

Introduction to RevBayes

Bastien Boussau
With massive borrowings from
Sebastian Hoehna
Tracy Heath
Michael Landis

What is RevBayes?

- Software for Bayesian statistical analyses
- Strong focus on phylogenetic models
- Strong focus on MCMC algorithms (Metropolis-Hastings, MCMCMC)
- C++ core for efficiency
- Interpreted R-like language for interactivity
- Built with probabilistic graphical models in mind

Useful pointers

- http://revbayes.github.io/
- http://revbayes.github.io/tutorials/
- https://revbayes.github.io/documentation/
- http://revbayes.github.io/download
- https://github.com/revbayes
- https://groups.google.com/g/revbayes-users

Graphical models in RevBayes

 Graphical models provide a simple way to represent probabilistic models

 They are also a powerful way to identify conditionally independent

variables:

 In RevBayes, objects are programmed in such a way that algorithms naturally benefit from conditional independence

The Rev language

- R-like
- Type inference
- Object-oriented
- Completions
- Case-sensitive
- Math functions:

```
exp(1)
ln(1)
sqrt(16)
power(2,2)
```

• Distributions:

```
dexp(x=1,lambda=1) # exponential distribution density function
qexp(0.5,1) # exponential distribution quantile function
rexp(n=10,1) # random draws from an exponential distribution
dnorm(-2.0,0.0,1.0) # normal distribution density function
rnorm(n=10,0,1) # random draws from a normal distribution
```

The Rev language: useful functions • Structure of a variable :

```
# printing the structure information of 'a'
str(a)
  _variable = a
  _RevType = Natural
  _RevTypeSpec = [ Natural, Integer, RevObject ]
  _value = 1
  _dagType = Constant DAG node
  _children = [ ]
  .methods = void function ()
```

- Type of a variable : type(a) Natural
- Help: ?mean
- Working directory: getwd()
- What's in my environment: ls()
- What commands are available? ls(all=TRUE)
- Sourcing a file: source("file")

Variable declaration in Rev

2 main types of variables:

Environment variable: name = « MyAnalysis »
Model variables: c <- 1
○ Deterministic variable: d := exp(c)
○ Stochastic variable: x ~ dnExponential(c)

A little practical exercise

Start revbayes
In the terminal:

rb

Let's explore the variable types

```
a<-1
b<-2
x~dnNormal(mean=a,sd=b)
print(x)
y:=x*x
z<-x*x
x.redraw();print("x: "+x);print("y: "+y);print("z: "+z)
x.redraw();print("x: "+x);print("y: "+y);print("z: "+z)
y.redraw()
z.redraw()</pre>
```

Let's explore the variable types

Variable declaration in Rev

- 2 main types of variables:
 - Environment variable: name = « MyAnalysis »
 - Model variables:
 - Constant variable: c <- 1
 Deterministic variable: d := exp(c)
 - \bigcirc Stochastic variable: x ~ dnExponential(c)
 - More fun with stochastic variables:

```
x  # print value of stochastic node 'x'
x.probability()  # print the probability if 'x'
x.lnProbability()  # print the log-probability if 'x'
str(x)  # printing all the information of 'x'
```

The Rev language: more details

```
• Vectors: v \leftarrow v(1,2,3) or: w \leftarrow [1,2,3] or: z[1] \leftarrow 1 z[2] \leftarrow 2
• Convenience functions: z[3] \leftarrow 3 z[3] \leftarrow 3
```

• Vectors are objects: seq(1,20,2)

v.methods()

- Control structures:
 - for loops
 - while loops

```
sum <- 0
for (i in 1:100) {
   sum <- sum + i
}
sum</pre>
```

