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# PyGeNN: A Python library for GPU-enhanced neural networks

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## ABSTRACT

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**Keywords: GPU, high-performance computing, parallel computing, benchmarking, computational neuroscience, spiking neural networks, Python**

## 1 INTRODUCTION

A wide range of spiking neural network (SNN) simulators are available, each with their own niche. NEST (Gewaltig and Diesmann, 2007) is widely used for large-scale point neuron simulations on distributed computing systems; NEURON (Carnevale and Hines, 2006) and Arbor (Akar et al., 2019) specialise in the simulation of complex multi-compartmental models; and CARLsim (Chou et al., 2018), NeuronGPU (Golosio et al., 2020) and GeNN (Yavuz et al., 2016) use Graphics Processing Units (GPUs) to accelerate point neuron models. For performance reasons, many of these simulators are written in C++ and, especially amongst the older simulators, users describe their models either using a Domain-Specific Language (DSL) or directly in C++. For programming language purists, a DSL may be an elegant way of describing an SNN network model and, for simulator developers, not having to add bindings to another language is convenient. However, both choices act as a barrier to potential users. Therefore, with both the computational neuroscience and machine learning communities gradually coalescing towards a Python-based ecosystem with a wealth of mature libraries for scientific computing (Hunter, 2007; Van Der Walt et al., 2011; Millman and Aivazis, 2011), exposing spiking neural network simulators to Python seems a pragmatic choice. NEST (Eppler et al., 2009), Neuron (Hines et al., 2009) and CARLsim (Balaji et al., 2020) have all taken this route and now offer a Python interface. Furthermore, newer simulators such as Arbor and Brian2 (Stimberg et al., 2019) have been designed from the ground up with a Python interface.

While we have recently demonstrated some very competitive performance results (Knight and Nowotny, 2018, 2020) using our GeNN simulator (Yavuz2016), it has not been usable directly from Python. GeNN can already be used as a backend for the Python-based Brian2 simulator (Stimberg et al., 2019) but, while Brian2GeNN (Stimberg et al., 2020) allows Brian2 users to harness the performance benefits GeNN

provides, it is not possible to expose all of GeNN's unique features to Python through the Brian2 API. Specifically, GeNN not only allows users to easily define their own neuron and synapse models but, also 'snippets' for offloading the potentially costly initialisation of model parameters and connectivity onto the GPU. Additionally, GeNN provides a lot of freedom for users to integrate their own code into the simulation loop. In this paper we describe the implementation of PyGeNN – a Python package which aims to expose the full range of GeNN functionality with minimal performance overheads. While implementing new neuron and synapse models in the majority of other GPU simulators requires extending the underling C++ code, using PyGeNN, models can be defined directly from Python. We then demonstrate this ability as well as the performance of PyGeNN using two large-scale models from the literature.

## 2 MATERIALS AND METHODS

### 2.1 GeNN

GeNN (Yavuz et al., 2016) is a library for generating CUDA code for the simulation of spiking neural network models. GeNN handles much of the complexity of using CUDA directly as well as automatically performing device-specific optimizations so as to to maximize performance.

GeNN consists of a main library – implementing the API used to define models as well as the generic parts of the code generator – and an additional library for each backend (currently there is a reference C++ backend for generating CPU code and a CUDA backend. An OpenCL backend is under development). Users describe their model by implementing a `modelDefinition` function within a C++ file. For example, a model consisting of 4 Izhikevich neurons with heterogeneous parameters, driven be a constant input current might be defined as follows:

```
void modelDefinition(ModelSpec &model)
{
    model.setDT(0.1);
    model.setName("izhikevich");

    NeuronModels::IzhikevichVariable::VarValues popInit(
        -65.0, -20.0, uninitialisedVar(), uninitialisedVar(),
        uninitialisedVar(), uninitialisedVar());

    model.addNeuronPopulation<NeuronModels::IzhikevichVariable>(
        "Pop", 4, {}, popInit);

    CurrentSourceModels::DC::ParamValues csParams(10.0);

    model.addCurrentSource<CurrentSourceModels::DC>(
        "CS", "Neurons", csParams, {});
}
```

