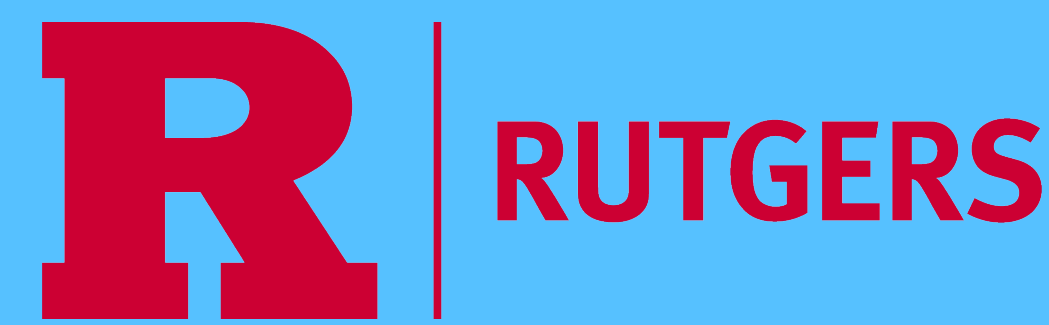


# Measuring Lipid Binding Affinities in Unbiased CG-MD Using the Density-Threshold Affinity



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Read our protocol paper!

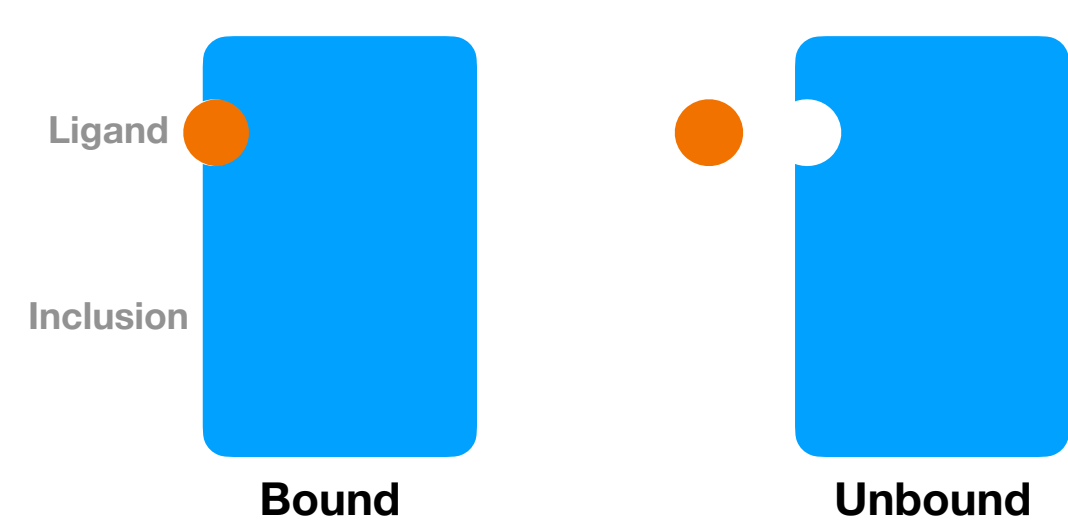
## Abstract

It is now well established that lipids bind specifically to membrane protein transmembrane domains (TMDs) and may allosterically modulate protein function, but measuring a lipid binding affinity experimentally remains a challenge. Coarse-grained molecular dynamics (CG-MD) simulations have been used extensively to study lipid-protein interactions due to the enhanced lipid diffusion and the longer accessible time-scales afforded by a CG model. Nonetheless, a number of conceptual challenges arise when MD trajectories are analyzed, including how to define the 'bound' state, and how to differentiate between a bulk lipid versus a specifically bound lipid of the same species. Most solutions to these problems have relied on measuring residence times to calculate off-rates, but these quantities are difficult to compare across force fields as well as to experimental data. We previously introduced the Density-Threshold Affinity (DTA), a method for determining the binding affinity of a lipid for a defined binding site by measuring thermodynamic quantities in unbiased CG-MD. The DTA quantifies the excess density of a defined lipid species in a binding site, compared against a bulk membrane patch of equal area. In the present work, we show how the DTA can be used to quantitatively rank binding sites as well as to determine which lipid species will out-compete the other membrane components for a particular binding site.

