metaSEM: An R Package for Meta-Analysis Using Structural Equation Modeling

Mike W.-L. Cheung

National University of Singapore

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Abstract

The **metaSEM** package provides functions to conducting univariate and multivariate meta-analysis using a structural equation modeling approach via the **OpenMx** package. It also implemented the two-stage structural equation modeling (TSSEM) approach to conducting fixed- and random-effects meta-analytic structural equation modeling (MASEM) on correlation or covariance matrices. This paper outlines the basic theories of its implementations. Examples are used to illustrate the procedures.

Keywords: meta-analysis, structural equation modeling, meta-analytic structural equation modeling, metaSEM, R.

1. Introduction

Meta-analysis is a popular technique to synthesizing research findings in social, behavioral, educational and medical sciences (e.g., Borenstein, Hedges, Higgins, and Rothstein 2009; Hedges and Olkin 1985; Hunter and Schmidt 2004; Whitehead 2002). There are several standalone packages to conducting meta-analysis, e.g., Comprehensive Meta-Analysis (Borenstein, Hedges, and Rothstein 2005) and Review Manager (The Nordic Cochrane Centre 2011). Many standard statistical packages, for instance, SPSS (Lipsey and Wilson 2000), SAS (Arthur, Bennett, and Huffcutt 2001) and STATA (Sterne 2009), have macros or packages to fitting some meta-analytic models. There are also several R packages for meta-analysis, for instance, meta (Schwarzer 2012), rmeta (Lumley 2009), mvmeta (Gasparrini 2012), metaLik (Guolo and Varin 2012) and metafor (Viechtbauer 2010).

The metaSEM package is yet another R package to conducting univariate and multivariate meta-analysis. It formulates meta-analytic models as structural equation models (Cheung 2008, 2011b) via the OpenMx package (Boker, Neale, Maes, Wilde, Spiegel, Brick, Spies, Estabrook, Kenny, Bates, Mehta, and Fox 2011). It also implemented the two-stage structural equation modeling (TSSEM) approach (Cheung and Chan 2005b, 2009) to fitting fixed- and random-effects meta-analytic structural equation modeling

(MASEM) on correlation or covariance matrices. The main functions in this package are:

- meta() and reml(): meta() fits univariate and multivariate meta-analysis with maximum likelihood (ML) estimation method while reml() estimates the variance components of the random-effects with restricted (residual) maximum likelihood (REML) estimation method. Mixed-effects meta-analysis can be fitted by using study characteristics as predictors. Equality constraints on the intercepts, regression coefficients and variance components can be imposed.
- meta3() and rem13(): They fit three-level meta-analysis by considering cluster effect. meta3() fits the three-level meta-analysis with ML estimation method while rem13() estimates the variance components with REML estimation method.
- tssem1() and tssem2(): tssem1() fits the first stage analysis of TSSEM by pooling correlation or covariance matrices with either a fixed- or random-effects model. tssem2(), which is a wrapper of wls(), conducts the second stage analysis of TSSEM by fitting structural models on the pooled correlation or covariance matrix.
- wls(): It fits a correlation or covariance structure analysis with weighted least squares (WLS) estimation method.

This paper was based on the **metaSEM** package version 0.7-1, the **OpenMx** package version 1.2.4-2063 and R version 2.15.1. The remaining sections are organized as follows. The next section presents basic ideas on structural equation models and how they are linked to meta-analytic models. Basic theory on the TSSEM are then presented. Several examples are used to illustrate these procedures. How to install the package is finally mentioned.

2. Structural Equation Modeling Based Meta-Analysis

In this section, basic theories in structural equation modeling (SEM) are introduced. Univariate and multivariate meta-analysis are then formulated as special cases of SEM (see Cheung 2008, 2011b).

2.1. Structural equation model

Structural equation modeling is a multivariate technique to fitting and testing hypothesized models. Let \mathbf{y} be a $p \times 1$ vector of sample of continuous data where p is the number of variables. It is hypothesized that the model for the first and the second moments are $\mu = \mu(\theta)$ and $\Sigma = \Sigma(\theta)$, respectively, where θ is a vector of parameters. That is, the model implied mean vector and the model implied covariance matrix are functions of θ .

The -2*log-likelihood of the *i*th case is:

$$-2*logL_i(\theta; \mathbf{y}_i)_{\mathrm{ML}} = p_i*log(2\pi) + log|\mathbf{\Sigma}_i(\theta)| + (\mathbf{y}_i - \mu_i(\theta))^{\mathrm{T}} \mathbf{\Sigma}_i(\theta)^{-1} (\mathbf{y}_i - \mu_i(\theta)), \quad (1)$$

where p_i is the number of filtered variables with complete data in the *i*th case, $\mu_i(\theta)$ and $\Sigma_i(\theta)$ are the model implied mean vector and the model implied covariance matrix for the *i*th case, respectively. Since there is a subscript *i* in these quantities, the model implied mean vector and covariance matrix may vary across cases. In order words, this model handles incomplete data automatically by selecting only the complete data in the log-likelihood function.

To obtain the parameter estimates, we may take the sum of the -2*log-likelihood over all cases and minimize it. This is known as the full information maximum likelihood (FIML or simply ML) estimation method. Iterative methods are used to obtain the parameter estimates. When it is convergent, the asymptotic covariance matrix (thus the standard errors) of the parameter estimates may be obtained from the inverse of the Hessian matrix. The parameter estimates divided by their standard errors (SEs) follow a z distribution under the null hypothesis. Likelihood ratio statistic may also be used to compare nested models.

2.2. Univariate fixed-effects model

Let us assume that there is only one effect size y_i in the *i*th study. y_i can be any effect size, such as odds ratio, raw mean difference, standardized mean difference, correlation coefficient or its Fisher's z transformed score. When the sample sizes are reasonably large, y_i can be assumed to be normally distributed with a variance of v_i (see e.g., Borenstein $et^{\tilde{a}l}$. 2009, for the formulas of common effect sizes). The univariate fixed-effects model for the *i*th study is:

$$y_i = \beta_{\text{fixed}} + e_i, \tag{2}$$

where β_{fixed} is the common effect under a fixed-effects model and $\text{Var}(e_i) = v_i$ is the known sampling variance.

To fit the univariate fixed-effects meta-analysis in SEM, we may use the following model:

$$\mu_i(\theta) = \beta_{\text{fixed}} \tag{3}$$

and

$$\Sigma_i(\theta) = v_i \tag{4}$$

Since v_i is known, the only parameter in the model is β_{fixed} .

2.3. Univariate random-effects model

A random-effects model allows studies to have their own study specific effect. The model for the ith study is:

$$y_i = \beta_{\text{random}} + u_i + e_i, \tag{5}$$

where β_{random} is the average effect under a random-effects model and $\text{Var}(u_i) = \tau^2$ is the heterogeneity variance that has to be estimated. To fit the univariate random-effects meta-analysis in SEM, we may use the following model:

$$\mu_i(\theta) = \beta_{\text{random}} \tag{6}$$

and

$$\Sigma_i(\theta) = \tau^2 + v_i. \tag{7}$$

 v_i and $\tau^2 + v_i$ are known as the conditional and the unconditional variances in the literature of meta-analysis, respectively. In this model we have to estimate both $\beta_{\rm random}$ and τ^2 .

Quantifying heterogeneity To test the homogeneity of the effect sizes, we may compute a Q statistic (Cochran 1954):

$$Q = \sum_{i=1}^{k} w_i (y_i - \hat{\beta}_{\text{fixed}})^2, \tag{8}$$

where $w_i = 1/v_i$. Under the null hypothesis of homogeneity of effect sizes, the Q statistic has an approximate chi-square distribution with (k-1) degrees of freedom.

Although the Q statistic may be used to test the null hypothesis of homogeneity of effect sizes, it does not indicate the degree of heterogeneity. One popular index to quantifying the heterogeneity of effect sizes is the I^2 proposed by Higgins and Thompson (2002). The general formula is:

$$I^2 = \frac{\hat{\tau}^2}{\hat{\tau}^2 + \tilde{v}} \tag{9}$$

where \tilde{v} is a typical within-study sampling variance.

Takkouche, Cadarso-Suárez, and Spiegelman (1999) suggested to use the harmonic mean of v_i as the typical within-study sampling variance, i.e.,

$$\tilde{v}_{\text{HM}} = \frac{1}{\sum_{i=1}^{k} v_i}.\tag{10}$$

Higgins and Thompson (2002) preferred to define the *typical* within-study sampling variance based on the Q statistic:

$$\tilde{v}_{Q} = \frac{(k-1)\sum_{i=1}^{k} v_{i}}{(\sum_{i=1}^{k} v_{i})^{2} - \sum_{i=1}^{k} v_{i}^{2}}.$$
(11)

One reason for using this typical within-study sampling variance is that the I^2 can be simplified to

$$I_{\mathbf{Q}}^{2} = Q - (k - 1)/Q. \tag{12}$$

Besides these two estimators, Xiong, Miller, and Morris (2010) also discussed an estimator on I^2 that is based on the arithmetic mean:

$$\tilde{v}_{\rm AM} = \sum_{i=1}^{k} v_i / k. \tag{13}$$

All of these estimators are available in the **metaSEM** package.

2.4. Univariate mixed-effects model

The mixed-effects meta-analysis extends the random-effects meta-analysis by using study characteristics as predictors. Assuming that \mathbf{x}_i is a $(m+1) \times 1$ vector of predictors including a constant of 1 where m is the number predictors in the ith study, the mixed-effects model is:

$$y_i = \beta' \mathbf{x}_i + u_i + e_i, \tag{14}$$

where β is a vector of regression coefficients including the intercept. To fit the univariate mixed-effects meta-analysis in SEM, we may use the following model for the conditional mean and variance:

$$\mu_i(\theta|\mathbf{x}_i) = \beta'\mathbf{x}_i \tag{15}$$

and

$$\Sigma_i(\theta|\mathbf{x}_i) = \tau^2 + v_i. \tag{16}$$

Since \mathbf{x}_i is specified via definition variables (see e.g., Cheung 2010), the means and covariance matrix of \mathbf{x} are not estimated. \mathbf{x} is treated as a design matrix rather than random variables. This approach is consistent with conventional meta-analysis.

Explained variance The percentage of explained variance by the inclusion of predictors can be calculated by comparing the $\hat{\tau}_0^2$ without predictor and the $\hat{\tau}_1^2$ with predictors (Raudenbush 2009):

$$R^2 = \frac{\hat{\tau}_0^2 - \hat{\tau}_1^2}{\hat{\tau}_0^2}. (17)$$

By definition, \mathbb{R}^2 is non-negative. When the calculated \mathbb{R}^2 is negative, it is truncated to zero.

2.5. Multivariate mixed-effects model

Let us assume that there are p effect sizes with m predictors in k studies. The model for the multivariate effect sizes in the ith study is:

$$\mathbf{y}_i = \mathbf{B}\mathbf{x}_i + \mathbf{u}_i + \mathbf{e}_i,\tag{18}$$

where \mathbf{y}_i is a $p \times 1$ effect sizes, \mathbf{B} is a $p \times (m+1)$ regression coefficients including the intercepts, \mathbf{x}_i is a $(m+1) \times 1$ predictors including 1 in the first column, \mathbf{u}_i is a $p \times 1$ study

specific random effects, and \mathbf{e}_i is a $p \times 1$ sampling error. We assume that $\mathsf{Var}(\mathbf{e}_i) = \mathbf{V}_i$ is known and given in the *i*th study and $\mathsf{Var}(\mathbf{u}_i) = \mathbf{T}^2$ is the variance component of the between-study heterogeneity that has to be estimated.

