

1 A NON-INVASIVE SEX IDENTIFICATION OF BLOOD
2 COCKLES TEGILLARCA GRANOSA (LINNAEUS, 1758)
3 USING MACHINE LEARNING

4 A Special Problem Proposal
5 Presented to
6 the Faculty of the Division of Physical Sciences and Mathematics
7 College of Arts and Sciences
8 University of the Philippines Visayas
9 Miag-ao, Iloilo

10 In Partial Fulfillment
11 of the Requirements for the Degree of
12 Bachelor of Science in Computer Science by

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20 October 28, 2024

Abstract

22 From 150 to 200 words of short, direct and complete sentences, the abstract should
23 be informative enough to serve as a substitute for reading the entire SP document
24 itself. It states the rationale and the objectives of the research. In the final Special
25 Problem document (i.e., the document you'll submit for your final defense), the
26 abstract should also contain a description of your research results, findings, and
27 contribution(s).

28 Suggested keywords based on ACM Computing Classification system can be
29 found at https://dl.acm.org/ccs/ccs_flat.cfm

30 **Keywords:** Keyword 1, keyword 2, keyword 3, keyword 4, etc.

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Chapter 1

Introduction

1.1 Overview

The Philippines is a global center of marine biodiversity and has established aquaculture as a significant contributor to total fishery production (Aypa & Bacongus, 2000; BFAR, 2019). As the 11th largest seafood producer in the world, the country produces over 4 million tonnes of seafood annually. Aquaculture is deeply integrated into Filipinos' livelihoods, encompassing fish cultivation and the production of various aquatic species, including mollusks. Among these are blood clams (*Tegillarca granosa*) which hold considerable economic and environmental significance.

Maintaining a balanced male-to-female ratio of blood cockles is crucial to prevent overharvesting and ensure sustainable production because an imbalanced ratio can lead to overexploitation and can impact the population's sustainability. However, there is limited literature on *T. granosa* that has a thorough understanding of its sex-determining mechanisms, particularly concerning sexual dimorphism in morphological and morphometric characteristics (Breton et al., 2017).

Currently, sex determination methods for blood cockles are invasive, including dissection, and histological examinations which often result in the death of the specimens. While there is growing literature on aquaculture commodities sex identification using machine learning and deep learning, there is a notable scarcity of research specifically addressing *T. granosa* [citation].

This study, titled "A Non-Invasive Sex Identification of *T. granosa* using Machine Learning," aims to provide a comprehensive analysis of blood cockles by

95 leveraging their morphological and morphometric characteristics. By integrating
96 machine learning and computer vision techniques, the study seeks to identify dis-
97 tinct features that indicate sexual dimorphism between male and female blood
98 cockles.

99 1.2 Problem Statement

100 Accurately identifying the sex of *T. granosa* is important in order to promote sus-
101 tainable aquaculture and biodiversity by maintaining a balanced male-to-female
102 ratio. A balanced ratio helps prevent overharvesting. Although sex identification
103 is important for blood cockle population management and sustainable aquacul-
104 ture, there is a notable lack of research in creating non-invasive methods to identify
105 the sex of *T. granosa*. Many of the latest studies and approaches are based on
106 invasive methods like dissection or histological analysis, which are impractical for
107 large-scale aquaculture operations focused on conservation.

108 The existing invasive methods for identifying the sex of *T. granosa* often re-
109 quire dissection, a technique that involves cutting open the shell to visually inspect
110 the gonads (Erica, 2018). This causes harm and death to the specimens. In some
111 cases, histological examination is used to examine tissue samples through a mi-
112 croscope, leading to further destruction of the organism (May et al., 2021). These
113 methods are time-consuming, labor-intensive, and can pose a threat to population
114 management, especially when it is essential to maintain a balanced sex ratio for
115 breeding programs. Moreover, invasive methods also require technical skills to ex-
116 ecute properly. Aquaculture operations, particularly in resource-limited settings,
117 face challenges in accessing laboratory equipment like microscopes and staining
118 tools which complicates the process.

119 A less invasive approach employed by aquaculturists is to monitor spawning
120 behavior in which individuals are separated and stimulated to reproduce in order
121 to determine their sex through the release of gametes (Miranda & Ferriols, 2023).
122 Although it is indeed less invasive than dissection, spawning still involves inducing
123 stress in blood cockles and may not be completely effective for fast identification
124 in large populations.

125 Given the limitations of both invasive and less invasive methods highlight the
126 need for a more advanced approach. An alternative, non-invasive method involv-
127 ing machine and deep learning technologies might solve these issues by providing
128 a fast, accurate, and effective solution without harming or stressing the blood
129 cockles.

1.3 Research Objectives

1.3.1 General Objective

The general objective of this study is to develop a non-invasive method for identifying the sex of *Tegillarca granosa* using machine and deep learning integrated with computer vision technologies. This method aims to provide accurate and streamlined sex identification without causing harm to the specimens, thus supporting sustainable aquaculture practices.

1.3.2 Specific Objectives

To achieve the general objective of developing a non-invasive sex identification of *T. granosa* using machine and deep learning, the following specific objectives have been established:

1. To collect and organize a comprehensive dataset of *T. granosa* which will include high-quality images and relevant morphological measurements that will serve as the basis for the machine-learning model.
2. To preprocess the collected data to perform quality control and consistency checks. This will include techniques such as color thresholding, segmentation, and image hole filling and dilating.
3. To develop and implement machine learning models that can classify the sex of *T. granosa* based on the collected dataset, implementing algorithms such as support vector machines (SVM) for pre-evaluation, and deep learning models such as Squeezenet and Unet.
4. To evaluate the performance of the models used using performance metrics such as accuracy, precision, recall, and F1-score to ensure the effectiveness and reliability of the models.
5. To compare the developed models against existing methods, such as dissection and spawning, and assess their potential for real-world application in aquaculture operations.

1.4 Scope and Limitations of the Research

This study focuses on developing a non-invasive method for identifying the sex of *Tegillarca granosa* using machine learning, deep learning, and computer vision technologies. The goal is to provide an accurate and efficient means of sex identification without causing harm to the specimens, contributing to sustainable aquaculture practices.

The researchers will work with 500 spawned blood cockles taken from Panay island, specifically Zarraga Iloilo and Ivasan Capiz, equally divided between 250 males and 250 females, obtained through temperature shock. The researchers will personally gather linear measurements, including length, width, height, rib count, length of the hinge line, and distance between the umbos using the vernier caliper. Images and corresponding views of the specimens will also be collected by the researchers under the supervision of the University Researchers Associate from the Institute of Aquaculture, College of Fisheries and Ocean Sciences.

Data collection will take place at the hatchery facility of the University of the Philippines Visayas. Data gathering will be conducted in batches, depending on the availability of spawned samples.

The method developed in this study is specific to *Tegillarca granosa* and may not be generalized to other species. The model is trained exclusively for *Tegillarca granosa* and morphological features including length, width, height, rib count, length of the hinge line, and distance between the umbos may not be shared by other shellfish species.

1.5 Significance of the Research

This study will give us significant advancement in non-invasive sex identification methods in *T. granosa* providing innovative solutions that could solve the challenges in identifying sex and reshape approaches to aquaculture. The significance of this study extends to the following:

Research Institution. The result of this study focusing on the sex-identification mechanism of bivalves, specifically *Tegillarca granosa*, will provide valuable insights into universities and research centers that focus on fisheries and coastal management such as the UPV Institute of Agriculture that aim to develop sustainable development and develop suitable culture techniques.

189 *Fishermen.* By developing a non-invasive method in sex identification, this
190 study can help long-term harvest efficiency and maintain the ratio of the harvest
191 which can help prevent overexploitation of the *T. granosa*.