## Conceptual problems when defining binding in hydrophobic contexts

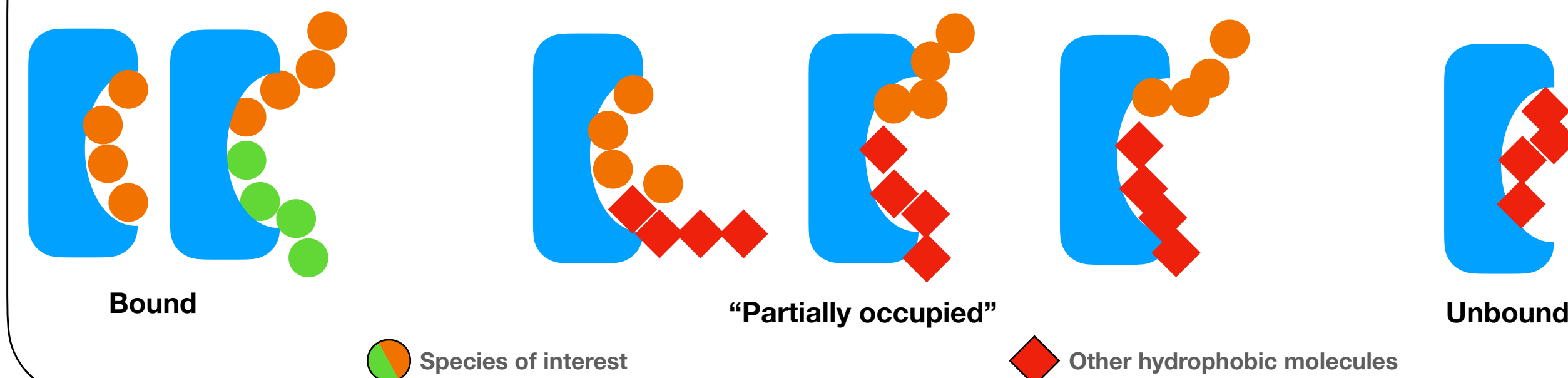
### Problem 1: Partial Occupancy

#### "Classic" two-state binding



$$\Delta G = -RT \ln \frac{P_{occ}}{P_{unocc}}$$

#### Binding of chain-like molecules in a hydrophobic environment

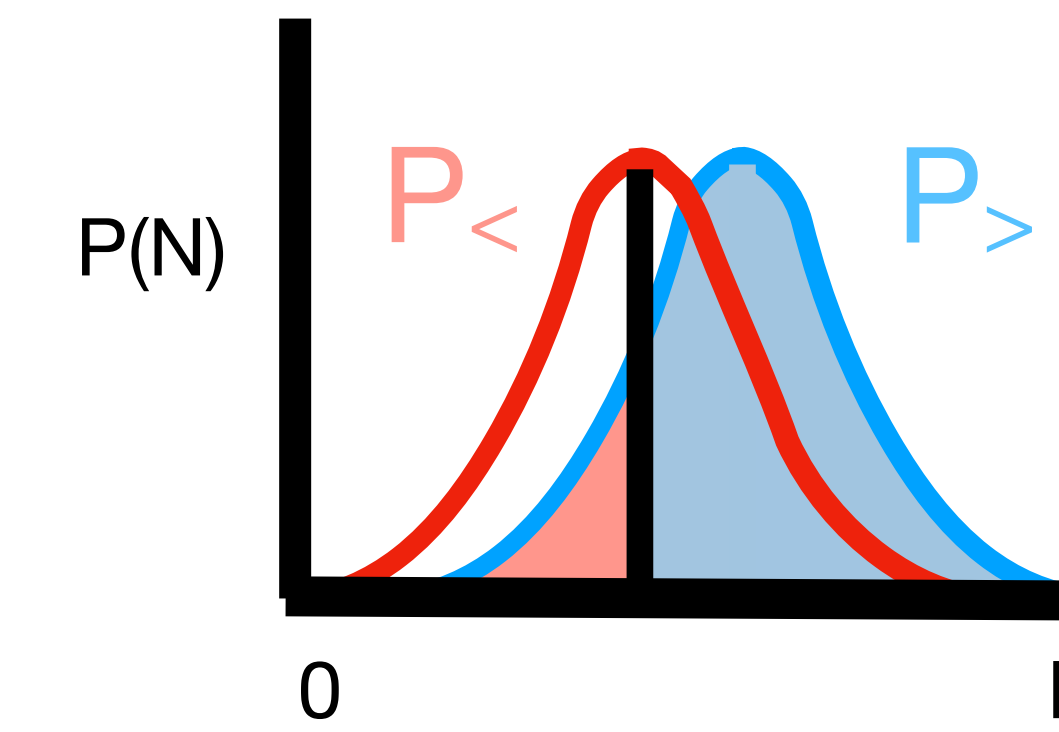
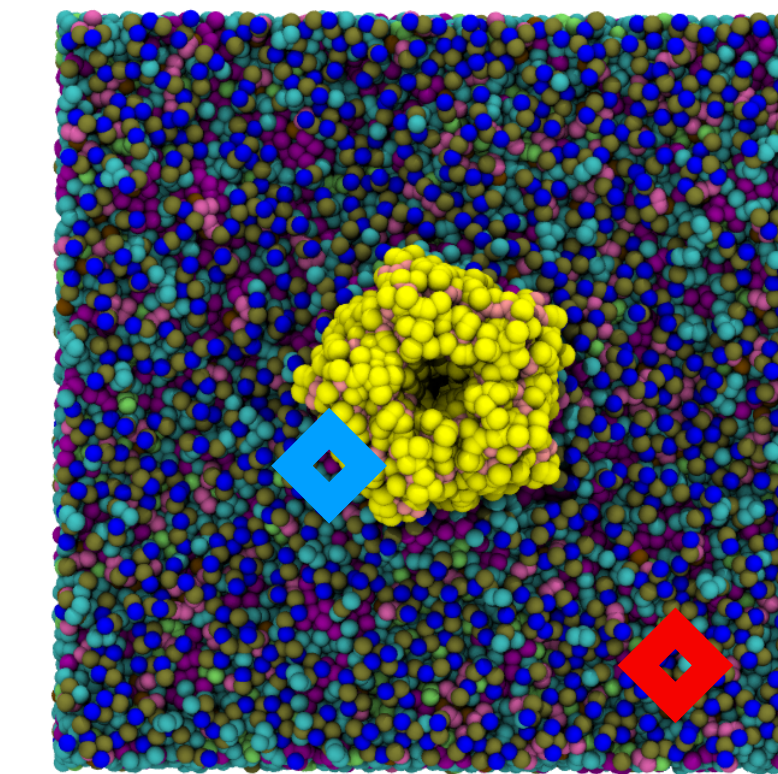