The *genn-buildmodel* command line tool is then used to compile this file; link it against the main GeNN library and the desired backend library; and finally run the resultant executable to generate the source code required to build a simulation dynamic library (a .dll file on Windows or a .so file on Linux and Mac). This dynamic library can then either be statically linked against a simulation loop provided by the user or dynamically loaded by the users simulation code. To demonstrate this latter approach, this example uses the `SharedLibraryModel` helper class supplied with GeNN to dynamically loads the previously defined

model, initialise the heterogenous neuron parameters and print each neuron's membrane voltage every timestep:

```
#include "sharedLibraryModel.h"

int main()
{
    SharedLibraryModel<float> model("./", "izhikevich");
    model.allocateMem();
    model.initialize();
    float *aPop = model.getScalar<float>("a");
    float *bPop = model.getScalar<float>("b");
    float *cPop = model.getScalar<float>("c");
    float *dPop = model.getScalar<float>("d");
    aPop[0] = 0.02; bPop[0] = 0.2;  cPop[0] = -65.0;    dPop[0] = 8.0;  // RS
    aPop[1] = 0.1;  bPop[1] = 0.2;  cPop[1] = -65.0;    dPop[1] = 2.0;  // FS
    aPop[2] = 0.02; bPop[2] = 0.2;  cPop[2] = -50.0;    dPop[2] = 2.0;  // CH
    aPop[3] = 0.02; bPop[3] = 0.2;  cPop[3] = -55.0;    dPop[3] = 4.0;  // IB
    model.initializeSparse();

    float *vPop = model.getScalar<float>("VPop");
    while(model.getTime() < 200.0f) {
        model.stepTime();
        model.pullVarFromDevice("Pop", "V");
        printf("%f, %f, %f, %f, %f\n", t, VPop[0], VPop[1], VPop[2], VPop[3]);
    }
    return EXIT_SUCCESS;
}
```

## 2.2 SWIG

In order to use GeNN from Python, both the model creation API and the `SharedLibraryModel` functionality need to be 'wrapped' so they can be called from Python. While this is possible using the API built into Python itself, a wrapper function would need to be manually implemented for each GeNN function to be exposed which would result in a lot of maintenance overhead. Instead, we chose to use SWIG (Beazley, 1996) to automatically generate wrapper functions and classes. SWIG generates Python modules based on special interface files which can directly include C++ code as well as special 'directives' which directly control SWIG:

```
%module(package="package") package
%include "test.h"
```

where the `%module` directive sets the name of the generated module and the package it will be located in and the `%include` directive parses and automatically generates wrapper functions for a C++ header file. We use SWIG in this manner to wrap both the model building and `SharedLibraryModel` APIs described in section 2.1. However, key parts of GeNN's API such as the `ModelSpec::addNeuronPopulation` method employed in section 2.1, rely on C++ templates which are not directly translatable to Python. Instead, valid template instantiations need to be given a unique name in Python using the `%template` SWIG directive:

```
%template(addNeuronPopulationLIF) ModelSpec::addNeuronPopulation<NeuronModels::LIF>;
```