192 *Coastal Communities.* The result of this study would be beneficial for the
193 coastal communities that are reliant on their source of income with aquaculture
194 commodities like blood cockles. Maintaining the diversity and aspect ratio of
195 male and female may increase the market value of blood cockle production since
196 cockle aquaculture faces significant obstacles worldwide due to the fluctuating
197 seed supplies and scarcity of broodstock from the wild.

198 *Future Researchers.* The result of this study would serve as the basis for studies
199 that involve sex identification in bivalves such as *T. granosa*. Some technologies
200 are yet to be explored in machine learning, deep learning, and computer vision
201 technologies that can lead to higher accuracy and distinguish the presence of
202 sexual dimorphism in the *T. granosa*.

Chapter 2

Review of Related Literature

Aquaculture is the fastest-growing industry in animal food production and has great potential as a sustainable solution to global food security, nutrition, and development (European Fishmeal and Fish Oil Producers, 2024). Aquaculture is deeply integrated into the livelihoods of Filipinos, not only through fish cultivation but also through the production of other aquatic species, including mollusks, oysters, clams, scallops, and mussels (Breton et al., 2017). Mollusks, particularly blood clams *Tegillarca granosa*, have economic and environmental significance. It has been a collective effort to maintain an ideal male-to-female ratio to avoid overharvesting and maintain the optimal ratio to preserve the population and production of the blood cockles.

The members of the Arcidae Family, including *T. granosa* are important sources of food and livelihood. Cockle aquaculture meets rising demands, however, it faces significant challenges due to fluctuating seed supplies (Miranda and Ferriols, 2022). To solve the problem, researchers exert a considerable amount of effort, developing a broader understanding of bivalves including their sex-determining mechanism due to their notable importance in terms of diversity, environmental benefits, and economic and market importance (Breton et al., 2017). Despite the promising idea of identifying sex, there is limited research reported in terms of sexual dimorphism, making it harder to distinguish through its morphological and morphometric characteristics.

By addressing the challenges in the sex identification of *T. granosa*, it would be able to address one problem at a time. Currently, no recent documented publications that integrate machine learning and computer vision in characterizing sexual dimorphism, reducing complexity, variability in sex determination, and differentiation mechanisms in bivalves, including *T. granosa* specifically.

2.1 Background on *Tegillarca granosa* and Their Importance

Tegillarca granosa (Linnaeus, 1758) is also known as blood cockles or blood clam. In the Philippines, it is commonly known as a Litob, a marine bivalve species from the family Arcidae. Litob is widely distributed in the world including Southeast Asia. They can be found in the intertidal mudflats adjacent to the mangrove forest (Srisunont, Nobpakhun, Yamalee, Srisunont, 2020).



Figure 2.1: Dorsal view of *Tegillarca granosa* shell.

T. granosa shell is medium-sized, fairly thick, ovate, and convex with both valves being equal in size but asymmetrical from the hinge. The top edge of the dorsal margin is straight while the front is rounded and slopes downward with its back being obliquely rounded with a concave bottom edge. It has a narrow diamond-shaped ligament near the hinge with 3-4 dark chevron markings although some may be incomplete. The shell's outer layer or the periostracum is smooth and brown with a straight hinge line and 40-68 fine short teeth arranged in a straight line. The beak or the prosogyrate curves forward with the shell having 18-21 raised ribs with blunt nodules, having spaces between them. The inner shell is white with crenulations along the valves' ventral, anterior, and posterior margins. The posterior adductor scar is elongated and squarish while the anterior adductor scar is similar but smaller in size. The mantle covering the bulk of *T. granosa*'s visceral mass is thin but the edges are thick and muscular. It bears the impression of the crenulated shell edges. Their foot is large with a ventral grove with no byssus or thread-like attachment. The *T. granosa*'s soft body is blood red (Narasimham, 1988).

T. granosa is one of the most well-known marine bivalves given that they are a protein-rich food, known for their rich flavor, substantial nutritional benefits, a good source of vitamins, low in fat, and contains a considerable amount of iron,

important in combating anemia (Zha et al., 2022). Blood cockles were collected by locals inhabiting the brackish mudflats during the low tides for consumption and sold in the market as a source of livelihood (Miranda, Ferriols, 2023). *T. granosa* is not only valuable for its market and food purposes, but also facilitates an important role in marine ecosystems as a food source for various organisms like wading birds, intertidal-feeding fish, and crustaceans such as shore crabs and shrimps (Burdon, et al., 2014). Blood cockles can act as sentinel species and a bioindicator of marine pollutants such as heavy metals (Ishak et al., 2016) and polycyclic aromatic hydrocarbons (PAHs) (Sany et al., 2014). Additionally, cockle shells can be utilized to create a cost-effective catalyst for biodiesel production by providing calcium oxide (Boey et al., 2011).

Determining the sex of bivalves is important for three reasons namely: diversity, environmental benefits, and economic significance (Breton et al., 2011). Firstly, with the estimated 25, 000 living species under class Bivalvia, it would be a suitable resource to develop a broader understanding of their evolution of the sex and sex determination mechanism (Breton et al., 2011). Second, studying sex determination is important since bivalves are utilized as bioindicators of environmental health. This would pave the way for understanding bivalves' life cycle and population dynamics in determining different factors that affect them (Campos et al., 2012). Thirdly, the immediate and practical reason to unveil the sex determination mechanism is the economic and nutritional importance of bivalves as a large population of people rely on fish and shellfish as sources of food and nutrition (Naylor et al., 2000). Additionally, male and female aquaculture commodities have different growth and economic values. Male Nile tilapia, for example, grow faster and have lower feed conversion rates than females, female Kuruma prawns (*Penaeus japonicus*) are generally larger than males at the time of harvest (Budd et al., 2015).

Clearly, much more work is required to understand the mechanisms underlying sexual dimorphism in bivalves, specifically *T. granosa*. Just like the other aquaculture commodities, sex affects not just reproduction but it can affect market preference, and underlying economic value, making the determination of sex important for meeting consumer demands. These are the increasing significance of the *T. granosa* despite the lack of reviewed articles in the Philippines.

2.2 Current Methods of Sex Identification in *Tegillarca granosa*

The current sex identification methods in *Tegillarca granosa* range from invasive histological techniques to less invasive methodologies like temperature-induced spawning. Each approach comes with its pros and cons regarding accuracy, feasibility, and impact on natural populations. Induced spawning and larval rearing are considered as the less invasive techniques used to study *Tegillarca granosa*. In the Philippines, limited research has been done on the *Tegillarca granosa* (Linnaeus, 1758), and this study, titled Initial Attempts on Spawning and Larval Rearing of the Blood Cockle, *Tegillarca granosa* in the Philippines, is conducted by Denise Vergara Miranda and Victor Marco Emmanuel Nuestro Ferriols (2023). The researchers conducted experiments on induced spawning and larval rearing, discovering that the eggs of female *T. granosa* were salmon pink, while the sperm released by males looked milky. After spawning, the researchers successfully generated 6, 531, 000 fertilized eggs.

They highlighted the importance of *T. granosa* and other anadarinids as a food source that was established worldwide, especially in Malaysia and Korea. However, in the Philippines, the bivalve aquaculture of the clam species is still limited. The experiment which focuses on the culture and rearing of *T. granosa* was attempted by subjecting the wild broodstocks to a series of temperature fluctuations to induce the spawning of gametes. This is currently the most natural and least invasive method for bivalves (Aji, 2011). The study of Miranda and Ferriols aimed to pave the way to the sustainable production of *T. granosa* seeds for aquaculture production and stock enhancement despite the scarcity of documented hatchery culture of *T. granosa* from larvae to adults that is available in the Philippines.