The -2*log-likelihood of the above model is:

$$-2*logL_i(\mathbf{B}, \mathbf{T}^2; \mathbf{y}_i)_{\mathrm{ML}} = p_i*log(2\pi) + log|\mathbf{T}^2 + \mathbf{V}_i| + (\mathbf{y}_i - \mathbf{B}\mathbf{x}_i)^{\mathrm{T}}(\mathbf{T}^2 + \mathbf{V}_i)^{-1}(\mathbf{y}_i - \mathbf{B}\mathbf{x}_i),$$
(19)

where p_i is the number of complete effect sizes in the *i*th study.

In applied research, different studies may report different numbers of effect sizes, that is, p_i may vary across studies. The above -2*log-likelihood may handle missing effect sizes by selecting the complete effect sizes only in the above equation. It is expected that there is no missing data in \mathbf{x}_i . When there are missing data in \mathbf{x}_i , the whole study will be deleted before the analysis.

The I^2 and R^2 in univariate meta-analysis are also calculated for each effect size in multivariate meta-analysis.

2.6. Restricted Maximum Likelihood (REML) Estimation Method

Since both the fixed- and the random-effects are estimated simultaneously, it is well-known that $\hat{\mathbf{T}}_{\mathrm{ML}}^2$ based on the ML estimation in Eq. 19 is negatively biased. It is because it does not take the uncertainty in estimating $\hat{\mathbf{B}}_{\mathrm{ML}}$ into account. If the unbiasedness of the variance component is crucial in the research questions, we may obtain the variance component $\hat{\mathbf{T}}_{\mathrm{REML}}^2$ based on the REML estimation method (Cheung 2011a; Harville 1977; Patterson and Thompson 1971).

The -2*log-likelihood of the model is:

$$-2*logL_i(\mathbf{T}^2; \mathbf{y}_i)_{\text{REML}} = p_i*log(2\pi) + log|\mathbf{T}^2 + \mathbf{V}_i| + (\mathbf{y}_i - \alpha \mathbf{X}_i)^{\text{T}} (\mathbf{T}^2 + \mathbf{V}_i)^{-1} (\mathbf{y}_i - \alpha \mathbf{X}_i) + |\mathbf{X}_i^{\text{T}} \mathbf{V}_i^{-1} \mathbf{X}_i|,$$
(20)

where $\alpha = (\mathbf{X}^{\mathrm{T}}\mathbf{V}^{-1}\mathbf{X})^{-1}\mathbf{X}^{\mathrm{T}}\mathbf{V}^{-1}\mathbf{y}$, and \mathbf{X} , \mathbf{V} and \mathbf{y} are the stacked matrices and vector of the correspondent matrices and vectors.

Since the fixed effects ${\bf B}$ is not involved in the above -2*log-likelihood function, it has to be calculated later.

2.7. Three-level meta-analysis

Observed effect sizes may be related or dependent. For example, effect sizes reported by the same research team may be more similar when comparing to effect sizes reported by other research teams. Effect sizes reported by studies from the same country may be more similar when comparing to studies across countries. If the degree of dependence is known, multivariate meta-analysis introduced before may be applied. When the degree of dependence is unknown, a three-level meta-analytic model may be used (e.g., Konstantopoulos 2011). The model is:

$$y_{ij} = \beta' \mathbf{x}_{ij} + u_{(2)ij} + u_{(3)j} + e_{ij}, \tag{21}$$

where y_{ij} is the effect size for the *i*th effect size in the *j*th cluster, β is a vector of regression coefficients including the intercept, \mathbf{x}_{ij} is the predictors including 1 in the first element for the *i*th study at the *j*th cluster, $u_{(2)ij}$ and $u_{(3)j}$ are the random-effects at level-2 and level-3, respectively, and e_{ij} is the known sampling variance of the effect size.

To fit the three-level meta-analytic model in SEM, we may use the following model for the conditional mean and variance:

$$\mu_{ij}(\theta|\mathbf{x}_{ij}) = \beta' \mathbf{x}_{ij} \tag{22}$$

and

$$\Sigma_{ij}(\theta|\mathbf{x}_{ij}) = \tau_{(2)}^2 + \tau_{(3)}^2 + v_{ij}, \tag{23}$$

where $\mathsf{Var}(u_{(2)ij}) = \tau_{(2)}^2$ and $\mathsf{Var}(u_{(3)j}) = \tau_{(3)}^2$ are the heterogeneity at level-2 and level-3, respectively. Similar to those listed in Eqs. 20, $\tau_{(2)}^2$ and $\tau_{(3)}^2$ may also be estimated with REML estimation method. This approach is implemented in rem13().

Quantifying heterogeneity and explained variance Similar to the I^2 defined in Eq. 9 for random-effects meta-analysis, we may define the degree of heterogeneity for three-level meta-analysis in level 2 and level 3 as:

$$I_{(2)}^2 = \frac{\hat{\tau}_{(2)}^2}{\hat{\tau}_{(2)}^2 + \hat{\tau}_{(3)}^2 + \tilde{v}}$$
 (24)

and

$$I_{(3)}^2 = \frac{\hat{\tau}_{(3)}^2}{\hat{\tau}_{(2)}^2 + \hat{\tau}_{(3)}^2 + \tilde{v}}$$
 (25)

where \tilde{v} is the *typical* within-study sampling variance defined in Eqs. 10, 11 and 13. When there are predictors, we may calculate the R^2 for the level 2 and level 3 similar to that defined in Eq. 17:

$$R_{(2)}^2 = \frac{\hat{\tau}_{0(2)}^2 - \hat{\tau}_{1(2)}^2}{\hat{\tau}_{0(2)}^2}.$$
 (26)

and

$$R_{(3)}^2 = \frac{\hat{\tau}_{0(3)}^2 - \hat{\tau}_{1(3)}^2}{\hat{\tau}_{0(3)}^2}.$$
 (27)

When the estimates are negative, they are truncated to zero.

2.8. Examples

Three example data sets are used to demonstrate the procedures of fitting univariate and multivariate meta-analyses. The first data set was taken from Becker (1983) who reported 10 studies on sex differences in conformity using the fictitious norm group

paradigm. di and vi are the standardized mean difference and its sampling variance, respectively. Becker hypothesized that the logarithm of the number of items (items) predicted the effect size.

The second data set is adapted from Berkey, Hoaglin, Antczak-Bouckoms, Mosteller, and Colditz (1998). They summarized five published trials comparing surgical and non-surgical treatments for medium-severity periodontal disease, one year after treatment. Publication year *pub_year* was hypothesized as a predictor.

The third data set was reported by Konstantopoulos (2011) that was based on Cooper, Valentine, Charlton, and Melson (2003). It described fifty-six effect sizes clustered in 11 districts. Year of publication was used as a predictor.

Univariate random-effects model The function meta() is used to conduct the analyses. The arguments y and v are used to specify the effect sizes and its sampling variances (and covariances for multivariate meta-analysis), respectively. By default, a random-effects meta-analysis is fitted. After running the analysis, summary() is used to extract the results. The estimated fixed- and random-effects are represented by the Intercept1 to Interceptp and by the Tau2_1_1 to Tau2_p_p parameters where p is the number of effect sizes, especially. coef() and vcov() may be used to extract the parameter estimates and their asymptotic sampling covariance matrix, respectively.

Before interpreting the results, it is necessary to check whether the optimization is successful. The OpenMx status1 returns the status from the optimizer. The optimization can be considered as fine if the code is either 0 or 1. Users may refer to OpenMx's Common Errors (and how to avoid them) for more details.

From the following analyses, the Q statistic (df = 9) is 30.65, p < .001. The estimated heterogeneity variance is 0.0774 while the I^2 based on the Q statistic is .6718. The average effect size with its 95% Wald confidence interval (CI) based on the random-effects model is 0.1747 (-0.0475, 0.3970).

```
R> ## Load the library
R> library(metaSEM)
R> ## Show the first few studies of the data set
R> head(Becker83)
                 vi percentage items
  study
           di
1
      1 -0.33 0.03
                             25
                                    2
2
         0.07 0.03
                             25
                                    2
3
      3 -0.30 0.02
                             50
                                    2
                                   38
4
         0.35 0.02
                            100
5
      5
         0.69 0.07
                            100
                                   30
         0.81 0.22
                            100
                                   45
R> ## Random-effects meta-analysis with ML
```

R> summary(random1 <- meta(y=di, v=vi, data=Becker83))

Running Meta analysis with ML

```
Call:
meta(y = di, v = vi, data = Becker83)
95% confidence intervals: z statistic approximation
Coefficients:
           Estimate Std.Error
                                  lbound
                                            ubound z value
Intercept1 0.174734 0.113378 -0.047482 0.396950 1.5412
           0.077376  0.054108  -0.028674  0.183426  1.4300
Tau2_1_1
           Pr(>|z|)
             0.1233
Intercept1
Tau2_1_1
             0.1527
Q statistic on homogeneity of effect sizes: 30.64949
Degrees of freedom of the Q statistic: 9
P value of the Q statistic: 0.0003399239
Heterogeneity indices (based on the estimated Tau2):
                             Estimate
Intercept1: I2 (Q statistic)
                               0.6718
Number of studies (or clusters): 10
Number of observed statistics: 10
Number of estimated parameters: 2
Degrees of freedom: 8
-2 log likelihood: 7.928307
OpenMx status1: 0 ("0" and "1": considered fine; other values indicate problems)
```

Univariate mixed-effects model We may conduct a mixed-effects meta-analysis by including predictors. The argument \mathbf{x} is used to specify the predictors. If there are more than one predictor, $\mathtt{cbind}()$ is used to specify them. The estimated regression coefficients are represented by the $\mathtt{Slope}i_j$ parameter where i and j represent the ith effect size and jth predictor. The following analysis indicates that log(items) is a significant predictor with the estimated regression coefficient and its 95% CI of 0.2109 (0.1225, 0.2992) with $R^2=1$.

```
R> ## Mixed-effects meta-analysis with "log(items)" as the predictor R> summary( meta(y=di, v=vi, x=log(items), data=Becker83) )
```

Running Meta analysis with ML

```
Call:
```

```
meta(y = di, v = vi, x = log(items), data = Becker 83)
```

```
95% confidence intervals: z statistic approximation
Coefficients:
              Estimate
                       Std.Error
                                       lbound
                                                   ubound
Intercept1 -3.2015e-01 1.0981e-01 -5.3539e-01 -1.0492e-01
            2.1088e-01 4.5084e-02 1.2251e-01 2.9924e-01
Slope1_1
            1.0000e-10 2.0095e-02 -3.9386e-02 3.9386e-02
Tau2_1_1
           z value Pr(>|z|)
Intercept1 -2.9154 0.003552 **
Slope1_1
           4.6774 2.905e-06 ***
Tau2_1_1
           0.0000 1.000000
Signif. codes: 0 '*** 0.001 '** 0.01 '* 0.05 '.' 0.1 ' 1
Q statistic on homogeneity of effect sizes: 30.64949
Degrees of freedom of the Q statistic: 9
P value of the Q statistic: 0.0003399239
Explained variances (R2):
                           у1
Tau2 (no predictor)
                       0.0774
Tau2 (with predictors) 0.0000
                       1.0000
Number of studies (or clusters): 10
Number of observed statistics: 10
Number of estimated parameters: 3
Degrees of freedom: 7
-2 log likelihood: -4.208024
OpenMx status1: 0 ("0" and "1": considered fine; other values indicate problems)
```

Univariate fixed-effects model Mathematically, fixed-effects meta-analysis is a special case of the random-effects meta-analysis by fixing the heterogeneity variance of the random-effects at 0. The argument RE.constraints, which expects a matrix as input, is used to constrain the variance component of the random effects. For example, the estimated common effect and its 95% Wald CI is 0.1006 (-0.0180, 0.2192) under a fixed-effects model.