$$\Delta G = -RT \ln \frac{P_{occ}^?}{P_{unocc}^?}$$

### Problem 2: Ligand and Solvent, Simultaneously

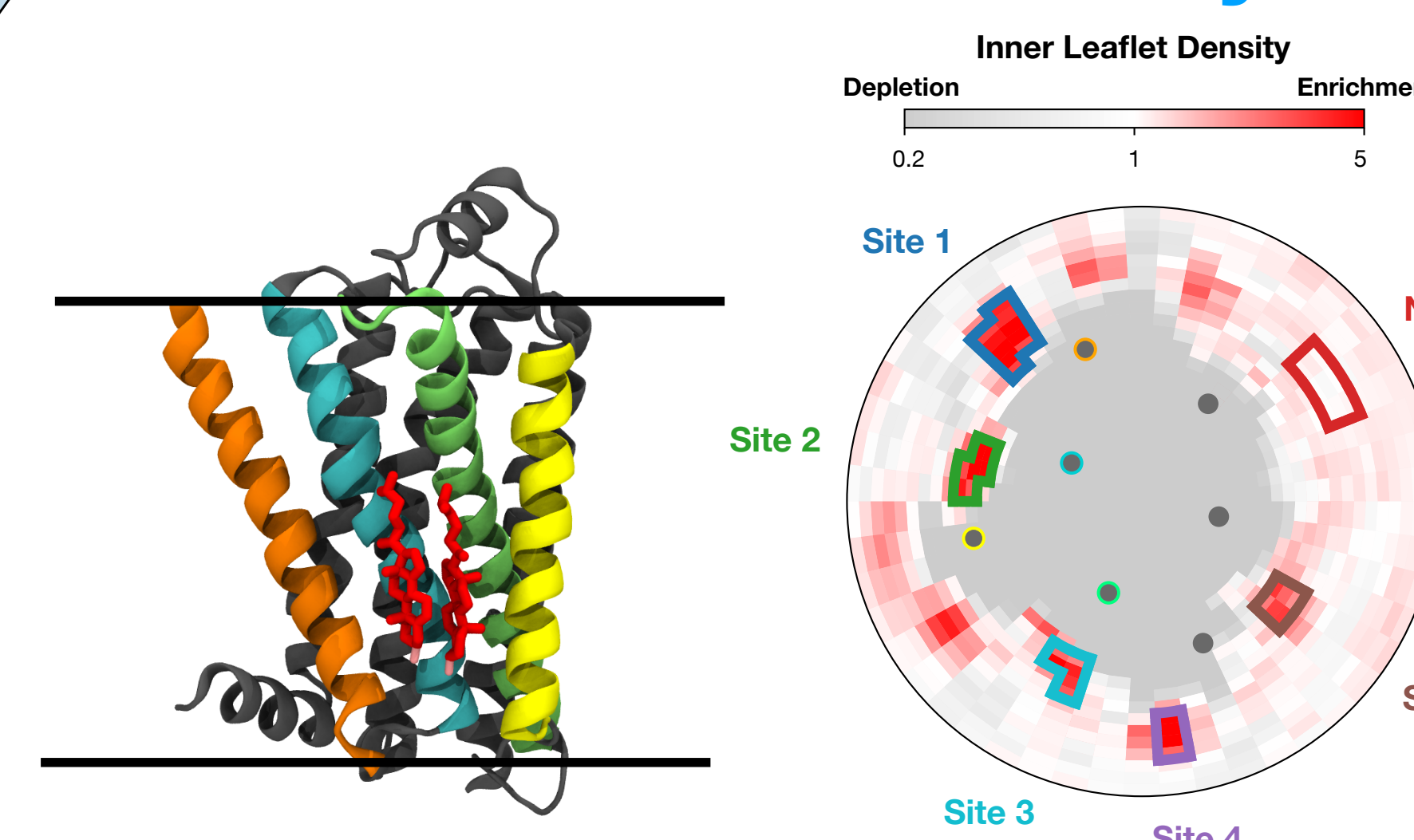
Is the molecule in the site because it is bound?  
Or is it in the site because it is solvent?