In the study entitled, The earliest example of sexual dimorphism in bivalves — evidence from the astartid *Nicaniella* (Lower Jurassic, southern Germany), the researchers utilized Principal Component Analysis and Fourier Analysis as a non-invasive method that investigates sexual expression in the *Nicaniella rakoveci*. In the study, researchers discovered that the bivalves with crenulations were found to have a different shell shape, which made them more inflated than those without crenulations. This suggests that when they became females, they adapted to hold more eggs, rather than for protection from predators as previously thought. The formation of crenulations is likely part of the genetic process that controls both the sex change and the changes in shell structure (Baran Karapunar et al., 2021). Overall, the findings demonstrate that the genetic mechanisms for sex change and shell morphology in bivalves existed as early as the Early Jurassic, contributing

327 to our understanding of bivalve diversity and evolution. Thus, the researchers
328 concluded that crenulations serve as a morphological marker for identifying the
329 sex and reproductive stage of these bivalves (Baran Karapunar et al., 2021).

330 On the other hand, invasive techniques such as histological analysis offer a
331 more thorough but harmful method for determining the sex of *T. granosa*. A
332 study on the Spawning Period of Blood Cockle *Tegillarca granosa* (Linnaeus,
333 1758) in Myeik Coastal. 240 blood cockle samples were examined for sex and
334 gonad maturity stages using histological examination, with shell lengths ranging
335 from 26-35mm and shell weights from 8.1-33g. For histological analysis, the whole
336 soft tissues were removed from the shell and the flesh containing most parts of
337 the gonads was fixed in formalin, dehydrated in an upgraded series of ethanol,
338 and cleared in xylene. This invasive method allows for precise identification of
339 the gonadal maturation stages based on the cellular and structural changes in the
340 gonads.

341 The classification of the gonad stages used was by Yurimoto et al. (2014).
342 There are five maturation stages of gonadal development: immature (Stage I),
343 developing (Stage II), mature (Stage III), spawning (Stage IV), and spent (Stage
344 V) stages. The sex of the *T. granosa* was confirmed by the color of the gonad and
345 by conducting a histological examination of the gonads. During the immature
346 stage, sex determination was indistinguishable due to the difficulties of observing
347 the germ cells. In the developing stage, the spermatocytes and a few spermatids
348 can be seen for males, and immature oocytes are attached to the tube wall for
349 the female. In the mature stage, the follicles are full of spermatozoa with their
350 tails pointing towards the center of the tube for the male and the female are full
351 of mature oocytes that are irregular or polygonal in shape with the oval nucleus.
352 Upon reaching spawning, some spermatozoa are released, causing the empty space
353 in the follicle wall for males and females there is a decrease in the number of
354 mature oocytes and it exhibits nuclear disappearance due to the breakdown of
355 the germinal vesicle. Lastly, the spent stage is where the genital tube is deformed
356 and devoid of spermatocytes which have completely spawned. In the female, the
357 genital tube is deformed and degenerated making it empty. The morphology of
358 the cockle gonad shows that the area of the gonad increases according to the
359 increased levels of gonad maturity. The coloration of the gonad tissue layer in the
360 blood cockle varies from orange-red to pale orange in females and from white to
361 grayish-white in males for different maturity stages (Maung, Phyu, Tum, 2021).

362 Although the histological examination is the most reliable method for obtain-
363 ing accurate information on the reproductive biology and sex determination of
364 *T. granosa*, it has limitations. Given its invasive nature, this approach requires
365 the dissection and destruction of specimens, making it unsuitable for continuous
366 monitoring and conservation efforts. Moreover, the current understanding of sex

determination in bivalves and mollusks is poor, and no chromosomes that can be differentiated based on their morphology have been discovered (Afiati, 2007). There exists a study that can provide insight into the sex-determining factor in bivalves but *N. schoberti* is more difficult to analyze concerning potential sexual dimorphism. Thickening the edges of the shell increases its inflation, which means the shell can hold more space inside. This extra space helps protandrous females accommodate more eggs.

2.3 Machine Learning and Deep Learning in Biological Studies

Machine learning has the potential to improve the quality of life of human beings and has a wide range of applications in terms of research and development. The term machine learning refers to the invention and algorithm evaluation that enables pattern recognition, classification, and prediction based on models generated from available data (Tarca AL, Carey VJ, Chen X-w, Romero R, Drăghici S, 2007). The study of machine learning methods has advanced in the last several years including biological studies. In biological studies, machine learning has been used for discovery and prediction. This section will explore existing machine learning studies that are applied in biological sciences highlighting the identification of sex in shells, bivalves, and mollusks.

2.3.1 Deep Learning for Phenotype Classification in Ark Shells

In the study, the researchers utilized three (3) convolutional neural network (CNN) models: the Visual Geometry Group Network (VGGnet), the Inception Residual Network (ResNet), and the SqueezeNet (Kim et al., 2024). These deep learning models are utilized to the ark shells namely *Anadara kagoshimensis*, *Tegillarca granosa*, and *Anadara broughtonii* to identify the phenotype classification.

The researchers classified the ark shells based on radial rib count where they investigated the difference in the number of radial ribs between three species and were counted. Their CNN-based model that classifies images of three ark shells can provide a theoretical basis for bivalve classification and enable the tracking of the entire production process of ark shells from catching to selling with the support of big data, which is useful for improving food safety, production efficiency, and economic benefits (Kim et al., 2024).

400 **2.3.2 Geometric Morphometrics and Machine Learning for** 401 **Species Delimitation**

402 In *Geometric morphometrics and machine learning challenge currently accepted*
403 *species limits of the land snail Placostylus (Pulmonata: Bothriembryontidae) on*
404 *the Isle of Pines, New Caledonia*, the shell size was quantified using centroid size
405 from the Procrustes analysis, and both the shape and size information were used in
406 training the machine learning model. Their study concluded that the researchers
407 support utilizing both methods: supervised and unsupervised machine learning,
408 rather than choosing either of them individually. In general, their research con-
409 tributes to the growing number of studies that have combined geometric morpho-
410 metrics, with the aid of machine learning which is helpful in biological innovation
411 and breakthrough (Dubey et al., 2006; Bocxlaer & Schultheiß, 2010; Mapp et al.,
412 2017; Nattier et al., 2017; Soda et al., 2017; Fang et al., 2018).

413 **2.3.3 Contour Analysis in Mollusc Shells Using Machine** 414 **Learning**

415 Tuset et al., (2020) in their study, *Recognising mollusc shell contours with en-*
416 *larged spines: Wavelet vs Elliptic Fourier analyses*, mentioned Gastropod shells
417 have large spines and sharp shapes which differ based on environmental, taxo-
418 nomic, and evolutionary influences. The researchers stated that classic morpho-
419 metric methods may not accurately depict morphological features of the shell,
420 especially when using the angular decomposition of the contour. The current
421 research examined and compared the robustness of the contour analysis using
422 wavelet transformed and Elliptic Fourier descriptors for gastropod shells with en-
423 larged spines. For that, the researchers analyzed two geographical and ecologically
424 separated populations of *Bolinus brandaris* from the NW Mediterranean Sea. Re-
425 sults showed that contour analysis of gastropod shells with enlarged spines can
426 be analyzed using both methodologies, but the wavelet analysis provided better
427 local discrimination. From an ecological perspective, shells with various sizes of
428 spines in both areas indicate a broad adaptability of the species.