Running Meta analysis with ML

```
Call:
meta(y = di, v = vi, data = Becker83, RE.constraints = matrix(0,
   ncol = 1, nrow = 1)
95% confidence intervals: z statistic approximation
Coefficients:
           Estimate Std.Error
                                  lbound
                                            ubound z value
Intercept1 0.100640 0.060510 -0.017957 0.219237 1.6632
          Pr(>|z|)
Intercept1 0.09627.
Signif. codes: 0 '*** 0.001 '** 0.01 '* 0.05 '.' 0.1 ' 1
Q statistic on homogeneity of effect sizes: 30.64949
Degrees of freedom of the Q statistic: 9
P value of the Q statistic: 0.0003399239
Heterogeneity indices (based on the estimated Tau2):
                             Estimate
Intercept1: I2 (Q statistic)
Number of studies (or clusters): 10
Number of observed statistics: 10
Number of estimated parameters: 1
Degrees of freedom: 9
-2 log likelihood: 17.86043
OpenMx status1: 0 ("0" and "1": considered fine; other values indicate problems)
```

Multivariate random-effects model Multivariate meta-analysis can be fitted by specifying the multivariate effect sizes and its sampling covariance matrix in the arguments y and v with cbind(), respectively. Only the lower triangle of the sampling covariance matrix arranged by column major is used in v. For example, if V =

```
 \begin{bmatrix} V_{11} \\ V_{21} & V_{22} \\ V_{31} & V_{32} & V_{33} \end{bmatrix}, \text{ we may use v=cbind(V11,V21,V31,V22,V32,V33) in meta()}.
```

The Q statistic (df=8) of the following example is 128.2267, p<.001. The estimated variance component is $\begin{bmatrix} 0.0070 \\ 0.0095 & 0.02614 \end{bmatrix}$. The I^2 based on the Q statistic for PD and AL are .6021 and .9250, respectively. The pooled effect sizes with their 95% Wald CIs based on the random-effects model for PD and AL are 0.3448 (0.2397, 0.4500) and -0.3379 (-0.4972, -0.1787), respectively.

```
R> ## Show the data set R> Berkey98
```

```
trial pub_year no_of_patients
                                PD
                                      AL var_PD cov_PD_AL
                            14 0.47 -0.32 0.0075
           1983
                                                   0.0030
1
     1
2
     2
           1982
                            15 0.20 -0.60 0.0057
                                                   0.0009
3
     3
                           78 0.40 -0.12 0.0021
           1979
                                                   0.0007
4
     4
           1987
                           89 0.26 -0.31 0.0029
                                                   0.0009
5
     5
           1988
                           16 0.56 -0.39 0.0148
                                                   0.0072
 var_AL
1 0.0077
2 0.0008
3 0.0014
4 0.0015
5 0.0304
R> ## Multivariate meta-analysis with a random-effects model
R> summary( meta(y=cbind(PD, AL), v=cbind(var_PD, cov_PD_AL, var_AL),
               data=Berkey98) )
Running Meta analysis with ML
Call:
meta(y = cbind(PD, AL), v = cbind(var_PD, cov_PD_AL, var_AL),
   data = Berkey98)
95% confidence intervals: z statistic approximation
Coefficients:
            Estimate Std.Error
                                    lbound
Intercept1 0.3448392 0.0536312 0.2397239 0.4499544
Intercept2 -0.3379381 0.0812479 -0.4971812 -0.1786951
Tau2_1_1
           0.0070020 0.0090497 -0.0107351 0.0247391
Tau2_2_1
           0.0094607 \quad 0.0099698 \quad -0.0100797 \quad 0.0290010
Tau2_2_2
           z value Pr(>|z|)
Intercept1 6.4298 1.278e-10 ***
Intercept2 -4.1593 3.192e-05 ***
Tau2_1_1
          0.7737
                   0.4391
Tau2_2_1
           0.9489
                     0.3427
Tau2_2_2 1.4737
                     0.1406
Signif. codes: 0 '***, 0.001 '**, 0.01 '*, 0.05 '., 0.1 ', 1
Q statistic on homogeneity of effect sizes: 128.2267
Degrees of freedom of the Q statistic: 8
P value of the Q statistic: 0
Heterogeneity indices (based on the estimated Tau2):
```

```
Estimate
```

```
Intercept1: I2 (Q statistic)    0.6021
Intercept2: I2 (Q statistic)    0.9250
```

Number of studies (or clusters): 5 Number of observed statistics: 10 Number of estimated parameters: 5

Degrees of freedom: 5

-2 log likelihood: -11.68131

OpenMx status1: 0 ("0" and "1": considered fine; other values indicate problems)

Multivariate mixed-effects model As an illustration, we use pub_year as a predictor. To make the intercept more interpretable, we center the publication year at 1979, the first record of publication year in the data set. The estimated regression coefficients and their 95% CIs on PD and AL are 0.0064 (-0.2050, 0.2177) and -0.0706 (-0.3883, 0.2471), respectively. The R^2 in predicting PD and AL are .0000 and .0433, respectively. When there are multiple parameters, it is preferable to test all parameters simultaneously, rather than individually. The likelihood ratio statistic on testing both regression coefficient is $\chi^2(df=2)=0.3273, p=.8490$. Thus, the null hypothesis of both regression coefficients are zero is not rejected.

Sometimes, we may want to test the equality of the regression coefficients and see if they are different. We may impose the equality constraint on the regression coefficients by using the same label in the argument coef.constraints. The average regression coefficient is 0.0017 (-0.1991, 0.2025). The likelihood ratio statistic on testing the equality of the regression coefficients is $\chi^2(df=1)=0.3270, p=.5674$. There is no evidence that one regression coefficient is stronger than the other.

Running No constraint

R> summary(mult2)

```
Call:
```

```
meta(y = cbind(PD, AL), v = cbind(var_PD, cov_PD_AL, var_AL),
    x = scale(pub_year, center = 1979), data = Berkey98, model.name = "No constraint")
```

95% confidence intervals: z statistic approximation Coefficients:

```
Estimate Std.Error lbound ubound Intercept1 0.3440001 0.0857659 0.1759021 0.5120982
```

```
Slope1_1
           0.0063540 0.1078235 -0.2049762 0.2176842
Intercept2 -0.2918175 0.1312796 -0.5491208 -0.0345141
Slope2_1 -0.0705888 0.1620965 -0.3882921 0.2471146
Tau2_1_1
          0.0080405 0.0101206 -0.0117955 0.0278766
Tau2_2_1
           Tau2_2_2
          z value Pr(>|z|)
Intercept1 4.0109 6.048e-05 ***
Slope1_1
           0.0589
                  0.95301
Intercept2 -2.2229
                   0.02622 *
Slope2_1 -0.4355 0.66322
Tau2_1_1
          0.7945
                  0.42692
Tau2_2_1
         0.8853
                   0.37599
Tau2_2_2 1.4646 0.14303
---
Signif. codes: 0 '*** 0.001 '** 0.01 '* 0.05 '.' 0.1 ' ' 1
Q statistic on homogeneity of effect sizes: 128.2267
Degrees of freedom of the Q statistic: 8
P value of the Q statistic: 0
Explained variances (R2):
                            у1
                                  у2
Tau2 (no predictor)
                     0.0070020 0.0261
Tau2 (with predictors) 0.0080405 0.0250
R2
                     0.0000000 0.0433
Number of studies (or clusters): 5
Number of observed statistics: 10
Number of estimated parameters: 7
Degrees of freedom: 3
-2 log likelihood: -12.00859
OpenMx status1: 0 ("0" and "1": considered fine; other values indicate problems)
R> ## Multivariate meta-analysis with both regression coefficients fixed at 0
R> mult0 <- meta(y=cbind(PD, AL), v=cbind(var_PD, cov_PD_AL, var_AL), data=Berkey98,
               x=scale(pub_year, center=1979),
               model.name="Both regression coefficients fixed at 0",
               coef.constraints=matrix(c("0", "0"), nrow=2))
Running Both regression coefficients fixed at 0
R> summary(mult0)
Call:
```

meta(y = cbind(PD, AL), v = cbind(var_PD, cov_PD_AL, var_AL),

```
x = scale(pub_year, center = 1979), data = Berkey98, coef.constraints = matrix(c("0",
        "0"), nrow = 2), model.name = "Both regression coefficients fixed at 0")
95% confidence intervals: z statistic approximation
Coefficients:
            Estimate Std.Error
                                    lbound
Intercept1 0.3448392 0.0536312 0.2397239 0.4499544
Intercept2 -0.3379381 0.0812479 -0.4971812 -0.1786951
Tau2_1_1
          0.0070020 0.0090497 -0.0107351 0.0247391
Tau2_2_1
           0.0094607 0.0099698 -0.0100797 0.0290010
Tau2_2_2 0.0261445 0.0177409 -0.0086270 0.0609161
          z value Pr(>|z|)
Intercept1 6.4298 1.278e-10 ***
Intercept2 -4.1593 3.192e-05 ***
Tau2_1_1
         0.7737
                     0.4391
Tau2_2_1
           0.9489
                     0.3427
Tau2_2_2
           1.4737
                     0.1406
Signif. codes: 0 '*** 0.001 '** 0.01 '* 0.05 '.' 0.1 ' 1
Q statistic on homogeneity of effect sizes: 128.2267
Degrees of freedom of the Q statistic: 8
P value of the Q statistic: 0
Explained variances (R2):
                            у1
                                   y2
Tau2 (no predictor)
                      0.007002 0.0261
Tau2 (with predictors) 0.007002 0.0261
R2
                      0.000000 0.0000
Number of studies (or clusters): 5
Number of observed statistics: 10
Number of estimated parameters: 5
Degrees of freedom: 5
-2 log likelihood: -11.68131
OpenMx status1: 0 ("0" and "1": considered fine; other values indicate problems)
R> ## Likelihood ratio test on testing both regression coefficients are 0
R> anova(mult2, mult0)
          base
                                            comparison ep
1 No constraint
                                                  <NA>
2 No constraint Both regression coefficients fixed at 0 5
  minus2LL df
                    AIC
                         diffLL diffdf
                                                 р
```

```
1 -12.00859 3 -18.00859
                              NA
                                     NA
                                              NA
2 -11.68131 5 -21.68131 0.3272789
                                      2 0.8490481
R> ## Multivariate meta-analysis with an equality constraint on the slopes
R> mult3 <- meta(y=cbind(PD, AL), v=cbind(var_PD, cov_PD_AL, var_AL), data=Berkey98,
              x=scale(pub_year, center=1979), model.name="With equality constraint",
              coef.constraints=matrix(c("0.3*Equal_Slope", "0.3*Equal_Slope"), nrow=.
Running With equality constraint
R> summary(mult3)
Call:
meta(y = cbind(PD, AL), v = cbind(var_PD, cov_PD_AL, var_AL),
   x = scale(pub_year, center = 1979), data = Berkey98, coef.constraints = matrix(c(
       "0.3*Equal_Slope"), nrow = 2), model.name = "With equality constraint")
95% confidence intervals: z statistic approximation
Coefficients:
             Estimate Std.Error
                                    lbound
            0.3437612 \quad 0.0849829 \quad 0.1771979 \quad 0.5103245
Intercept1
Equal_Slope 0.0016748 0.1024443 -0.1991123 0.2024619
Intercept2 -0.3390010 0.1041005 -0.5430344 -0.1349677
           0.0070474 0.0094638 -0.0115013 0.0255962
Tau2_1_1
Tau2_2_1
            Tau2_2_2
           z value Pr(>|z|)
           4.0451 5.231e-05 ***
Intercept1
Equal_Slope 0.0163 0.986956
Intercept2 -3.2565 0.001128 **
Tau2_1_1
           0.7447 0.456471
Tau2_2_1
            0.9006 0.367800
Tau2_2_2
           1.4492 0.147278
Signif. codes: 0 '*** 0.001 '** 0.01 '* 0.05 '.' 0.1 ' 1
Q statistic on homogeneity of effect sizes: 128.2267
Degrees of freedom of the Q statistic: 8
P value of the Q statistic: 0
Explained variances (R2):
                            у1
                                   у2
Tau2 (no predictor)
                     0.0070020 0.0261
```