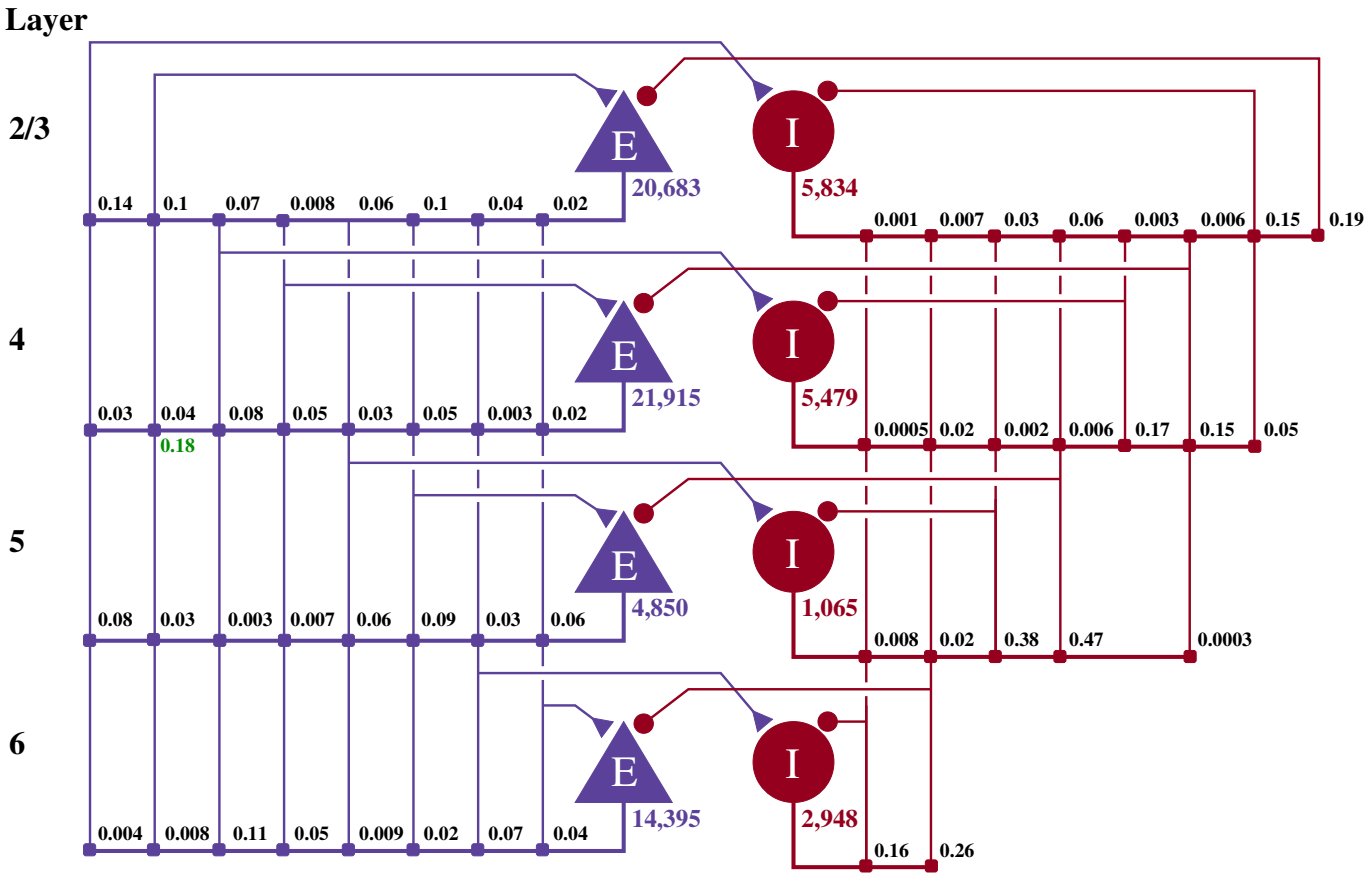


**Figure 1.** Illustration of the microcircuit model. Blue triangles represent excitatory populations, red circles represent inhibitory populations and the numbers beneath each symbol shows the number of neurons in each population. Connection probabilities are shown in small bold numbers at the appropriate point in the connection matrix. All excitatory synaptic weights are normally distributed with a mean of $0.0878\,\mathrm{nA}$ (unless otherwise indicated in green) and a standard deviation of $0.008\,78\,\mathrm{nA}$. All inhibitory synaptic weights are normally distributed with a mean of $0.3512\,\mathrm{nA}$ and a standard deviation of $0.035\,12\,\mathrm{nA}$.

Having to manually add these directives whenever a model is added to GeNN would be exactly the sort of maintenance overhead we were trying to avoid by using SWIG. Instead, when building the Python wrapper, we search the GeNN header files for the macros used to declare models in C++ and automatically generate SWIG `%template` directives.

As previously discussed, a key feature of GeNN is the ease with which it allows users to define their own neuron and synapse models as well as 'snippets' defining how variables and connectivity should be initialised. Beneath the syntactic sugar described in our previous work (Knight and Nowotny, 2018), new models can be defined in C++ by defining a new class derived from, for example, the `NeuronModels::Base` class. The ability to extend this system to Python was a key requirement of PyGeNN and, by using SWIG 'directors', C++ classes can be made inheritable from Python using a single SWIG directive:

```
%feature("director") NeuronModels::Base;
```