## The Density-Threshold Affinity: $\Delta G_{bind}$

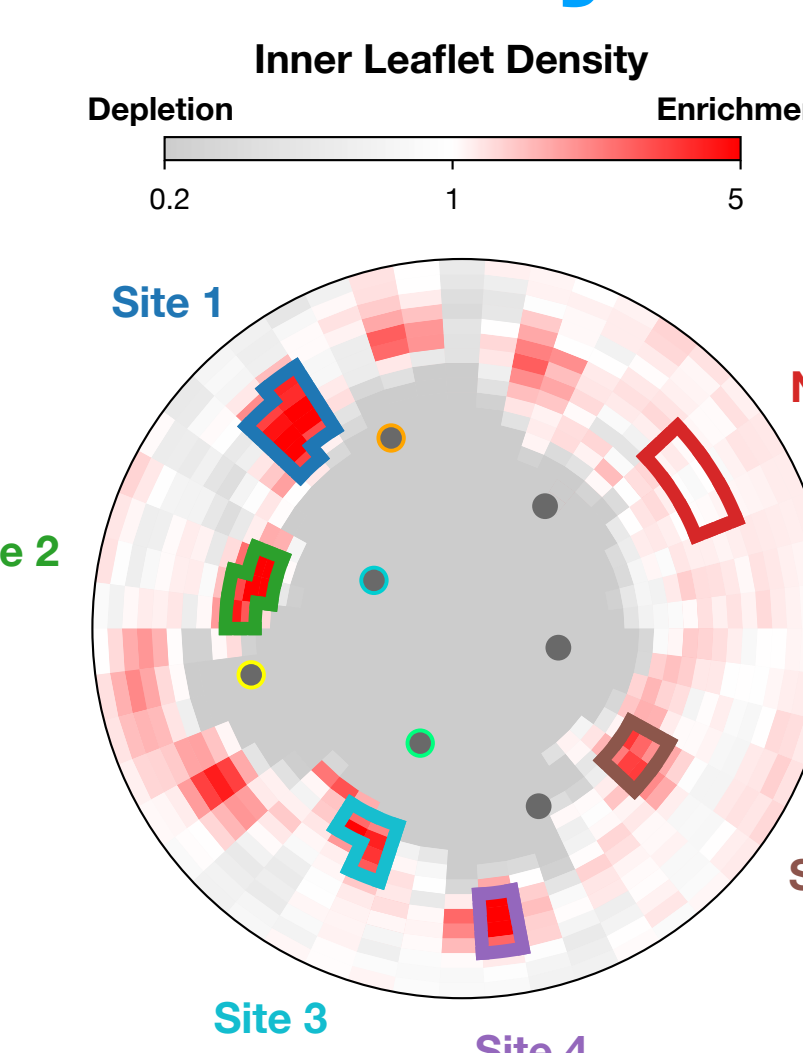


$$\Delta G = -RT \ln \frac{P_{>}}{P_{<}}$$

## Cholesterol affinity for GPCR: binding sites identified & ranked



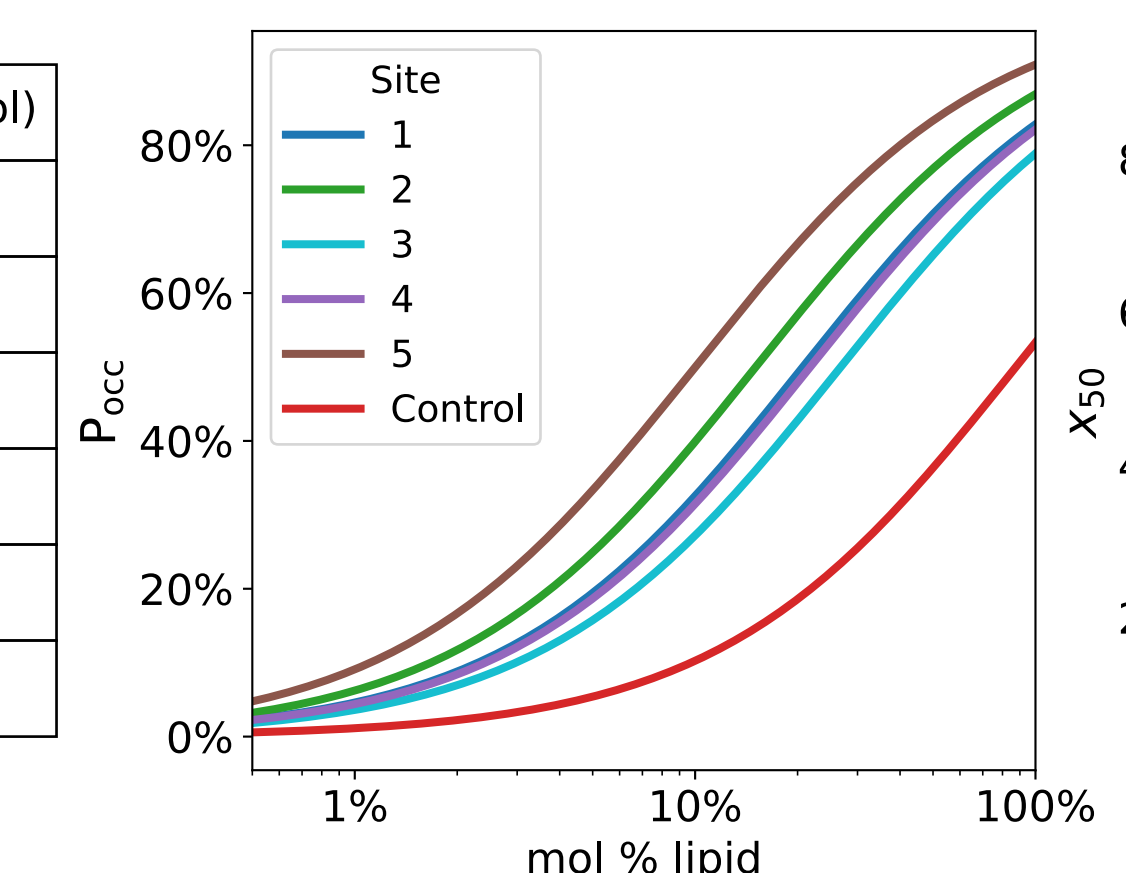
**Fig 1A:** The  $\beta_2$  Adrenergic Receptor ( $\beta_2$ -AR, pdb id 3D4S)<sup>1</sup> with intracellular domain removed. Approximate membrane surface indicated by black lines. Cholesterol beads (red) are depicted in their crystallographic sites<sup>1</sup> as an illustration, but the structural cholesterol molecules were not coarse-grained alongside the protein.