429 **2.3.4 Machine Learning for Shape Analysis of Marine Or-** 430 **ganisms**

431 In the study of Lishchenko and Jones (2021), titled *Application of Shape Analyses*
432 *to Recording Structures of Marine Organisms for Stock Discrimination and Taxo-*

433 *nomic Purposes*, they utilized geometric morphometrics (GM) as an approach to
 434 the traditional method of collecting linear measurements with the application of
 435 multivariate statistical methods and outline analysis in recording the structures
 436 of marine organisms. The main taxonomic categories (mollusks, teleost fish, and
 437 elasmobranchs) with their hard bodies have been used as an indication of age and
 438 a determinable time-scale and structure continue to go through life (Arkhipkin,
 439 2005; Kerr and Campana, 2014). This study has explored variations in the mor-
 440 phometry of recording structures in stock discrimination and systematics. The
 441 researchers utilized the principal component analysis rather than the traditional
 442 approach, which helps simplify the data without losing important information.
 443 They utilized landmark-based geometric morphometrics which has three different
 444 types namely: discrete juxtaposition of tissue, maxima or curvature or other mor-
 445 phogenetic processes, and lastly, the extremal points are constructed landmarks.

446 Generalized Procrustes Analysis (GPA) is a common superimposition tech-
 447 nique in landmark-based geometric morphometrics that aligns landmarks via
 448 translation, scaling, and rotation to eliminate non-shape deviations (Zelditch et
 449 al., 2004). However, there is a limit to the amount of smooth areas that may
 450 be captured, and it is possible to overlook significant shape details. Utilization
 451 of the semi-landmarks enhanced the shape description (Adams et al., 2004). The
 452 researchers observed that using an outline-based approach would be more effective
 453 than using a landmark-based approach. Another approach is the Fourier analysis
 454 which is a curve-fitting approach commonly used due to its well-known mathemat-
 455 ical background and how general functions can be decomposed into trigonometric
 456 or exponential functions with definite frequencies. It has two main approaches
 457 namely: Polar Transform (PT) in which it expresses the outline using equally
 458 spaced radii and Elliptical Fourier Analysis (EFA) which separately analyzes the
 459 x and y coordinates of the shape. The PT works for simple rounded outlines
 460 and has the tendency to miss details in more complex shapes, unlike EFA which
 461 can handle complex, convoluted outlines (Zahn and Roskies, 1972; Doering and
 462 Ludwig, 1990; Ponton, 2006). Many researchers view EFA as the most effective
 463 Fourier method for providing a comprehensive and detailed description of record-
 464 ing structures (Mérigot et al., 2007; Ferguson et al., 2011; Leguá et al., 2013;
 465 Mahé et al., 2016).

466 Landmark-based methods used in the study showed that there are detectable
 467 differences between male and female octopuses. However, the accuracy of deter-
 468 mining sex based on these differences was low, similar to the results obtained
 469 with traditional morphometric techniques. The study involved a relatively small
 470 sample size of 160 individuals, and the structure being analyzed (the stylet, or
 471 internalized shell) varies significantly between individuals. Although the results
 472 aligned with findings from other studies that attempted to identify gender differ-

473 ences in cephalopods, the researchers concluded that the approach might not be
474 accurate enough for reliable sex determination.

475 **2.3.5 Deep Learning for Landmark-Free Morphological Fea-** 476 **ture Extraction**

477 In another study, *a deep learning approach for morphological feature extraction*
478 *based on variational auto-encoder: an application to mandible shape*, the Morpho-
479 VAE machine learning approach was used to conduct a landmark-free shape ana-
480 lysis. Morpho-Vae reduces dimensions by concentrating on morphological features
481 that distinguish data with different labels using an image-based deep learning
482 framework that combines unsupervised and supervised machine learning. After
483 utilizing the method in primate mandible images, the morphological features re-
484 veal the characteristics to which family they belonged. Based on the result, the
485 method applied provides a versatile and promising tool for evaluating a wide range
486 of image data of biological shapes including those missing segments.

487 **2.3.6 Machine Learning for Sex Differentiation in Abalone**

488 In the study, *Towards Abalone Differentiation Through Machine Learning*, re-
489 searchers identified a problem in abalone farming which is having to identify the
490 sex of abalone to apply measures for its growth or preservation. The researchers
491 classified abalone sex using machine learning. Researchers trained the machine to
492 classify different types of classes which are male, female, and immature. Based
493 on the result, demonstrated the impact of utilizing linear classifiers.

494 Similarly, in the study, *Data scaling performance on various machine learning*
495 *algorithms to identify abalone sex*, the researchers of the University of India (2022),
496 focused on the data scaling performance of various machine learning algorithms to
497 identify the abalone sex, specifically using min-max normalization and zero-mean
498 standardization. The different machine learning algorithms are the Supervised
499 Vector Machine (SVM), Random Forest, Naive Bayesian, and Decision Tree. Their
500 study aims to utilize machine learning in terms of identifying the trends and
501 distribution patterns in the abalone dataset. Eight features of the abalone dataset
502 (length, diameter, height, whole weight, shucked weight, viscera weight, shell
503 weight, ring) were used to determine the three sexes of Abalone. Their data has
504 been grouped based on sex which are Female, Male, and Infant. They utilized
505 the Synthetic Minority Oversampling Technique (SMOTE) in data balancing for
506 the preprocessing of the data. Followed by data scaling or normalization where

507 it converts numeric values in a data set to a general scale without distorting
 508 differences in the range of values. Then they classified by splitting the data into
 509 training and testing sets (Arifin et al., 2022).

510 The researchers found out that the Naive Bayes performs more consistently
 511 than other algorithms when the abalone dataset was applied to both min-max and
 512 zero-mean normalization has the highest to lowest average accuracy, respectively
 513 62.37% (Random Forest), 59.49% (SVM kernel RBF), 57.20% (Decision Tree),
 514 56.59% (SVM linear kernel), and 53.39% (Naïve Bayesian). Overall, despite the
 515 decrease in the performance with the normalization, the Random Forest achieves
 516 the highest average balanced accuracy of 74.87%, sensitivity of 66.43%, and speci-
 517 ficity of 83.31%. Liu et al. found that Random Forest not only can handle large
 518 and complex datasets and can run in parallel using multiple random forests but
 519 is also more accurate than other algorithms because it selects the best features to
 520 improve model performance (Arifin et al., 2022).

521 2.3.7 Machine Learning for Geographical Traceability in 522 Bivalves

523 In the study, *BivalveNet: A hybrid deep neural network for common cockle (Ceras-*
 524 *toderma edule) geographical traceability based on shell image analysis*, the re-
 525 searchers incorporated computer vision and machine learning technologies for an
 526 efficient determination of blood cockle harvesting origin based on the shell geomet-
 527 ric and morphometric analysis. It aims to improve the traceability methodologies
 528 in these organisms and its potential as a reliable traceability tool. Thirty *Cerasto-*
 529 *derma edule* samples were collected along the five locations on the Atlantic West
 530 and South Portuguese coast with individual images processed using lazy snapping
 531 segmentation, spectro-textural-morphological phenotype extraction, and feature
 532 selection through hybrid Principal Component Analysis and Neighborhood Com-
 533 ponent Analysis (Concepcion et al., 2023).

534 The researchers developed a non-invasive image-based traceability technique,
 535 an alternative to the chemical and biochemical analysis of the bivalves. It was
 536 able to incorporate machine learning methods to promote lesser human interven-
 537 tion The researchers discovered that BivalveNet emerged as the superior model for
 538 bivalves with 96.91% accuracy which is comparable to the accuracy of the destruc-
 539 tive methods with 97% and 97.2% accuracy rate. The result of the study aided
 540 the researchers in concluding that there is a possibility of on-site evaluation of the
 541 bivalve through the implementation of a mobile app would allow the public and
 542 official entities to obtain information regarding the provenance of seafood prod-
 543 ucts' traceability because of its non-invasive and image-based aspects (Concepcion

et al., 2023).