## 2.3 PyGeNN

While GeNN *could* be used from Python via the wrapper generated using the techniques described in the previous section, the resultant code would be unpleasant to use directly. For example, rather than

being able to specify neuron parameters using a native Python data structure such as a list or dictionary, you have to use a wrapped type such as `DoubleVector([0.25, 10.0, 0.0, 0.0, 20.0, 2.0, 0.5])`. To provide a more user-friendly and pythonic interface, we have built PyGeNN on top of the wrapper generated by SWIG. PyGeNN combines the separate model building and simulation stages of building a GeNN model in C++ into a single API, likely to be more familiar to users of existing Python-based model description languages such as PyNEST (Eppler et al., 2009) or PyNN (Davison et al., 2008). By combining the two stages together, PyGeNN can provide a unified dictionary-based API for initialising homogeneous and heterogeneous parameters as shown in this re-implementation of the previous example:

```
from pygenn import genn_wrapper, genn_model

model = genn_model.GeNNModel("float", "izhikevich")
model.dT = 0.1

izk_init = {"V": -65.0,
            "U": -20.0,
            "a": [0.02,     0.1,    0.02,   0.02],
            "b": [0.2,      0.2,    0.2,    0.2],
            "c": [-65.0,    -65.0,  -50.0,  -55.0],
            "d": [8.0,      2.0,    2.0,    4.0]}

pop = model.add_neuron_population("Pop", 4, "IzhikevichVariable", {}, izk_init)
model.add_current_source("CS", "DC", "Neurons", {"amp": 10.0}, {})

model.build()
model.load()

v = pop.vars["V"].view
while model.t < 200.0:
    model.step_time()
    model.pull_state_from_device("Neurons")
    print("%t, %f, %f, %f, %f" % (model.t, v[0], v[1], v[2], v[3]))
```

Initialisation of variables with homogeneous values – such as the neuron's membrane voltage – is performed by GeNN and those with heterogeneous values – such as the `a`, `b` and `c` parameters – are initialised by PyGeNN when the model is loaded. While the PyGeNN API is more pythonic and, hopefully, more user-friendly than the C++ interface, it still provides users with the same low-level control over the simulation. Furthermore, by using SWIG's numpy (Van Der Walt et al., 2011) interface, the host memory allocated by GeNN can be accessed directly from Python using the `pop.vars["V"].view` syntax meaning that no potentially additional expensive copying of data is required. **(TODO: DEFINING NEW NEURON MODELS, PARAMETERS AND VARIABLES)**

## 2.4 Spike recording system

Internally, GeNN stores the spikes emitted by a neuron population during one simulation timestep in an array containing the indices of the neurons that spiked alongside a counter of how many spikes have been emitted. Previously, recording spikes in GeNN was very similar to the recording of voltages shown in the previous example code – the array of neuron indices was simply copied from the GPU to the CPU every timestep. However, especially when simulating models with a small simulation timestep, such frequent

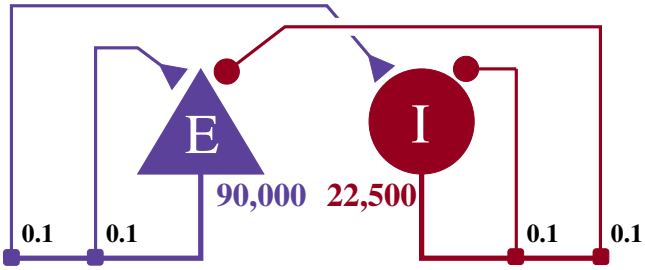


**Figure 2.** Illustration of the balanced random network model. The blue triangle represents the excitatory population, the red circle represents the inhibitory population, and the numbers beneath each symbol show the number of neurons in each population. Connection probabilities are shown in small bold numbers at the appropriate point in the connection matrix. All excitatory synaptic weights are initialised to $0.045\,61\,\mathrm{nA}$ and all inhibitory synaptic weights are initialised to $0.228\,05\,\mathrm{nA}$. **(TODO: THOMAS: UPDATE THE 800 AND 200 AND PUT SOME SORT OF SYMBOL FOR 3-FACTOR LEARNING ON THE E->E AND E->I.)**