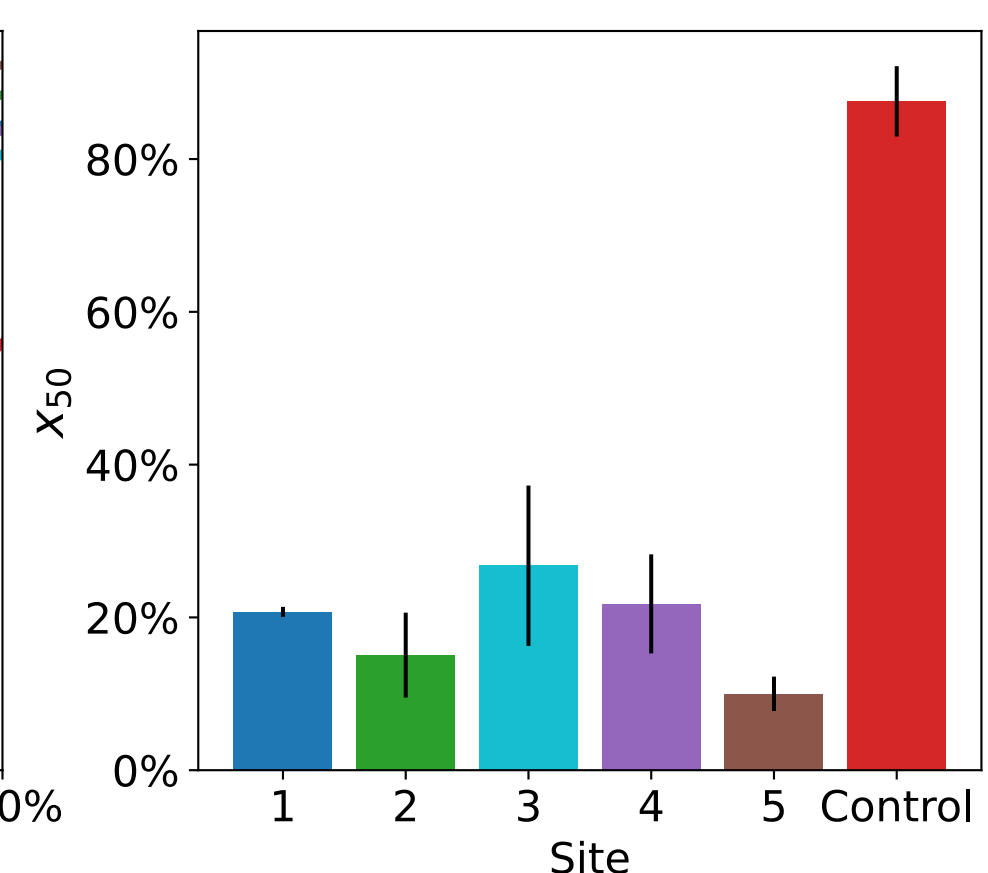
**Fig 1B:** Density enrichment of cholesterol in the inner leaflet within 2.5 nm of the protein center. Protein helices are indicated by grey circles, highlighted to match coloring in Fig 1A. Potential binding sites identified from this analysis are outlined in color and analyzed further on subsequent panels.

Site	$\Delta G_{bind}$ (kcal/mol)
1	$-1.0 \pm 0.1$
2	$-1.2 \pm 0.3$
3	$-0.8 \pm 0.3$
4	$-1.0 \pm 0.2$
5	$-1.5 \pm 0.2$
Control	$-0.1 \pm 0.1$

**Fig 1C:** Binding affinities ( $\Delta G_{bind}$ ) and standard error of the mean measured in each site. Standard error computed with N=3 replicas. Cells are colored to match site outlines in Fig 1B.



