

# AgeStrucNe User Manual

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# AgeStrucNe

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## 0.1 Introduction

The AgeStrucNe GUI interface offers a user interface to allow easy access to simuPOP-based simulations [1] and the LDNe-based Nb and Ne estimations, using version 2 of the LDNe program [3]. The program integrates this functionality as implemented in Tiago Antao's python program, AgeStructureNe, available at <https://github.com/tiagoantao/AgeStructureNe.git>.

We also offer an interface for plotting Nb and Ne estimations, and regressions based on the estimations. Our program offers a separate interface for each of three functions: population simulation, Nb and Ne estimation, and estimate visualization. The genepop file output from a simulation can be loaded into an Nb estimation interface, and in turn, the output from an Nb estimation can be loaded into a visualization interface. The Nb estimation interface can also use any genepop file for input.

For program questions, please contact us through the program's email account at [agestrucne@gmail.com](mailto:agestrucne@gmail.com).

## 0.2 Installation

The program can be installed using one of several approaches, depending on the host computer's platform. Please see the Installation section in the README.md available at

<https://github.com/popgengui/agestrucne/blob/data/README.md>

## 0.3 Starting the program

The program is launched at a terminal using a python 2.7, 3.5, or 3.6 executable, invoking the "agestrucne" executable. Please see the README.md file for instructions on installing the program.

Once you have downloaded the "data" branch of our program's github repository, you can, for example, open a terminal, change your current directory to the agestrucne folder supplied by the download, and load a configuration file into the Simulation interface (Section 0.4).

## 0.4 Loading interfaces

To load one or more of the three interfaces for performing simulations, Nb/Ne estimations, or plotting programs, from the main menu click on the New menu (Figure 0.1). You can load any number of interfaces and run them simultaneously, though caution is warranted, since a computer can be taxed to a standstill by the over-allocation of CPUs and/or memory (Figure 0.1).

## 0.5 Running a simulation

### 0.5.1 Load an interface

Use the add menu (Figure 0.1) to load a new simulation interface. Steps for preparing the interface to run a simulation follow.

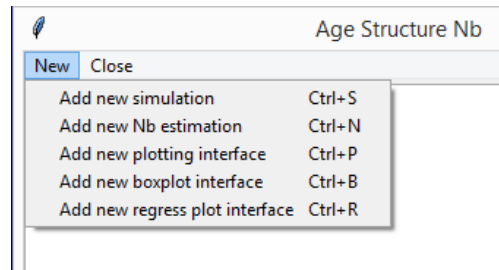


Figure 0.1: Adding an interface

### 0.5.2 Load a configuration file.

The initial simulation interface requires the user to load a configuration file (Figure 0.2). If you have not already downloaded our collection of configuration files, you can get them at our programs github repository, downloading our data branch at <https://github.com/popgengui/agestrucne/tree/data>. The files are inside the “configuration\_files” subdirectory, inside the main program directory, “agestrucne.” You can also load your own configuration (see our provided configuration files for a formatted example and also see the section Manually configuration files). Note that you can also open these files and change the parameters manually, if you prefer it to setting them in the interface

### 0.5.3 Adjust simulation parameters

With a configuration file loaded (Section 0.5.2) you can change the values in the editable controls. These are detailed below in the Simulation Input section.

### 0.5.4 Start the simulation

Click the button labeled run simulation, and the simulation will start. The buttons text now changes to say cancel simulation, and next to it a new label notes that a simulation is in progress. While the simulation is in progress, the parameter controls are disabled.

### 0.5.5 Running a simulation from a terminal

The module `pgdrivesimulation.py` provides a command line interface to run a simulation. At a Linux or DOS terminal, you can invoke the command with the form,

```
<python> <pgdrivesimulation.py> <options>
```

Where `<python>` is either the `python3` or `python2.7` executable, `<pgdrivesimulation.py>` includes the path to the module (found in the distribution’s main directory), and `<options>` is a list of parameters specified using the option flags. You can see the list of option flags by invoking the command with out any arguments. To see the details for each argument, execute:

```
<python> <pgdrivesimulation.py> -h
```

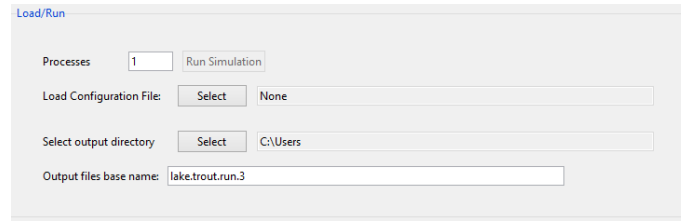


Figure 0.2: Simulation interface, Load/Run section.

Note that there are both required and optional arguments. The latter are only a select number of those offered in the GUI interface (Section 0.6).

## 0.6 Simulation input

The simulation interface is divided into controls inside sub-frames, based on category.

### 0.6.1 The Load/Run section

This section offers parameters related to input and output files (Figure 0.2).

#### 0.6.1.1 Processes

Valid values are between 1 and the total number of available (logical) cores in your computer. Multiple processes are only useful if you have set the Replicates parameter (see the Simulation subframe details below) to a value greater than one.

#### 0.6.1.2 Configuration File

Press the “Select” button next to the label, “Load Configuration File” to load a configuration file into the interface. We have included configuration files for many species. These can be found in the “configuration\_files” subdirectory inside the main program folder.

#### 0.6.1.3 Output directory

Press the select button next to the label, Select output directory, select a folder for the output files written by the simulation

#### 0.6.1.4 Output files base name

You can type in a base name for the simulation output files. The simulation will prepend this to the \*.genepop, \*.conf, \*\_age\_totals.tsv and \*\_nb\_values.tsv output files (Section 0.8).

### 0.6.2 Configuration Info section

This parameter group simply shows you the input file information and has no settable parameters (Figure 0.3).

Figure 0.3: Simulation interface, Configuraton info.

N0 (Newborns)	32
Nb	206
Nb/Nc	0.986
Nb/Ne	0.267
Nb Tolerance	0.05
Ages	32
Female relative fecundity	0.0 0.0 0.0 0.0 0.0 0.0 0.0 0.0 0.0
Male relative fecundity	0.0 0.0 0.0 0.0 0.0 0.0 0.0 0.0 0.0
Force skip	0
Reproductive cycles	100
<input type="checkbox"/> Monogamous	
Litter	None
Probability male birth	0.5
Population size	1505
Female survival	0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85
Male survival	0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85

Figure 0.4: Simulation interface, population section

### 0.6.2.1 Configuration file name

This gives the file name of the loaded configuration file.

### 0.6.2.2 Model name

This gives the name of the model parameterized by the configuration file. In our example configuration files, the model name is usually a species common name. This parameter was used when the program depended on separate life table and configuration files. Currently the configuration files contain all of the life table information needed for the simulations, so that the model name can be any string you find suitable, to indicate, for example, that a given configuration file has parameters similar to another with the same name.

## 0.6.3 The Population section

offers many parameter settings that characterize the populations size and fecundity (Figure 0.4).

### 0.6.3.1 N0 (Newborns)

. This gives the number of newborns added at each simulated reproductive cycle. This value is not editable directly, but is calculated using several values, all of which are editable. These including Nb, Nb/Nc , Female, Male Survival,

and the probability of male birth. The  $N_0$  is recalculated whenever any of these values changes, using the following procedure:

1. Assign an  $N_c$  value, as  $N_b$  divided by  $N_b/N_c$ .
2. Assign a `current_male_proportion` equal to the Probability of male birth.
3. Assign a `current_female_proportion` equal to  $1 - \text{the Probability of male birth}$ .
4. Assign a `cumulative_proportion`=1.
5. For each age value `age_val` giving a male and female survival rate:
  - a Update, `current_male_proportion`=`current_male_proportion` x `male_survival` at `age_val`.
  - b Update, `current_female`=`current_female` x `female_survival` at `age_val`.
  - c Update `cumulative_proportion` =`cumulative_proportion` + `current_male_proportion`.
  - d Update `cumulative_proportion`=`cumulative_proportion` + `current_female_proportion`.
6. Set  $N_0 = N_c / \text{cumulative\_proportion}$ , rounding it to the nearest integer.

