01$ ,  $***P < 0.001$ , Wilcoxon rank-sum test), consistent with this manipulation lowering visual gain. **g**, In contrast, optogenetic inhibition of visTRN neurons preferentially diminished performance on auditory trials, consistent with inappropriate visual gain increase ( $n = 12$  sessions from 3 mice).

diminished visTRN firing rate modulation was observed in both ‘attend to vision’ and ‘attend to audition’ trials, PFC disruption had a greater effect on ‘attend to audition’ trials. This result may highlight a role of the PFC in distractor suppression.

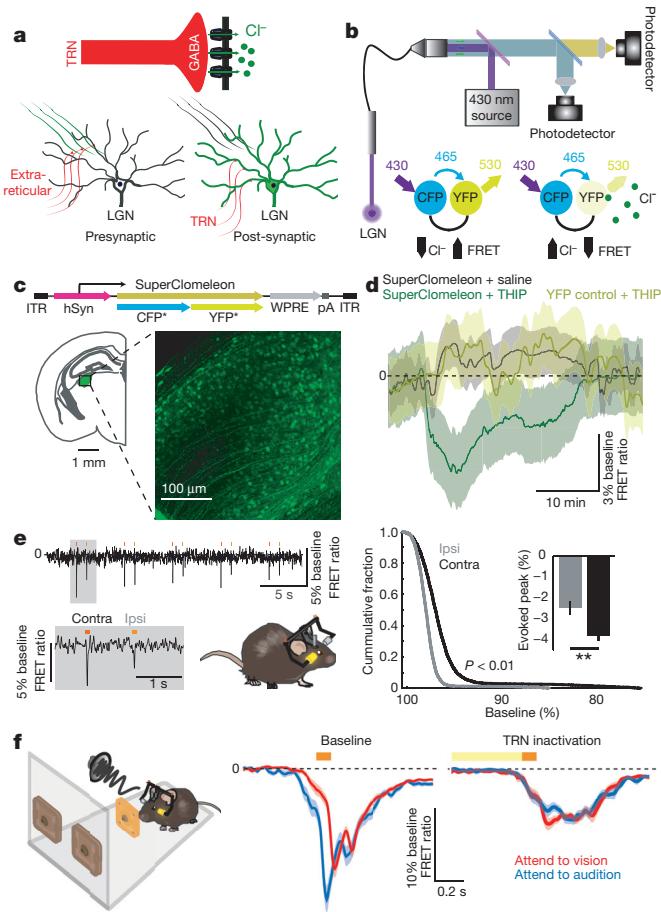
To examine whether the physiological effects observed in the visTRN were causal to behaviour, we manipulated this subnetwork bidirectionally. Although activating the visTRN resulted in similar effects to driving LGN inhibition in VGAT-ChR2 mice (Fig. 3f,  $n = 12$  sessions, 3 mice), the effect size was smaller, probably reflecting the weaker nature of the genetic manipulation (Extended Data Fig. 8). This result supports the notion that elevated visTRN firing reduces visual thalamic gain. In contrast, inhibiting visTRN function using the light-activated  $\text{Cl}^-$  pump eNpHR3.0 diminished performance on ‘attend to audition’ trials, suggesting it inappropriately enhanced visual thalamic gain when it needed to be suppressed (Fig. 3g,  $n = 12$  sessions, 3 mice).



**Figure 4 | Direct evidence for visual thalamic gain modulation in divided attention.** **a**, Cartoon depiction of multi-electrode targeting of the LGN at 2.06 mm posterior to Bregma in freely behaving mice. **b**, An example of differential modulation of a single LGN cell spiking under the two anticipatory task conditions. Note that contralateral eye stimulation (with respect to recording electrodes) resulted in more robust visual drive. Moreover, the cell discharged more spikes during anticipation and presentation when attention was directed towards vision. Contra, contralateral; Ipsi, ipsilateral. **c, d**, Group analysis of the phenomenon in **b** ( $n = 161$  cells from 4 mice, Wilcoxon rank-sum test). **e**, Similarly, enhanced visual responses were observed at the level of visual evoked potentials. Top left, example visual evoked potential (VEP); bottom left, cumulative distribution of VEP amplitudes, showing higher values for ‘attend to vision’ trials ( $P < 0.01$ , Kolmogorov–Smirnov test). Right, average VEP from 4 mice (684 visual trials and 633 auditory trials from 29 sessions; dashed line indicates baseline potential). Shaded errors are 95% confidence intervals. Orange blocks denote visual stimulus presentation.

To determine whether modulation of visTRN firing rate affected visual processing, we investigated LGN spiking in response to well-controlled visual stimuli. We implanted mice with multi-electrode arrays targeted to the LGN (Fig. 4a). To minimize trial-to-trial variability related to slight changes in head and eye position, we changed the position of visual stimuli from wall-mounted to head-mounted LEDs. LGN neurons showed enhanced baseline and evoked activity when attention was directed to vision (Fig. 4b (example), Fig. 4c, d, (population,  $n = 161$  cells, 4 mice)), consistent with baseline and evoked neuroimaging results observed in human LGN<sup>21</sup>. Differences in evoked responses were also observed in the visual evoked potential (Fig. 4e,  $n = 684$  visual trials and  $n = 633$  auditory trials, 4 mice). These physiological effects were not observed during error trials (Extended Data Fig. 9).

Suppression of LGN spiking in ‘attend to audition’ trials could be a result of direct feedforward inhibition or reduction in feedforward excitation. In contrast to many extrareticular inhibitory inputs<sup>22</sup>, the visTRN is known to exert direct feedforward inhibition on LGN neurons<sup>23</sup> (Fig. 5a). As such, we sought to measure LGN inhibition directly. To do so, we leveraged a recently developed technique known as fibre photometry, which has been used to measure bulk changes of intracellular  $\text{Ca}^{2+}$  concentration ( $[\text{Ca}^{2+}]_i$ ) (ref. 24). We modified two aspects of conventional photometry to allow interrogation of intracellular  $\text{Cl}^-$  concentration ( $[\text{Cl}^-]_i$ ), a proxy for synaptic inhibition and an otherwise inaccessible measure. First was the genetic labelling of neurons with the  $\text{Cl}^-$  indicator SuperClomeleon<sup>25</sup>. This fluorescence resonance energy transfer (FRET) indicator contains cyan fluorescent



**Figure 5 | Measuring bulk intracellular  $[\text{Cl}^-]$  in vivo shows dynamic changes in LGN inhibition during behaviour.** **a**, Possible mechanisms for modulation of LGN firing rate: extrareticular inputs can change activity by presynaptic inhibition of feedforward excitation, whereas visTRN inhibits LGN directly. **b**, FRET photometry setup and schematic of CFP-to-YFP FRET. **c**, Cloning of SuperClomeleon, a FRET-based  $\text{Cl}^-$  indicator, into an adeno-associated virus (AAV) followed by *in vivo* expression in the LGN. hSyn, human synapsin promoter; ITR, inverted terminal repeat sequences; pA, polyadenylation signal; WPRE, woodchuck hepatitis virus post-transcriptional regulatory element. **d**, Pharmacological confirmation of the technical feasibility of SuperClomeleon FRET for  $\text{GABA}_A$ -mediated increase in intracellular  $[\text{Cl}^-]$  by injection of the  $\text{GABA}_A$  agonist THIP. Note that the YFP control mice did not show similar signals ( $n = 3$  mice per condition; shaded errors are 95% confidence intervals). **e**, Mice showed stronger visual-evoked SuperClomeleon FRET responses for the contralateral eye, as would be predicted ( $n = 3$  mice,  $P < 0.05$ , Wilcoxon rank-sum test). Yellow bars mark the display of the light stimuli. **f**, Left, cartoon depiction of photometry in the cross-modal task, during which the visual stimulus was signalled through a head-mounted LED as in Fig. 4 (see Supplementary Video 1 for illustration); middle, differential visual-evoked  $[\text{Cl}^-]$  LGN responses in relation to the modality anticipated (363 correct visual trials and 274 correct auditory trials from 6 mice). Shaded errors are 95% confidence intervals. Note that 'attend to audition' trials showed an earlier increase in  $[\text{Cl}^-]$  (decreased SuperClomeleon FRET) and the separation between the two traces started before stimulus onset, consistent with differential anticipatory changes of visTRN activity; right, optogenetic TRN inactivation eliminates this differential response (101 correct visual and 82 correct auditory trials from 3 mice). Orange bars indicate visual stimulus presentation and the yellow bar denotes optogenetic TRN inactivation.

protein (CFP) as FRET donor and yellow fluorescent protein (YFP) as FRET acceptor. Under conditions of elevated  $[\text{Cl}^-]$ , YFP is quenched and FRET is reduced (Fig. 5b). Second was the light path; we excited SuperClomeleon with 430-nm light and collected both CFP and YFP emission data to perform subsequent ratiometric measurements off-line (Fig. 5b). To use this technology *in vivo*, we generated a viral

construct harbouring SuperClomeleon (AAV-hSyn-SuperClomeleon) and injected it into the LGN (Fig. 5c).

We validated SuperClomeleon FRET as a measure of inhibition by two methods. First, we reasoned that pharmacological activation of  $\text{GABA}_A$  receptors would increase  $[\text{Cl}^-]$  and reduce YFP emission. Indeed, intraperitoneal injection of the  $\text{GABA}_A$  agonist 4,5,6,7-tetrahydroisoxazolo[5,4-c]pyridin-3-ol (THIP, 8 mg kg<sup>-1</sup>) reduced SuperClomeleon FRET in a pharmacokinetically predicted manner<sup>26</sup> (Fig. 5d,  $n = 3$  mice). Second, we expected visual drive to result in inhibition of LGN neurons that balanced excitation. This was indeed the case; these signals were larger when visual stimuli were delivered to the side contralateral to the recorded LGN than when delivered to the ipsilateral side (Fig. 5e,  $n = 3$  mice). These rapid events were not observed in YFP control mice (Extended Data Fig. 10a). SuperClomeleon visual transients were sensitive to the  $\text{GABA}_A$  receptor antagonist flumazenil in a dose-dependent manner (Extended Data Fig. 10b), confirming that these signals reflect GABAergic inhibition.

Having validated chloride photometry, we asked whether changes in visual gain associated with sensory selection were explained by opposing changes in LGN feedforward inhibition. We found this to be the case; visual-evoked chloride photometry showed significantly larger responses in 'attend to audition' trials than in 'attend to vision' trials (Fig. 5f,  $n = 363$  visual trials and  $n = 274$  auditory trials, 6 mice; example trials shown in Supplementary Video 1). Signal kinetics in these two conditions were distinct: an earlier reduction in FRET ratio was seen in auditory selection trials compared to visual selection trials, consistent with the differential baseline spiking observed in visTRN neurons. Optogenetic visTRN inactivation eliminated this differential inhibitory response (Fig. 5f and Extended Data Fig. 10c,  $n = 3$  mice,  $>82$  trials). Overall, our data support the model that thalamic gain control can be explained by feedforward inhibition and that the TRN is the source of this inhibition. More generally, to our knowledge, this experiment constitutes the first measurement of inhibitory dynamics in freely behaving animals.