synchronization between the CPU and GPU is costly – especially if a higher-level language such as Python is involved. Furthermore, biological neurons typically spike at a low rate (in the cortex, the average firing rate is only around $3\,\mathrm{Hz}$ (Buzsáki and Mizuseki, 2014)) meaning that the amount of spike data transferred every timestep is typically very small. To address both of these sources of inefficiency, we have added a new data structure to GeNN which stores spike data for many timesteps on device. To reduce the memory required for this data structure and to make its size independent of neural activity, the spikes emitted by a population of $N$ neurons in a single simulation timestep are stored in a $N\mathrm{bit}$ bitfield where a '1' represents a spike and a '0' the absence of one. Spiking data over multiple timesteps is then represented by bitfields stored in a circular buffer. Using this approach, even the spiking output of relatively large models, running for many timesteps can be stored in a small amount of memory. For example, the spiking output of a model with $100 \times 10^3$ neurons running for $10 \times 10^3$ simulation timesteps, required less than $120\,\mathrm{MB}$ – a small fraction of the memory on a modern GPU. While efficiently handling spikes stored in a bitfield is a little trickier than working with a list of neuron indices, GeNN provides an efficient C++ helper function for saving the spikes stored in a bitfield to a text file and a numpy-based method for decoding them in PyGeNN.

## 2.5 Cortical microcircuit model

Potjans and Diesmann (2014) developed this model of $1\,\mathrm{mm}^3$ of early-sensory cortex. The model consists of $77\,169$ LIF neurons, divided into separate populations representing the excitatory and inhibitory population in each of 4 cortical layers (2/3, 4, 5 and 6) as illustrated by figure 1. Neurons in each population are connected randomly with numbers of synapses derived from an extensive review of the anatomical literature. Within each synaptic projection, all synaptic strengths and transmission delays are normally distributed and, in total, the model has approximately $0.3 \times 10^9$ synapses. As well as receiving synaptic input, each neuron in the network also receives an independent Poisson input current, representing input from neighbouring un-modelled cortical regions. For a full description of the model parameters, please refer to Potjans and Diesmann (2014, tables 4 and 5) and for a description of the strategies used by GeNN to parallelise the initialisation and subsequent simulation of this network, please refer to Knight and Nowotny (2018, section 2.3). This model requires simulation using a relatively small timestep of $0.1\,\mathrm{ms}$, making the overheads of copying spikes from the GPU every timestep particularly significant.

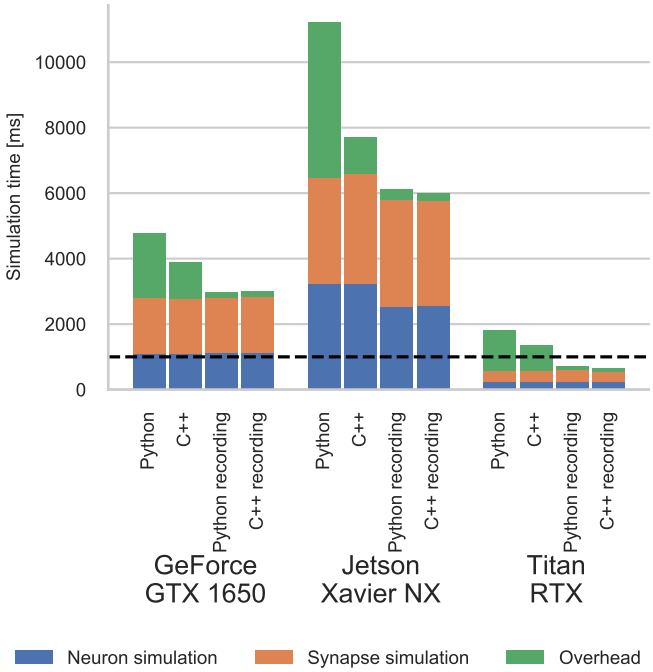


**Figure 3.** Simulation times of the microcircuit model running on various GPU hardware for $1\,\mathrm{s}$ of biological time. 'Overhead' refers to time spent in simulation loop but not within CUDA kernels. The dotted horizontal line indicates realtime performance