#### **0.6.3.2 $N_b/N_c$**

This is the effective number of breeders in one reproductive cycle divided by the census size.

#### **0.6.3.3 $N_b/N_e$**

This is the ratio of the effective number of breeders in one reproductive cycle to the effective population size per generation. This value is not used in the simulation itself, but is written to the output `genepop` file, and can be used in the  $N_b$  estimation interface to make a bias correction in the LDNe estimation of  $N_b$  (see section 0.10.3.3).

#### **0.6.3.4 $N_b$**

This is the target effective number of breeders in the simulated population.

#### **0.6.3.5 $N_b$ Tolerance**

This determines the threshold for allowable “true”  $N_b$  values (see section 0.6.3.4) for simulated populations calculated using the parentage analysis without parents (PWoP) procedure ([4]). For example, if the  $N_b$  is set at 600, and the  $N_b$  Tolerance is set at 0.02, allowable  $N_b$  values would be in the range from 588-612 for simulated populations using PWoP.



**0.6.3.6 Ages**

This value gives the number of age classes for the population to be simulated. Note that this is disabled, and that the length of the lists for Female, Male Fecundity and Female, Male Survival values (see below) are set to length Ages minus one for the former and Ages minus two for the latter. The age value and changes in these lists, therefore, need to be edited in a configuration file (Section 0.7).

**0.6.3.7 Female and Male Survival**

These are lists whose  $i^{th}$  value gives the probability of survival for an individual of the  $i^{th}$  age category.

**0.6.3.8 Female and Male Fecundity**

These are lists whose  $i^{th}$  item gives the probability of reproducing for individuals of the  $i^{th}$  age category.

**0.6.3.9 Force Skip**

This value gives a probability, for each non-zero value,  $f_a$ , in the female fecundity list, that during a given reproductive cycle  $r$  the value will be replaced with zero. Such replacement means that females belonging to the age class  $a$ , given by  $f_a$ , for cycle  $r$ , are infertile. This parameter is set (assigned a non-zero value) in only a few of the configuration files we copied from the AgeStructureNe program, and we have not made it editable in our interface. In your own custom configuration files you can set it to any value 0 through 100 (the value shown in the interface will be the files value divided by 100).

**0.6.3.10 Litter**

If not a “None” value, it will be a list of integers, affecting litter sizes. Note that we do not allow interface editing of these parameters, but note that, as above for the Force Skip setting, you can enter this parameter value in a configuration file. This should be a list, and can have one of 2 valid configurations:

- a The list can have a single value  $l$ , and  $l < 0$ , then at each reproductive cycle the maximum possible number of offspring available to each reproducing female is given by  $l * -1$ .
- b Otherwise, the list should have (positive) integers. In this case these integers proportionally allot litter sizes, as given by their indices in the list. In particular, at each reproductive cycle, as a female is chosen to mate:
  - i An age,  $a$ , is chosen randomly.
  - ii A female  $f_a$  is chosen randomly from the females of age  $a$ .
  - iii A list index  $i$  (i.e. one of 1,2,3, ...  $n$ , where  $n$  is the number of items in the litter list), is selected by weighted probability, proportionally according to the ratio of each list value to the sum of the list values.

Genome

Mutation frequency	0.0
Number of microsatellites	100
Number of SNPs	0
Starting Msat allele total	10

Figure 0.5: Simulation interface, genome section

- iv Female  $f_a$  is then the mother of the next  $i$  offspring (i.e. the female selection steps are skipped for the next  $i$  pairings, since  $f_a$  is the female of the pair). Thus, she will parent the next  $i$  offspring, unless the  $j^{th}$  of her offspring assignments produces the maximum total offspring for the cycle (i.e. N0 is reached), and  $j < i$ .

#### 0.6.3.11 Reproductive cycles

This shows the total number of reproductive cycles that will be simulated.

#### 0.6.3.12 Monogamous

When this box is checked, monogamy is enforced.

#### 0.6.3.13 Probability of male birth

This value is used during reproductive cycles to determine the sex of new individuals. As noted above in the description of the N0 (see 0.6.3.1), it is used also in the N0 calculation, and so the latter is recalculated when this value is changed. When the Cull method (0.6.5.1) is set to `equal_sex_ratio`, this parameter is automatically set to 0.5, and its entry box is disabled.

#### 0.6.3.14 Population size

This value determines the number of individuals that will be created in the simulations initial population. In subsequent cycles, the size will change according to the reproductive parameters, notably N0 (0.6.3.1).

### 0.6.4 The Genome section

Parameters in the Genome section determine the simulated individuals allelic content (Figure 0.5).

#### 0.6.4.1 Mutation frequency

If non-zero, this value is applied to microsatellites (not to SNPs). It will be used to set the simuPOP simulation `StepwiseMutaters` rate parameter.

Figure 0.6: Simulation interface, genome section

#### 0.6.4.2 Number of microsatellites

Microsatellites are simulated as diploid. Note that in cases in which you specify both  $n$  microsatellites and  $m$  SNPs, in the output genepop file, the first  $n$  loci are the microsatellites and the last  $m$  loci are the SNPs.

#### 0.6.4.3 Number of SNPs

SNPs are simulated as diploid.

#### 0.6.4.4 Starting Msat allele total

This value gives the initial number of microsatellite alleles for each microsatellite in the initial population. For each microsatellite, the initial genotype frequencies are drawn from the Dirichlet distribution. With the number of microsatellites set to 10, for example, each microsatellite will have 10 alleles with frequencies given by the Dirichlet distribution of order 10, with alphas uniformly set to 1.0. The maximum allowed number of alleles is 100.

### 0.6.5 The Simulation section

These parameters determine several per-cycle behaviours (Figure 0.6).

#### 0.6.5.1 Cull method

Cull method indicates one of two possible per-cycle methods of removing individual from the population.

1. Survival rates. Individuals of age greater than zero are removed from the population by comparing a random number against the probability for that individuals age and sex, in the survival list (Section 0.6.3.7).
2. Equal sex ratio. For each cohort with age  $a, a > 0$ , (i.e. excepting newborns), divide the individuals by sex,  $s$ , into two lists. From each list with  $t_s$  individuals, randomly cull  $n_s$  individuals from each list where

$$n_s = \text{floor}(p_s), \text{ where } p_s = (t_s(1 - \text{survival}_s[a]))$$

and  $survival_s[a]$  is the survival rate (Section 0.6.3.7) for individuals of age  $a$  and sex  $s$ . One more individual from each is culled by the probability given by the fractional part of  $p_s$ .

#### 0.6.5.2 Filter recorded pops by heterozygosity

When checked, the genepop file output will be restricted to the populations as filtered using the Heterozygosity filter parameters. See section 0.6.5.3.

#### 0.6.5.3 Het filter parameters

If the Het filter checkbox is checked, apply a filter to each pop of the form  $m, x, t$ , where,

- Mean expected heterozygosity is calculated as

$$mean(H_{L_1}, H_{L_2}, H_{L_3} \dots H_{L_N})$$

for  $N$  loci, and

$$H_{L_i} = 1 - \sum_{j=1}^n freq(a_j)^2$$

and  $freq(a_j)$  is the frequency of the  $j^{th}$  of the  $n$  alleles of loci  $L_i$ .