Seminal studies have shown the thalamus to be more than a cortical relay<sup>27,28</sup>. By providing a mechanistic circuit dissection of thalamic involvement in divided attention, we extend these studies in two directions. First, our findings in mice show the generality of thalamic modulation of attention across mammalian brains. Second, we provide a first description, with causal circuit dependence, of how prefrontal top-down control changes thalamic inhibitory dynamics to modulate sensory gain. The specific involvement of prelimbic cortex in this behaviour, which we further demonstrate through combined optogenetics and chloride photometry (Extended Data Fig. 10d), does not eliminate the possibility that the OFC and the ACC may be engaged in other types of top-down control, potentially via cortico-cortical interactions<sup>5</sup>. In addition to regulating sensory gain, prelimbic control of thalamic inhibition may regulate the degree by which relay nuclei participate in large-scale functional interactions<sup>17</sup>.

The ability to directly measure  $[\text{Cl}^-]$  dynamics provided access to a critical biological variable:  $\text{GABA}_A$ -mediated synaptic inhibition. Although photometry has already been introduced into neuroscience for measurement of  $[\text{Ca}^{2+}]_i$  in cell bodies and terminals<sup>25</sup>, FRET-based chloride photometry has not been performed previously. In this study, developing chloride photometry was essential for establishing a direct physiological link between visTRN and LGN spiking (technical discussion in Supplementary Information).

Thirty years ago, Francis Crick proposed that the TRN functions as a 'searchlight', directing the internal spotlight of attention to thalamocortical circuits that process ongoing behavioural demands<sup>19</sup>. Owing to technical limitations, this transformative model has been difficult to test, particularly under conditions where the attentional spotlight shifts. Our study combined novel and established technology to provide mechanistic details for Crick's 'searchlight hypothesis', thereby contributing to understanding the circuit mechanisms of sensory selection.

**Online Content** Methods, along with any additional Extended Data display items and Source Data, are available in the online version of the paper; references unique to these sections appear only in the online paper.

Received 12 April; accepted 18 August 2015.

Published online 21 October 2015.

1. Miller, E. K. & Cohen, J. D. An integrative theory of prefrontal cortex function. *Annu. Rev. Neurosci.* **24**, 167–202 (2001).
2. Buschman, T. J. & Miller, E. K. Top-down versus bottom-up control of attention in the prefrontal and posterior parietal cortices. *Science* **315**, 1860–1862 (2007).
3. Fritz, J., Shamma, S., Elhilali, M. & Klein, D. Rapid task-related plasticity of spectrotemporal receptive fields in primary auditory cortex. *Nature Neurosci.* **6**, 1216–1223 (2003).
4. Rodgers, C. C. & DeWeese, M. R. Neural correlates of task switching in prefrontal cortex and primary auditory cortex in a novel stimulus selection task for rodents. *Neuron* **82**, 1157–1170 (2014).
5. Zhang, S. *et al.* Selective attention. Long-range and local circuits for top-down modulation of visual cortex processing. *Science* **345**, 660–665 (2014).
6. Glickfeld, L. L., Histed, M. H. & Maunsell, J. H. Mouse primary visual cortex is used to detect both orientation and contrast changes. *J. Neurosci.* **33**, 19416–19422 (2013).
7. Hubel, D. H. & Wiesel, T. N. Receptive fields, binocular interaction and functional architecture in the cat's visual cortex. *J. Physiol. (Lond.)* **160**, 106–154 (1962).
8. Newsome, W. T., Britten, K. H. & Movshon, J. A. Neuronal correlates of a perceptual decision. *Nature* **341**, 52–54 (1989).
9. Hoover, W. B. & Vertes, R. P. Anatomical analysis of afferent projections to the medial prefrontal cortex in the rat. *Brain Struct. Funct.* **212**, 149–179 (2007).
10. Vong, L. *et al.* Leptin action on GABAergic neurons prevents obesity and reduces inhibitory tone to POMC neurons. *Neuron* **71**, 142–154 (2011).
11. Halassa, M. M. *et al.* Selective optical drive of thalamic reticular nucleus generates thalamic bursts and cortical spindles. *Nature Neurosci.* **14**, 1118–1120 (2011).
12. Zhao, S. *et al.* Cell type-specific channelrhodopsin-2 transgenic mice for optogenetic dissection of neural circuitry function. *Nature Methods* **8**, 745–752 (2011).
13. Fritz, J. B., David, S. V., Radtke-Schuller, S., Yin, P. & Shamma, S. A. Adaptive, behaviorally gated, persistent encoding of task-relevant auditory information in ferret frontal cortex. *Nature Neurosci.* **13**, 1011–1019 (2010).
14. Letzkus, J. J. *et al.* A disinhibitory microcircuit for associative fear learning in the auditory cortex. *Nature* **480**, 331–335 (2011).
15. McAlonan, K., Cavanaugh, J. & Wurtz, R. H. Guarding the gateway to cortex with attention in visual thalamus. *Nature* **456**, 391–394 (2008).
16. Purushothaman, G., Marion, R., Li, K. & Casagrande, V. A. Gating and control of primary visual cortex by pulvinar. *Nature Neurosci.* **15**, 905–912 (2012).
17. Saalmann, Y. B., Pinski, M. A., Wang, L., Li, X. & Kastner, S. The pulvinar regulates information transmission between cortical areas based on attention demands. *Science* **337**, 753–756 (2012).
18. Pinault, D. The thalamic reticular nucleus: structure, function and concept. *Brain Res. Brain Res. Rev.* **46**, 1–31 (2004).
19. Crick, F. Function of the thalamic reticular complex: the searchlight hypothesis. *Proc. Natl Acad. Sci. USA* **81**, 4586–4590 (1984).
20. Halassa, M. M. *et al.* State-dependent architecture of thalamic reticular subnetworks. *Cell* **158**, 808–821 (2014).
21. O'Connor, D. H., Fukui, M. M., Pinski, M. A. & Kastner, S. Attention modulates responses in the human lateral geniculate nucleus. *Nature Neurosci.* **5**, 1203–1209 (2002).
22. Chen, C. & Regehr, W. G. Presynaptic modulation of the retinogeniculate synapse. *J. Neurosci.* **23**, 3130–3135 (2003).
23. Cox, C. L., Huguenard, J. R. & Prince, D. A. Nucleus reticularis neurons mediate diverse inhibitory effects in thalamus. *Proc. Natl Acad. Sci. USA* **94**, 8854–8859 (1997).
24. Gunaydin, L. A. *et al.* Natural neural projection dynamics underlying social behavior. *Cell* **157**, 1535–1551 (2014).
25. Grimley, J. S. *et al.* Visualization of synaptic inhibition with an optogenetic sensor developed by cell-free protein engineering automation. *J. Neurosci.* **33**, 16297–16309 (2013).
26. Cremer, T. & Ebert, B. Plasma and CNS concentrations of Gaboxadol in rats following subcutaneous administration. *Eur. J. Pharmacol.* **562**, 47–52 (2007).
27. Casagrande, V. A., Sáry, G., Royal, D. & Ruiz, O. On the impact of attention and motor planning on the lateral geniculate nucleus. *Prog. Brain Res.* **149**, 11–29 (2005).
28. Mitchell, A. S. *et al.* Advances in understanding mechanisms of thalamic relays in cognition and behavior. *J. Neurosci.* **34**, 15340–15346 (2014).

**Supplementary Information** is available in the online version of the paper.

**Acknowledgements** We thank J. A. Movshon, W. Ma, R. W. Tsien, G. Fishell and D. Rinberg for helpful comments on the manuscript and G. J. Augustine for providing us with the *SuperClomeleon* construct and for helpful discussion around its use. The work was supported by the Swiss National Science Foundation (P2ZAP3 151786) to R.D.W. and the Simons Foundation, the Sloan Foundation, the Brain and Behavior Research Foundation and the US National Institutes of Health (R00NS078115) to M.M.H.; M.M.H. is additionally supported by the Feldstein Medical Foundation, a Klingenstein-Simons Fellowship and a Biobehavioral Research Award for Innovative New Scientists (BRAINS) R01 (R01 MH107680) from the National Institute of Mental Health.

**Author Contributions** M.M.H. conceived and designed all aspects of the study. R.D.W. devised the training paradigm for the cross-modal task and L.I.S. performed all associated programming. R.D.W. collected electrophysiological data. T.J.D. provided fibre photometry training, advice and rig designs; L.I.S. extended the method to FRET-based photometry, built the rig and collected data. R.D.W. analysed behavioural data and L.I.S. analysed psychophysical, electrophysiological and photometry data. M.N. generated the retrograde lentiviruses in-house, performed *SuperClomeleon* cloning into an AAV backbone and acquired confocal images. K.D. provided support for fibre photometry training. M.M.H. supervised the experiment, directed the analysis and wrote the manuscript. All authors read the final version of the manuscript.

**Author Information** Reprints and permissions information is available at [www.nature.com/reprints](http://www.nature.com/reprints). The authors declare no competing financial interests. Readers are welcome to comment on the online version of the paper. Correspondence and requests for materials should be addressed to M.M.H. (michael.halassa@nyumc.org).

**METHODS**

**Animals.** VGAT-ChR2 mice were purchased from the Jackson Laboratory and maintained on a C57Bl6/J background. VGAT-Cre mice were backcrossed to C57Bl6/J mice for at least six generations. For experiments in Fig. 1, a total of fifteen animals were trained, ten of which were later used to establish psychometric functions (four for divided attention; six for reversal learning). For Fig. 2, four VGAT-ChR2 mice were used for disruption of PFC and primary sensory cortices and three mice were used for inactivating LGN. In Fig. 3, four VGAT-Cre mice were used for electrophysiological recordings from optogenetically identified visTRN neurons, of which two were used for combined electrophysiological recordings with optogenetic PFC inactivation (Fig. 3c–e). An additional six mice were used for optogenetic activation or inhibition of visTRN (three per manipulation) during behaviour (Fig. 3f–g). Four wild-type mice were used for LGN recordings (Fig. 4). For fibre photometry experiments (Fig. 5), six mice were injected with AAV-hSyn-SuperClomeleon for behavioural and pharmacological experiments and three YFP control mice were used. Including all animals used for Extended Data figures, a total of 28 male mice, 1.5–6 months old, were trained on the cross-modal task. All experimental procedures involving animals were performed according to the guidelines of the Institutional Animal Care and Use Committee at the New York University Langone Medical Center and the US National Institutes of Health.