## 2.6 Pavlovian conditioning model

The cortical microcircuit model described in the previous section is ideal for exploring the performance of short simulations of relatively large models. However, the performance of longer simulations of smaller models is equally vital.**(TODO: DETERMINE E.G. PERCENTAGE MODELS E.G. ON OPENSOURCEBRAIN WHICH ARE SMALL)**. Such models can be particularly troublesome for GPU simulation as, not only might they not offer enough parallelism to fully occupy the device but, each timestep can be simulated so quickly that the overheads of launching kernels etc can dominate. Additional overheads can be incurred when models require injecting external stimuli throughout the simulation. **(TODO: SOMETHING ABOUT NEUROMORPHIC SYSTEMS OFTEN BEING REAL-TIME / BS ACCELERATED TIME)** Longer simulations are particularly useful when exploring synaptic plasticity so, to explore the performance of PyGeNN in this scenario, we simulate a model of Pavlovian conditioning using a three-factor STDP learning rule (Izhikevich, 2007). In this experiment, $100$ random groups of $50$ neurons (each representing stimuli $S_1 ... S_{100}$) are chosen from amongst the neurons in a $800$ neuron excitatory population and a $200$ neuron inhibitory population. Excitatory neurons are modelled as regular-spiking Izhikevich neurons (Izhikevich, 2003) and inhibitory neurons as fast-spiking Izhikevich neurons (Izhikevich, 2003). Stimuli are presented to the network in a random order, separated by intervals sampled from $U(100, 300)\mathrm{ms}$. The neurons associated with an active stimulus are stimulated for a single $1\,\mathrm{ms}$ simulation timestep with a current of $40.0\,\mathrm{nA}$, in additional to the random background current of $U(-6.5, 6.5)\mathrm{nA}$, delivered to each neuron throughout the simulation. $S_1$ is arbitrarily chosen as the Conditional Stimuli (CS) and, whenever this stimuli is presented, a reward in the form of an increase in dopamine is delivered to all the plastic synapses in the network after a delay sampled from $U(0, 1000)\mathrm{ms}$. This delay period is large enough to allow a few irrelevant stimuli to be presented which act as distractors. **(TODO: TALK MORE ABOUT MODEL/LEARNING RULE/REFER BACK TO OLD PAPER)**. The simplest way to implement this

stimulation regime is to add a current source to the excitatory and inhibitory neuron populations which adds the uniformly-distributed input current to an externally-controllable per-neuron current. In PyGeNN, the following model can be defined to do just that:

```
stim_noise_model = genn_model.create_custom_current_source_class(
    "stim_noise",
    param_names=["n"],
    var_name_types=[("iExt", "scalar", VarAccess_READ_ONLY)],
    injection_code=
        """
        $(injectCurrent, $(iExt) + ($(gennrand_uniform) * $(n) * 2.0) - $(n));
        """)
```

where the `n` parameter sets the magnitude of the background noise, the `$(injectCurrent, I)` function injects a current of $I$nA into the neuron and `$(gennrand_uniform)` uses cuRAND **(TODO: CITE)** to sample from $U(0, 1)$. Once a current source population using this model has been instantiated and a memory view to `iExt` obtained in the manner described in section 2.3, in timesteps when stimulus injection is required, current can be injected into the list of neurons contained in `stimuli_input_set` with:

```
curr_ext_view[stimuli_input_set] = 40.0
curr_pop.push_var_to_device("iExt")
```

The same approach can then be used to zero the current afterwards. However, as almost $20\,000$ stimuli will be injected over the course of a $1$ h simulation, in order to reduce potential overheads, we can offload the stimuli delivery entirely to the GPU using the following slightly more complex model:

```
stim_noise_model = genn_model.create_custom_current_source_class(
    "stim_noise",
    param_names=["n", "stimMagnitude"],
    var_name_types=[("startStim", "unsigned int"),
                    ("endStim", "unsigned int", VarAccess_READ_ONLY)],
    extra_global_params=[("stimTimes", "scalar*")],
    injection_code=
        """
        scalar current = ($(gennrand_uniform) * $(n) * 2.0) - $(n);
        if($(startStim) != $(endStim) && $(t) >= $(stimTimes)[$(startStim)]) {
            current += $(stimMagnitude);
            $(startStim)++;
        }
        $(injectCurrent, current);
        """)
```

This model retains the same logic for generating background noise but, additionally, uses a simple sparse matrix data structure to store the times at which each neuron should have current injected. **(TODO: FIGURE)** The `startStim` and `endStim` variables point to the subset of the `stimTimes` array used by each neuron's current source and, once the simulation time `$(t)` passes the time pointed to by `startStim`, current is injected and `startStim` is advanced. This array is stored in a 'extra global parameter' which is a read-only memory area that can be allocated and populated from PyGeNN, in this case by 'stacking' together a list of lists of spike times:

```
curr_pop.set_extra_global_param("stimTimes", np.hstack(neuron_stimuli_times))
```