- $m$  is the minimum mean expected heterozygosity,
- $x$  is the maximum mean het, and
- $t$  is the total number of populations to record.

The output genepop file will then record only populations whose mean Het falls inside the range. Further, it will stop recording them once  $t$  populations are recorded (or the last cycle is completed, if  $t$  pops have not satisfied the Het range limits).

#### 0.6.5.4 Nb and census adjustment

This parameter offers one or more specifications that will change the target Nb and the number of individuals in the population by a fixed rate and at a range of cycles (one or more). Entries are of the form  $min - max : rate$ , specifying a change in Nb and census size applied at cycle numbers  $min$  through  $max$ . The values conform to  $min \leq 2 \leq max$ , and  $rate \geq 0.0$ . No adjustment is made with  $rate = 0.0$ . For example, to reduce the Nb and the total number of individuals by a tenth at cycle 3 (remaining in effect for the remaining cycles, unless another adjustment is added to the list), you would edit the entry to read, 3-3:0.1. The adjustments are different, depending whether  $rate$  is less than or greater than 1.0:

- If  $rate$  is less than 1.0, the target Nb value, and each age class in the current census is reduced by the proportion given by  $rate$ . Note that the change in Nb will result in a change to N0 as described above the in section 0.6.3.1.
- If  $rate > 1.0$ , the target Nb value will be multiplied by  $rate$ , with a resulting recalculation of N0. No change will be made to the current census.

### 0.6.5.5 Replicates

This value sets the number of independent simulations run with the current parameter set. These can be run in parallel if you specify more than one process in the Processes parameter.

### 0.6.5.6 Skip breeding probability

If this value is not set to “None,” it should be a list of percentages. It effects the number of available females of a given age at a given cycle number  $c$ . The  $i^{th}percentp_i$  gives the probability ( $p/100$ ) that a female of  $age = i$ , is not able to breed in cycle  $c$ . Like the Litter (Section 0.6.3.10) and Force Skip parameters (Section 0.6.3.9), this parameter is not settable in the interface, but can be included in your configuration file.

### 0.6.5.7 Cycles of burn-in

This integer  $n$  is valid in the range  $1 \leq n \leq r$ , with  $r$  giving the total reproductive cycles (Section 0.6.3.11). This value tells simulation that the Nb tolerance test (section 0.6.3.5) should not be performed for the first  $n$  cycles. The default value for this parameter equals the number of Ages in the model to allow any individuals of the initial population to cycle out of the population.

### 0.6.5.8 Start recording at cycle

This integer value  $c$  will result in the genepop file containing only the populations of cycles  $c$  through  $r$ , where  $r = \text{total Reproductive cycles}$ . This can greatly reduce the size of the output genepop file, when you are interested only in the last  $r - c$  cycles, but want to simulate many cycles before recording, and when you have large populations and many loci to simulate.

## 0.7 Manually editing configuration files

Our program provides a collection of simulation configuration files available at <https://github.com/popgengui/agestrucne/tree/data>. You can directly edit a \*.conf file, which can sometimes be more convenient than using the GUI interface to change parameter values. If you open one of the supplied configuration files in the subdirectory, “configuration\_files,” youll see the parameter=value pairs, each below a section header inside square brackets. Some of the parameter names differ from their functional equivalents in the interface (see Table 0.1 ).

## 0.8 Simulation output

When a simulation is complete the message “simulation in progress” will disappear from the interface and editable entry boxes will no longer be grayed-out. A completed simulation delivers a genepop file for each replicate, named using the Output files base name parameter shown in the The Load/Run section of the simulation input (Section 0.6). The base name is extended with a replicate number and a “genepop” extension, so that, for example, if your simulation output base name is bulltrout, and you specified 3 replicates, the output file for the

Short name in file	Interface Name	Link to description
NbNc	Nb/Nc	(Section 0.6.3.2)
NbNe	Nb/Ne	(Section 0.6.3.3)
N0	N0 (Newborns)	(Section 0.6.3.1)
Nb	Nb	Set to “None” value(Section 0.6.3.4)
NbVar	Nb tolerance	(Section 0.6.3.5)
ages	Ages	(Section 0.6.3.6)
dataDir	Ignored	Ignored
doNegBinom	Not documented	Use “False.” Feature not documented.
fecundityFemale	Female relative fecundity	(Section 0.6.3.8)
fecundityMale	Male relative fecundity	(Section 0.6.3.8)
forceSkip	Force skip	(Section 0.6.3.9)
gens	Reproductive cycles	(Section 0.6.3.11)
isMonog	Monogamous	(Section 0.6.3.12)
lbd	Ignored	Ignored
litter	Proportional Litter Sizes	(Section 0.6.3.10)
maleProb	Probability male birth	(Section 0.6.3.13)
model_name	Model	(Section 0.6.2.2)
numSNPs	Number of SNPs	(Section 0.6.4.3)
mutFreq	Mutation frequency	(Section 0.6.4.1)
numMSats	Number of microsatellites	(Section 0.6.4.2)
popSize	Population size	(Section 0.6.3.14)
reps	Replicates	(Section 0.6.5.5)
skip	Skip breeding probability	(Section 0.6.5.6)
startAlleles	Starting Msat allele total	(Section 0.6.4.4)
startLambda	Cycles of burn-in	(Section 0.6.5.7)
startSave	Start recording at cycle	(Section 0.6.5.8)
survivalFemale	Female survival	(Section 0.6.3.7)
survivalMale	Male survival	(Section 0.6.3.7)
gammaAFemale	Not documented	Ignored when neg. binom. is False
gammaBFemale	Not documented	Ignored when neg. binom. is False
gammaAMale	Not documented	Ignored when neg. binom. is False
gammaBMale	Not documented	Ignored when neg. binom. is False
cull_method	Cull method	(Section 0.6.5.1)
nbadjustment	Nb and census adjustment	(Section 0.6.5.4)
do_het_filter	Filter recorded pops by heterozygosity	(Section 0.6.5.2)
het_filter	Het filter parameters	(Section 0.6.5.3)

Table 0.1: Short param names as found in files, and their descriptions

3rd replicate would be named “bulltrout.r3.genepop”. Also, there are three files produced during the first replicate only, all prefixed with the output base name. One with the extension “conf”, lists the parameter settings for the simulation (and, hence, for all replicates), another has extension “\_age.counts\_by\_gen.tsv,” and a third file has extension “\_nb.values\_calc\_by\_gen.tsv”. Details on the output files follow.

1. The conf file shows the parameter settings used in the simulation (except the number of replicates, which it always sets to one). This file can be loaded into another instance of the Simulation Input (see the Load/Run parameter) and another simulation with matching parameters can be run. Conveniently, if it represents many customized settings on a former configuration file, small changes to it can be made to run a simulation similar, but without having to re-enter all of the settings used to create it.