Tegillarca granosa is known for having no sexual dimorphism. However, through several related studies, the researchers can apply how family shells of *Tegillarca granosa* have been identified based on its morphological and morphometric characteristics, and the methods used in machine learning in identifying its sex.

2.4 Limitations on Sex Identification in *Tegillarca granosa*

To date, no distinction has been made between the male and female *T. granosa* in sexing methodology. In cockle aquaculture without clearly apparent sexual dimorphism, sexing can be performed using invasive methods such as chemical stimulation, dissection, and gonad-stripping. Induced spawning, specifically temperature shocking, is the most natural and least invasive method for bivalves (Aji, 2011). However, the method (Wong & Lim, 2018) of immersing cockles in water from hot to cold with a specific temperature requires deliberate and careful manipulation of the temperature over a specific period and would require constant management and monitoring.

Recent studies involved non-invasive methods, with a specific emphasis on morphological characteristics as indicators of sex differentiation. However, Tatsuya Yurimoto et al. (2014) stated that the existing methods for determining the sex of bivalves and mollusks in general are somewhat limited (Afati, 2007). At present, there is no recorded evidence of sexual dimorphism in *Tegillarca granosa*. Gonochoristic is the classification given to *Tegillarca granosa* (Lee, 1997). However, Lee et al. (2012) reported that the sex ratio varied with shell length, suggesting that sex might alter. Hermaphrodites can exhibit either sequential (asynchronous) or simultaneous (synchronous or functional) characteristics. Sequential hermaphrodites switch genders after being male or female for one or multiple yearly cycles. (Heller 1993; Gosling 2004; Collin 2013). Sex change and consecutive hermaphroditism have been observed in different bivalve species, including Ostreidae, Pectinidae, Veneridae, and Patellidae. However, macroscopically differentiating bivalve sex is challenging. The only way it may be identified is through histological analysis of gonad remains but to do so there is an act of killing the organism (Coe Citation1943; Gosling Citation2004). Verification of sex change in bivalves to classify whether male or female while they are alive is challenging since they need to be re-confirmed and re-evaluated to be the same individual after a year.

579 Lee et al. (2012) found out that *T. granosa*, a species in Arcidae, has been
580 discovered to be a sequential hermaphrodite, with the sex ratio changing with an
581 increase in the shell size. In bivalves, sex changes usually happen when the gonad
582 is not differentiated between spawning seasons. (Thompson et al. 1996). But
583 in *T. granosa*, after the spawning season, sex changes during its inactive phase.
584 Results showed a 15.1% sex change ratio, with males having a higher sex change
585 ratio (21.2%) than females (6.2%). The 1+ year class had a higher ratio (17.8%)
586 than the 2+ year class (12.1%). Thus, this study indicates that *T. granosa* is
587 a sequential hermaphrodite. The results of the study demonstrated that the bi-
588 valve's age affects the sex ratio and degree of sex change, but additional in-depth
589 investigation is required to determine the role that genetic and environmental
590 factors play in these changes. No literature in the study of mollusks specifically
591 addresses the machine learning algorithm used to determine the sex of *T. granosa*
592 bivalves in various models. Nevertheless, various techniques such as shape ana-
593 lysis, morphometric analysis, Wavelet, and Fourier analysis, as well as different
594 deep learning models like VGNet, ResNet, and SqueezeNet in CNN networks are
595 utilized for phenotype classification, while different machine learning algorithms
596 could serve as the foundation for this research project

597 2.5 Synthesis of the Study

598 This section of the paper summarizes the technologies used in the different studies
599 related to the pursuit of the study entitled, Non-invasive Sex Identification of *T.*
600 *granosa* using machine learning.

Literature	Technology / Method Used	Description of Problem	Pros	Cons
Initial Attempts on Spawning and Larval Rearing of the Blood Cockle, <i>Tegillarca granosa</i> in the Philippines	Temperature shock	No recent studies are available on the production and rearing of <i>T. granosa</i> in the Philippines.	Employed less invasive techniques which minimize the stress in <i>T. granosa</i> and can lead to better survival rates.	Time-consuming as the entire process from fertilization to the spat stage took 120 days.
The earliest example of sexual dimorphism in bivalves—evidence from the astartid <i>Nicautella</i> (Lower Jurassic, southern Germany)	Morphometric analysis, microscope imaging, principal component analysis (PCA), and Fourier shape analysis	To address the observed shell dimorphism in the Early Jurassic bivalve <i>Nicautella rakoveci</i> , namely the presence or lack of crenulations on the ventral shell margin, and whether these variations represent sexual dimorphism and sequential hermaphroditism.	The methods used reveal significant morphological differences with regard to sexual dimorphism.	There could be misinterpretation of the shape differences of bivalves due to the constraints and resolution of technologies used.
Spawning Period of Blood Cockle <i>Tegillarca Granosa</i> (Linnaeus, 1758) in Myeik Coastal	Histological examination	The need to understand the reproductive period of <i>T. granosa</i> in Myeik to ensure sustainable aquaculture and to prevent over-exploitation.	Method used allows for accurate sex identification based on the histological characteristics and color of the gonads.	Invasive technique used to determine the sex of <i>T. granosa</i> through gonad histological analysis.
Deep learning-based phenotype classification of three ark shells: <i>Anadara kooshimensis</i> , <i>Tegillarca granosa</i> , and <i>Anadara broughtonii</i>	Convolutional neural network (CNN) models, VGGNet, Inception-ResNet, SqueezeNet	Traditional methods of recognizing and classifying ark shell species based on shell traits are time-consuming and inaccurate.	Automated classification of the three ark shells using a deep learning model obtained an accuracy of 92.4%.	Challenges may arise with certain ark shells that share similar morphology.
Geometric morphometrics and machine learning challenge currently accepted species limits of the land snail <i>Placostylus</i> (Pulmonata: Bothriembryonidae) on the Isle of Pines, New Caledonia	Neural network analysis (supervised learning) and Gaussian mixture models (unsupervised learning)	To determine whether the shape and size of the snail's shells can distinguish between two <i>Placostylus</i> species, particularly in groups that appear to be hybrids.	Combining geometric morphometrics and machine learning effectively answers biological issues, providing insights into species classification and possible hybridization.	Difficulty classifying intermediate phenotypes, with potential for overfitting and misclassification in both learning methods.
Recognizing mollusc shell contours with enlarged spines: Wavelet vs Elliptic Fourier analyses	Wavelet functions and Elliptic Fourier descriptors	Addresses the difficulty of accurately defining phenotypic diversity in gastropod shells.	Advanced contour analysis methods allow accurate differentiation of gastropod shell forms.	Cannot clarify the causes of phenotypic variation in the two populations studied.
Application of Shape Analyses to Recording Structures of Marine Organisms for Stock Discrimination and Taxonomic Purposes	Landmark- and outline-based Geometric Morphometric methods	To address difficulties in differentiating between stocks of marine organisms to prevent misidentification that could affect conservation and management.	Shape analysis improves taxonomic classification precision and offers close distinction between related species or organisms.	Landmark-based methods can be sensitive to landmark placement.
A deep learning approach for morphological feature extraction based on variational auto-encoder: an application to mandible shape	Morphological regulated variational AutoEncoder (Morpho-VAE)	The need for reliable, landmark-free methods, such as a modified variational autoencoder, to extract and decipher complex shapes from image data.	Employs dimension reduction and feature extraction, making it a user-friendly tool for biology non-experts.	Limited sample size in certain families presented challenges.
Towards Abalone Differentiation Through Machine Learning	Machine learning algorithms	Identifying the sex of abalones is challenging for producers applying specific growth or preservation strategies.	Machine learning algorithms accurately classify abalone sex into three categories: male, female, and immature.	Selected features may not fully capture the complexity of abalone morphology.
BivalveNet: A hybrid deep neural network for common cockle (<i>Cerastoderma edule</i>) geographical traceability based on shell image analysis	EfficientNet-B0, ResNet101, MobileNetV2, InceptionV3	Addresses the difficulty of accurately tracing bivalve harvesting origins using computer vision and machine learning algorithms to enhance seafood traceability and combat food fraud.	Non-invasive, image-based tools for bivalve traceability provide faster, cheaper, and equally accurate alternatives to traditional chemical analysis methods.	Small sample size (only 30 cockles) limits model reliability.