# 3 RESULTS

In the following subsection we will analyse the performance of the models introduced in sections 2.5 and 2.6 on a representative selection of NVIDIA GPU hardware:

- Jetson Xavier NX – a low-power embedded system with a GPU based on the Volta architecture with 8 GB of shared memory.
- GeForce GTX 1050Ti – a low-end desktop GPU based on the Pascal architecture with 4 GB of dedicated memory.
- GeForce GTX 1650 – a low-end desktop GPU based on the Turing architecture with 4 GB of dedicated memory.
- Titan RTX – a high-end workstation GPU based on the Turing architecture with 24 GB of dedicated memory.

All of these systems run Ubuntu 18 apart from the GeForce 1050 Ti-equipped system which runs Windows 10.

## 3.1 Cortical microcircuit model performance

Figure 3 shows the simulation times for the full-scale microcircuit mode and, as one might predict, the Jetson Xavier NX is slower than the two desktop GPUs. However, considering that it only consumes a maximum of 15 W compared to 75 W or 320 W for the GeForce GTX 1650 and Titan RTX respectively, it still performs impressively.

The time taken to actually simulate the models ('Neuron simulation' and 'Synapse simulation') are the same when using Python and C++ as all GeNN optimisation options are exposed to PyGeNN. However, both the PyGeNN and C++ simulations spend a significant amount of every simulation step copying spike data off the device and storing it in a suitable data structure ('Overhead'). Because Python is an interpreted language, such operations are inherently slower – this is particularly noticeable on devices with a slower CPU such as the Jetson Xavier NX. Unlike the desktop GPUs, the Jetson Xavier NX's 8 GB of memory is shared between the GPU and the CPU meaning that data doesn't have to be copied between GPU and CPU memory and can instead by accessed by both. **(TODO: RUN XAVIER NX WITHOUT RECORDING AND WITHOUT ZERO COPY)** While, using this shared memory for recording spikes, reduces the overheads associated with copying data off the device, because the GPU and CPU caches are not coherent, caching must be disabled on this memory which reduces the performance of the neuron kernel. However, when the spike recording system described in section 2.4 is used, spike data is kept in GPU memory until the end of the simulation and this overhead is reduced by around a factor of 10. This now means that, on the high-end desktop GPU, this simulation now runs faster than real-time – previously only achievable using a specialised neuromorphic system (Rhodes et al., 2020) and significantly faster than other recently published GPU simulators (Golosio et al., 2020).

## 3.2 Pavlovian conditioning performance

Figure 4 shows the results of an example simulation of this model. At the beginning of each simulation (Figure 4A), the neurons representing every stimuli respond equally. However, after 1 h of simulation, the