Tau2 (with predictors) 0.0070474 0.0262

```
R2 0.0000000 0.0000

Number of studies (or clusters): 5
```

Number of observed statistics: 10 Number of estimated parameters: 6

Degrees of freedom: 4

-2 log likelihood: -11.68158

OpenMx status1: 0 ("0" and "1": considered fine; other values indicate problems)

R> ## Likelihood ratio test on the equality of regression coefficients R> anova(mult2, mult3)

Multivariate fixed-effects model A multivariate fixed-effects meta-analysis can be conducted by fixing the variance component at a zero matrix. The following code illustrates the syntax.

Running Meta analysis with ML

```
Call:
```

95% confidence intervals: z statistic approximation Coefficients:

```
Estimate Std.Error lbound ubound z value
Intercept1 0.307219 0.028575 0.251212 0.363225 10.751
Intercept2 -0.394377 0.018649 -0.430929 -0.357825 -21.147
Pr(>|z|)
Intercept1 < 2.2e-16 ***
Intercept2 < 2.2e-16 ***
```

Signif. codes: 0 '*** 0.001 '** 0.01 '* 0.05 '.' 0.1 ' 1

```
Q statistic on homogeneity of effect sizes: 128.2267
Degrees of freedom of the Q statistic: 8
P value of the Q statistic: 0
Heterogeneity indices (based on the estimated Tau2):
                              Estimate
Intercept1: I2 (Q statistic)
Intercept2: I2 (Q statistic)
                                     0
Number of studies (or clusters): 5
Number of observed statistics: 10
Number of estimated parameters: 2
Degrees of freedom: 8
-2 log likelihood: 90.88326
OpenMx status1: 0 ("0" and "1": considered fine; other values indicate problems)
REML The reml() function may be used to estimate the variance component with
the REML estimation method. It should be noted that it does not estimate the fixed-
effects. The fixed-effects estimates can be obtained via the meta() function by specifying
the estimated variance component from reml() as fixed values in the RE.constraints
argument. This approach is consistent with the idea of REML that removes the fixed-
effects parameter when estimating the variance component.
R> ## Random-effects meta-analysis with REML
R> summary( VarComp <- reml(y=di, v=vi, data=Becker83) )
Running Variance component with REML
Call:
reml(y = di, v = vi, data = Becker83)
95% confidence intervals: z statistic approximation
Coefficients:
          Estimate Std.Error
                                 lbound
                                            ubound z value
Tau2_1_1 0.091445 0.064228 -0.034439 0.217329 1.4238
         Pr(>|z|)
Tau2_1_1 0.1545
Number of studies (or clusters): 10
Number of observed statistics: 9
Number of estimated parameters: 1
Degrees of freedom: 8
-2 log likelihood: -6.110579
OpenMx status: 0 ("0" and "1": considered fine; other values indicate problems)
```

```
R> ## Extract the variance component
R> VarComp_REML <- matrix( coef(VarComp), ncol=1, nrow=1 )</pre>
R> ## Meta-analysis by treating the variance component as fixed
R> summary( meta(y=di, v=vi, data=Becker83, RE.constraints=VarComp_REML) )
Running Meta analysis with ML
Call:
meta(y = di, v = vi, data = Becker83, RE.constraints = VarComp_REML)
95% confidence intervals: z statistic approximation
Coefficients:
            Estimate Std.Error
                                  lbound
                                            ubound z value
Intercept1 0.180189 0.117535 -0.050176 0.410555 1.5331
           Pr(>|z|)
Intercept1
             0.1253
Q statistic on homogeneity of effect sizes: 30.64949
Degrees of freedom of the Q statistic: 9
P value of the Q statistic: 0.0003399239
Heterogeneity indices (based on the estimated Tau2):
                             Estimate
Intercept1: I2 (Q statistic)
                               0.7075
Number of studies (or clusters): 10
Number of observed statistics: 10
Number of estimated parameters: 1
Degrees of freedom: 9
-2 log likelihood: 7.986161
OpenMx status1: 1 ("0" and "1": considered fine; other values indicate problems)
R> ## Estimate variance components with REML
R> summary( reml(y=di, v=vi, x=log(items), data=Becker83) )
Running Variance component with REML
Call:
reml(y = di, v = vi, x = log(items), data = Becker 83)
95% confidence intervals: z statistic approximation
Coefficients:
           Estimate Std.Error
                                   lbound
                                              ubound
Tau2_1_1 0.0052656 0.0212014 -0.0362884 0.0468196
```

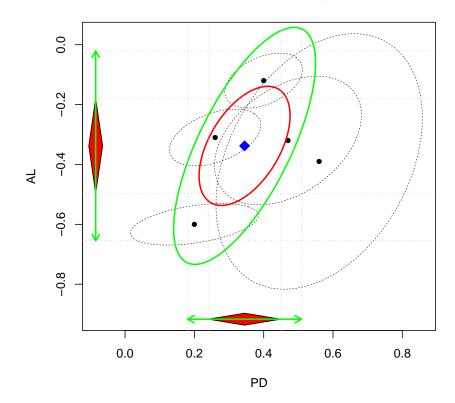
```
z value Pr(>|z|)
Tau2_1_1 0.2484
                   0.8039
Number of studies (or clusters): 10
Number of observed statistics: 8
Number of estimated parameters: 1
Degrees of freedom: 7
-2 log likelihood: -10.84567
OpenMx status: 0 ("0" and "1": considered fine; other values indicate problems)
R> ## Estimate variance components with REML
R> summary( reml(y=cbind(PD, AL), v=cbind(var_PD, cov_PD_AL, var_AL), data=Berkey98)
Running Variance component with REML
Call:
reml(y = cbind(PD, AL), v = cbind(var_PD, cov_PD_AL, var_AL),
    data = Berkey98)
95% confidence intervals: z statistic approximation
Coefficients:
          Estimate Std.Error
                                lbound
                                          ubound z value
Tau2_1_1 0.011733 0.013645 -0.015011 0.038477 0.8599
Tau2_2_1 0.011916 0.014416 -0.016340 0.040172 0.8266
Tau2_2_2 0.032651 0.024402 -0.015176 0.080479 1.3380
        Pr(>|z|)
Tau2_1_1 0.3899
Tau2_2_1 0.4085
Tau2_2_2
         0.1809
Number of studies (or clusters): 5
Number of observed statistics: 8
Number of estimated parameters: 3
Degrees of freedom: 5
-2 log likelihood: -18.86768
OpenMx status: 0 ("0" and "1": considered fine; other values indicate problems)
```

Plots of multivariate effect sizes If multivariate meta-analysis is conducted, pairwise plots on the pooled effect sizes and their confidence ellipses can be obtained via the plot() function. By default, 95% confidence intervals on the average effect sizes and confidence ellipses on the random effects are plotted. For example,

Running Meta analysis with ML

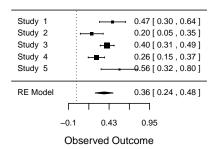
R> plot(Berkey98.fit, main="Multivariate meta-analysis", axis.label=c("PD", "AL"))

Multivariate meta-analysis



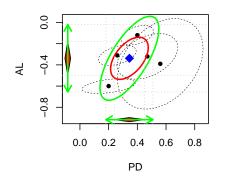
By combining with the forest plots provided by the **metafor** package, we may combine the univariate and multivariate natures of the effect sizes in a single figure. This will be very useful in displaying results of multivariate meta-analysis.

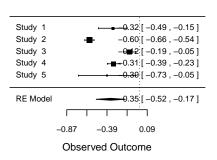
Forest plot for PD



Multivariate meta-analysis

Forest plot for AL





Three-level meta-analysis The meta3() function may be used to fit three-level meta-analytic models. It is assumed that effect sizes within cluster are dependent. The Q statistic (df=55) of the following example is 578.86, p<.001. The I^2 at level-2 and level-3 are .3440 and .6043, respectively. The estimated coefficient of Year of publication is 0.0051, p=.5518. The R^2 at level-2 and level-3 are .0000 and .0221, respectively.

```
R> ## ML estimation method
R> ## No predictor
R> summary( meta3(y=y, v=v, cluster=District, data=Cooper03) )
```

Running Meta analysis with ML

```
Call: meta3(y = y, v = v, cluster = District, data = Cooper03)
```

95% confidence intervals: z statistic approximation

```
Coefficients:
           Estimate Std.Error
                                 lbound
                                           ubound
Intercept 0.1844554 0.0805411 0.0265977 0.3423131
Tau2_2
         0.0328648 0.0111397 0.0110314 0.0546982
          0.0577384 0.0307423 -0.0025154 0.1179921
Tau2_3
         z value Pr(>|z|)
Intercept 2.2902 0.022010 *
Tau2_2
         2.9502 0.003175 **
Tau2_3 1.8781 0.060362 .
Signif. codes: 0 '*** 0.001 '** 0.01 '* 0.05 '.' 0.1 ' ' 1
Q statistic on homogeneity of effect sizes: 578.864
Degrees of freedom of the Q statistic: 55
P value of the Q statistic: 0
Heterogeneity indices (based on the estimated Tau2):
                 Estimate
I2_2 (Q statistic)
                   0.3440
I2_3 (Q statistic)
                   0.6043
Number of studies (or clusters): 11
Number of observed statistics: 56
Number of estimated parameters: 3
Degrees of freedom: 53
-2 log likelihood: 16.78987
OpenMx status1: 0 ("0" and "1": considered fine; other values indicate problems)
R> ## Year as a predictor
R> summary( meta3(y=y, v=v, cluster=District, x=scale(Year, scale=FALSE), data=CooperO3) )
Running Meta analysis with ML
Call:
meta3(y = y, v = v, cluster = District, x = scale(Year, scale = FALSE),
   data = Cooper03)
95% confidence intervals: z statistic approximation
Coefficients:
           Estimate Std.Error
                                 lbound
                                           ubound
          Slope_1
Intercept 0.1780268 0.0805219 0.0202067 0.3358469
Tau2_2
         0.0329390 0.0111620 0.0110618 0.0548162
Tau2_3
```