2. The age counts file is a table with tab-delimited fields that gives a count of total individuals for each age class, for each reproductive cycle. The first line in the file gives column headers, the first “generation,” referring to reproductive cycle number, a zero-based count of reproductive cycles, and the rest listing age classes simply as 1,2,3t, t = total age classes. This file is created only for the first simulation replicate.
3. The Nb values file is a table with tab-delimited fields giving the PWoP Nb values calculated during the simulation, and used to compare to the target Nb value +/- the Nb Tolerance value. The first column gives the zero based reproductive cycle number and the second the PWoP-based Nb value that passed the tolerance test, and represents the accepted population for that cycle. This file is created only for the first simulation replicate.
4. The genepop file conforms to the genepop file standards given at [http://genepop.curtin.edu.au/help\\_input.html](http://genepop.curtin.edu.au/help_input.html). The header line notes the name of the \*.gen file it came from, which simply names an intermediate file from which it derived its population information. It also gives the value of Nb/Ne. If the value is non-zero, it can be loaded automatically into the Nb/Ne estimation interface (see the Parameters section 0.10.3 of the Nb/Ne Estimation interface description). The second line of the genepop file gives the name of the first loci, which is simply its ordinal, “ $l_0$ .” Each consecutive loci,  $l_0, l_1, l_2 \dots l_{L-1}$  (where  $L$  gives the total number of microsatellites plus the total number of SNPs) is listed on a separate line. Note that the first  $M$  loci will represent the microsatellites, and the last  $S$  loci will represent the SNPs, with  $M$  and  $S$  the totals given in the genome parameters (Section 0.6.4) of the Simulation Input. Thereafter the file consists of separate “pop” sections, each representing a reproductive cycle. The first  $n$  cycles (as numbered 1, 2, 3... $n$ ) will not be in the file if the Start at cycle number parameter is set to  $n + 1$ . The population for each cycle is listed, in order of cycle number. Each is demarked by a line with “pop” as its sole entry. Individuals, one to a line, follow each “pop” entry. Each individual has an ID with multiple fields delimited by a semicolon, giving, individual id number;sex (1 = male, 2 = female);id of father;id of mother;age class. These are followed by a comma, and then a space-delimited set of alleles for each locus named in the lines 2 total number of loci. Note that these allele entries represent diploidy, and use 3-digit allele numbering so that, for each loci, allele one is named by the first 3 digits, and allele 2 by the last 3.

## 0.9 Running an Nb or Ne Estimation

The Nb (and Ne) estimation interface performs an LD based Nb or Ne estimation from genepop file input as supplied by the user. While it was developed in concert with the simuPOP-based simulation output from our programs interface, it will perform estimations on any genepop file input. To run estimations:

- a Load a new Nb interface with the add menu’s “Add new Nb estimation” option (Figure 0.1) and set the parameters with the provided controls. For details see Section 0.10

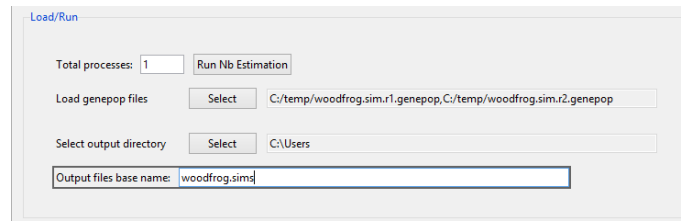


Figure 0.7: Nb/Ne estimation interface, Load/Run section

- b Load a genepop file..
- c Adjust the estimation parameters. The parameters are detailed in Section 0.10
- d Click the button labeled “Run Nb Estimation”, and the computations will start. The buttons text now changes to say “cancel simulation,” and next to it a new label will note that “estimations in progress.” As in the other interfaces, while the estimations are in progress, the parameter controls are disabled.

### 0.9.1 Running an Nb or Ne estimation from the command line

The module `pgdriveneestimator.py` offers a command line interface for running LDNe2 estimations. At a Linux or DOS terminal you can type a command of the form,

```
<python> <pgdriveneestimator.py> <options>
```

where `<python>` is the `python3` or `python2.7` executable, `<python>` gives the path to the module (in the distributions main directory), and `<options>` is a list of parameters given using the option flags. To see the list of options, invoke the command without any options. To see a detailed list of options, invoke the command with,

```
<python> <pgdriveneestimator.py> -h
```

## 0.10 Nb/Ne Estimations input

The interface provides for multiple subsampling schemes of both individuals and loci within the input genepop file pop sections. The sub-sections of the interface follow.

### 0.10.1 The Load/Run section

This section (Figure 0.7) offers an interface to load input and name the output files.



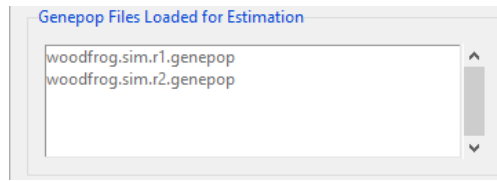


Figure 0.8: Nb/Ne estimation interface, genepop files loaded section

#### 0.10.1.1 Total processes

The program will run estimations on the individual “pop” sections in parallel using the number of processes set here. It is usually advisable, unless your computer has many process already running, to use most if not all of your available (virtual) processing cores, to speed up the estimation run parameter. The program defaults to using half of the available cpu’s (or virtual cores) available on your machine.

#### 0.10.1.2 Load genepop files button

Clicking on this button produces a file loading interface to locate and load one or more genepop file(s). Note that when you load multiple genepop files, the parameter settings will be applied to all. In particular, activating an Nb bias adjustment will apply it to all the files, so that only data with which it is compatible should be loaded. This also applies to other parameters, such as the population number range and loci number range parameters (Sections 0.10.4.1, 0.10.4.2, 0.10.5.1, and 0.10.5.2).

#### 0.10.1.3 Select output directory

By clicking on the button and choosing your preferred folder you select where the estimation output files will be written. Note that this will also be used as a temporary directory in which intermediate files will be written inside new directories with the “tmp” prefix, ending in random characters. These files will be removed on completion of the simulation. Sometimes, if the estimation run is cancelled or otherwise is interrupted, they will not be removed, but can be manually deleted from your directory.

#### 0.10.1.4 Output files base name

The text entered here will become the prefix for the output files (Section 0.11).

### 0.10.2 The Genepop Files Loaded section

This section has a single box that shows you the names of the loaded genepop files (Figure 0.8). It is not an editable section.

### 0.10.3 The Parameters section

This section supplies the main parameters, including the choice of subsampling in pop sections and/or loci (Figure 0.9).

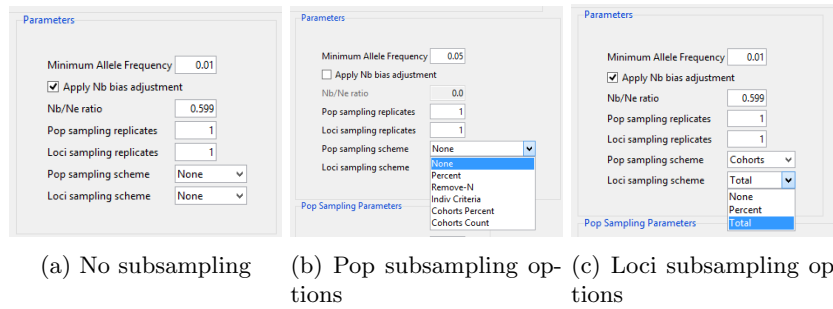


Figure 0.9: Nb/Ne estimations interface, parameters section

### 0.10.3.1 Minimum allele frequency

This value sets the threshold below which the LDNe program will ignore an allele in its LDNe calculation. The interface defaults to 0.05, as a value that is commonly used, and which reduces upward bias [3]

### 0.10.3.2 Nb bias adjustment check box

Checking this allows you to apply a bias adjustment to the estimations [3]. See Section 0.10.3.3.

### 0.10.3.3 Nb/Ne ratio

This parameter is the value used for the bias adjustment, when it is checked. A zero value or an un-checked box means no bias adjustment will be done. Note that when you load a genepop file generated by the simulation interface, and you check the box labeled “applyNb bias adjustment”, the program will load the Nb/Ne value as set in the simulation interface. You can accept it or enter another value. If no value is available in the genepop file, then you will need to enter a non-zero value to make any bias adjustment.