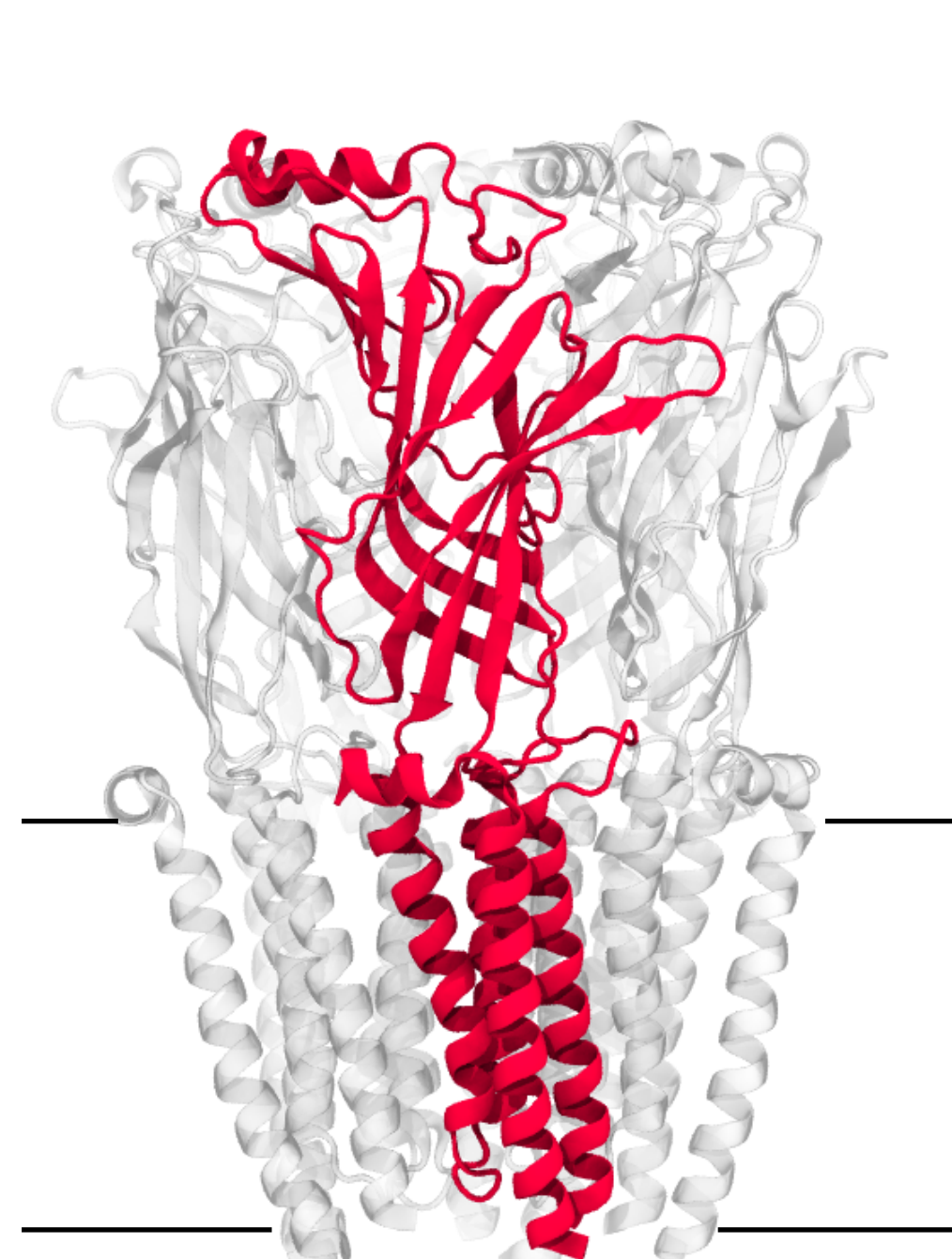
**Fig 1D:** Titration curve for each site. Lines are colored to match site outlines in Fig 1B.



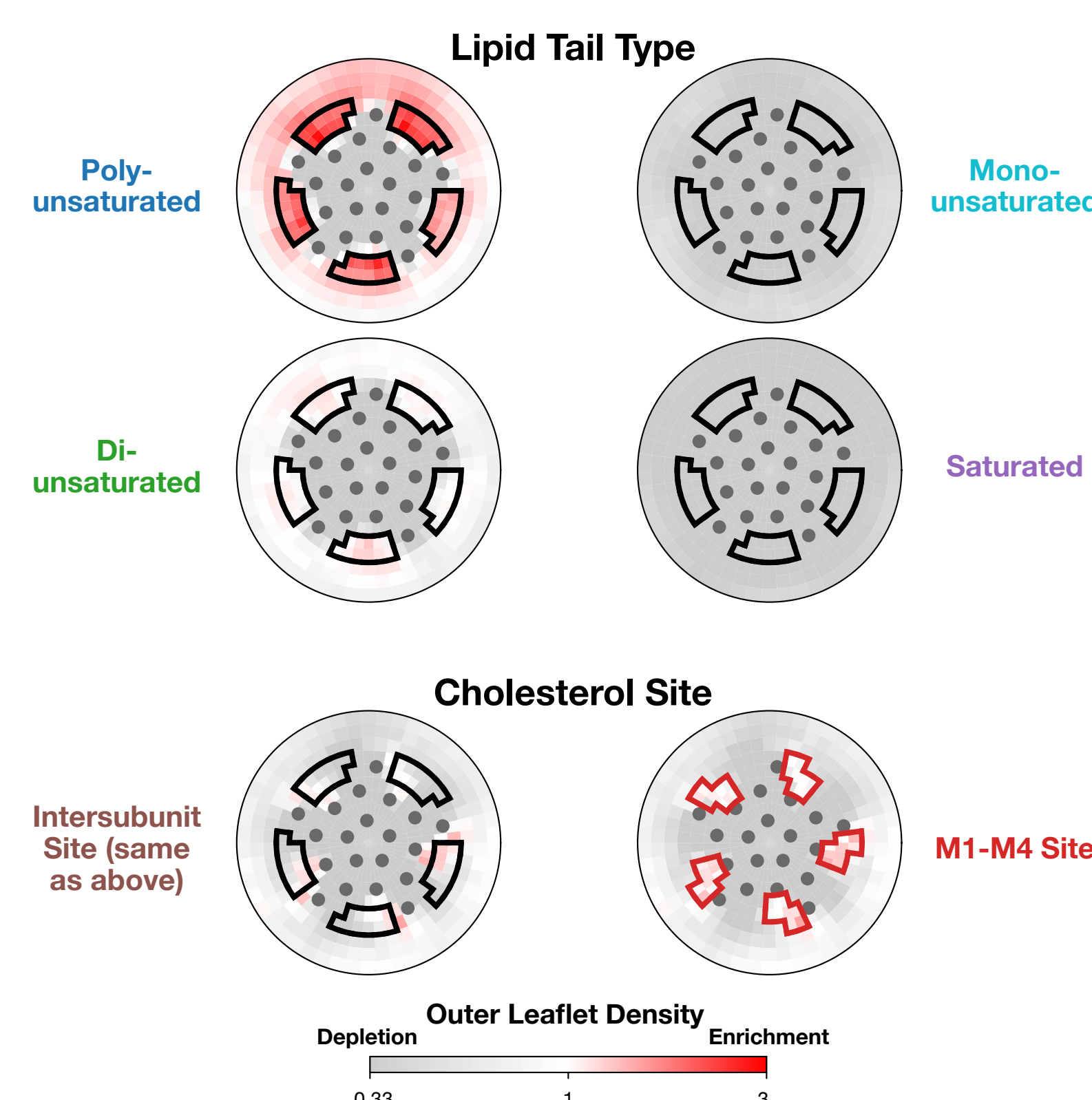
**Fig 1E:** Molecular percentage at which the site is expected to be occupied 50% of the time ( $x_{50}$ ). Bars are colored to match sites outlined in Fig 1B.