**Behavioural training and testing.** *Behavioural setup.* Experiments were conducted in a custom-built trapezoidal testing chamber (base 1, 12 cm; base 2, 25 cm; height, 25 cm) positioned over a grid floor. The testing chamber contained three nose-pokes, each of which consisted of an infrared LED/infrared phototransistor pair (Digikey, Thief River Falls, Minnesota) for response detection. Activation of a central nose-poke located on the grid floor, 6 cm away from the reward wall, was required for trial initiation. Two headphone speakers (Skullcandy, Park City, Utah) embedded in the floor delivered biasing cues binaurally. Two white LEDs (Mouser, El Cajon, California) were mounted 6.5 cm apart on the base wall below two additional nose-pokes. Liquid reward consisting of 10 µl evaporated milk was delivered directly to these wall-mounted nose-pokes via a single-syringe pump (New Era Pump Systems, Farmingdale, New York). Access to these response nose-pokes was restricted by a rotating, servo-controlled (Tower Hobbies, Champaign, Illinois) disc (radius, 7 cm). Rewards could be accessed from these nose-pokes only when two holes in the rotating disc were aligned with the underlying nose-pokes. Trial logic was controlled by custom software running on an Arduino Leonardo microcontroller (Ivrea, Italy).

**Training.** Mice were food restricted to 85–90% of their *ad libitum* body weight before training. Training consisted of multiple levels. First, mice were habituated to the test box and allowed to collect reward freely. Reward availability was signalled by the rotation of the aforementioned wall-mounted disc. The location of reward (left or right poke) was indicated by either a visual or an auditory stimulus.

For ‘attend to vision’ (visual) trials, the rewarded response poke was indicated by illumination of the LED mounted underneath it. In ‘attend to audition’ (auditory) trials, an upsweep (10–14 kHz, 500 ms) indicated a reward on the left and a downsweep (16–12 kHz, 500 ms) indicated a reward on the right. To facilitate discrimination learning, sweeps were initially presented in a directional manner.

Trials were given in single-modality blocks of six, with alternating block type (that is, six visual trials followed by six auditory trials; Extended Data Fig. 1, top row). The stimulus was presented until the animal collected the reward. An individual trial was terminated 20 s after reward collection and a new trial became available 5 s later. Second, mice learnt to poke to receive a reward. All other parameters remained constant. An incorrect poke had no negative consequence. By the end of this training stage, all mice collected at least 20 rewards per 30-minute session.

Third, mice were trained to initiate individual trials, allowing for the establishment of a temporal window in which they could anticipate subsequent delivery of the stimuli. For successful initiation, mice had to break the infrared beam briefly (50 ms) in the initiation poke to trigger stimulus presentation and rotation of the wall-mounted disc. Mice were informed about trial availability and modality type by brown noise (10-kHz low-pass-filtered white noise, visual trial) or blue noise (11-kHz high-pass-filtered white noise, auditory trial) delivered binaurally. At this stage, modality types were arranged in a non-conflicting block design (Extended Data Fig. 1, top row). Correct poking resulted in reward delivery, whereas incorrect poking resulted in immediate termination of the trial by disc rotation, blocking access to reward. Rewards were available for 15 s following correct poking, followed by a 5-s intertrial interval (ITI). Incorrect poking was punished with a time-out, which consisted of a 30-s ITI. Mice could not initiate new trials during an ITI. To avoid development of side preferences, the target stimulus would appear at the same location as it did on the previous trial following an incorrect response. After one week of training on this stage, mice successfully associated the target stimuli

with the appropriate reward location (Extended Data Fig. 1, top row). At this stage, directionality of sound stimuli did not affect performance.

Fourth, mice had to resolve sensory conflict. Auditory and visual target stimuli were always presented in a conflicting manner (Extended Data Fig. 1, middle row). The brown and blue noise cues indicated the modality to be selected. During a session, four different trial types were presented in blocks in repeating order: (1) three auditory trials, (2) three visual trials, (3) six conflict trials with an auditory target and (4) six conflict trials with a visual target. To prevent modality preferences, an incorrect response resulted in the repetition of the same trial type, thereby specifically increasing the block length of the trial types with weak performance. This training stage was introduced to teach mice to attend only to the target modality during a conflict trial. Over the course of this training stage (1 week), the duration of the target stimuli was successively shortened to 3 s, 1 s and 0.5 s. In parallel, the time that mice had to break the infrared barrier in the initiation nose-poke was continually increased and randomized to a final range of 0.5–0.7 s, rendering the precise presentation time of target stimuli unpredictable. Once mice performed successfully on conflict trials (Extended Data Fig. 1, middle row) the single-modality trials were removed and block length was reduced to three trials. This change in the training paradigm was made to facilitate learning of the trial-type cueing (brown and blue noise).

On the fifth and final stage of training, all block structure was removed and trial type was randomized (Extended Data Fig. 1, bottom row). We used three measures to ensure that mice followed the trial-type cueing and did not employ simple alternating strategies. In addition to computing overall accuracy (Extended Data Fig. 1, graphs on left), we quantified the number of consecutive correct trials (Extended Data Fig. 1, middle column) and calculated the fraction of correct modality switches (Extended Data Fig. 1, graphs on right). At this final stage, rewards were available for only 5 s.

**Psychophysics.** For experiments determining the visual detection psychometric function, the ratio between visual and auditory trials was adjusted from the typical 1/1 to 4/1 to facilitate the acquisition of a larger number of visual trials while maintaining the divided-attention nature of the task. In addition, visual stimulus duration was shortened to 0.1 s and the light was randomly displayed at one of five different intensities (0.15, 0.3, 0.6, 0.9, 1.2 lm). To establish the comparison between psychometric functions of visual-only and divided-attention trials (Fig. 1c), we trained mice that reached criterion (>70% accuracy) on the cross-modal task to perform a visual-only task. For one week, mice were trained on a visual-only task every other day; trials containing only visual target stimuli were cued by broadband white noise. Subsequently, visual-only trials were introduced into the cross-modal task at a 1/4 ratio and in a random interleaved manner. Mice were found to differentially anticipate visual-only and visual target with auditory conflict trials (Fig. 1c), whereas they continued to perform equally well on conflict trials with an auditory target (Extended Data Fig. 3).

To separate the effect of anticipating a conflicting stimulus (top-down) from the presence or absence of a distracting stimulus itself (bottom-up), we performed two experiments. In the first experiment, mice performed the cross-modal task with 70% conflict trials and 30% in which conflict was expected but auditory distraction was removed (Fig. 1d). In the second experiment, mice that had been trained on the cross-modal task had the biasing cues replaced with broadband white noise and the modality rewarded was changed on a session-by-session basis such that mice would deduce it based only on reward history (Fig. 1e).

**Behavioural analysis and determination of visual detection threshold.** Performance in behavioural tests was assessed based on the fraction of correct responses relative to chance level or guess rate (50%,  $\gamma$ ). The visual detection threshold ( $\alpha$ ) and maximum performance ( $\lambda$ ) were estimated by fitting performance across stimulation intensities with a logistic function<sup>29,30</sup>:

$$F(x; \alpha, \beta, \lambda, \gamma) = \gamma + \frac{(1 - \gamma - \lambda)}{1 + \exp(-\beta(x - \alpha))}$$

where  $x$  corresponds to the five stimulus levels expressed as a percentage of maximum stimulus intensity. The fraction of correct trials as a percentage of all trials was summed across sessions and the overall performance as a function of stimulus intensity was fit using maximum likelihood estimation<sup>30</sup> implemented in the Palamedes psychophysical toolbox (<http://www.palamedestoolbox.org/>). Estimation of the distribution of the  $\alpha$  parameter was made via non-parametric bootstrap analysis of curve fits (Fig. 1). To adjust for variable lapse rates (Extended Data Fig. 2), the fraction of correct trials was normalized so that the minimum and maximum performance rates corresponded to 50% and 100%, respectively<sup>31</sup>. Curve fitting and estimation of the  $\alpha$  parameter then proceeded as described above. Model selection for the number of psychophysical parameters was based on the Akaike information criterion<sup>20,32</sup>.

**Optogenetics in behaviour.** For experiments with optical stimulation (Fig. 2), testing conditions were equivalent to the final stage of training. Laser trains of

either blue (for ChR2 activation) or yellow (for eNpHR3.0 activation) light consisting of 50-Hz 18-ms pulses (90% duty cycle) at an intensity of 5–6 mW (measured at the tip of the optic fibres) were delivered on every other trial. On laser trials, stimulation occurred either during the anticipatory period (0.5–0.7 s) or during stimulus presentation (0.5 s). Because behaviour and recording systems were automated and stimulus sequence and optogenetic manipulations varied on a trial-by-trial basis, researchers were not blinded to the conditions. In the case of multiple sequential pharmacological or optogenetic manipulations in the same animals, tests were performed in a predefined, pseudorandom order. For comparisons of multiple groups, Kruskal–Wallis one-way analysis of variance was used to assess variance across groups before pairwise comparisons. Power analysis based on effect size estimates was used to determine sample size required for statistical significance with a power of  $\beta = 0.7$ ; more than three samples were required to detect significant differences.

**Electrophysiology and optical chloride measurements in behaviour.** For combined TRN recordings with optogenetic PFC disruption (Fig. 3d, e), laser trains of blue light (as described earlier) were delivered during the anticipatory period on every other trial. For electrophysiological recordings of LGN units and fibre photometry measurements, visual stimuli were presented through illumination of diffusion-coated wide-angle 3-mm flat-top LED lights (LightHouseLEDS, Washington), fixed directly on the head of the mouse and centred 8 mm from the eyes. The LEDs mounted on the base wall of the behavioural box were turned off in this condition. These changes allowed emitted light to activate  $\sim 150^\circ$  of the visual angle when the eye is centred at rest<sup>33</sup>.