How do we set up inference with MCMC for a simple phylogenetic model in RevBayes ?

```
########
# Data #
########
# We read the sequence alignment:
data = readDiscreteCharacterData("...")
n branches = 2 * data.ntaxa() - 3
```

```
#######
# Data #
#######
# We read the sequence alignment:
data = readDiscreteCharacterData("...")
n branches = 2 * data.ntaxa() - 3
# Model of sequence evolution #
# Uniform prior on topologies
topology ~ dnUniformTopology(...)
# Exponential priors on branch lengths
for (i in 1:n branches) {
bls[i] ~ dnExponential(10)
# Putting branch lengths and topology together
psi := treeAssembly(topology, bls)
# We define a JC rate matrix:
0 \le fnJC(4)
# The sequences are drawn from a CTMC running
along the tree
seq ~ dnPhyloCTMC( tree=psi, Q=Q, type="DNA" )
# We condition the CTMC on the sequence
alignment.
seq.clamp( data )
# We declare the model as one big object that
we are going to use in the MCMC:
my model = model(psi)
```

```
#######
# Data #
#######
# We read the sequence alignment:
data = readDiscreteCharacterData("...")
n branches = 2 * data.ntaxa() - 3
######################################
# Model of sequence evolution #
# Uniform prior on topologies
topology ~ dnUniformTopology(...)
# Exponential priors on branch lengths
for (i in 1:n branches) {
 bls[i] ~ dnExponential(10)
# Putting branch lengths and topology together
psi := treeAssembly(topology, bls)
# We define a JC rate matrix:
0 \le fnJC(4)
# The sequences are drawn from a CTMC running
along the tree
seq ~ dnPhyloCTMC( tree=psi, Q=Q, type="DNA" )
# We condition the CTMC on the sequence
alignment.
seq.clamp( data )
# We declare the model as one big object that
we are going to use in the MCMC:
my model = model(psi)
```

```
#########
# Moves #
#########
# We create a vector of moves to store them
all:
moves = VectorMoves()
# Move on the topology
moves.append(mvNNI(topology, weight=10.0))
# We define moves on the branch lengths, one
for each branch.
for (i in 1:n_branches) {
   moves.append(mvScaleBactrian(bls[i],
tune=TRUE))
}
```

```
#######
# Data #
#######
# We read the sequence alignment:
data = readDiscreteCharacterData("...")
n branches = 2 * data.ntaxa() - 3
# Model of sequence evolution #
# Uniform prior on topologies
topology ~ dnUniformTopology(...)
# Exponential priors on branch lengths
for (i in 1:n branches) {
bls[i] ~ dnExponential(10)
# Putting branch lengths and topology together
psi := treeAssembly(topology, bls)
# We define a JC rate matrix:
O \leq fnJC(4)
# The sequences are drawn from a CTMC running
along the tree
seq ~ dnPhyloCTMC( tree=psi, Q=Q, type="DNA" )
# We condition the CTMC on the sequence
alignment.
seq.clamp( data )
# We declare the model as one big object that
we are going to use in the MCMC:
my model = model(psi)
```

```
########
# Moves #
#########
# We create a vector of moves to store them
all:
moves = VectorMoves()
# Move on the topology
moves.append(mvNNI(topology, weight=10.0))
# We define moves on the branch lengths, one
for each branch.
for (i in 1:n branches) {
   moves.append(mvScaleBactrian(bls[i],
tune=TRUE))
###################
# MCMC analysis #
#################
# Now we define monitors to keep track of
what's happening during the MCMC.
# One monitor to store the parameter
distributions into a file:
monitors[1] = mnModel(filename="...",
printgen=10, separator = TAB)
# We create an MCMC object:
analysis = mcmc(my model, monitors, moves,
# We run the MCMC for 20,000 iterations:
analysis.run(20000)
```

```
########
# Data #
              DATA
########
# We read th
data = readI
n branches = 2 * data.ntaxa() - 3
# Model of sequence evolution #
# Uniform prior on topologies
topology ~ dnUniformTopology(...)
# Exponential priors on branch lengths
for (i in 1:n branches) {
bls[i] ~ dnExponential(10)
# Putting branch lengths and topology together
psi := treeAssembly(topology, bls)
# We define a JC rate matrix:
0 < - fnJC(
# The sequ
                                   running
along the
seq ~ dnPh
                                   ="DNA" )
# We condi
alignment.
seq.clamp( data )
# We declare the model as one big object that
we are going to use in the MCMC:
my model = model(psi)
```

```
Moves #
             MOVES
                                    e them
 We crea
all:
moves = VectorMoves()
# Move on the topology
moves.append(mvNNI(topology, weight=10.0))
# We define moves on the branch lengths, one
for each branch.
for (i in 1:n branches) {
  moves.append(mvScaleBactrian(bls[i],
tune=TRUE))
###################
# MCMC analysis #
##################
# Now we define monitors to keep track of
what's ha
# One mon
             MCMC
distribut
monitors[
printgen=10, separator = TAB)
# We create an MCMC object:
analysis = mcmc(my model, monitors, moves,
# We run the MCMC for 20,000 iterations:
analysis.run(20000)
```

Advice on organizing an analysis

- create a folder for the analysis
- put data in a folder « data »
- put the scripts in a folder « scripts »
- store the output files in « analyses » or « output »
- to run an analysis from the terminal:
 rb scripts/myscript.rev
- or, from within rb: source("scripts/myscript.rev")