1. Hanson, et al., Structure, 2008

## Differential binding of lipid tails to pLGIC intersubunit site in oocyte-mimetic membrane



**Fig 2A:** The  $\alpha$ -7 nicotinic acetylcholine receptor (nAChR) (pdb id 8v89)<sup>4</sup> with intracellular domain removed, shown in membrane/lateral view. Black bars indicate approx. membrane position.

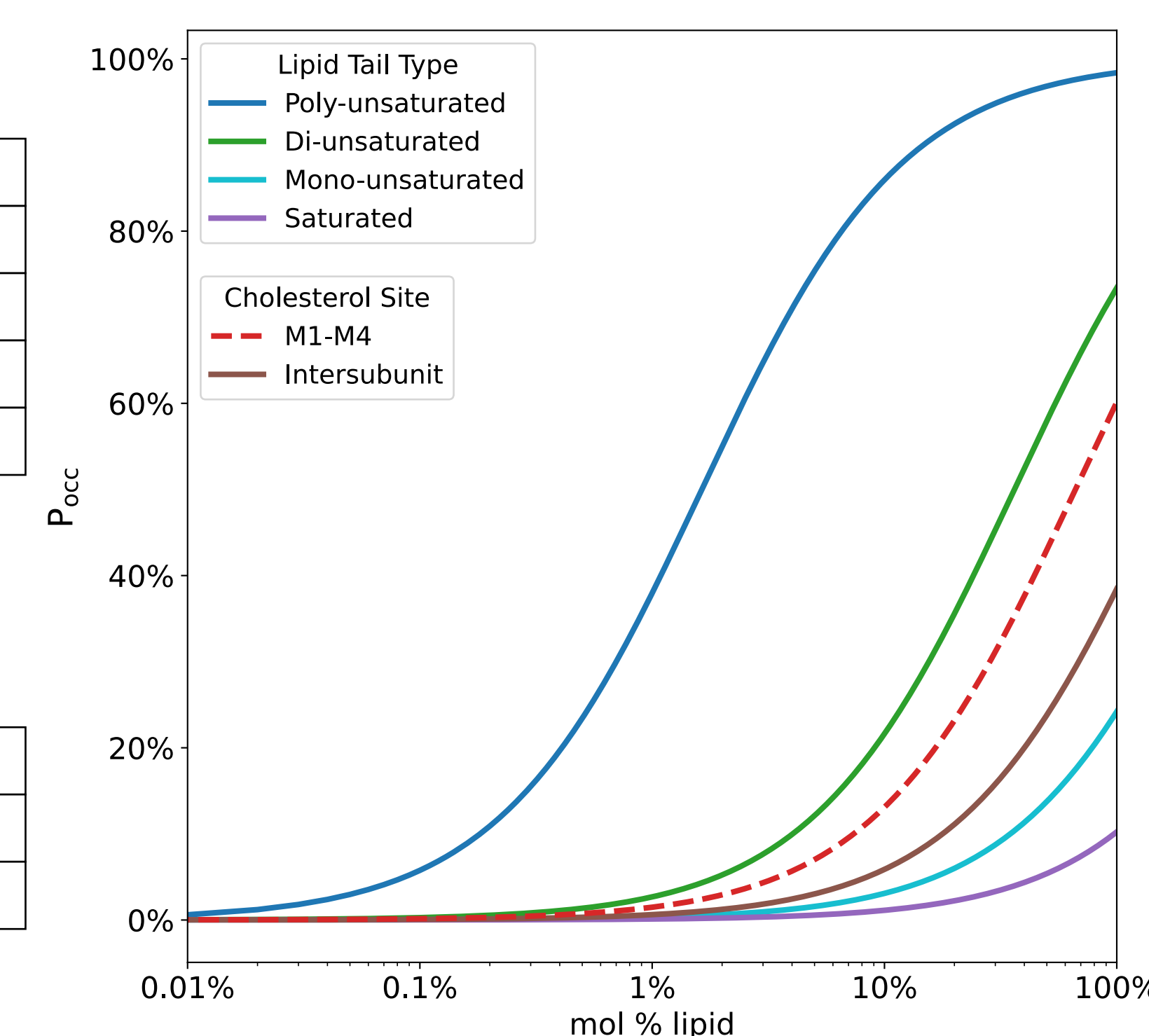


**Fig 2B:** Density enrichment for different lipid tail types (top) and of cholesterol (bottom). The five symmetric sites are treated as independent replicas and standard error is computed with N=20 (5 subunits  $\times$  4 replicas).