**Viruses.** For retrograde optogenetic tagging and TRN manipulation, FuGB2-pseudotyped retrograde lentiviruses (RG-LV) were used as described previously<sup>20</sup>. visTRN neurons were labelled through injection (0.4–0.6  $\mu\text{l}$ ) of RG-LV-EF1 $\alpha$ -DIO-ChR2-GFP (for activation) or RG-LV-EF1 $\alpha$ -DIO-eNpHR3.0-eYFP (for inactivation) into the primary visual thalamus (anterior–posterior (A–P),  $-2.1 \text{ mm}$ ; medial–lateral (M–L),  $\pm 2 \text{ mm}$ ; dorsal–ventral (D–V),  $2.5 \text{ mm}$ ) using a quintessential stereotactic injector (QSI, Stoelting, Wood Dale, Illinois). Coordinates are referenced to Bregma. For combined electrophysiological recordings with optogenetic PFC disruption during behaviour,  $0.4 \mu\text{l}$  AAV2-hSyn-DIO-ChR2-GFP (titre,  $10^{12}$  vector core per ml) was injected into the PFC (A–P,  $2.6 \text{ mm}$ ; M–L,  $\pm 0.25 \text{ mm}$ ; D–V,  $-1.25 \text{ mm}$ ). To measure chloride flux in the LGN, the transgene *SuperClomeleon* (gift from G. J. Augustine<sup>35</sup>) was cloned into the AgeI and EcoRI restriction sites of an AAV-hSyn-SSFO-eYFP plasmid to obtain AAV-hSyn-SuperClomeleon. The SuperClomeleon recombinant AAV was packaged as serotype 2 (University of North Carolina, Vector core facility; titre,  $10^{12}$  vector core per ml) and  $0.6$ – $0.7 \mu\text{l}$  virus was injected into the visual thalamus. Following injections, mice were allowed to recover for 2–4 weeks to allow for virus expression.

**Optic fibre implant experiments.** Mice were anaesthetized using 1% isoflurane and mounted on a stereotactic frame. For cortical inactivation experiments, up to three pairs of 4–5-mm-long optic fibres (Doric Lenses, Quebec, Canada) were inserted bilaterally to target up to three different brain areas per mouse (prelimbic cortex,  $2.6 \text{ mm A-P}$ ,  $\pm 0.25 \text{ mm M-L}$ ,  $-1.25 \text{ mm D-V}$ ; primary visual cortex,  $-3.5 \text{ mm A-P}$ ,  $\pm 2.50 \text{ mm M-L}$ ,  $-0.50 \text{ mm D-V}$ ; primary auditory cortex,  $-2.8 \text{ mm A-P}$ ,  $\pm 4.00 \text{ mm M-L}$ ,  $-2.00 \text{ mm D-V}$ ; AAC,  $0.5 \text{ mm A-P}$ ,  $\pm 0.25 \text{ mm M-L}$ ,  $-1.00 \text{ mm D-V}$ ; lateral OFC,  $2.6 \text{ mm A-P}$ ,  $\pm 1.50 \text{ mm M-L}$ ,  $-2.00 \text{ mm D-V}$ ; primary visual thalamus,  $-2.1 \text{ mm A-P}$ ,  $\pm 2.00 \text{ mm M-L}$ ,  $-2.50 \text{ mm D-V}$ ; visTRN,  $-1.6 \text{ mm A-P}$ ,  $\pm 2.20 \text{ mm M-L}$ ,  $-3.00 \text{ mm D-V}$ ). Two or three stainless steel screws were implanted into the skull to anchor the implant and were fixed with dental cement. Animals were allowed to recover and training resumed one week later. For ChR2 activation a 473-nm laser was used and a 561-nm laser was used for eNpHR3.0 activation (Omicron-Laserage, Dudenhofen, Germany).

**Drive construction.** Custom drive housings were designed using 3D CAD software (SolidWorks, Concord, Massachusetts) and printed in Accura 55 plastic (American Precision Prototyping, Tulsa, Oklahoma) as described previously<sup>20,34</sup>. Prior to implantation, each drive was loaded with 8–12 independently movable microdrives carrying up to 3 nichrome ( $12.5 \mu\text{m}$ ) and/or tungsten ( $25 \mu\text{m}$ ) stereotrodes (California Fine Wire Company, Grove Beach, California). Stereotrodes were pinned to custom-designed 32- or 64-channel electrode interface boards (EIB; Sunstone Circuits, Mulino, Oregon) along with a common reference wire (A-M systems, Carlsborg, Washington). For optogenetic tagging, an optical fibre was embedded adjacent to the stereotrode array. In these cases, the optic fibre extended 3.5 mm from the base of the drive so that it could be stereotactically positioned above the TRN during implantation. Targeting of the TRN or LGN was achieved by guiding stereotrodes and optic fibres through a square array of polyimide sleeves attached to the base of the drive body.

**Drive and fibre implantation for optical activity measurements.** Prior to surgical implantation, mice were anaesthetized with 1% isoflurane and placed in a stereotactic frame. Stainless steel screws were implanted into the skull to provide

electrical and mechanical stability for the drives. For drive implantations, craniotomies ( $\sim 3 \text{ mm} \times 2 \text{ mm}$ ) were drilled, centred at  $-2 \text{ mm A-P}$  and  $2.5 \text{ mm M-L}$  for TRN recordings ( $15^\circ$  angled implantation) and at  $-2.3 \text{ mm A-P}$  and  $-2.5 \text{ mm M-L}$  for LGN recordings. The dura mater was carefully removed and drives were centred at the craniotomy coordinates using a custom stereotactic arm. Drive bodies were slowly lowered into the craniotomy until stereotrode tips were  $\sim 500 \mu\text{m}$  below the cortical surface and optical fibres were positioned just above the TRN ( $2.5 \text{ mm D-V}$ ). For fibre-photometry-based optical recording, low-internal-fluorescence optic fibres ( $400 \mu\text{m}$  diameter) (Doric Lenses, Canada) were implanted just dorsal to the LGN ( $-2.2 \text{ mm A-P}$ ,  $2.15 \text{ mm M-L}$  and  $2.6 \text{ mm D-V}$ ) following virus injection.

**Electrophysiological recordings.** After mice had recovered from implantation surgery, recordings were made using a Neuralynx multiplexing digital recording system (Neuralynx, Bozeman, Montana). Signals were acquired using a 32- or 64-channel digital headstage connected to the implanted EIB. Signals from each electrode were amplified, filtered between 0.1 Hz and 9 kHz and digitized at 30 kHz. Local field potential signals were obtained from a single wire per stereotrode. Following implantation, stereotrode sets were incrementally lowered from the cortex into the target thalamic structure over the course of 1–2 weeks (Extended Data Fig. 5). Spike sorting was performed offline following acquisition based on relative spike amplitude and energy within electrode pairs using the MClust toolbox (<http://redishlab.neuroscience.umn.edu/mclust/MClust.html>). Following manual clustering, cross-correlation and autocorrelation analyses were used to confirm adequate separation.

Optogenetically tagged visTRN units were identified based on ChR2-mediated response to stimulation using a 473-nm analogue-modulated laser (Omicron-Laserage, Dudenhofen, Germany)<sup>20</sup>. Laser light was delivered by a  $200-\mu\text{m}$  optic fibre targeted to the TRN (Extended Data Fig. 5) connected to a fibre optic patch cord ( $200-\mu\text{m}$  core, Doric Lenses, Quebec, Canada). The laser intensity was set at  $\sim 8 \text{ mW}$  optical output power measured at the patch cord terminus. Fibres were polished before implantation so that the power at the tip was  $\geq 50\%$  maximum, resulting in  $\sim 4$ – $5 \text{ mW}$  laser light being delivered to the brain. Only neurons that showed clear transient responses to laser stimulation were included in the analysis.

**Analysis of firing rate.** Changes in firing rate during task performance were assessed for 138 identified visTRN neurons recorded from four animals and 119 LGN neurons in two animals. Peri-event time histograms aligned to trial initiation and to stimulus presentation were computed using a 5-ms bin width for individual neurons in each recording session<sup>4</sup>. Separate histograms were created for correct and incorrect trials within auditory and visual target stimuli and convolved with a Gaussian kernel (8 ms half-width at half-height) to create a spike density function<sup>35,36</sup>. The average firing rate across trials was determined during the anticipation window before stimulus presentation. The evoked response amplitude was estimated by averaging the firing rate within a 100-ms window starting 20 ms after stimulus onset. Window duration was chosen based on the latency-to-peak response for point stimuli in the mouse LGN<sup>37</sup>.

For normalized rate changes in TRN neurons, firing rates during the attentional window in each trial were compared with the baseline firing rate (5-s window, 0.5 s before task initiation). Statistical comparison of firing rate changes was used to identify neurons with significant task-associated changes in firing rate via non-parametric comparison of firing rate during the attentional window and the baseline period<sup>38</sup>. The test statistic ( $W$ ) was calculated based on ranking of all trials ( $N$ ) and comparison was performed using the sign function (sgn):

$$W = \left| \sum_j^N [\operatorname{sgn}(x_{2,j} - x_{1,j}) \cdot R_j] \right|$$

where  $x_1$  and  $x_2$  were the attentional window and baseline firing rate, respectively, and  $R$  denotes the rank. The threshold for significance was set at 0.05 and significantly modulated units were defined as neurons in which the test statistic was less than the critical value for the sample size ( $W_{\alpha=0.05}$ ). Comparison of firing rates across trial types (for example, visual versus auditory correct) was performed using the Wilcoxon rank-sum test. Homogeneity of variance for firing rates across conditions was determined using the Fligner–Killeen test of homoscedasticity.

**Analysis of visual evoked potentials.** Visual evoked potentials (VEPs) were computed from the broadband LGN local field potential (LFP; 0.1 Hz–10 kHz). The particular stereotrode used for VEP analysis in behaviour was selected based on the amplitude of responses in post-task recordings during which there were many more trials included. Task-related VEPs were averaged during correct auditory and visual trials across recording sessions. To determine peak response, the lowest negative-potential offsets associated with the visual response<sup>39</sup> (0–250-ms window) were identified on a trial-by-trial basis. Signals from individual trials were smoothed with a 25-ms half-width filter over the response window before obtaining the peak offset<sup>40</sup>.