601 Recent developments and breakthroughs in machine learning offer hopeful so-
602 lutions for biological issues. Research findings indicate that various machine learn-
603 ing techniques such as CNNs, geometric morphometrics, and deep learning mod-
604 els. They are deemed to be effective for identifying phenotypes and determining
605 the gender of various aquaculture commodities, such as mollusks and abalones.
606 These techniques provide a starting point for creating new, non-invasive ways to
607 differentiate male and female *T. granosa*, potentially addressing the drawbacks of
608 manual and invasive methods. Thus, machine learning to examine morphological
609 and morphometric features may streamline the process of sex identification.

610 Nevertheless, the use of machine learning to determine the sex of *T. granosa*
611 has not been fully explored. It lacks up-to-date and significant related literature
612 on using machine learning to identify sex in *T. granosa*, particularly given the
613 species' possible sequential hermaphroditism and lack of obvious external sexual
614 distinctions.

Chapter 3

Research Methodology

This chapter discusses the materials and methods to be employed in the study, focusing on the development requirements and the software and languages utilized. This will also entail the overall workflow in conducting the study, Non-Invasive Methods in Determining the Sex of *Tegillarca granosa* (blood cockles) using machine learning technologies. The different machine/deep learning algorithms will be thoroughly discussed to ensure a comprehensive understanding of the entity of the research endeavor and its processes.

Dr. Victor Emmanuel Ferriols, the director of the Institute of Aquaculture, will oversee the overall workflow and conduct of this experiment. The researchers will also be guided by the research associates, LC Mae Gasit and Allena Esther Artera. Consequently, the whole dataset collection process will be done at the University of the Philippines Visayas hatchery facility.

3.1 Sample Collection

A total of 1000 adult *T. granosa* that have already spawned will be used in this experiment wherein their sex was already classified as male or female. The sample sizes are going to range from 34 to 61 mm and will be sourced from the coastal area in the municipality of Zaraga, Iloilo, Philippines, as well as from fish markets in the municipality of Ivisan, Capiz, Philippines. The research and experimentation will be done at the University of the Philippines Visayas hatchery facility in Miagao, Iloilo, Philippines. The samples will be placed in 200 L fiberglass reinforced plastic (FRP) tanks containing filtered seawater with 35 ppt salinity (Ferriols, Miranda, 2023) and will be subjected to spawning to categorize male from female

639 *T. granosa*. The samples will undergo a series of temperature fluctuations to
640 induce the spawning of gametes as described in the study of Ferriols and Miranda
641 (2023). This method, induced spawning, is the most natural and least invasive
642 method for bivalves compared to other methods (Aji, 2021). Thus, after the
643 spawning, there would be 500 classified males and 500 classified females.

644 3.2 Ethical Considerations

645 Ethical approval was not required for this study involving animals, as per local leg-
646 islation and institutional guidelines, because the experiments were conducted only
647 on species that are commonly used as food and intended for human consumption.

648 3.3 Creating *T. granosa* Dataset

649 For the initial preparation of the experiment, the researchers will collect primary
650 observations for 100 samples of *T. granosa*. For the actual experimentation, the
651 researchers will collect the dataset by batch eventually comprising 1000 samples
652 of *T. granosa*. The images captured for the dataset will be saved in png format
653 with a file naming convention of the sample’s sex, the orientation or view of the
654 shell, and its corresponding number out of the total 1000 samples. Female *T.*
655 *granosa* samples will begin with 0 in their file name, while males will begin with
656 1, followed by the views captured such as (1) dorsal, (2) ventral, (3) anterior,
657 (4) posterior, (5) left lateral, and (6) right lateral, and lastly, a unique sample
658 number. For example, “010001” will be the file name for the first female sample
659 taken from the dorsal view and “110001” for the first male sample also taken from
660 the dorsal view. The dataset will be organized in a CSV file that lists each image’s
661 file name along with their shell’s width, height, length, rib count, length of the
662 hinge line, and distance between their umbos. This dataset will be essential for
663 machine learning model training and testing.

664 3.4 Morphological Characteristics Collection

665 Morphology refers to the biological form and represents one of the most visually
666 recognizable phenotypes across all organisms (Tsutsumi et al., 2023). Morphology
667 is a term that describes structural characteristics by measuring specific compo-
668 nents, namely, dimensions such as shapes, sizes, and colors. As stated by the

researchers, quantifying and characterizing the shape is essential to understanding and visualizing the variations in *T. granosa*'s morphology. In this study, the researchers are going to measure the height, width, and length of *T. granosa*. The dimensions will be recorded using a Vernier caliper to the nearest 0.01 mm. The length of the *T. granosa* refers to the measurement from the anterior to the posterior of the shell, the width will be measured through the shell's widest point from the left to the right valve and lastly, the height will be measured from the base of the shell to the shell's apex. The height of the gap between the valves near the hinge will also be measured. The authors Reymont and Kennedy (1998), indicated that the use of counts of the shell ribs as supplementary information increases identification accuracy. Thus, the researchers will also take into account the difference in the rib count of the male and female *T. granosa* and the ratio will be calculated since the sizes of the blood clams may vary. Sex ratio, size frequency distribution, and relative growth rates were used to investigate sexual dimorphism.

3.5 Image Acquisition and Pre-Processing

In this study, there would be three major phases for the image processing to be employed namely (1) color thresholding, (2) segmentation, and (3) image hole filling and dilating. The researchers constructed a controlled environment for capturing the samples utilizing a box-like structure of (?) meters with a green background surface. This setup was designed to maintain uniform captures of the images, and a consistent measurement between the sample and the camera, fixing the camera at 50 cm above the *T. granosa*. Placing a ring light to the left of the box, and using a camera with flash to ensure the image quality, eliminate shadows and clarity of the sample during the image acquisition process. For color thresholding, the researchers utilized the red, green, blue (RGB), hue saturation value (HSV), luminance, blue chromaticity, red chromaticity (YCbCr), and (Luminance, a, b)** (CIElab) images obtained from the smartphone considering their wide availability across various stages in the bivalve industry using the MATLAB Colour Thresholding Toolbox in determining which among the four-color spectra may generate the cleanest version of the training images with absence of any blobs (Jayasundara et al., 2023). Google Pixel 3 XL will be utilized with the following specifications: 2960 x 1440 for the resolution, 4,032 x 3,024 pixels (12.2 MP) for the dimensions, f/1.8 for the fstop, 28mm (wide), $\frac{1}{2}$.55", 1.4 μ m, dual pixel PDAF, OIS. [insert reference] After thresholding, the lazy snapping technique will be implemented by manually drawing the background and the foreground lines that represent the black pixels and the bivalve pixels. The lazy snapping algorithm will be configured using the 20 000 superpixels which can divide the *T. granosa*'s

707 images into 20, 000 irregularly shaped geometric pixels that will be based on the
708 CIElab gradients through K-means clustering with $K = 3$. For the last step, the
709 researchers will perform image hole filling and dilating to ensure that no blobs
710 are remaining that can contribute to noise which can affect the correctness of the
711 extracted feature by taking into consideration the 200-pixel blobs that are discon-
712 nected from the largest object in its binary form. This will result in black pixels
713 made by binary filling and dilating to remove the blobs. [reference] Image process-
714 ing will be performed on the MATLAB [version[-]] installed on the [laptop] with
715 specs. The images will be saved based on how it was stated on the collection of
716 the image dataset. To ensure consistent comparisons for the analysis, the images
717 were captured in different angles including dorsal, ventral, lateral, and anterior
718 and posterior taken in uniform angles to provide visual coverage of the *T. granosa*
719 sample.