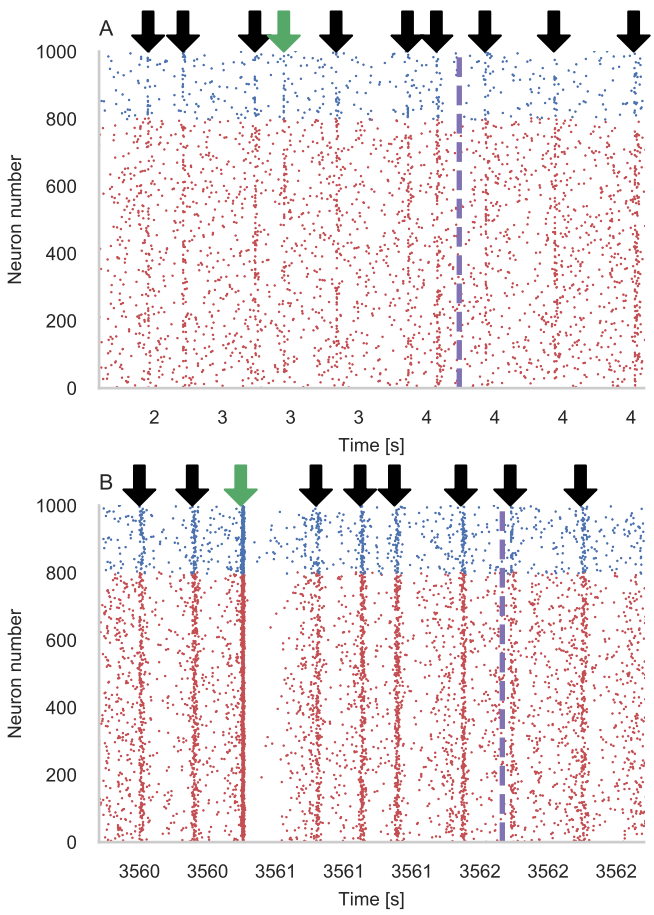


**Figure 4.** Results of Pavlovian conditioning experiment. Raster plots showing activity centred around first delivery of Conditional Stimulus (CS) during initial (A) and final (B) 50 s of simulation. Downward green arrows indicate times at which CS is delivered and downward black arrows indicate times when other, un-rewarded stimuli are delivered. Vertical dashed lines indicate times at which dopamine is delivered.

response to the CS becomes much stronger (Figure 4B) – showing that these neurons have been selectively associated even in the presence of the distractors and delayed reward.

Figure 5 shows the performance of simulations of this model, running on a selection of desktop GPUs using PyGeNN with and without the recording system described in section 2.4 and the optimized stimuli-delivery described in section 2.6. These PyGeNN results are compared to a C++ simulation which takes advantage of both optimizations. Interestingly the Titan RTX and GTX 1650 perform identically in this benchmark. This is because, as discussed previously, this model is simply not large enough to fill the 4608 CUDA cores present on the Titan RTX. Therefore, as the two GPUs share the same Turing architecture and have very similar clock speeds (1350 MHz–1770 MHz for the Titan RTX and 1485 MHz–1665 MHz for the GTX 1650), the two GPUs perform very similarly. Furthermore, on these two systems, while using the recording system significantly improves performance, the impact of delivering stimuli on the GPU is minimal. However, the GTX 1050 Ti performs rather differently. Although the clock speed of this device is not significantly lower (1290 MHz–1392 MHz, its performance is significantly worse.

"concurrent execution of integer and floating point operations" **(TODO: EXAMINE GPU TIMELINE IN NSIGHT SYSTEMS)**

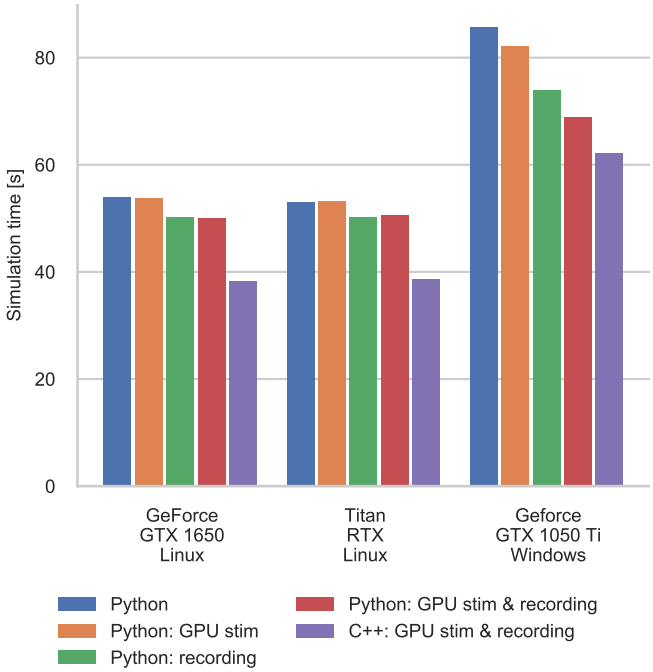


**Figure 5.** Simulation times of the Pavlovian Conditioning model running on various GPU hardware for 1 h or biological time.

## 4 DISCUSSION

discuss!

- Turing architecture is great for GeNN!
- PyGeNN as an intermediate layer - PyNN, ML
- Cost of C++ - Python calls in models

## CONFLICT OF INTEREST STATEMENT

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

## AUTHOR CONTRIBUTIONS

JK and TN wrote the paper. TN is the original developer of GeNN. AK was the original developer of PyGeNN. JK is currently the primary developer of both GeNN and PyGeNN and was responsible for implementing the spike recording system. JK performed the experiments and the analysis of the results that are presented in this work.

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## DATA AVAILABILITY STATEMENT

The datasets [GENERATED/ANALYZED] for this study can be found in the [NAME OF REPOSITORY] [LINK].

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