### 0.10.3.4 Pop sampling replicates

When set to an integer  $n$ , for each estimate, for each pop sampling parameter, the estimate is repeated  $n$  times. While you can set this to any value for any subsampling scheme, note that it is sensible only when your subsampling parameter involves a random sample of individuals (Section 0.10.3.6) more than 1 loci subsampling parameter subsampling schemes (Section 0.10.3.7), or both. If there is no random subsampling, the replicates will be performed, with identical results.

### 0.10.3.5 Loci sampling replicates.

When set to  $n$ , for each pop sampling replicate, for each loci subsampling parameter value, the estimate is repeated  $n$  times. As with the Pop sampling replicates parameter, if the loci subsampling scheme has no randomized subsample (Section 0.10.3.7), then the estimates will be identical.

### 0.10.3.6 Pop sampling scheme

This drop-down box offers the following subsampling schemes (Figure 0.9b).

1. None. This scheme uses all individuals with the pop section for the Nb or Ne estimation, unless the value for Indiv max per pop reduces the pop size to  $m$  randomly selected individuals. If the pop section has fewer individuals than the value given by Indiv min per pop, the pop section is skipped, with a message written to the Messages file (Figure 0.10a).
2. Percent. When you select this scheme the pop sampling interface (Section 0.10.4) shows a percent box with two buttons below it (Figure 0.10b), “Add Value,” and “Trim.” You can edit any box currently in the list. Clicking “Add Value” will append a box to the list, its default value taken from its nearest neighbor. For each percentage  $p$  in the list, each pop section will be reduced to  $p$  percent of its total individuals, unless its census is not in the pop number range (Sections 0.10.4.1 and 0.10.4.2), in which case the population will be skipped, and a message written to the messages file (Section 0.11.1). For each subsample the individuals are randomly selected. Note that any multiple loci subsampling values and/or Pop sampling replicates and Loci sampling replicates will result in an estimate for each percentage value, repeated for each of the loci subsampling and replicate values.
3. Remove-N. This scheme also offers an editable list, which behaves as described for the Percent scheme (Figure 0.10c). For this scheme, when you enter an integer  $N$ , the pop subsample will be that given by its total minus  $N$ , the removed  $N$  individuals randomly selected except in the case of  $N = 1$ , in which case each of the *remove* – 1 cases will be estimated (i.e. there will be  $t$  estimations for a pop section with  $t$  individuals. Pop sections are skipped if the total individuals in the population are not in the pop number range (see Sections 0.10.4.1, and 0.10.4.2). Note, as with the Percent scheme, estimates for each  $N$  value will be repeated for loci sample values, pop sampling replicates, and Loci sampling replicates.
4. Cohorts Percent. Because this scheme selects individuals by age, it requires input genepop files produced by the Simulation output (Section 0.8). If your input is empirical data and you wish to calculate Nb, it is assumed that each pop in the genepop file is a single cohort and all individuals should be used in the Nb calculation. When Cohorts is selected the interface shows, besides its usual pop number and min/max pop size parameters, an entry labelled “Indiv max age” (Figure 0.10d). For each population, individuals outside the ages given by  $[0.0, \text{max age}]$  are excluded. However, because the simulations produce genepop files with no individuals aged less than 1.0, the interval is effectively  $[1.0, \text{max age}]$ . For each pop, and for each percentage value in the list, subsampling steps are:
  - a. For each age value in  $[0.0, \text{maxage}]$ , count the total individuals  $t_a$  in the age group  $a$ .
  - b. Find the smallest of the age group totals,  $t_{\text{smallest}}$ .

- c. Randomly subsample  $t_{smallest}$  individuals from each age group to get a total sample size  $s$ .
  - d.  $s$  is in the range [Indiv min per pop, Indiv max per pop] then, for each percentage  $p$  in the percentage list, randomly select  $p$  percent of the collected individuals. If  $s$  is outside the range an error occurs and the analysis is terminated.
5. Cohorts Count. This scheme behaves very similarly to the Cohorts Percent (above, 4), with nearly identical parameters (Figure 0.10e). However, instead of sampling percentages, for each sample value  $c_i$  in the list of sampling values,  $c_i$  individuals will be randomly selected from each age class. If for any pop section, any of the age classes has a count of individuals less than  $c_i$ , an error is raised and the estimations terminate.
  6. Individual Criteria. This scheme allows you to select a range of cohorts by contiguous age group. Like the cohorts schemes, it requires your input to be genepop files produced by the Simulation output. When Individual Criteria is selected the interface shows, besides its the pop number and min/max pop size parameters, two entries for a minimum and maximum age (Figure 0.10f). The program pools all individuals within the minimum and maximum age range (inclusive), and calculates the estimation using the pool. If the total of pooled individuals is outside the range given by [Indiv min per pop Indiv max per pop], an error is thrown and the analysis is terminated.

#### 0.10.3.7 Loci sampling scheme

1. None. All loci will be used in the estimations, from the  $i^{th}$  to the  $j^{th}$ ,  $i$  and  $j$  given by the Loci number start and Loci number end (Figure 0.11a). If the Max Loci count  $m$  is less than the total loci, then  $m$  loci will be randomly selected.
2. Percent. For each percentage  $p$  listed in the “percentages” boxes (Figure 0.11b), an estimation is calculated using a random selection of  $p$  percent of the loci from those within the range given by Loci number start to Loci number end.
3. Total. For each total  $t$  listed in the “totals” boxes (Figure 0.11c), an estimation is calculated using a random of  $t$  loci from those within the range given by Loci number start to Loci number end.

#### 0.10.4 Pop sampling parameters section

In this section (Figure 0.10), you set the pop section sampling parameters, which are presented according to the scheme selected in the Pop sampling scheme parameter.

##### 0.10.4.1 Pop number start

When this is set to integer  $n$ , the estimates will skip pop section numbers (as ordered in the genepop file) in the range  $[1, n - 1]$ .

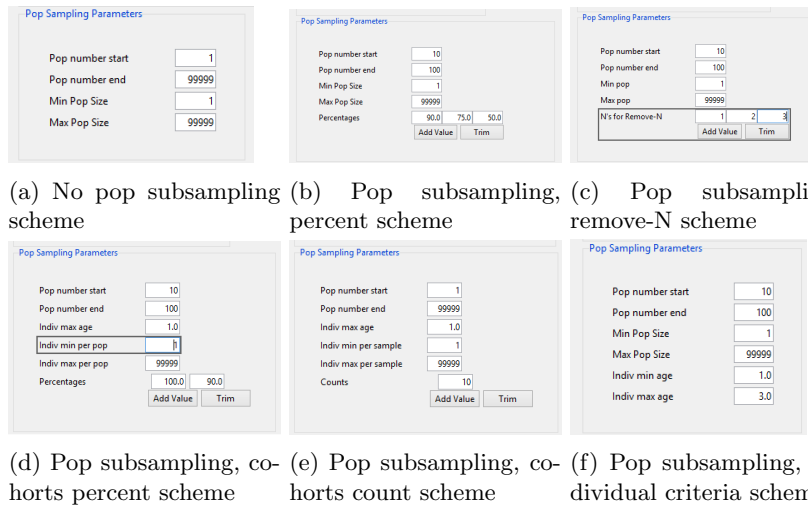


Figure 0.10: Nb/Ne estimations interface, pop subsampling parameters

#### 0.10.4.2 Pop number end

If the genepop file has  $t$  total pop sections, then, when this parameter is set to integer  $n$ , the estimates will skip pop section numbers in the range  $[n + 1, t]$ .