720 3.6 Machine/ Deep Learning Technologies

721 This section of the paper will discuss the technologies to be used in training, and
722 testing the model as well as associated techniques and algorithms. Since obtaining
723 the induced samples was done per batch, the researchers will conduct an initial
724 run with a support vector machine before delving into more complex methods
725 such as deep learning models.

726 3.7 Support Vector Machine for Pre-evaluation

727 The shape of recording structures was first analyzed by collecting measurements
728 of linear distances and applying multivariate statistical methods to these data
729 (traditional linear measurement method) (Rohlf and Marcus, 1993). Geometric
730 morphometric (GM) methods are an alternative way of analyzing and quantifying
731 shape, which in theory retains more detail about the geometry of the structure
732 than could be obtained from linear measurements (Adams et al., 2004). Machine
733 learning techniques such as decision tree classification, support vector machines
734 (SVMs), and artificial neural networks (ANNs) have been applied to the analysis
735 of bivalve shell geometry and morphology to classify shells based on morpholog-
736 ical features, including shell shape, size, and texture, among others (Kiel, 2021).
737 The results of these studies have shown that machine learning algorithms can
738 accurately classify bivalve shells and provide insights into the relationships be-
739 tween shell morphology and various environmental factors. Following this, the
740 researchers are going to conduct a pre-evaluation of the linear measurements for

741 100 samples of *T. granosa* using a Support Vector Machine in order to quantify
742 whether the linear measurements can be a determining factor in determining the
743 sex of the samples before proceeding to more complex methods.

744 3.8 Deep Learning for Image-Based Classifica- 745 tion

746 After collecting a sufficient number of images and identifying initial patterns,
747 convolutional neural networks (CNNs) will be used. CNNs, models like VGGNet,
748 ResNet, and Inception have been effectively applied in phenotype classification
749 (Kim et al., 2024). In this study, the deep learning model will be specifically
750 adapted for the sex identification of *T. granosa* based on shell images. CNNs
751 will analyze the images and learn important details about their shapes that can
752 help identify whether they are male or female. Unlike the approach of using
753 three models taken by Kim et al. (2024), the researchers will focus on just one
754 model that has shown the best performance in their study which is SqueezeNet.
755 SqueezeNet is particularly advantageous because it reduces the number of pa-
756 rameters and amount of memory required to store the model without sacrificing
757 accuracy (Koonce, 2021; Sayed et al., 2021). Its ability to achieve high accuracy
758 in classifying shell images makes it a suitable choice for distinguishing between
759 male and female *T. granosa*. Python and Keras libraries will be used to train and
760 test the model. The dataset will be divided into training (), validation (), and
761 testing. Performance metrics such as accuracy, precision, recall, and F1-score will
762 be used to evaluate the model’s effectiveness.

763 Chapter 4

764 Preliminary Results/System 765 Prototype

766 This chapter presents the preliminary results or the system prototype of your SP.
767 Include screenshots, tables, or graphs and provide the discussion of results.

References

- Aji, L. P. (2011). Review: Spawning induction in bivalve. *Jurnal Penelitian Sains*, 14, 14207.
- Arifin, W. A., Ariawan, I., Rosalia, A. A., Lukman, L., & Tufailah, N. (2021). Data scaling performance on various machine learning algorithms to identify abalone sex. *Jurnal Teknologi Dan Sistem Komputer*, 10(1), 26–31. doi: 10.14710/jtsiskom.2021.14105
- Bahtiar, B., Purnama, M. F., Kasim, M., & Ishak, E. (2022). Population dynamics of blood clams tegillarca granosa in kendari bay, southeast sulawesi, indonesia. *Biodiversitas Journal of Biological Diversity*, 23(10). doi: 10.13057/biodiv/d231015
- Barrera-Hernandez, R., Barrera-Soto, V., Martinez-Rodriguez, J. L., Ríos-Alvarado, A. B., & Ortiz-Rodríguez, F. (2023). Towards abalone differentiation through machine learning. In *Towards abalone differentiation through machine learning* (pp. 108–118). Springer. doi: 10.1007/978-3-031-34222-6_9
- Boey, P.-L., Maniam, G. P., Hamid, S. A., & Ali, D. M. H. (2011). Utilization of waste cockle shell (anadara granosa) in biodiesel production from palm olein: Optimization using response surface methodology. *Fuel*, 90(7), 2353–2358. doi: 10.1016/j.fuel.2011.03.002
- Breton, S., Capt, C., Guerra, D., & Stewart, D. (2017, June). *Sex determining mechanisms in bivalves*. Preprints.org. doi: 10.20944/preprints201706.0127.v1
- Breton, S., Stewart, D. T., Shepardson, S., Trdan, R. J., Bogan, A. E., Chapman, E. G., ... Hoeh, W. R. (2010). Novel protein genes in animal mtdna: A new sex determination system in freshwater mussels (bivalvia: Unionoida)? *Molecular Biology and Evolution*, 28(5), 1645–1659. doi: 10.1093/molbev/msq345
- Brotohadikusomo, A. (1994, November). *Ecology of two species of blood clams anadara*. Proquest.
- Budd, A., Banh, Q., Domingos, J., & Jerry, D. (2015). Sex control in fish: Approaches, challenges and opportunities for aquaculture. *Journal of Marine*

800 *Science and Engineering*, 3(2), 329–355. doi: 10.3390/jmse3020329

801 Campos, A., Tedesco, S., Vasconcelos, V., & Cristobal, S. (2012). Proteomic
802 research in bivalves: Towards the identification of molecular markers of
803 aquatic pollution. *Proteomic Research in Bivalves*, 75(14), 4346–4359. doi:
804 10.1016/j.jprot.2012.04.027

805 *Fao 2024 report: Sustainable aquatic food systems important for global food*
806 *security – european fishmeal*. (2024). [https://effop.org/news-events/](https://effop.org/news-events/fao-2024-report-sustainable-aquatic-food-systems-important-for-global-food-security/)
807 [fao-2024-report-sustainable-aquatic-food-systems-important-for](https://effop.org/news-events/fao-2024-report-sustainable-aquatic-food-systems-important-for-global-food-security/)
808 [-global-food-security/](https://effop.org/news-events/fao-2024-report-sustainable-aquatic-food-systems-important-for-global-food-security/).