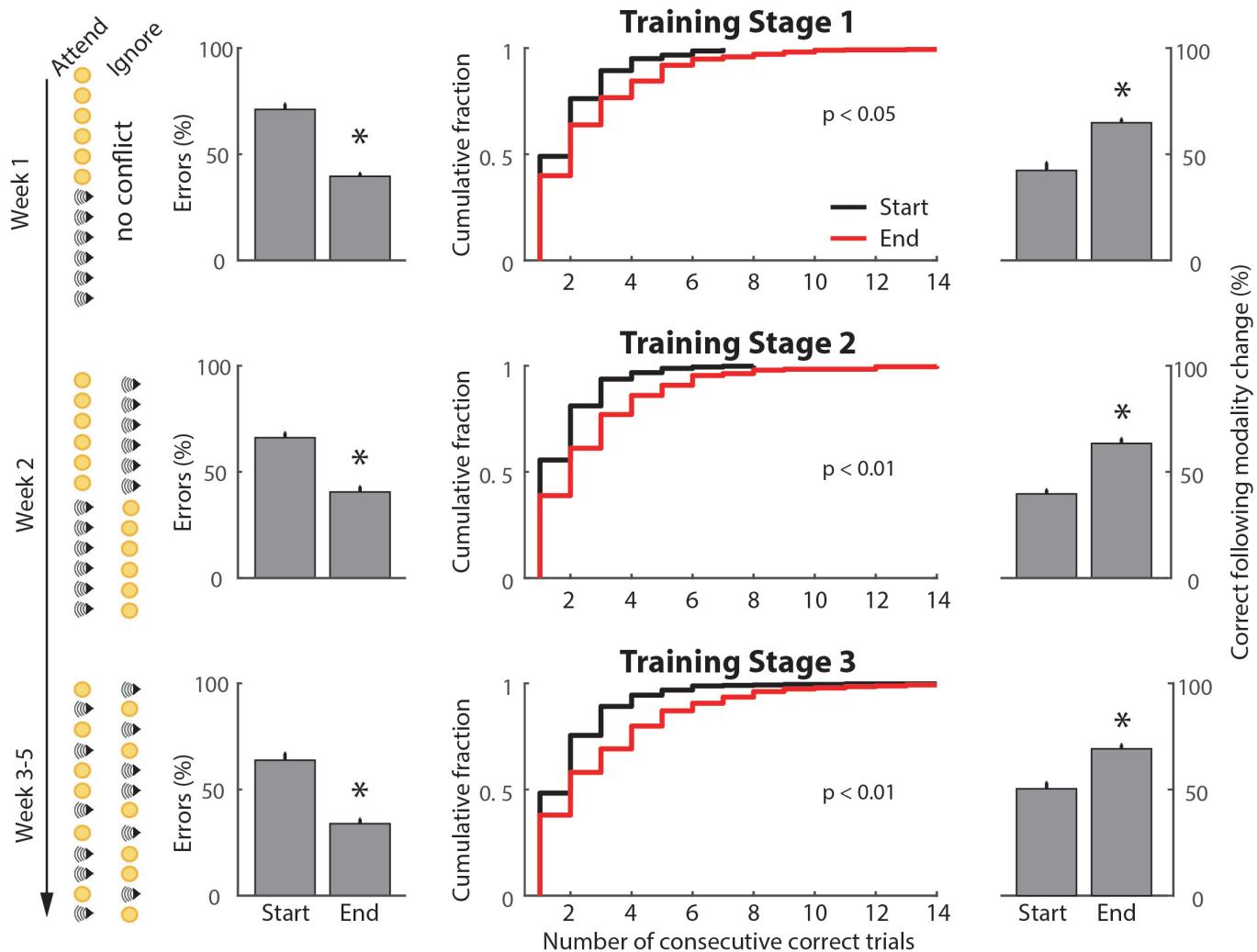
**Histology.** Mice were euthanized and transcardially perfused with PBS followed by 4% paraformaldehyde. Brains were dissected, post-fixed overnight at 4 °C and sectioned using a vibratome (LEICA, Buffalo Grove, Illinois).

For GFP enhancement, immunofluorescent staining was carried out on 50-μm-thick sections using chicken anti-GFP (1/1,000, GFP-1020, Aves). Sections were incubated overnight with primary antibody in PBS-T (10% normal goat serum and 0.05% Tween20) at 4 °C. Detection of primary antibodies was carried out with Alexa-Fluor-conjugated secondary antibodies (1/1,000, A-11039, Invitrogen). All sections were imaged on a Zeiss LSM510 META confocal microscope.

**Fibre-photometry-based optical chloride measurements.** FRET-based measurement of chloride was performed during behaviour using a custom-designed fibre photometry system<sup>24</sup>. A fibre-coupled LED (Thorlabs, Newton, New Jersey) light source, filtered using a 434-nm clean-up filter (MF434-17 Thorlabs, Newton, New Jersey), was used for CFP excitation. Excitation light was split via a long-pass dichroic mirror (DMLP425, Thorlabs, Newton, New Jersey) and coupled to a 400-μm, 0.48-NA (pharmacology) optic patch cord (Doric lenses, Canada) linked to a 400-μm chronically implanted optical fibre. Excitation and emission light were conveyed by a single patch cord linking the fibre photometry system to the implanted fibre. SuperClomeleon CFP and YFP emissions<sup>25</sup> were separated using a single-edge beam splitter (FF511-Di01, Semrock, Rochester, New York). Each emission wavelength was independently focused onto a separate femtowatt silicon photoreceiver (Newport, Irvine, California) using custom optics (12.7-mm focal length plano-convex lens mounted in Thorlabs SM1NR05 lens tube). The light signal was digitized and recorded using a TDT signal acquisition system (Tucker-Davis Technologies, Alachua, Florida). Signal bandwidth was limited to <750 Hz based on the photoreceiver response characteristics. The fluorescence ratio was calculated across the recording period. To minimize the effect of slow fluctuations, normalized delta fluorescence ( $\Delta f/F$ ) was calculated for evoked responses relative to the baseline fluorescence level before each event (1-s window). Traces were smoothed with a convolution filter (50 ms half-width). Peak response (Extended Data Fig. 10) was estimated as the minimum over a 500-ms window following stimulus onset. For pharmacological activation of GABA<sub>A</sub> receptors with 4,5,6,7-tetrahydroisoxazolo[5,4-c]pyridin-3-ol (THIP), baseline fluorescence was estimated over 5 minutes before injection. For visual stimulation, light pulses of 100-ms duration were displayed to the ipsi- or contralateral side of the recorded LGN.

Effects of the GABA<sub>A</sub> receptor antagonist flumazenil on visual evoked responses were quantified by comparing the average peak response from 5 minutes before injection (baseline) to one within a 5-minute time window around the maximal response suppression (maximal drug effect) and at the end of the recording session (recovery, at least 100 min after injection). For optogenetic manipulations of frontal cortical structures, smaller-diameter patch cords (200 μm, 0.37 NA) were used to allow movement and prevent tangling. For these recordings, power analysis was performed to determine sample size required to detect significant differences with a power of  $\beta = 0.7$  based on the observed differential signal in correct auditory and visual trials under baseline conditions. Analysis indicated that more than four independent samples would be required to detect a change in these differential responses.

29. Ress, D. & Heeger, D. J. Neuronal correlates of perception in early visual cortex. *Nature Neurosci.* **6**, 414–420 (2003).
30. Levitan, C. A., Ban, Y. H., Stiles, N. R. & Shimojo, S. Rate perception adapts across the senses: evidence for a unified timing mechanism. *Sci. Rep.* **5**, 8857 (2015).
31. Mareschal, I., Calder, A. J., Dadds, M. R. & Clifford, C. W. Gaze categorization under uncertainty: psychophysics and modeling. *J. Vis.* **13**, 18 (2013).
32. Hastie, T., Tibshirani, R. & Friedman, J. *The Elements of Statistical Learning: Data Mining, Inference and Prediction* 2nd edn, Ch. 7 (Springer, 2009).
33. Wallace, D. J. et al. Rats maintain an overhead binocular field at the expense of constant fusion. *Nature* **498**, 65–69 (2013).
34. Brunetti, P. M. et al. Design and fabrication of ultralight weight, adjustable multi-electrode probes for electrophysiological recordings in mice. *J. Vis. Exp.* **91**, e51675 (2014).
35. Fries, P., Neuenschwander, S., Engel, A. K., Goebel, R. & Singer, W. Rapid feature selective neuronal synchronization through correlated latency shifting. *Nature Neurosci.* **4**, 194–200 (2001).
36. Szucs, A. Applications of the spike density function in analysis of neuronal firing patterns. *J. Neurosci. Methods* **81**, 159–167 (1998).
37. Piscopo, D. M., El-Danaf, R. N., Huberman, A. D. & Niell, C. M. Diverse visual features encoded in mouse lateral geniculate nucleus. *J. Neurosci.* **33**, 4642–4656 (2013).
38. De Araujo, I. E. et al. Neural ensemble coding of satiety states. *Neuron* **51**, 483–494 (2006).
39. Ridder III, W. H. & Nusinowitz, S. The visual evoked potential in the mouse—origins and response characteristics. *Vision Res.* **46**, 902–913 (2006).
40. Izaki, Y., Fujiwara, S. E. & Akema, T. Rat prefrontal response and prestimulation local field potential power in vivo. *Neuroreport* **19**, 255–258 (2008).