#### 0.10.4.3 Indiv min per pop

For the None, Percent, and Remove-N sampling schemes, a pop section must contain at least this many individuals, or it will be skipped. Skipped populations will be noted in the Messages file. For the Cohorts and Individual Criteria sampling schemes, the total cohort sample must meet or exceed this minimum, or an error occurs and the estimation run is terminated.

#### 0.10.4.4 Indiv max per pop

For the None sampling scheme, if total individuals in a pop section exceed this value  $i_{\max}$ , then  $i_{\max}$  individuals will be randomly selected from the pop. For the Percent and Remove-N schemes, pop sections with individuals totaling more than this value will be skipped, with a message written to the Messages file. For the Cohorts scheme, if sample size exceeds this value, then an error occurs and the estimation run is terminated; for the Individual Criteria scheme, the sample is reduced to the value by random selection of individuals.

#### 0.10.4.5 Scheme-specific parameters

See the descriptions in Section 0.10.3.6.

### 0.10.5 Loci sampling parameters section

In this part of the interface you can limit the loci used to determine the Nb or Ne estimation.

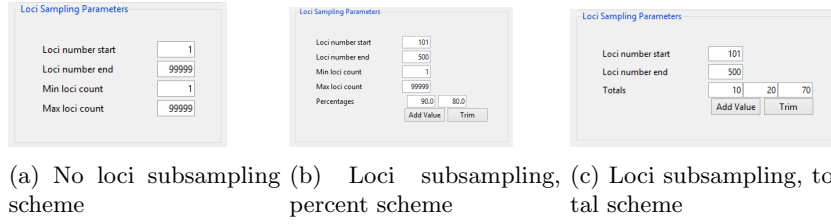


Figure 0.11: Nb/Ne estimations interface, loci subsampling parameters

#### 0.10.5.1 Loci number start

Loci used in the estimation will be drawn from the  $i^{th}$  to the  $j^{th}$  Loci as ordered in the genepop file. This value gives the  $i^{th}$ . Note that this range allows you, for example, to run a simulation in which the first  $m$  loci are microsatellites and the last  $s$  loci are SNPs, and then use the same genepop file in two different estimations, one based on microsatellites using loci range 1 to  $m$ , and the second based on SNPs using range loci  $m + 1$  to  $m + s$ .

#### 0.10.5.2 Loci number end

This value gives the  $j^{th}$  loci in the range as described for Loci number start.

#### 0.10.5.3 Min Loci count

This value sets a minimum for the total loci to be used in the estimation. If the loci sample in the genepop file is less than this value, the program generates an error message and the run is terminated.

#### 0.10.5.4 Max Loci count

This sets a maximum value  $m$  on the number of loci to be used in the estimation. If the range gives a larger total, then  $m$  loci will be randomly selected from those in the range, Loci number start to Loci number end.

#### 0.10.5.5 Scheme specific parameters

See the descriptions in Section 0.10.3.7.

### 0.11 Nb estimation output

#### 0.11.1 Messages file

With extension \*.msgs, this file shows the parameter settings used in the estimations, and also logs error messages.

#### 0.11.2 Estimates table

With extension \*.tsv, this file gives tab-delimited quantities associated with the Nb or Ne estimations (Table 0.2)

Output file column	Description
original_file	Source genepop (gp) file from which the pop sections are derived.
pop	The $i^{th}$ reproductive cycle as ordered in the source gp file.
census	The total individuals in the pop.
indiv_count	The total individuals used in the estimation.
sample_value	The parameter, if any, used in the pop subsampling (e.g. percent).
replicate_number	The pop replicate number.
loci_sample_value	The parameter value, if any, used in the loci subsampling (e.g. percent).
loci_replicate_number	The loci replicate number.
min_allele_freq	An LDNe2 parameter [no citation yet for LDNe2], the min allele frequency required to include an allele in the estimation.
est_type	Refers to the estimation type, currently only implemented for “ld”, that is, the estimation is based on the LDNe method.
est_ne	The Ne or Nb estimation [no citation].
95ci_low	The lower of a 95% confidence interval for the estimate, based on the jackknife method described in [no citation yet for LDNe2].
95ci_high	The upper 95% confidence interval for the estimate, based on the jackknife method described in [no citation yet for LDNe2].
overall_rsquared	A weighted mean of the allelic pairwise estimators $\hat{r}_\Delta$ (see [no citation yet for LDNe2]).
expected_rsquared	The expected value of the estimator $\hat{r}_\Delta$ (see [no citation yet for LDNe2]).
indep_comparisons	The number of independent comparisons used to determine the estimate (see [no citation yet for LDNe2]).
harmon_mean_samp_size	Sample size to calculate the LDNe estimate ([no citation yet for LDNe2]).
alt_ci_low	Lower, parametric 95% confidence interval ([no citation yet for LDNe2]).
alt_ci_high	Upper, parametric 95% confidence interval ([no citation yet for LDNe2]).
nbne	If not “None,” then this is the Nb/Ne ratio used in a bias adjustment for the estimate (see [2]).
ne_est_adj	The estimate as bias-adjusted (see [2]). If no bias adjustment was calculated, then this value will be identical to that in the est_ne column.
mean_het	The mean expected heterozygosity of the population, calculated as defined in Section 0.6.5.3. The calculation employs all individuals in the pop (no subsampling is applied), and only the loci with the range given by the start loci number parameter (Section 0.10.5.1) and the end loci number parameter (Section 0.10.5.2).

Table 0.2: Nb or Ne estimation output values

## 0.12 Visualization Interfaces

The program offers 3 interfaces that plot the Ne/Nb estimations as output by the program’s LDNe output (Section 0.11).

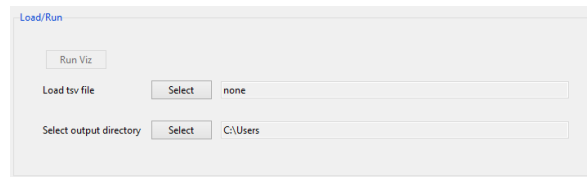


Figure 0.12: The viz interface, load/run section.

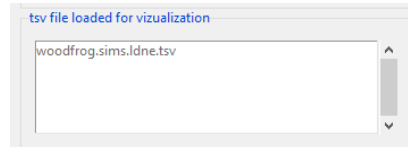


Figure 0.13: The viz interface, tsv file loaded section.

### 0.12.1 The plotting interface

This interface will be deprecated as its functionality is replaced by the boxplot (Section 0.12.2) and regression (Section 0.12.3) interfaces. Currently it is the only vizualization interface that allows customized axis labelling and output giving outlier information. However, note that it does not allow customized data grouping, and plots subsample groups separately by population, so that you will create many plots if you have many populations in your input genepop file(s). It currently is loaded by clicking “Add” on the program’s main menu (Figure 0.1), and selecting “Add new plotting interface.”

#### 0.12.1.1 Load/Run section

This section of the viz interface (Figure 0.12) has buttons that prompt for input and output file information. Note that this interface is to be deprecated after more complete versions of the boxplot and regression plot interfaces are complete.

**Load tsv file** This button prompts for a \*tsv file as output by the Ne estimation program (Section 0.11.2).

**Load output directory** This button prompts for the name of a directory to which viz output files will be written.

#### 0.12.1.2 Tsv file loaded section

This read-only interface shows the name of the loaded tsv file (Section 0.13).

#### 0.12.1.3 Viz type section

This section (Figure 0.14) offers a choice of plot types of the estimates (as extracted from the \*tsv files adjusted estimate column “ne\_est\_adj” (Table 0.2).