z value Pr(>|z|)

```
Slope_1
          0.5950 0.551814
Intercept 2.2109 0.027042 *
Tau2_2
          2.9510 0.003168 **
Tau2_3
         1.8800 0.060104 .
___
Signif. codes: 0 '*** 0.001 '** 0.01 '* 0.05 '.' 0.1 ' ' 1
Q statistic on homogeneity of effect sizes: 578.864
Degrees of freedom of the Q statistic: 55
P value of the Q statistic: 0
Explained variances (R2):
                       Level 2 Level 3
Tau2 (no predictor)
                       0.032865 0.0577
Tau2 (with predictors) 0.032939 0.0565
R2
                       0.000000 0.0221
Number of studies (or clusters): 11
Number of observed statistics: 56
Number of estimated parameters: 4
Degrees of freedom: 52
-2 log likelihood: 16.43629
OpenMx status1: 0 ("0" and "1": considered fine; other values indicate problems)
R> ## REML estimation method
R> ## No predictor; with LBCI
R> summary( reml3(y=y, v=v, cluster=District, data=Cooper03, intervals.type="LB") )
Running Variance component with REML
Call:
reml3(y = y, v = v, cluster = District, data = CooperO3, intervals.type = "LB")
95% confidence intervals: Likelihood-based statistic
Coefficients:
      Estimate Std.Error
                            lbound
                                     ubound z value
Tau2_2 0.032737
                      NA 0.016264 0.062842
                                                 NA
Tau2_3 0.065062
                     NA 0.022234 0.207846
                                                 NA
      Pr(>|z|)
Tau2_2
             NA
Tau2_3
             NA
Number of studies (or clusters): 56
Number of observed statistics: 55
Number of estimated parameters: 2
```

```
Degrees of freedom: 53
-2 log likelihood: -81.14044
OpenMx status: 0 ("0" and "1": considered fine; other values indicate problems)
R> ## Year as a predictor
R> summary( reml3(y=y, v=v, cluster=District, x=scale(Year, scale=FALSE), data=Cooper03) )
Running Variance component with REML
Call:
reml3(y = y, v = v, cluster = District, x = scale(Year, scale = FALSE),
   data = Cooper03)
95% confidence intervals: z statistic approximation
Coefficients:
         Estimate Std.Error
                                 lbound
                                            ubound z value
Tau2_2 0.0326502 0.0110529 0.0109870 0.0543134 2.9540
Tau2_3 0.0722656 0.0405349 -0.0071813 0.1517125 1.7828
      Pr(>|z|)
Tau2_2 0.003137 **
Tau2_3 0.074619 .
Signif. codes: 0 '*** 0.001 '** 0.01 '* 0.05 '.' 0.1 ' ' 1
Number of studies (or clusters): 56
Number of observed statistics: 54
Number of estimated parameters: 2
Degrees of freedom: 52
-2 log likelihood: -72.09405
OpenMx status: 0 ("0" and "1": considered fine; other values indicate problems)
```

3. Meta-analytic structural equation modeling

MASEM combines ideas of meta-analysis and SEM by pooling correlation (or covariance) matrices and testing structural models on the pooled correlation (or covariance) matrix. There are two stages in conducting a MASEM. In the first stage the correlation (or covariance) matrices are pooled together. In the second stage, the pooled correlation (or covariance) matrix is used to fit structural equation models.

Cheung and Chan (2005b, 2009) proposed a two-stage structural equation modeling (TSSEM) based on a fixed-effects model. The **metaSEM** package implemented this TSSEM approach. Moreover, the TSSEM approach has been extended to the random-effects model by using a multivariate meta-analysis (Cheung 2011b) in the first stage

analysis. Regardless of whether a fixed- or random-effects model is used, the tssem2() function will handle this automatically. In other words, parameter estimates, standard errors and goodness-of-fit indices in the stage 2 analysis has already taken the stage 1 model into account.

3.1. Stage 1 analysis

The objective of the stage 1 analysis is to obtain a pooled correlation (or covariance) matrix. Under the fixed-effects model, it is assumed that all population correlation (or covariance) matrices are the same while there are study specific correlation (or covariance matrices) under the random-effects model. To simplify the presentation, I will mainly focus on analysis of correlation matrices. Generalization to analysis of covariance matrices is straight forward (see Cheung and Chan 2009)

Fixed-effects model The population correlation matrix in the *i*th study can be decomposed as:

$$\Sigma_i(\theta) = \mathbf{D}_i \mathbf{P}_i \mathbf{D}_i \tag{28}$$

where $\Sigma_i(\theta)$ is the model implied covariance matrix, \mathbf{D}_i is the diagonal matrix of standard deviations, and \mathbf{P}_i is the correlation matrix. Under the assumption of homogeneity of correlation matrices, we may obtain a common correlation matrix by imposing the constraint $P = P_1 = P_2 = \ldots = P_k$ where D_i may vary across studies. When there are missing correlations, the missing data are filtered out. If we want to obtain a common covariance matrix under the assumption of homogeneity of covariance matrices, we may also add the constraint $D = D_1 = D_2 = \ldots = D_k$.

Random-effects model When a random-effects model is used, the correlation matrices are treated as vectors of multivariate effect sizes. The model for the sample correlation vector $\mathbf{r}_i = vechs(R_i)$ in the *i*th study by taking the column-wise non-redundant elements from R_i is:

$$\mathbf{r}_i = \rho_{\text{random}} + \mathbf{u}_i + \mathbf{e}_i \tag{29}$$

where ρ_{random} is the average correlation vector under a random-effects model, $\text{Var}(\mathbf{u}_i) = \mathbf{T}^2$ is the variance components of the random effects, and $\text{Var}(\mathbf{e}_i) = V_i$ is the conditional sampling covariance matrix. Multivariate meta-analysis listed in Eqs. 18 and 19 are used to conduct the stage 1 analysis with random-effects model.

3.2. Stage 2 analysis

After the stage 1 analysis with either a fixed- or a random-effects model, a vector of pooled correlations $\bar{\mathbf{r}}$ and its asymptotic covariance matrix \mathbf{V} are estimated. A correlation structural model $\rho(\hat{\gamma})$ is fitted with weighted least squares (WLS) estimation method by minimizing the following fit function (Cheung and Chan 2005b, 2009):

$$F(\hat{\gamma}) = (\bar{\mathbf{r}} - \rho(\hat{\gamma}))^{\mathrm{T}} \mathbf{V}^{-1} (\bar{\mathbf{r}} - \rho(\hat{\gamma})). \tag{30}$$

Likelihood-ratio statistics and various goodness-of-fit indices may be used to judge whether the proposed structural model is appropriate while SEs may be used to test the significance of individual parameter estimates.

3.3. Examples

An example from Cheung (2009b) is used to illustrate the procedures. In this example, Digman (1997) reported a second-order factor analysis on a five-factor model with 14 studies. He proposed that there were two second-order factors on the five-factor model: an **alpha** factor for *agreeableness*, *conscientiousness*, and *emotional stability*, and a **beta** factor for *extroversion* and *intellect*.

Fixed-effects model The tssem1() function is used to pool the correlation matrices with a fixed-effects model in the first stage by specifying method='FEM' in the argument. tssem2() is then used to fit a factor analytic model on the pooled correlation matrix with the inverse of its asymptotic covariance matrix as the weight matrix. When a correlation structure is fitted, it is necessary to ensure that the diagonals of the model implied matrix are all fixed at ones. We may impose this constraint by specifying diag.constraints=TRUE in the argument.

The fit indices for testing the homogeneity of the correlation matrices in Stage 1 analysis are $\chi^2(df=130,N=4,496)=1,499.73,p<.001$, CFI=0.6825, TLI=0.6581, SRMR=0.1750 and RMSEA=0.1812. This indicates that it is not reasonable to assume that the correlation matrices are homogeneous. Sub-group analysis or random-effects model that will be illustrated later are more appropriate.

The structural model in the stage 2 analysis is specified via the reticular action model (RAM) formulation (McArdle and McDonald 1984). As an exercise, we continue to fit the stage 2 model even though the homogeneity assumption of the correlation matrices is questionable. The fit indices for fitting the structural model in Stage 2 are $\chi^2(df=4,N=4496)=65.06,p<.001$, CFI=0.9802, TLI=0.9506, SRMR=0.0284 and RMSEA=0.0583. Although the goodness-of-fit indices look good, we should be cautious in interpreting them because of the poor goodness-of-fit indices in Stage 1 analysis.

R> ## Show the first 2 studies in Digman97 R> head(Digman97\$data, n=2)

```
$`Digman 1 (1994)`
             С
                 ES
                         Ε
          0.62 0.41 -0.48 0.00
    1.00
          1.00 0.59 -0.10 0.35
С
    0.62
ES
    0.41
          0.59 1.00
                     0.27 0.41
   -0.48 -0.10 0.27
                      1.00 0.37
         0.35 0.41
                     0.37 1.00
```

```
$`Digman 2 (1994)`
          C ES
                            Ι
      Α
                     Ε
   1.00 0.39 0.53 -0.30 -0.05
   0.39 1.00 0.59 0.07 0.44
ES 0.53 0.59 1.00 0.09 0.22
E -0.30 0.07 0.09 1.00 0.45
I -0.05 0.44 0.22 0.45 1.00
R> ## Show the first 2 sample sizes in Digman97
R> head(Digman97$n, n=2)
[1] 102 149
R> ## Stage 1 analysis
R> fixed1 <- tssem1(Digman97$data, Digman97$n, method="FEM")
Running TSSEM1 Analysis of Correlation Matrix
R> summary(fixed1)
Call:
tssem1FEM(my.df = my.df, n = n, cor.analysis = cor.analysis,
    model.name = model.name, cluster = cluster, suppressWarnings = suppressWarnings)
Coefficients:
      Estimate Std.Error z value Pr(>|z|)
S[1,2] 0.363116 0.013390 27.1182 < 2.2e-16 ***
S[1,3] 0.390176 0.012903 30.2402 < 2.2e-16 ***
S[1,4] 0.103751 0.015070 6.8848 5.786e-12 ***
S[1,5] 0.092246 0.015071 6.1208 9.309e-10 ***
S[2,3] 0.415999 0.012539 33.1756 < 2.2e-16 ***
S[2,4] 0.135208 0.014799 9.1365 < 2.2e-16 ***
S[2,5] 0.141213 0.014890 9.4836 < 2.2e-16 ***
S[3,4] 0.244505 0.014175 17.2494 < 2.2e-16 ***
S[3,5] 0.138167 0.014857 9.2995 < 2.2e-16 ***
S[4,5] 0.424514 0.012395 34.2477 < 2.2e-16 ***
Signif. codes: 0 '*** 0.001 '** 0.01 '* 0.05 '.' 0.1 ' ' 1
Goodness-of-fit indices:
                                    Value
Sample size
                                4496.0000
Chi-square of target model
                                1499.7340
```

```
DF of target model
                                     130.0000
p value of target model
                                       0.0000
Chi-square of independence model 4454.5995
DF of independence model
                                     140.0000
RMSEA
                                       0.1812
SRMR
                                       0.1750
TLI
                                       0.6581
CFI
                                       0.6825
ATC
                                    1239.7340
BIC
                                     406.3114
OpenMx status: 0 ("0" and "1": considered fine; other values indicate problems)
R> ## Model specification with RAM formulation
R> ## S matrix
R > Phi <- matrix(c(1, "0.3*cor", "0.3*cor", 1), ncol=2, nrow=2)
R > S1 \leftarrow bdiagMat(list(diag(c("0.2*e1","0.2*e2","0.2*e3","0.2*e4","0.2*e5")), Phi))
R> ## Not necessary but useful in understanding the model
\mathbb{R} dimnames(S1) <- list(c("A", "C", "ES", "E", "I", "Alpha", "Beta"),
                         c("A", "C", "ES", "E", "I", "Alpha", "Beta"))
R> S1
      Α
                С
                          ES
                                    Ε
                                             Ι
      "0.2*e1" "0"
                          "0"
                                    "0"
                                             "0"
Α
                                             "0"
      "0"
                "0.2*e2" "0"
                                    "0"
С
                "0"
                          "0.2*e3" "0"
                                             "0"
      "0"
ES
      "0"
                "0"
                          "0"
                                    "0.2*e4" "0"
Ε
      "0"
                "0"
                          "0"
                                    "0"
Ι
                                             "0.2*e5"
Alpha "0"
                "0"
                          "0"
                                    "0"
                                             "0"
                "0"
                          "0"
                                    "0"
                                             "0"
      "0"
Beta
      Alpha
                 Beta
      "0"
                 "0"
Α
      "0"
                 "0"
С
      "0"
                 "0"
ES
Ε
      "0"
                 "0"
                 "0"
      "0"
Ι
Alpha "1"
                 "0.3*cor"
Beta "0.3*cor" "1"
R> ## Convert it into mxMatrix class
R> S1 <- as.mxMatrix(S1)</pre>
R> ## A matrix
R> Lambda <- matrix(c(".3*Alpha_A", ".3*Alpha_C", ".3*Alpha_ES",
```