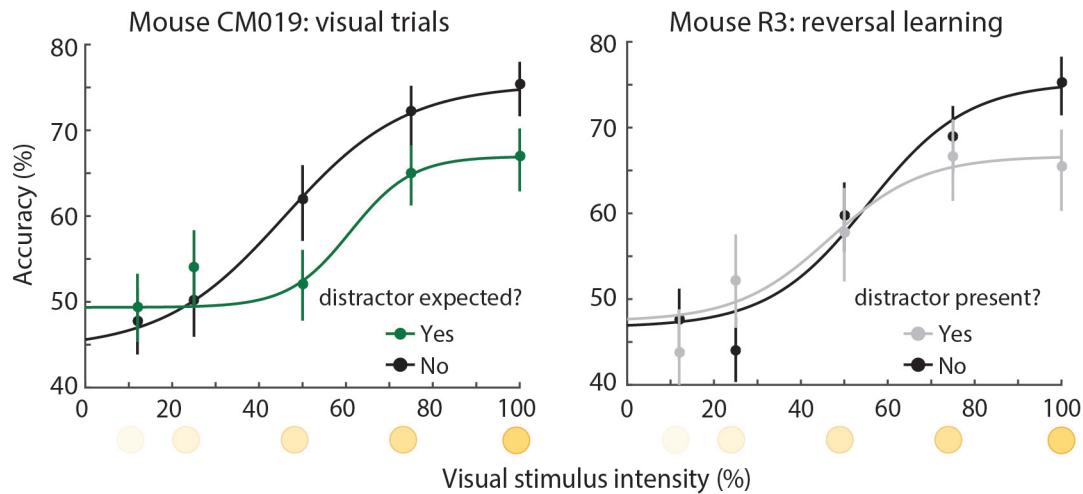


#### Extended Data Figure 1 | Cross-modal task training and performance validation

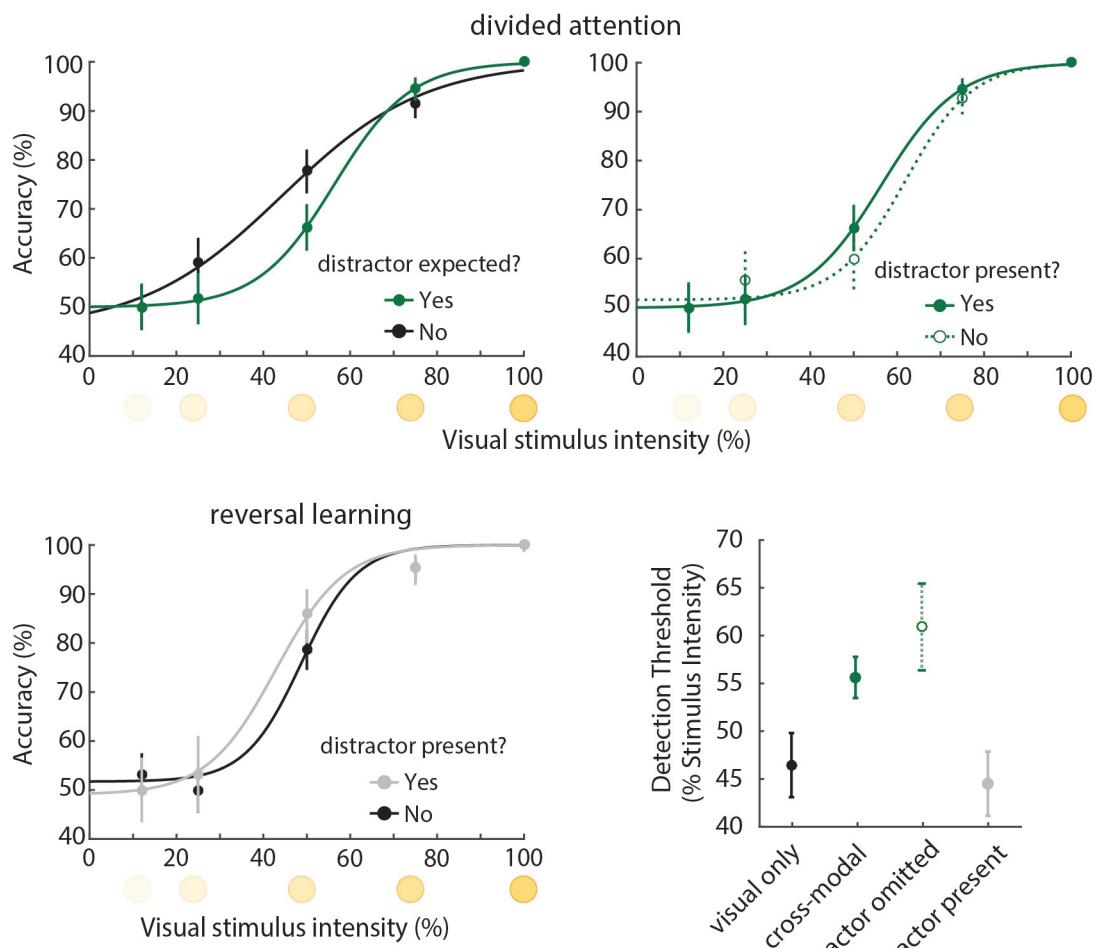
**validation.** Quantification of performance across training stages for the cross-modal task. The trial sequence for each training stage is indicated on the left. Improved performance was observed in the last three days of training relative to the first three for each stage. Bar graphs on the left (column 1) show the

reduction in the error fraction ( $n = 15$  mice,  $*P < 0.05$ , Wilcoxon rank-sum test), column 2 shows the number of consecutive correct responses ( $P$ -values shown, Kolmogorov–Smirnov test) and bar graphs on the right (column 3) show the probability of a correct response following a modality shift ( $*P < 0.05$ , Wilcoxon rank-sum test).

## Single Mouse Data

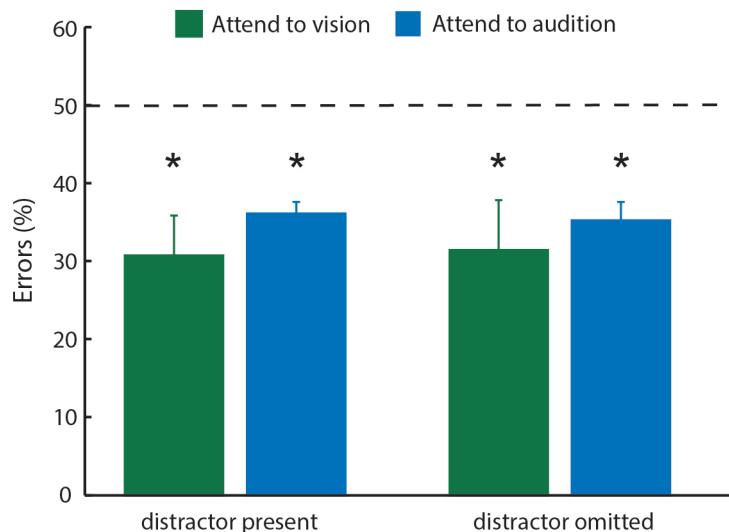


## Group Data, Lapse rate corrected



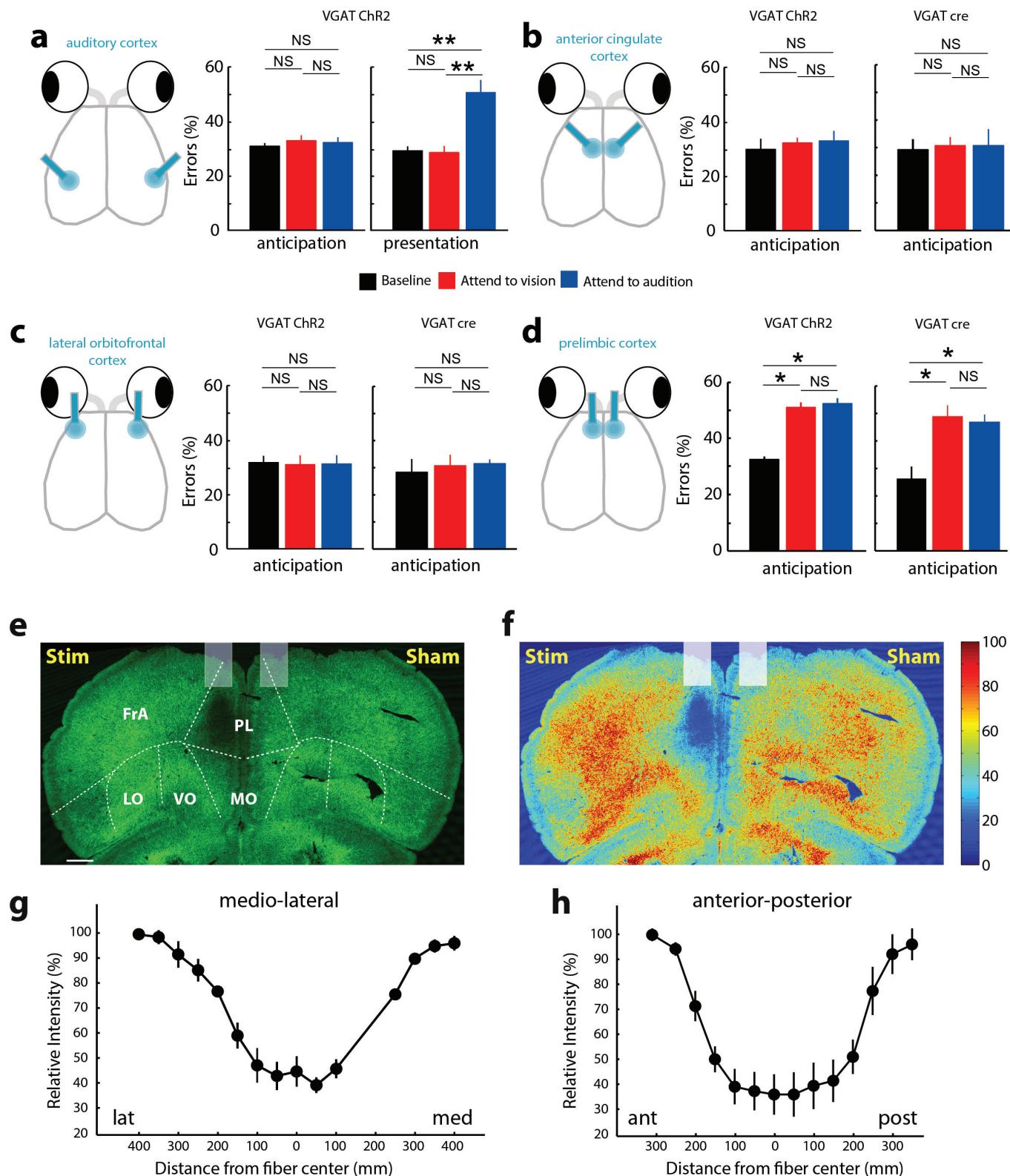
**Extended Data Figure 2 | Effects of cross-modal divided attention in the mouse.** Top row, single-mouse examples of visual detection performance during cross-modal divided attention and reversal learning. Comparison of performance under visual-only (black) and cross-modal (green) conditions are shown on the left. Although neither condition contained sensory conflict, the mere expectation of one increased detection threshold ( $\geq 124$  trials per

condition). Detection threshold was not affected by the presence of an auditory distractor during reversal learning ( $\geq 90$  trials per condition), as shown on the right. Middle and bottom rows, group data normalized to peak performance (lapse rate), showing that the effects of divided attention on detection threshold were persistent. Bootstrap estimation of visual detection threshold shows a similar pattern as data in Fig. 1 (error bars are 95% confidence intervals).