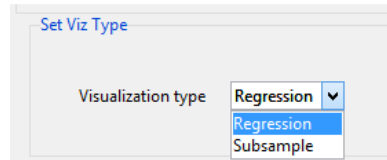


Figure 0.14: The viz interface, plot type selection section.

### Two types of plotting are offered:

- Regression. This type of visualization produces a series of three images:
  1. Regression plot. Using the python scipy stats package, this plot shows regression lines, one for each replicate in the source \*tsv file, of estimates vs population number. [Need a more thorough description here.]
  2. Box plot. This plot shows a box and whisker plot of the distribution of the  $r$  estimates for each population, given  $r$  simulation replicates.
  3. Scatter plot. This plot shows the same distribution as that shown in the box plot, but only as a column of points, one for each estimate of the replicates.

It also produces 2 text files (with default names, “stats\_out.txt” and “stats\_out.outliers.txt”

- stats out file. This file lists summary and individual slope values and individual confidence intervals for the plotted replicates.
- stats outliers file. This file lists the outlier values that were not plotted.

Note that as indicated in the (Figure 0.17) interface, if there are multiple pop and/or loci subsample values (for example the pop “percent” scheme described in section 0.10.3.6), then you must select a single pop and a single loci subsample value for which to produce the plots.

- Subsample. This type of visualization is tailored to Nb or Ne estimation output files with relatively few populations and replicates, but with many subsampling values( see, for example, the “percent” subsampling schemes described in Sections 0.10.3.6 and 0.10.3.7). It produces one image for each replicate/population combination in the loaded \*tsv file. Each image shows a box and whisker plot for each combination of pop/loci subsample value for the given population/replicate combination.

#### 0.12.1.4 Regression plotting section

In this section (Figure 0.17) of the Viz interface you set the parameters for the regression series of images.

(a) Selecting a pop subsample value. (b) Selecting a loci subsample value.

Figure 0.15: The viz interface, regression plotting section.

**Pop subsample value** This selection only appears when the loaded \*.tsv file has entries with more than one pop subsample value (e.g. the “percent” scheme for pop subsampling, described in Section 0.10.3.6). The selected value here results in plotting based only on the entries with a matching value in the tsv column, “sample\_value” (Table 0.2).

**Loci subsample value** This selection only appears when the loaded \*.tsv file has entries with more than one loci subsample value (e.g. the “percent” scheme for loci subsampling, described in Section 0.10.3.7). The selected value here results in plotting based only on the entries with a matching value in the tsv column “loci\_sample\_value” 0.2.

**Plot title** All plots will show this as the title.

**x label** All plots will show this as the label for the x axis.

**y label** All plots will show this as the label for the y axis.

**Regression file, Boxplot file, Scatterplot file** For each plot image (Section 0.12.1.3) you can select one of the drop-down options or enter text to do the following:

- Show. Select this and the image will be shown on your screen.
- none. Select this and the image will not be produced.
- Enter text with a \*.png or \*.pdf extension, and the plot image will be saved as the corresponding image type, named as entered.
- Enter text without a \*.png or \*.pdf extension, and the plot image will be saved as a \*.png image, with the text entered as the prefix.

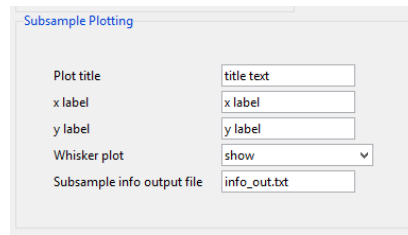


Figure 0.16: The viz interface, subsample plotting interface.

**Stats output file** This entry names stats-out and outliers text files (Section 0.12.1.3).

#### 0.12.1.5 Subsample plotting section

In this section (Figure 0.16) of the Viz interface you set the parameters for plotting subsamples (pop and loci) when the same input file and pop number have multiple estimations. Note that the number of plots generated for this series equals the number of files used times the number of pops in the input input. Each whisker plot shows the distribution of estimations for the subsample values indicated on the x-axis labels.

(Table 0.2).

**Plot title** All plots will show this as the title.

**x label** All plots will show this as the label for the x axis.

**y label** All plots will show this as the label for the y axis.

**Whisker plot** The plot will be shown or saved as a file according to the following:

- Show. Select this and the image will be shown on your screen.
- none. Select this option and enter text according to the following.
  - Enter text with a \*.png or \*.pdf extension, and the plot image will be saved as the corresponding image type, named as entered.
  - Enter text without a \*.png or \*.pdf extension, and the plot image will be saved as a \*.png image, with the text entered as the prefix.

### 0.12.2 Boxplot Interface

This interface is available by clicking on the “Add” menu in the main menu (Figure 0.1), then clicking “Add new boxplot interface.” After using the Button labeled “Select to load a \*.tsv file (Section 0.11), you can group and filter input values to get a boxplot of the LDNe Nb/Ne estimations according to the groups and filters (Figure 0.17a).

### 0.12.2.1 Grouping.

The interface offers four Group-by fields allowing you to group the data using up to four value sets according to the input fields as given in the Ne/Nb estimation output (See Table 0.2):

- genepop file from original\_file column in the Ne/Nb estimation output tsv file.
- pop number, from the pop column.
- total indiv. sampled, from the indiv\_count column.
- pop subsample value, from the sample\_value column.
- pop sampling replicate, from the replicate\_number column.
- loci subsample value, from the loci\_sample\_value column.
- loci sampling replicate, from the loci\_replicate\_number column.

### 0.12.2.2 Filtering.

It also allows you to set one of the set of values for several input fields, so that output entries without that value will be excluded from the plot data. For example, if you have output from a run in which you set a percentage subsampling scheme for pops (Section 0.10.3.6), and you sampled by 50, 80, and 100% , you can select one of the percentages, and all entries used to make the boxplot will be those with the selected pop subsampling value.

### 0.12.2.3 Y-axis field.

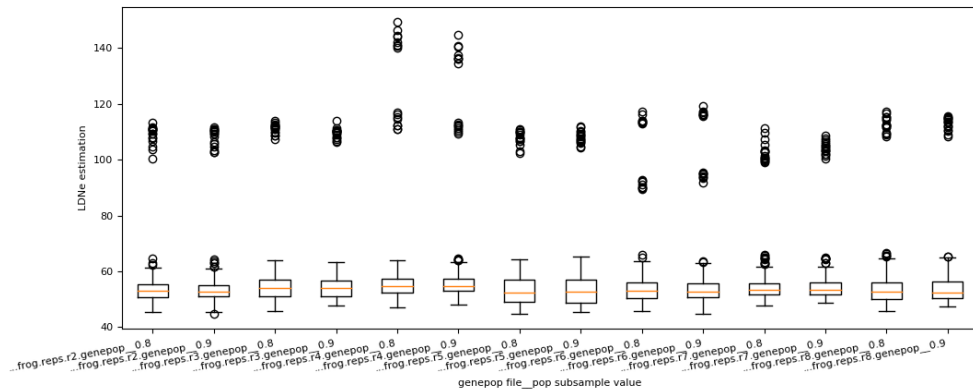
You can select the values for the y-axis field using the “Y-axis values” drop-down box. The following are the available output fields in the LDNe Nb/Ne estimation output (See Table 0.2):

- LDNe estimation, from the est\_ne column.
- low 95% CI, from the 95ci\_low column.
- high 95% CI, from the 95ci\_high column.
- adjusted LDNe estimation, from the ne\_est\_adj column.

### 0.12.2.4 Y-axis value limits.

You can restrict the range of Y values plotted using the sliders or their associated entry boxes. Note that in all cases values of Infinity, and “NA,” which can be present in the output fields, have been excluded.