rep(0,5),".3*Beta_E",".3*Beta_I"), ncol=2, nrow=5)

```
R> A1 <- rbind(cbind(matrix(0,ncol=5,nrow=5), Lambda),
               matrix(0, ncol=7, nrow=2) )
R> ## Not necessary but useful in understanding the model
R > dimnames(A1) <- list(c("A", "C", "ES", "E", "I", "Alpha", "Beta"),
                       c("A", "C", "ES", "E", "I", "Alpha", "Beta"))
R> A1
          C ES E I
                          Alpha
                                        Beta
      "0" "0" "0" "0" ".3*Alpha_A"
                                        "0"
Α
      "0" "0" "0" "0" ".3*Alpha_C"
С
                                         "0"
      "0" "0" "0" "0" "0" ".3*Alpha_ES" "0"
ES
      "0" "0" "0" "0" "0" "0"
Ε
                                        ".3*Beta_E"
      "0" "0" "0" "0" "0" "0"
                                        ".3*Beta_I"
Alpha "0" "0" "0" "0" "0" "0"
                                        "0"
Beta "0" "0" "0" "0" "0" "0"
                                        "0"
R> A1 <- as.mxMatrix(A1)</pre>
R > F1 \leftarrow create.Fmatrix(c(1,1,1,1,1,0,0), name="F1")
R> ## Stage 2 analysis
R> fixed2 <- tssem2(fixed1, Amatrix=A1, Smatrix=S1, Fmatrix=F1,
                   diag.constraints=TRUE, intervals.type="LB",
                   model.name="TSSEM2 Digman97")
Running TSSEM2 Digman97
R> summary(fixed2)
Call:
wls(Cov = tssem1.obj$pooledS, asyCov = tssem1.obj$acovS, n = tssem1.obj$total.n,
    Amatrix = Amatrix, Smatrix = Smatrix, Fmatrix = Fmatrix,
    diag.constraints = diag.constraints, cor.analysis = cor.analysis,
    intervals.type = intervals.type, model.name = model.name,
    suppressWarnings = suppressWarnings)
95% confidence intervals: Likelihood-based statistic
Coefficients:
             Estimate Std.Error lbound ubound z value
Amatrix[1,6] 0.56258
                             NA 0.53242 0.59286
                                                      NA
Amatrix[2,6] 0.60512
                             NA 0.57509 0.63532
                                                      NA
Amatrix[3,6] 0.71913
                             NA 0.68862 0.75031
                                                      NA
                             NA 0.71911 0.85587
Amatrix[4,7] 0.78200
                                                      NA
Amatrix[5,7] 0.55089
                             NA 0.49939 0.60231
                                                      NA
Smatrix[1,1] 0.68351
                             NA 0.64852 0.71653
                                                      NA
```

<pre>Smatrix[2,2]</pre>	0.63382	NA	0.59636	0.66927	NA
<pre>Smatrix[3,3]</pre>	0.48285	NA	0.43702	0.52580	NA
Smatrix[4,4]	0.38847	NA	0.26741	0.48290	NA
Smatrix[5,5]	0.69653	NA	0.63721	0.75061	NA
<pre>Smatrix[7,6]</pre>	0.36261	NA	0.31843	0.40650	NA
	Pr(> z)				
Amatrix[1,6]	NA				
Amatrix[2,6]	NA				
Amatrix[3,6]	NA				
Amatrix[4,7]	NA				
Amatrix[5,7]	NA				
<pre>Smatrix[1,1]</pre>	NA				
<pre>Smatrix[2,2]</pre>	NA				
<pre>Smatrix[3,3]</pre>	NA				
Smatrix[4,4]	NA				
<pre>Smatrix[5,5]</pre>	NA				
<pre>Smatrix[7,6]</pre>	NA				

Goodness-of-fit indices:

	Value
Sample size	4496.0000
Chi-square of target model	65.0605
DF of target model	4.0000
p value of target model	0.0000
Number of constraints imposed on "Smatrix" $$	5.0000
DF manually adjusted	0.0000
Chi-square of independence model	3101.3209
DF of independence model	10.0000
RMSEA	0.0583
SRMR	0.0284
TLI	0.9506
CFI	0.9802
AIC	57.0605
BIC	31.4167

OpenMx status1: 0 ("0" and "1": considered fine; other values indicate problems)

Fixed-effects model with sub-group analysis Studies may not share the same population correlation matrix. If the studies can be grouped into various subgroups, we may pool the correlation matrices separately for each subgroup (Cheung and Chan 2005a). This is similar to the subgroup analysis in conventional meta-analysis (Hedges and Olkin 1985). For example, Digman (1997) groups the 14 studies according to their sample characteristics. These include children, adolescents, young adults, and mature adults. This information is stored in the variable Digman97\$cluster. We may further

Coefficients:

Estimate Std.Error z value Pr(>|z|)S[1,2] 0.297484 0.015455 19.2483 < 2.2e-16 *** S[1,3] 0.370088 0.014552 25.4315 < 2.2e-16 ***

group these studies into younger participants versus older participants. Separate fixed-effects analysis may be applied into these two groups.

The following R code may be used to conduct the analysis. Users have to supply the cluster (a vector of labels) to the cluster argument in tssem1(). The correlation or covariance matrices will be pooled separately for each cluster. The structural models will also be fitted separately for each cluster.

```
R> #### Create a variable for different samples
R> #### Younger participants: Children and Adolescents
R> #### Older participants: others
R> sample <- ifelse(Digman97$cluster %in% c("Children", "Adolescents"),
                   yes="Younger participants", no="Older participants")
R> #### Show the sample
R> sample
 [1] "Younger participants" "Younger participants"
 [3] "Younger participants" "Younger participants"
 [5] "Younger participants" "Older participants"
 [7] "Older participants"
                            "Older participants"
 [9] "Older participants"
                            "Older participants"
[11] "Older participants"
                            "Older participants"
[13] "Older participants"
                            "Older participants"
R> ## Stage 1 analysis with clusters
R> fixed1.cluster <- tssem1(Digman97$data, Digman97$n, method="FEM",
                           cluster=sample)
Running TSSEM1 Analysis of Correlation Matrix
Running TSSEM1 Analysis of Correlation Matrix
R> #### Please note that the estimates for the younger participants are problematic.
R> summary(fixed1.cluster)
$`Older participants`
Call:
tssem1FEM(my.df = data.cluster[[i]], n = n.cluster[[i]], cor.analysis = cor.analysis,
    model.name = model.name, suppressWarnings = suppressWarnings)
```

```
S[1,4] 0.137688 0.016423 8.3836 < 2.2e-16 ***
S[1,5] 0.097971 0.016744 5.8510 4.888e-09 ***
S[2,3] 0.393709 0.014163 27.7975 < 2.2e-16 ***
S[2,4] 0.182984 0.016075 11.3829 < 2.2e-16 ***
S[2,5] 0.092664 0.016664 5.5608 2.685e-08 ***
S[3,4] 0.260756 0.015573 16.7440 < 2.2e-16 ***
S[3,5] 0.096063 0.016594 5.7889 7.083e-09 ***
S[4,5] 0.411753 0.013917 29.5853 < 2.2e-16 ***
Signif. codes: 0 '*** 0.001 '** 0.01 '* 0.05 '.' 0.1 ' '1
Goodness-of-fit indices:
                                    Value
Sample size
                                 3658.0000
Chi-square of target model
                                 823.8769
DF of target model
                                   80.0000
p value of target model
                                   0.0000
Chi-square of independence model 2992.9294
DF of independence model
                                   90.0000
RMSEA
                                   0.1513
SRMR.
                                   0.1528
TLI
                                   0.7117
CFI
                                   0.7437
AIC
                                  663.8769
BIC
                                  167.5032
OpenMx status: 0 ("0" and "1": considered fine; other values indicate problems)
$`Younger participants`
Call:
tssem1FEM(my.df = data.cluster[[i]], n = n.cluster[[i]], cor.analysis = cor.analysis,
    model.name = model.name, suppressWarnings = suppressWarnings)
Coefficients:
       Estimate Std.Error z value Pr(>|z|)
S[1,2] 0.604396 0.022189 27.2389 < 2.2e-16 ***
S[1,3] 0.465441 0.027579 16.8767 < 2.2e-16 ***
S[1,4] -0.030869 0.036048 -0.8563
                                    0.39181
S[1,5] 0.061581 0.034650 1.7772
                                    0.07554 .
S[2,3] 0.501309 0.026431 18.9666 < 2.2e-16 ***
S[2,4] -0.060834  0.034660 -1.7551
                                    0.07923 .
S[2,5] 0.321019 0.031157 10.3033 < 2.2e-16 ***
S[3,4] 0.175422 0.033776 5.1937 2.062e-07 ***
```

```
S[3,5] 0.305214 0.031680 9.6344 < 2.2e-16 ***
S[4,5] 0.478573 0.026966 17.7473 < 2.2e-16 ***
Signif. codes: 0 '*** 0.001 '** 0.01 '* 0.05 '.' 0.1 ' 1
Goodness-of-fit indices:
                                     Value
Sample size
                                  838.0000
Chi-square of target model
                                  344.1826
DF of target model
                                  40.0000
p value of target model
                                   0.0000
Chi-square of independence model 1461.6701
DF of independence model
                                   50.0000
RMSEA
                                   0.2131
SRMR
                                    0.1508
TLI
                                    0.7307
CFI
                                    0.7845
AIC
                                  264.1826
BIC
                                   74.9419
OpenMx status: 0 ("0" and "1": considered fine; other values indicate problems)
R> ## Stage 2 analysis with clusters
R> fixed2.cluster <- tssem2(fixed1.cluster, Amatrix=A1, Smatrix=S1, Fmatrix=F1,
                           diag.constraints=TRUE, intervals.type="LB")
Running TSSEM2 (Fixed Effects Model) Analysis of Correlation Structure
Running TSSEM2 (Fixed Effects Model) Analysis of Correlation Structure
R> summary(fixed2.cluster)
$`Older participants`
Call:
wls(Cov = tssem1.obj$pooledS, asyCov = tssem1.obj$acovS, n = tssem1.obj$total.n,
    Amatrix = Amatrix, Smatrix = Smatrix, Fmatrix = Fmatrix,
    diag.constraints = diag.constraints, cor.analysis = cor.analysis,
    intervals.type = intervals.type, model.name = model.name,
    suppressWarnings = suppressWarnings)
95% confidence intervals: Likelihood-based statistic
Coefficients:
              Estimate Std.Error
                                   lbound
                                              ubound
Amatrix[1,6] 0.512525 NA 0.476852 0.548377
```