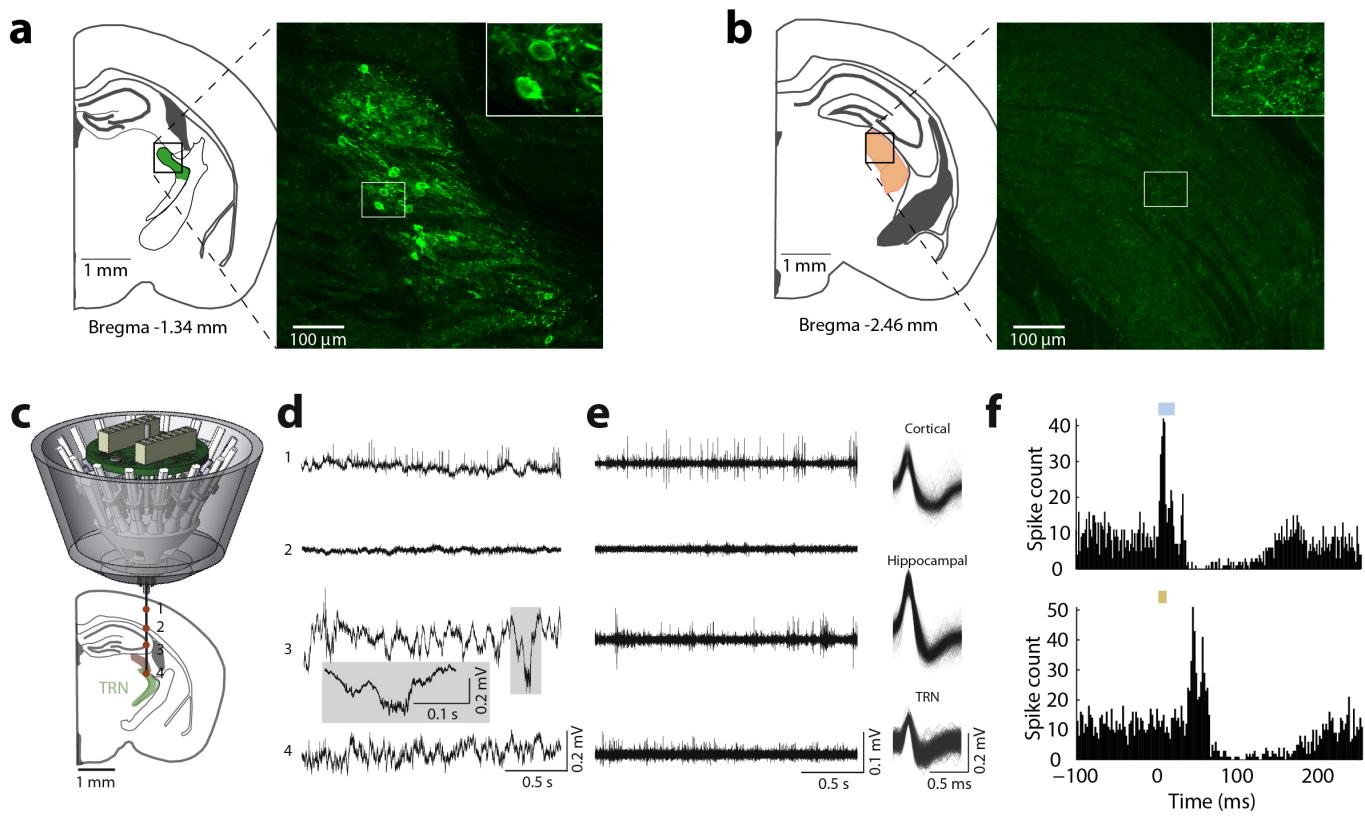
**Extended Data Figure 3 | Comparable performance on trial types and intact overall auditory performance despite auditory stimuli being eliminated on a subset of ‘attend to vision’ trials.** Left, performance was comparable on

auditory and maximum-intensity visual trials ( $n = 4$  mice, same as in Fig. 1d). Right, mice exhibited comparable overall performance when auditory stimuli were eliminated from a subset of ‘attend to vision’ trials.



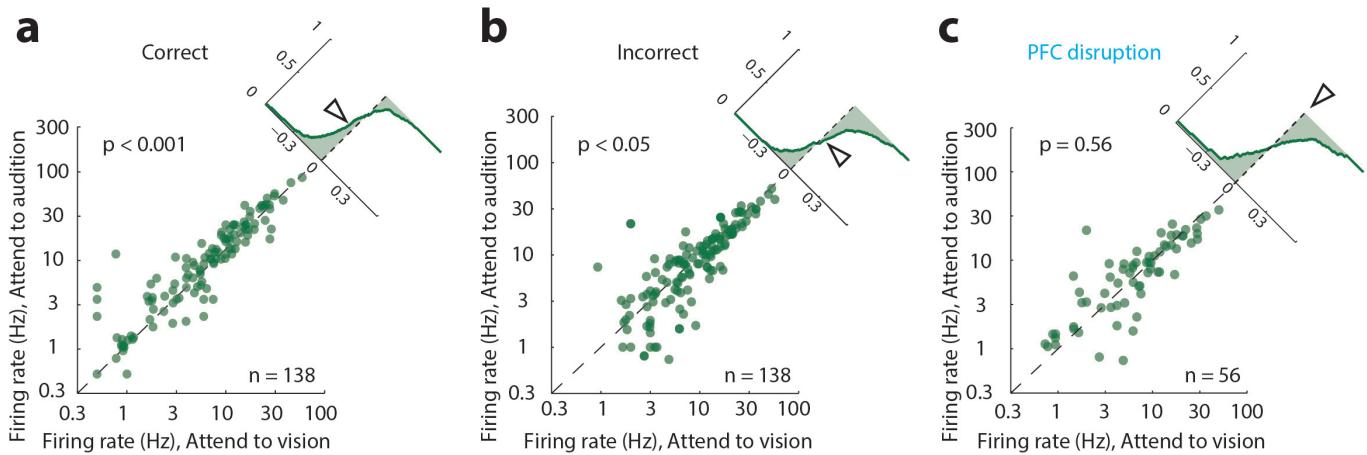
**Extended Data Figure 4 | Region- and timing-specific effects of optogenetic manipulation on cross-modal task performance.** **a**, Optogenetic disruption of auditory cortex during target stimulus anticipation disrupted performance specifically for auditory trials ( $n = 4$  mice,  $**P < 0.01$ , Wilcoxon rank-sum test). Disruption of AAC (b) or lateral OFC (c) in VGAT-ChR2 mice or following localized injection of a ChR2-expressing virus did not affect performance ( $n = 4$  mice (2 VGAT-ChR2 and 2 VGAT-Cre), 4 sessions per manipulation). **d**, In contrast, inactivation of prelimbic (PL) cortex led to robust reduction in performance in both types of manipulation ( $n = 8$  mice

(4 VGAT-ChR2 and 4 VGAT-Cre),  $*P < 0.05$  Wilcoxon rank-sum test). **e–h**, Photobleaching experiment to quantify the spread of laser light. A coronal section (e) shows GFP bleaching following two-hour exposure to laser stimulation (6 mW, 50 Hz, 90% duty cycle). **f–h**, Fluorescence intensity quantification shows that the extent of light spread is limited to 300  $\mu$ m around the tip of the optic fibre ( $n = 3$  mice). Ant, anterior; FrA, frontal association cortex; lat, lateral; LO, lateral orbitofrontal cortex; med, medial; MO, medial orbitofrontal cortex; post, posterior; Stim, stimulation; Sham, sham surgery control; VO, ventral orbitofrontal cortex. Scale bar in e, 200  $\mu$ m.



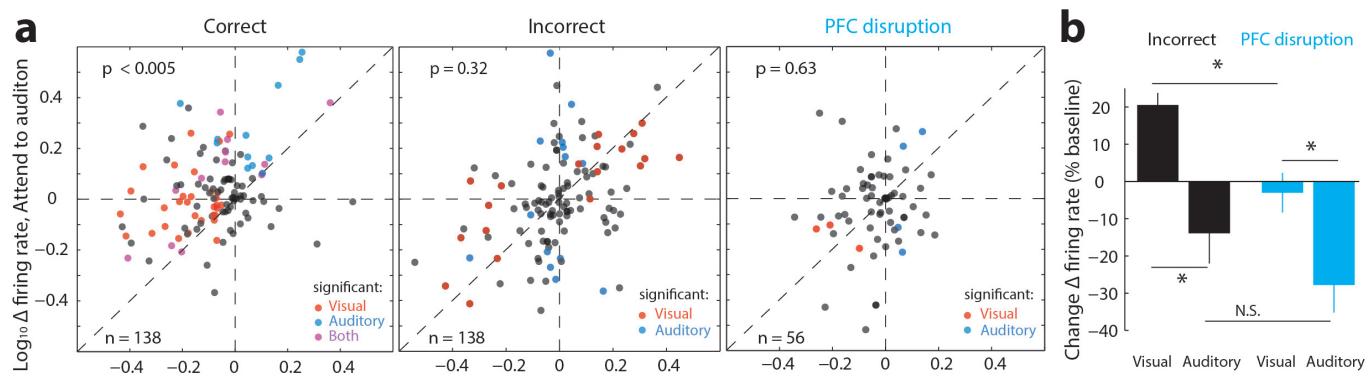
**Extended Data Figure 5 | Independently adjustable, multi-electrode recording of visTRN neurons.** **a, b,** Injection of DIO-ChR2-eYFP retrograde lentivirus into LGN labels visTRN neurons but not LGN interneurons. **a,** The histological image is the maximal projection of four 2- $\mu\text{m}$  confocal planes showing labelling of visTRN neurons (inset shows a zoom view of cell bodies) approximately 1.34 mm posterior to Bregma. **b,** Image as in **a**, but from LGN of the same animal, approximately 2.46 mm posterior to Bregma (inset shows a zoom view of terminals). **c,** Schematic of independently adjustable multi-electrode drive. **d,** An example of activity recorded from different depths during adjustment. Distinct patterns of physiological activity are observed along the

trajectory in the broadband local field potential signal (0.1 Hz–32 kHz). The numbers correspond to different recording sites (marked by the red dots on the schematic in **c**). **e,** High-pass-filtered signals (600 Hz–10 kHz) showing spiking activity, with isolated clustered units showing distinguishable waveform characteristics in distinct structures. **f,** Example peri-event time histograms of ChR2-mediated visTRN response. Top, response to laser activation (473 nm,  $\sim 4$  mW, stimulation, 20 ms). Bottom, response to visual stimuli (10-ms pulse). Blue and orange blocks indicate laser and visual stimulation, respectively.



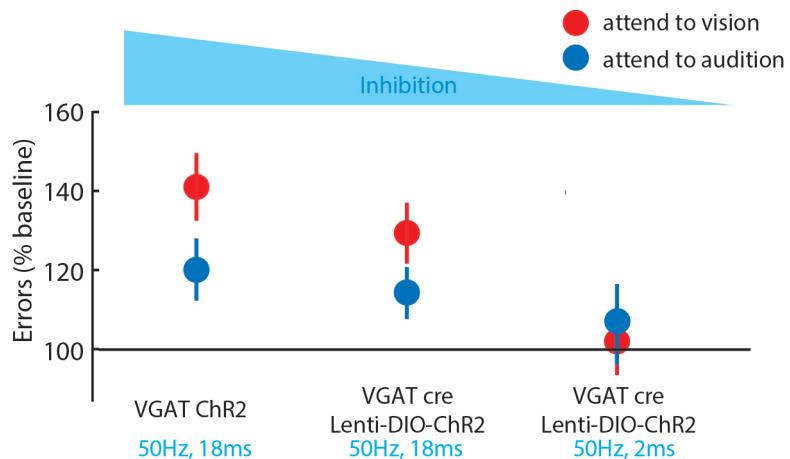
**Extended Data Figure 6 | Distinct changes in visTRN firing rate during natural errors compared to errors due to PFC disruption.** a–c, Scatter plots showing the change in absolute firing rate for visTRN neurons for correct (a), incorrect (b) or disrupted-PFC trials (c). Insets show the cumulative probability plot of separation from the unity line (no change). Although correct trials had a lower firing rate in ‘attend to vision’ than in ‘attend to audition’ trials ( $n = 138$ ,

$P < 0.001$  Wilcoxon signed-rank test), this pattern was reversed for incorrect trials ( $n = 138$ ,  $P < 0.05$ , Wilcoxon signed-rank test); this suggests that perhaps the animal was attending to the wrong modality. This reversal was not observed in trials with PFC disruption (despite mouse performance being at chance level).