(a) The boxplot interface, grouping on two input fields, genepop file and pop subsample value.



(b) The resulting boxplot.

Figure 0.17: The LDNe Ne/Nb boxplot interface.

### 0.12.2.5 The plot.

The plots produced (Figure 0.17b) result from a call to the matplotlib “boxplot” command, with its “x” parameter set to the grouped/filtered data, and the “labels argument” set to show the grouped field values (separated by double underscores). Otherwise, arguments to boxplot are not specified, and so use default arguments. You can use the “Save” button in the “Save Plot” subframe to save the plot to file. A \*.png image format will result if you use any file name, except when you add “.pdf,” which will then be the format of the saved file image. png and pdf are the only formats used.

## 0.12.3 Regression Interface

This interface is available by clicking on the “Add” menu in main menu (Figure 0.1), then clicking “Add new regress plot interface.” After loading a LDNe Nb/Ne estimation output file (Section 0.11.2), you can plot regression lines (Figures 0.18 and 0.19).

Estimation Regression Plots 0

\*tsv file

Load tsv file  C:/Users/ted/Documents/source\_code/python/neGui/temp\_test/frogs.estimate

Data Filters

genepop file  pop sampling replicate  loci sampling replicate  pop subsample value  loci subsample value

Y-axis Data

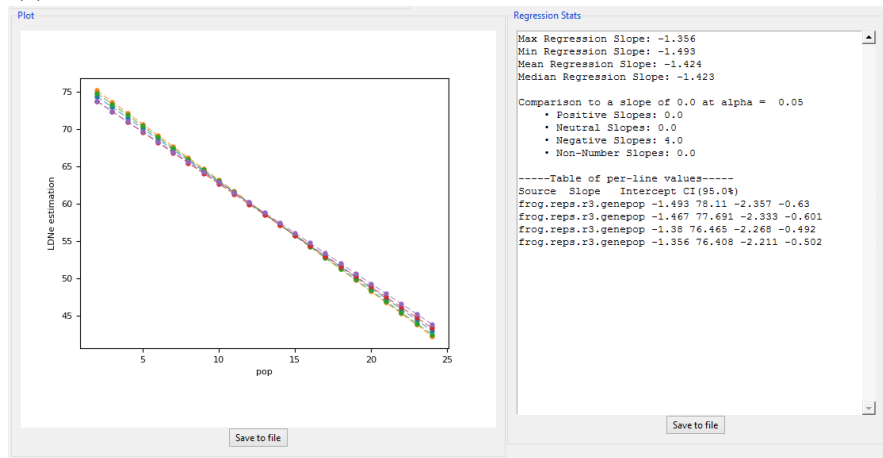
Y axis value  lower limit y axis values  upper limit y axis values

Set scale to  Set scale to

X-axis Data

X axis variable

(a) Settings, with Y axis limits set, and the genepop file filter set to one file only.



(b) The resulting plot.

Figure 0.18: The Nb/Ne estimation regression interface with pop number as the x-axis variable.

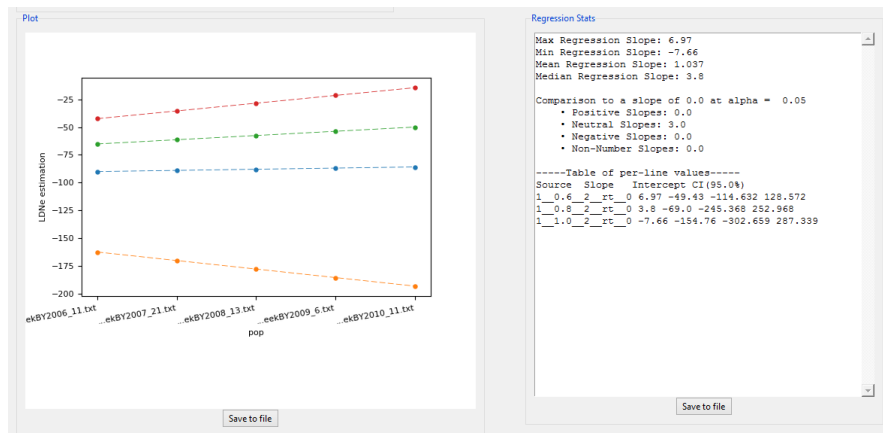
### 0.12.3.1 Filtering.

These selections offer the same filtering as do those for the boxplot (Section 0.12.2.2). In this case, restricting an input field to a single value reduces the number of lines regressed, such as regressing over pop values, but only for one of multiple genepop files (Figure 0.18a), or regressing over a series of genepop files, but only for the estimations associated with pop sampling replicate 2 (Figure 0.19a).

### 0.12.3.2 Y-axis field

The “Y axis value” drop-down box allows you to select which set of LDNe output values are used as the response variable in the regression. See the description for the boxplots in section 0.12.2.3.

(a) Settings, with Y limits set, and pop sampling replicate 2 selected. Note that there are 3 pop sampling values, and we select “All”, so that we get a regression line for each value.



(b) The resulting plot. Note that there is a line for each of the 3 pop subsampling values, as seen in the Regression Stats box.

Figure 0.19: The Nb/Ne estimation regression interface with file sort ordination as the x-axis variable.

### 0.12.3.3 Y-axis value limits

These controls allow you to set a minimum and maximum value, to establish a range of y-values to use for the regression. See the description in (Section 0.12.2.4).

### 0.12.3.4 X axis variable

You can regress using either the pop number (i.e. the values in the pop column in the output tsv file, see Table 0.2), or the genepop file (i.e. the file names in the “original\_file” column in the output tsv file, stripped of their directory paths. When you use file names as x-axis values the program will assign numeric values to the file names in one of two ways.

- Genepop file names are numbers. If your genepop files are named using all-numbers, they will become the numerical values used along the x-axis.

- Genepop file names are not all-numbers. In this case the files will be assigned integers  $1, 2, 3 \dots N$  according to a natural sorting algorithm, that will sort non-number parts of the name alphabetically, but will sort number-parts numerically. This means, for example, that files with form myfile\_1.txt, myfile\_11.txt, and myfile\_2.txt will be sorted in order, and assigned x-values 1,2,3:

1. myfile\_1.txt
2. myfile\_2.txt
3. myfile\_11.txt

This genepop-file based numbering was created to accommodate regression over a series of genepop files with a single pop entry, with each file representing, for example, an annual sample of a real population, and each file named text giving the population name and a number representing the year, or an otherwise ordinal number.

#### **0.12.3.5 The plot**

The plot shows a regression line for each combination of input fields, as well as a line based on an expected slope. The program currently does not label the expected line.

You can use the “Save” button to save the current plot to file, either as a png (the default image format when you use a name with no extension, or an extension other than “.pdf” which is the sole alternative format.

#### **0.12.3.6 The regression stats text box**

This text box shows regression statistics and quantities. You can save this text to file using the “Save” button.



# Bibliography

- [1] Bo Peng and Marek Kimmel. simuPOP: a forward-time population genetics simulation environment. *Bioinformatics*, 21(18):3686–3687, September 2005.
- [2] Robin S. Waples, Tiago Antao, and Gordon Luikart. Effects of Overlapping Generations on Linkage Disequilibrium Estimates of Effective Population Size. *Genetics*, 197(2):769–780, June 2014.
- [3] Robin S. Waples and Chi Do. ldne: a program for estimating effective population size from data on linkage disequilibrium. *Molecular Ecology Resources*, 8(4):753–756, July 2008.
- [4] Robin S. Waples and Ryan K. Waples. Inbreeding effective population size and parentage analysis without parents. *Molecular Ecology Resources*, 11:162–171, March 2011.