Amatrix[2,6]	0.550049		NA	0.514877	0.585392
Amatrix[3,6]	0.732091		NA	0.695584	0.770073
Amatrix[4,7]	0.967544		NA	0.868045	1.109618
Amatrix[5,7]	0.430460		NA	0.369165	0.486893
Smatrix[1,1]	0.737318		NA	0.699282	0.772612
Smatrix[2,2]	0.697447		NA	0.657314	0.734902
Smatrix[3,3]	0.464043		NA	0.406978	0.516169
Smatrix[4,4]	0.063858		NA	-0.231965	0.246579
Smatrix[5,5]	0.814705		NA	0.762935	0.863710
Smatrix[7,6]	0.349090		NA	0.292052	0.403236
	z value Pr	(> z)			
Amatrix[1,6]	NA	NA			
Amatrix[2,6]	NA	NA			
Amatrix[3,6]	NA	NA			
Amatrix[4,7]	NA	NA			
Amatrix[5,7]	NA	NA			
Smatrix[1,1]	NA	NA			
Smatrix[2,2]	NA	NA			
Smatrix[3,3]	NA	NA			
Smatrix[4,4]	NA	NA			
Smatrix[5,5]	NA	NA			
Smatrix[7,6]	NA	NA			

Goodness-of-fit indices:

	Value
Sample size	3658.0000
Chi-square of target model	21.9237
DF of target model	4.0000
p value of target model	0.0002
Number of constraints imposed on "Smatrix" $$	5.0000
DF manually adjusted	0.0000
Chi-square of independence model	2267.2438
DF of independence model	10.0000
RMSEA	0.0350
SRMR	0.0160
TLI	0.9801
CFI	0.9921
AIC	13.9237
BIC	-10.8950

<code>OpenMx</code> status1: 0 ("0" and "1": considered fine; other values indicate problems)

^{\$`}Younger participants`

Call:

```
wls(Cov = tssem1.obj$pooledS, asyCov = tssem1.obj$acovS, n = tssem1.obj$total.n,
   Amatrix = Amatrix, Smatrix = Smatrix, Fmatrix = Fmatrix,
   diag.constraints = diag.constraints, cor.analysis = cor.analysis,
   intervals.type = intervals.type, model.name = model.name,
   suppressWarnings = suppressWarnings)
```

95% confidence intervals: Likelihood-based statistic Coefficients:

	Estimate	Std.Error	lbound	ubound
Amatrix[1,6]	7.4764e-01	NA	7.0068e-01	7.9470e-01
Amatrix[2,6]	9.1196e-01	NA	8.7319e-01	9.5151e-01
Amatrix[3,6]	6.7721e-01	NA	6.2601e-01	7.2752e-01
Amatrix[4,7]	1.5236e-01	NA	1.4099e-02	3.4171e-01
Amatrix[5,7]	3.2896e+00	NA	1.5251e+00	2.3344e+02
Smatrix[1,1]	4.4104e-01	NA	3.6842e-01	5.0906e-01
Smatrix[2,2]	1.6833e-01	NA	9.4595e-02	2.3756e-01
Smatrix[3,3]	5.4138e-01	NA	4.7070e-01	6.0812e-01
Smatrix[4,4]	9.7679e-01	NA	8.8327e-01	9.9937e-01
Smatrix[5,5]	-9.8213e+00	NA	-9.9502e+05	-1.3113e+00
Smatrix[7,6]	1.1708e-01	NA	1.1658e-02	2.7501e-01
	z value Pr(>	z)		
Amatrix[1,6]	NA	NA		
Amatrix[2,6]	NA	NA		
Amatrix[3,6]	NA	NA		
Amatrix[4,7]	NA	NA		
Amatrix[5,7]	NA	NA		
Smatrix[1,1]	NA	NA		
Smatrix[2,2]	NA	NA		
Smatrix[3,3]	NA	NA		
Smatrix[4,4]	NA	NA		
Smatrix[5,5]	NA	NA		
Smatrix[7,6]	NA	NA		

Goodness-of-fit indices:

	Value
Sample size	838.0000
Chi-square of target model	144.8651
DF of target model	4.0000
p value of target model	0.0000
Number of constraints imposed on "Smatrix" $$	5.0000
DF manually adjusted	0.0000
Chi-square of independence model	2469.4301

```
DF of independence model 10.0000
RMSEA 0.2051
SRMR 0.1051
TLI 0.8568
CFI 0.9427
AIC 136.8651
BIC 117.9410
```

OpenMx status1: 0 ("0" and "1": considered fine; other values indicate problems)

Random-effects model TSSEM using a random-effects model may be requested by specifying the method='REM' argument in tssem1(). By default (RE.type="Symm"), a positive definite symmetric covariance matrix among the random-effects is used. For practical reasons, e.g., there are not enough studies, it may not be feasible to estimate the full variance components of the random effects. A diagonal matrix on the random-effects may be specified by using RE.type="Diag".

The fit indices for fitting the structural model in Stage 2 are $\chi^2(df = 4, N = 4,496) = 8.51, p < .001$, CFI=0.9911, TLI=0.9776, SRMR=0.0463 and RMSEA=0.0158. This indicates that the model fits the data quite well.

```
R> random1 <- tssem1(Digman97$data, Digman97$n, method="REM", RE.type="Diag")
```

Running TSSEM1 (Random Effects Model) Analysis of Correlation Matrix

R> summary(random1)

Call:

```
meta(y = ES, v = acovR, RE.constraints = diag(x = paste(RE.startvalues,
    "*Tau2_", 1:no.es, "_", 1:no.es, sep = ""), nrow = no.es,
    ncol = no.es), RE.lbound = RE.lbound, I2 = I2, model.name = model.name)
```

95% confidence intervals: z statistic approximation Coefficients:

```
Estimate Std.Error
                                      lbound
                                                 ubound
Intercept1
            3.9465e-01 5.4223e-02 2.8837e-01 5.0092e-01
Intercept2
            4.4009e-01 4.1258e-02 3.5923e-01 5.2096e-01
Intercept3
            5.4542e-02 6.1716e-02 -6.6418e-02 1.7550e-01
Intercept4
            9.8668e-02 4.6219e-02 8.0813e-03 1.8926e-01
Intercept5
           4.2966e-01 4.0155e-02 3.5096e-01 5.0837e-01
            1.2851e-01 4.0816e-02 4.8514e-02 2.0851e-01
Intercept6
Intercept7
            2.0526e-01 4.9591e-02 1.0806e-01 3.0245e-01
Intercept8
            2.3994e-01 3.1924e-02 1.7737e-01 3.0251e-01
            1.8910e-01 4.3014e-02 1.0480e-01 2.7341e-01
Intercept9
Intercept10 4.4413e-01 3.2547e-02 3.8034e-01 5.0792e-01
```

```
3.7207e-02 1.5000e-02 7.8079e-03 6.6606e-02
Tau2_1_1
Tau2_2_2
            2.0305e-02 8.4348e-03
                                    3.7735e-03 3.6837e-02
            4.8220e-02 1.9723e-02 9.5631e-03 8.6876e-02
Tau2_3_3
Tau2_4_4
            2.4610e-02 1.0624e-02 3.7872e-03 4.5434e-02
Tau2_5_5
            1.8725e-02 8.2474e-03
                                    2.5602e-03 3.4889e-02
Tau2_6_6
            1.8256e-02 8.7889e-03 1.0302e-03 3.5482e-02
Tau2_7_7
            2.9424e-02 1.2263e-02 5.3893e-03 5.3458e-02
Tau2_8_8
            9.6511e-03 4.8824e-03 8.1702e-05 1.9221e-02
Tau2_9_9
            2.0934e-02 9.1280e-03 3.0429e-03 3.8824e-02
            1.1151e-02 5.0467e-03 1.2591e-03 2.1042e-02
Tau2_10_10
           z value Pr(>|z|)
            7.2782 3.384e-13 ***
Intercept1
Intercept2 10.6668 < 2.2e-16 ***
Intercept3
           0.8838 0.376820
Intercept4
            2.1348 0.032776 *
Intercept5 10.7000 < 2.2e-16 ***
Intercept6
           3.1486 0.001641 **
Intercept7
           4.1390 3.488e-05 ***
            7.5159 5.662e-14 ***
Intercept8
Intercept9
            4.3963 1.101e-05 ***
Intercept10 13.6460 < 2.2e-16 ***
Tau2_1_1
            2.4805 0.013120 *
Tau2_2_2
            2.4073 0.016069 *
Tau2_3_3
            2.4448 0.014492 *
Tau2_4_4
            2.3164 0.020535 *
Tau2_5_5
            2.2704 0.023184 *
Tau2_6_6
            2.0772 0.037785 *
Tau2_7_7
            2.3995 0.016420 *
Tau2_8_8
            1.9767 0.048076 *
Tau2_9_9
            2.2933 0.021829 *
Tau2_10_10
            2.2095 0.027143 *
Signif. codes: 0 '*** 0.001 '** 0.01 '* 0.05 '.' 0.1 ' 1
Q statistic on homogeneity of effect sizes: 2380.994
Degrees of freedom of the Q statistic: 130
P value of the Q statistic: 0
Heterogeneity indices (based on the estimated Tau2):
                             Estimate
Intercept1: I2 (Q statistic)
                               0.9487
Intercept2: I2 (Q statistic)
                               0.9082
Intercept3: I2 (Q statistic)
                               0.9414
Intercept4: I2 (Q statistic)
                               0.8894
```

```
Intercept5: I2 (Q statistic) 0.9005
Intercept6: I2 (Q statistic) 0.8537
Intercept7: I2 (Q statistic) 0.9093
Intercept8: I2 (Q statistic) 0.7714
Intercept9: I2 (Q statistic) 0.8746
Intercept10: I2 (Q statistic) 0.8431
```

Number of studies (or clusters): 14 Number of observed statistics: 140 Number of estimated parameters: 20

Degrees of freedom: 120 -2 log likelihood: -110.8453

OpenMx status1: 0 ("0" and "1": considered fine; other values indicate problems)

Running TSSEM2 (Random Effects Model) Analysis of Correlation Structure

R> summary(random2)

Call:

95% confidence intervals: Likelihood-based statistic Coefficients:

```
Estimate Std.Error lbound ubound z value
Amatrix[1,6] 0.57255 NA 0.47370 0.67690
Amatrix[2,6] 0.59010
                          NA 0.49047 0.69487
                                                  NA
Amatrix[3,6] 0.77046
                           NA 0.65994 0.90431
                                                  NA
Amatrix[4,7] 0.69340
                           NA 0.56258 0.87183
                                                  NA
Amatrix[5,7] 0.64011
                           NA 0.50833 0.78644
                                                  NΑ
Smatrix[1,1] 0.67218
                           NA 0.54176 0.77561
                                                  NA
Smatrix[2,2] 0.65178
                           NA 0.51711 0.75944
                                                  NA
                           NA 0.18193 0.56452
Smatrix[3,3] 0.40640
                                                  NA
Smatrix[4,4] 0.51919
                           NA 0.23937 0.68352
                                                  NΑ
Smatrix[5,5] 0.59026
                           NA 0.38131 0.74161
                                                  NΑ
Smatrix[7,6] 0.39366
                           NA 0.30236 0.49030
                                                  NA
            Pr(>|z|)
Amatrix[1,6]
                  NA
```