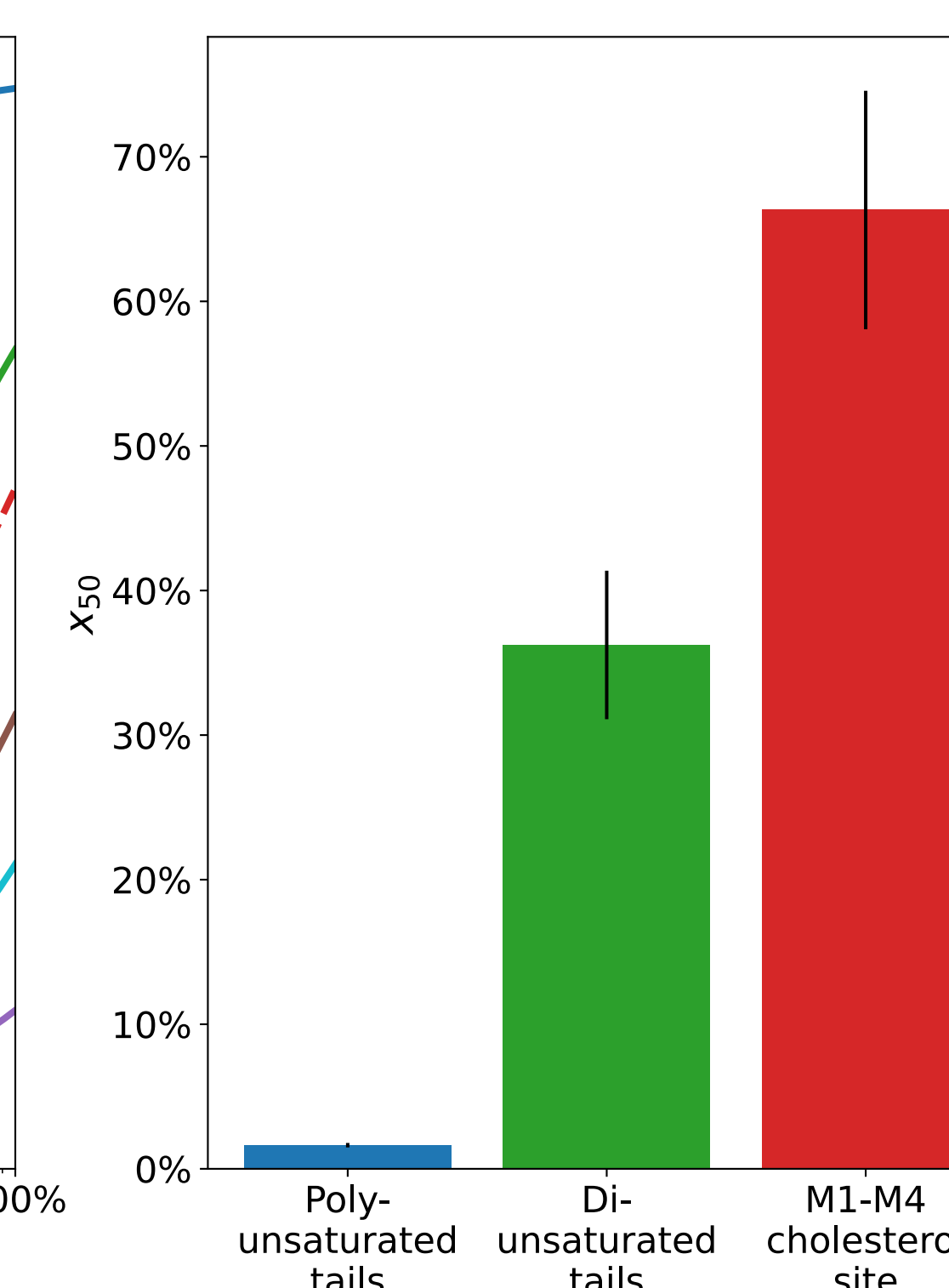
Lipid Tail Type	$\Delta G_{bind}$ (kcal/mol)
Poly-unsaturated	$-2.6 \pm 0.1$
Di-unsaturated	$-0.6 \pm 0.1$
Mono-unsaturated	$0.7 \pm 0.1$
Saturated	$1.4 \pm 0.1$

Cholesterol Site	$\Delta G_{bind}$ (kcal/mol)
M1-M4	$-0.3 \pm 0.1$
Intersubunit	$0.3 \pm 0.1$

**Fig 2C:** Binding affinities ( $\Delta G_{bind}$ ) and standard error of the mean measured in each site. The five symmetric sites are treated as independent replicas and standard error is computed with N=20 (5 subunits  $\times$  4 replicas).



**Fig 2D:** Titration curve for intersubunit site (solid lines) and for cholesterol in the M1-M4 site (dashed line).



**Fig 2E:** Molecular percentage at which the site is expected to be occupied 50% of the time ( $x_{50}$ ).

1. Hill, et al. Am. J. Physiol. 2005

1. Burke, et al. Cell, 2024

## Conclusion

## References

## Support

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