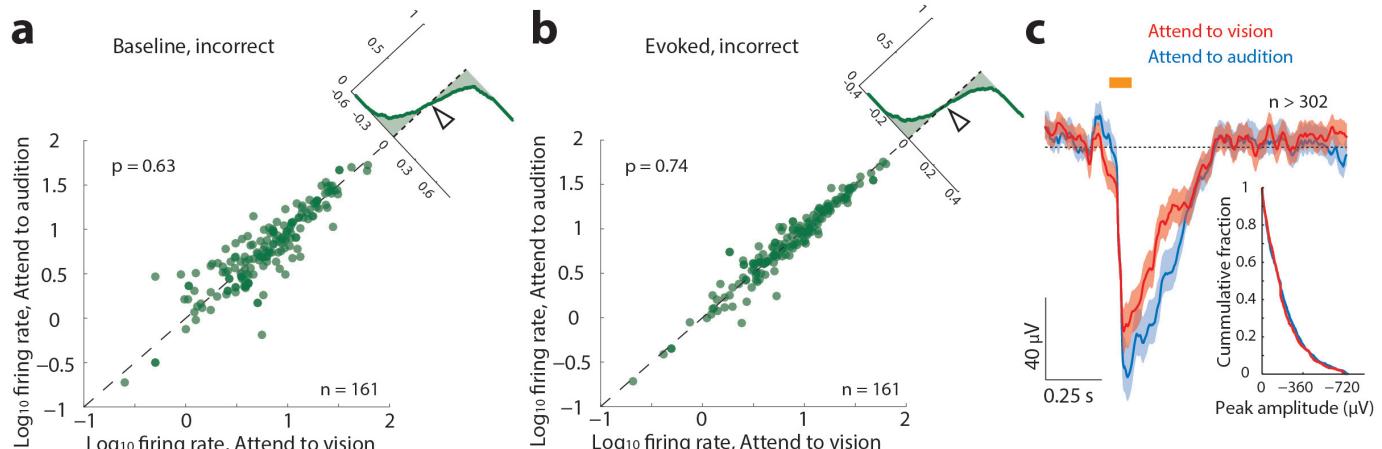
**Extended Data Figure 7 | The effect of PFC disruption on visTRN activity is distinct from naturally occurring errors.** **a**, Scatter plots of response from visTRN neurons, comparing the modulation of their firing rate (change from baseline) under the two distinct anticipatory conditions. Each sample is a single cell. Colours denote significance reached for each cell on a trial-by-trial basis (red, visual; blue, auditory; purple, both; rank-sum-test comparison to baseline). Note that in correct performance ( $n = 138$ , 4 mice,  $P < 0.005$ , Wilcoxon signed-rank test), ‘attend to vision’ resulted in a negative shift and

‘attend to audition’ resulted in a positive shift, consistent with examples shown in Fig. 3. During naturally occurring error trials, the modulation is partially reversed for both trial types, suggesting that at least a subset of errors are the result of attending to the wrong modality. In contrast, PFC disruption ( $n = 56$  cells, 2 mice) resulted in a weaker, non-uniform effect (‘attend to visual’ trials are less affected). **b**, Quantification of effects seen in **a**. N.S., not significant.



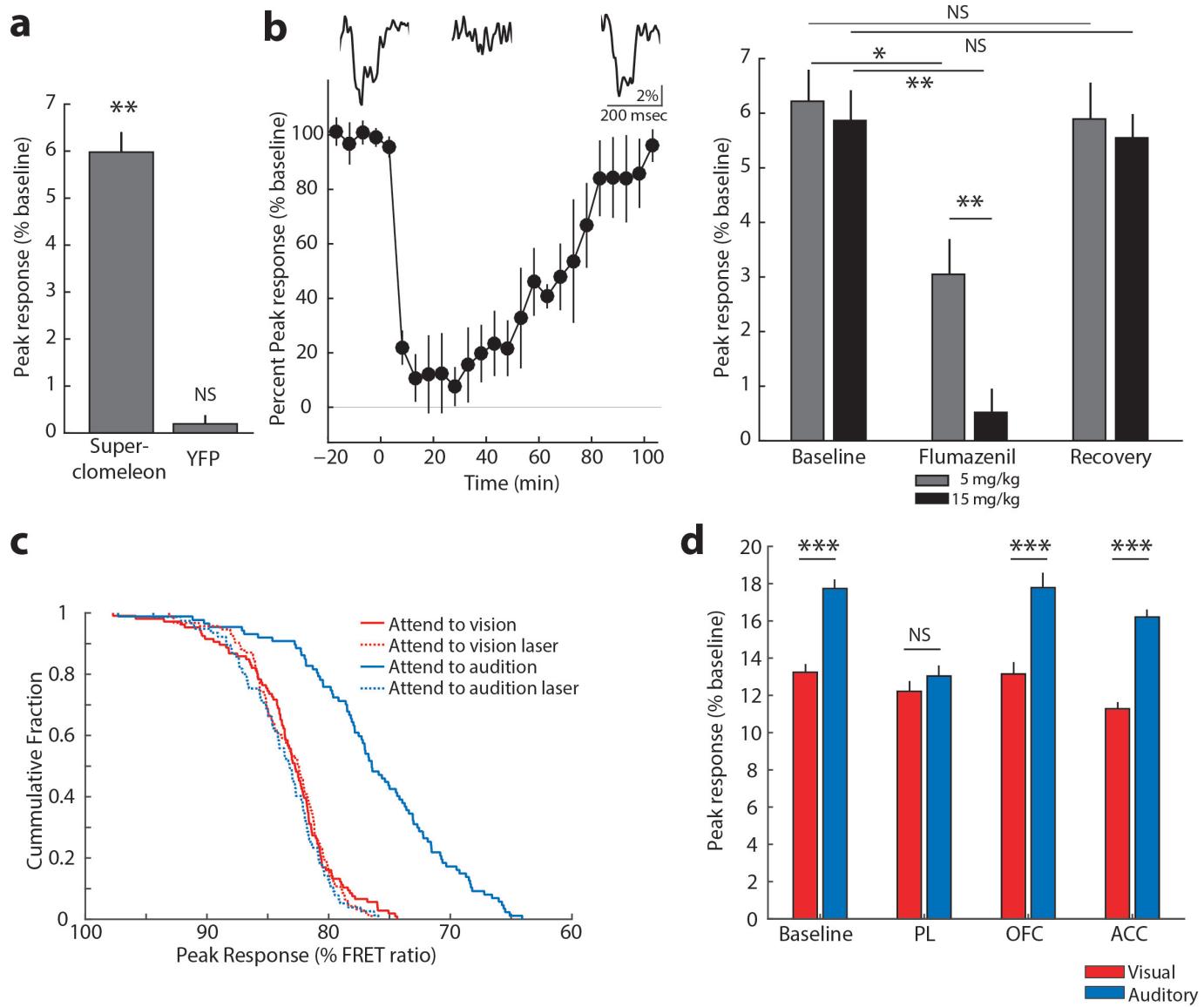
**Extended Data Figure 8 | The magnitude of behavioural disruption co-varies with the strength of optogenetic manipulation of the LGN or visTRN.** Activation of inhibitory terminals in the LGN with a 90% duty cycle laser (Fig. 2) resulted in maximal disruption of cross-modal performance. Activating

visually labelled TRN with identical stimulation parameters resulted in a quantitatively lower behavioural effect. Reducing the duty cycle of visTRN stimulation to 10% resulted in no effect on accuracy, as shown previously<sup>4</sup>.



**Extended Data Figure 9 | Attentional modulation by LGN is not observed on error trials.** **a, b**, No significant difference was observed in the average firing rate of LGN neurons during stimulus anticipation ( $P = 0.63$ , Wilcoxon signed-rank test,  $n = 161$  cells, 4 mice) or presentation ( $P = 0.74$ , Wilcoxon

signed-rank test,  $n = 161$ ) among trial types when behavioural outcomes were incorrect. **c**, Similar effects were observed for VEPs (visual,  $n = 324$  trials; auditory,  $n = 302$  trials; 4 mice).



**Extended Data Figure 10 | Light-evoked fast transients from chloride photometry measured in the LGN are GABA<sub>A</sub>-receptor dependent and sensitive to visTRN and prelimbic inactivation in the cross-modal task.**  
**a**, Peak SuperClomeleon FRET- and YFP-control responses to light stimuli (50 ms, 0.1 Hz) delivered to the eye contralateral to the recorded LGN ( $n > 90$  trials from 3 mice for SuperClomeleon and from 4 mice for YFP, \*\*\* $P < 0.001$ , Friedman test). **b**, Chloride photometry transients are sensitive to the GABA<sub>A</sub> receptor antagonist flumazenil in a dose-dependent manner. Left, intraperitoneal injection of 15 mg kg<sup>-1</sup> flumazenil resulted in a 90% peak reduction of light-evoked chloride photometry responses, which recovered over the course of 90–100 min as predicted by flumazenil pharmacokinetics. Insets show example traces of single events recorded during baseline, peak suppression and recovery. Right, quantification of the maximal suppressive effects and recovery of 5 mg kg<sup>-1</sup> and 15 mg kg<sup>-1</sup> flumazenil on chloride

photometry responses ( $n > 90$  trials from 3 mice, \* $P < 0.05$ , \*\* $P < 0.01$ , Friedman test). **c**, Cumulative distributions of unitary visual-evoked SuperClomeleon FRET peaks in response to light stimuli in the cross-modal task. Under baseline conditions, ‘attend to audition’ trials exhibited significantly larger amplitudes than ‘attend to vision’ trials, consistent with average data in Fig. 5f. Optogenetic silencing of visTRN neurons eliminated the difference between trial types and resulted in peak amplitudes comparable to baseline ‘attend to vision’ trials ( $n = 3$  mice,  $P < 0.005$  for ‘attend to audition’ trials vs all other trial types, Kolmogorov–Smirnov statistics with Bonferroni correction). **d**, Combined optogenetic and chloride photometry inactivation of different frontal cortical regions in the LGN while mice performed the cross-modal task. Only PL inactivation eliminates differential inhibition between visual and auditory trials ( $n = 6$  mice, \*\*\* $P < 0.001$ , Wilcoxon rank-sum test).