Amatrix[2,6]	NA
Amatrix[3,6]	NA
Amatrix[4,7]	NA
Amatrix[5,7]	NA
Smatrix[1,1]	NA
Smatrix[2,2]	NA
Smatrix[3,3]	NA
Smatrix[4,4]	NA
Smatrix[5,5]	NA
Smatrix[7,6]	NA

Goodness-of-fit indices:

	Value
Sample size	4496.0000
Chi-square of target model	8.5118
DF of target model	4.0000
p value of target model	0.0745
Number of constraints imposed on "Smatrix" $$	5.0000
DF manually adjusted	0.0000
Chi-square of independence model	514.5602
DF of independence model	10.0000
RMSEA	0.0158
SRMR	0.0463
TLI	0.9776
CFI	0.9911
AIC	0.5118
BIC	-25.1319

OpenMx status1: 0 ("0" and "1": considered fine; other values indicate problems)

4. Other Useful Functions

4.1. Analysis of Correlation or Covariance Structure with Weighted Least Squares

The wls() function may be used to fit a correlation (or covariance) structure with weighted least squares (WLS) estimation method. The following example fits a one-factor CFA model on the correlation matrix with WLS estimation method. It should be noted that only the off-diagonal elements are used when a correlation structure is fitted.

```
R> #### Analysis of correlation structure
R> R1 <- matrix(c(1.00, 0.22, 0.24, 0.18,
0.22, 1.00, 0.30, 0.22,
```

```
0.24, 0.30, 1.00, 0.24,
                  0.18, 0.22, 0.24, 1.00), ncol=4, nrow=4)
R> n <- 1000
R> acovR1 <- asyCov(R1, n)</pre>
R> ## One-factor CFA model
R> (A1 <- cbind(matrix(0, nrow=5, ncol=4),</pre>
                matrix(c("0.2*a1","0.2*a2","0.2*a3","0.2*a4",0),
                ncol=1)))
     [,1] [,2] [,3] [,4] [,5]
[1,] "0"
           "0"
                "0" "0" "0.2*a1"
[2,] "0"
           "0"
                "0" "0" "0.2*a2"
[3,] "0"
           "0"
                "0"
                    "0" "0.2*a3"
[4,] "0"
           "0"
                "0"
                     "0"
                           "0.2*a4"
[5,] "0"
           "0"
                "0" "0" "0"
R> A1 <- as.mxMatrix(A1)</pre>
R > (S1 \leftarrow diag(c("0.2*e1","0.2*e2","0.2*e3","0.2*e4",1)))
     [,1]
               [,2]
                         [,3]
                                  [,4]
                                            [,5]
[1,] "0.2*e1" "0"
                         "0"
                                  "0"
                                            "0"
               "0.2*e2" "0"
[2,] "0"
                                  "0"
                                            "0"
               "0"
                        "0.2*e3" "0"
                                            "0"
[3,] "0"
[4,] "0"
               "0"
                        "0"
                                  "0.2*e4" "0"
[5,] "0"
               "0"
                         "0"
                                  "0"
                                            "1"
R> S1 <- as.mxMatrix(S1)</pre>
R> ## The first 4 variables are observed while the last one is latent.
R > (F1 \leftarrow create.Fmatrix(c(1,1,1,1,0), name="F1"))
FullMatrix 'F1'
Clabels: No labels assigned.
@values
     [,1] [,2] [,3] [,4] [,5]
[1,]
        1
              0
                   0
                        0
[2,]
                        0
        0
              1
                   0
                              0
[3,]
        0
              0
                        0
                   1
                              0
[4,]
        0
              0
                        1
```

Ofree: No free parameters.

Olbound: No lower bounds assigned.

Qubound: No upper bounds assigned.

Running WLS Analysis of Correlation Structure

R> summary(wls.fit1)

Call:

wls(Cov = R1, asyCov = acovR1, n = n, Amatrix = A1, Smatrix = S1,
 Fmatrix = F1, diag.constraints = TRUE, cor.analysis = TRUE,
 intervals.type = "LB")

95% confidence intervals: Likelihood-based statistic Coefficients:

	Estimate	Std.Error	lbound	ubound	z	value
Amatrix[1,5]	0.42159	NA	0.34435	0.50017		NA
Amatrix[2,5]	0.52376	NA	0.44603	0.60472		NA
Amatrix[3,5]	0.57092	NA	0.49204	0.65446		NA
Amatrix[4,5]	0.42159	NA	0.34435	0.50017		NA
<pre>Smatrix[1,1]</pre>	0.82226	NA	0.74983	0.88141		NA
<pre>Smatrix[2,2]</pre>	0.72567	NA	0.63430	0.80105		NA
<pre>Smatrix[3,3]</pre>	0.67405	NA	0.57165	0.75790		NA
Smatrix[4,4]	0.82226	NA	0.74990	0.88149		NA
	Pr(> z)					
Amatrix[1,5]	NA					
Amatrix[2,5]	NA					
Amatrix[3,5]	NA					
Amatrix[4,5]	NA					
<pre>Smatrix[1,1]</pre>	NA					
<pre>Smatrix[2,2]</pre>	NA					
<pre>Smatrix[3,3]</pre>	NA					
Smatrix[4,4]	NA					

Goodness-of-fit indices:

	Value
Sample size	1000.0000
Chi-square of target model	0.0134
DF of target model	2.0000
p value of target model	0.9933

```
DF manually adjusted
                                                0.0000
Chi-square of independence model
                                              207.8683
DF of independence model
                                                6.0000
RMSEA
                                                0.0000
SRMR
                                                0.0012
TLI
                                                1.0295
CFI
                                                1.0000
ATC
                                               -3.9866
                                              -13.8021
BIC
OpenMx status1: 0 ("0" and "1": considered fine; other values indicate problems)
```

4.0000

4.2. Likelihood-based Confidence Intervals

Number of constraints imposed on "Smatrix"

Most CIs are based on the estimated standard errors. These are known as Wald CIs. Wald CIs are symmetric around the estimates. The Wald CIs might be outside of the meaningful boundaries, for example, a negative lower limit for the variance or larger than 1 for a correlation coefficient. A preferable approach is to construct the CIs based on the likelihood. This is known as the likelihood based CI (Cheung 2009a; Neale and Miller 1997). Likelihood based CIs on the parameter estimates can be required by specifying intervals.type='LB' argument. This is especially useful in constructing confidence intervals for the variance components.

```
R> ## Random-effects meta-analysis with ML
R> summary( meta(y=di, v=vi, data=Becker83, intervals.type="LB") )
Running Meta analysis with ML
Call:
meta(y = di, v = vi, data = Becker83, intervals.type = "LB")
95% confidence intervals: Likelihood-based statistic
Coefficients:
            Estimate Std.Error
                                  lbound
                                            ubound z value
Intercept1 0.174734
                            NA -0.052165 0.437627
                                                         NA
Tau2_1_1
            0.077376
                            NA 0.015124 0.302999
                                                         NA
           Pr(>|z|)
Intercept1
                 NA
Tau2_1_1
                 NA
Q statistic on homogeneity of effect sizes: 30.64949
Degrees of freedom of the Q statistic: 9
P value of the Q statistic: 0.0003399239
```

```
Heterogeneity indices (I2) and their 95% likelihood-based CIs:
                              lbound Estimate ubound
Intercept1: I2 (Q statistic) 0.28410 0.67182 0.8888
Number of studies (or clusters): 10
Number of observed statistics: 10
Number of estimated parameters: 2
Degrees of freedom: 8
-2 log likelihood: 7.928307
OpenMx status1: 0 ("0" and "1": considered fine; other values indicate problems)
R> ## Mixed-effects meta-analysis with "log(items)" as a predictor
R> summary( meta(y=di, v=vi, x=log(items), data=Becker83, intervals.type="LB") )
Running Meta analysis with ML
Call:
meta(y = di, v = vi, x = log(items), data = Becker83, intervals.type = "LB")
95% confidence intervals: Likelihood-based statistic
Coefficients:
              Estimate Std.Error
                                      lbound
                                                  ubound
Intercept1 -3.2015e-01
                         NA -5.4408e-01 -7.7598e-02
            2.1088e-01
                            NA 1.1838e-01 3.0789e-01
Slope1_1
            1.0000e-10
                            NA 1.0000e-10 5.7947e-02
Tau2_1_1
           z value Pr(>|z|)
Intercept1
                NA
                        NA
                NA
                         NA
Slope1_1
Tau2_1_1
               NA
                         NA
Q statistic on homogeneity of effect sizes: 30.64949
Degrees of freedom of the Q statistic: 9
P value of the Q statistic: 0.0003399239
Explained variances (R2):
                           у1
Tau2 (no predictor)
                       0.0774
Tau2 (with predictors) 0.0000
R2
                       1.0000
Number of studies (or clusters): 10
Number of observed statistics: 10
Number of estimated parameters: 3
Degrees of freedom: 7
```

```
-2 log likelihood: -4.208024
OpenMx status1: 0 ("0" and "1": considered fine; other values indicate problems)
```

4.3. Reading External Data Files

Data sets are most likely stored externally. **metaSEM** reads three types of data formats. The first type is full correlation/covariance matrices, for example, fullmat.dat is the same as the built-in data set Cheung09. Missing values are represented by NA (the default option). Suppose you have saved it at d:\fullmat.dat, you may read it by using the following command in R:

```
my.df <- readFullMat(file="d:/fullmat.dat")</pre>
```

The second type is lower triangle correlation/covariance matrices, for example, lowertriangle.dat. Missing values are represented by the strings NA. Suppose you have saved it at d:\lowertriangle.dat, you may read it by using the following command in R:

```
my.df <- readLowTriMat(file = "d:/lowertriangle.dat", no.var = 9, na.strings="NA")
```

The third type is vectors of correlation/covariance elements based on column vectorization. One row represents one study, for example, stackvec.dat. Suppose you have saved it at d:\stackvec.dat, you may read it by using the following R command:

```
my.df <- readStackVec(file="d:/stackvec.dat")</pre>
```

5. Installation

First of all, you need R to run it. The **metaSEM** depends on the **OpenMx** package. Since **OpenMx** is not available at CRAN, both **OpenMx** and **metaSEM** packages have to be installed separately. To install **OpenMx**, run the following command inside an R session:

```
install.packages('OpenMx', repos='http://openmx.psyc.virginia.edu/packages/')
```

See http://openmx.psyc.virginia.edu/installing-openmx for the details on how to install **OpenMx**. Moreover, **metaSEM** also depends on the **ellipse** package that can be installed by the following command inside an R session:

```
install.packages('ellipse')
```

5.1. Windows platform

Download the Windows binary of **metaSEM**. If the file is saved at d:\. Run the following command inside an R session:

install.packages(pkgs="d:/metaSEM_0.7-1.zip", repos=NULL)

Please note that $d:\$ in Windows is represented by either $d:\$ or $d:\$ in R.

5.2. Linux and Mac OS X platform

Download the source package of metaSEM. Run the following command (as Root) inside an R session:

install.packages(pkgs="metaSEM_0.7-1.tar.gz", repos=NULL, type="source")

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Affiliation:

Mike W.-L. Cheung Department of Psychology Faculty of Arts and Social Sciences National University of Singapore Singapore 117570

E-mail: mikewlcheung@nus.edu.sg

URL: http://courses.nus.edu.sg/course/psycwlm/internet/metaSEM/