809 Ishak, A. R., Mohamad, S., Soo, T. K., & Hamid, F. S. (2016). Leachate and
810 surface water characterization and heavy metal health risk on cockles in
811 kuala selangor. In *Procedia - social and behavioral sciences* (Vol. 222, pp.
812 263–271). doi: 10.1016/j.sbspro.2016.05.156

813 Kim, E., Yang, S.-M., Cha, J.-E., Jung, D.-H., & Kim, H.-Y. (2024). Deep
814 learning-based phenotype classification of three ark shells: *Anadara kagoshi-*
815 *mensis*, *tegillarca granosa*, and *anadara broughtonii*. *Frontiers in Marine*
816 *Science*, 11. doi: 10.3389/fmars.2024.1356356

817 Lee, J. S., Park, J. J., Shin, Y. K., Kim, H., & Jeon, M. A. (2014). Sex change
818 and sequential hermaphroditism in *tegillarca granosa* (bivalvia: Arcidae).
819 *Invertebrate Reproduction & Development*, 58(4), 314–318. doi: 10.1080/
820 07924259.2014.949014

821 May, K., Maung, C., Phyu, E., & Tun, N. (2021). Spawning period of blood cockle
822 *tegillarca granosa* (linnaeus, 1758) in myeik coastal areas. *J. Myanmar Acad.*
823 *Arts Sci*, 4.

824 Miranda, D. V., & Ferriols, V. M. E. N. (2023). Initial attempts on spawning and
825 larval rearing of the blood cockle, *tegillarca granosa* (linnaeus, 1758), in the
826 philippines. *Asian Fisheries Science*, 36(2). doi: 10.33997/j.afs.2023.36.2
827 .001

828 Narasimham, K. A. (1988). Taxonomy of the blood clams *anadara* (*tegillarca*)
829 *granosa* (linnaeus, 1758) and *a. (t.) rhombea* (born, 1780).

830 Naylor, R. L., Goldburg, R. J., Primavera, J. H., Kautsky, N., Beveridge,
831 M. C. M., Clay, J., ... Troell, M. (2000). Effect of aquaculture on world
832 fish supplies. *Nature*, 405(6790), 1017–1024. doi: 10.1038/35016500

833 Philippines, B. S. (2024). *Better seafood philippines*. Sustainable Fish-
834 eries Partnership. Retrieved from [https://sustainablefish.org/](https://sustainablefish.org/impact-initiatives/supporting-small-scale-fisheries/better-seafood-philippines/)
835 [impact-initiatives/supporting-small-scale-fisheries/better](https://sustainablefish.org/impact-initiatives/supporting-small-scale-fisheries/better-seafood-philippines/)
836 [-seafood-philippines/](https://sustainablefish.org/impact-initiatives/supporting-small-scale-fisheries/better-seafood-philippines/)

837 Quenu, M., Trewick, S. A., Brescia, F., & Morgan-Richards, M. (2020). Geometric
838 morphometrics and machine learning challenge currently accepted species
839 limits of the land snail *placostylus* (pulmonata: Bothriembryontidae) on the
840 isle of pines, new caledonia. *Journal of Molluscan Studies*, 86(1), 35–41.
841 doi: 10.1093/mollus/eyz031

- 842 Sany, S. B. T., Hashim, R., Rezayi, M., Salleh, A., Rahman, M. A., Safari, O.,
843 & Sasekumar, A. (2014). Human health risk of polycyclic aromatic hydro-
844 carbons from consumption of blood cockle and exposure to contaminated
845 sediments and water along the klang strait, malaysia. *Marine Pollution*
846 *Bulletin*, 84(1-2), 268–279. doi: 10.1016/j.marpolbul.2014.05.004
- 847 Srisunont, C., Nobpakhun, Y., Yamalee, C., & Srisunont, T. (2020). Influence
848 of seasonal variation and anthropogenic stress on blood cockle (*tegillarca*
849 *granosa*) production potential. *Influence of Seasonal Variation and Anthro-*
850 *pogenic Stress on Blood Cockle (Tegillarca Granosa) Production Potential*,
851 44(2), 62–82.
- 852 Tsutsumi, M., Saito, N., Koyabu, D., & Furusawa, C. (2023). A deep learning
853 approach for morphological feature extraction based on variational auto-
854 encoder: An application to mandible shape. *Npj Systems Biology and Ap-*
855 *plications*, 9(1), 1–12. doi: 10.1038/s41540-023-00293-6
- 856 Tuset, V. M., Galimany, E., Farrés, A., Marco-Herrero, E., Otero-Ferrer, J. L.,
857 Lombarte, A., & Ramón, M. (2020). Recognising mollusc shell contours
858 with enlarged spines: Wavelet vs elliptic fourier analyses. *Zoology*, 140,
859 125778–125778. doi: 10.1016/j.zool.2020.125778
- 860 Wong, T. M., & Lim, T. G. (2018). *Cockle (anadara granosa) seed produced in*
861 *the laboratory, malaysia*. (Handle.net) doi: 10.3366/in_3366.pdf
- 862 Yurimoto, T., Kassim, F. M., Man, A., & Fuseya, R. (2014). *Spawning season and*
863 *larval occurrence of blood cockle (anadara granosa) off the selangor coast,*
864 *peninsular malaysia*. (DOAJ: Directory of Open Access Journals)
- 865 Yusuff, F. M., Shari, M. A. M., Joni, A. A. M., Kusin, F. M., Mohamed, K. N.,
866 Zulkeflee, Z., ... Arshad, A. (2021). Health status comparison of blood
867 cockle (*tegillarca granosa*) between low and high yield farms in selangor and
868 johor. *IOP Conference Series: Earth and Environmental Science*, 934(1),
869 012048. doi: 10.1088/1755-1315/934/1/012048
- 870 Zha, S., Tang, Y., Shi, W., Liu, H., Sun, C., Bao, Y., & Liu, G. (2022). Im-
871 pacts of four commonly used nanoparticles on the metabolism of a ma-
872 rine bivalve species, *tegillarca granosa*. *Chemosphere*, 296, 134079. doi:
873 10.1016/j.chemosphere.2022.134079
- 874 (Aji, 2011) (Bahtiar, Purnama, Kasim, & Ishak, 2022) (Barrera-Hernandez, Barrera-
875 Soto, Martinez-Rodriguez, Ríos-Alvarado, & Ortiz-Rodríguez, 2023) (Philippines,
876 2024) (Boey, Maniam, Hamid, & Ali, 2011) (Breton, Capt, Guerra, & Stewart,
877 2017) (Breton et al., 2010) (Brotohadikusomo, 1994) (Budd, Banh, Domingos,
878 & Jerry, 2015) (Srisunont, Nobpakhun, Yamalee, & Srisunont, 2020) (Campos,
879 Tedesco, Vasconcelos, & Cristobal, 2012) (*FAO 2024 Report: Sustainable Aquatic*
880 *Food Systems Important for Global Food Security – European Fishmeal*, 2024)
881 (Ishak, Mohamad, Soo, & Hamid, 2016) (Kim, Yang, Cha, Jung, & Kim, 2024)
882 (Lee, Park, Shin, Kim, & Jeon, 2014) (Quenu, Trewick, Brescia, & Morgan-

883 Richards, 2020) (May, Maung, Phyu, & Tun, 2021) (Miranda & Ferriols, 2023)
884 (Narasimham, 1988) (Naylor et al., 2000) (Yurimoto, Kassim, Man, & Fuseya,
885 2014) (Sany et al., 2014) (Tsutsumi, Saito, Koyabu, & Furusawa, 2023) (Tuset et
886 al., 2020) (Arifin, Ariawan, Rosalia, Lukman, & Tufailah, 2021) (Wong & Lim,
887 2018) (Yusuff et al., 2021) (Zha et al., 2022)

888 **Appendix A**

889 **Appendix Title**

890 **Appendix B**

891 **Resource Persons**

892 **Mr. Firstname1 Lastname1**

893 Role1

894 Affiliation1

895 emailaddr1@domain.com

896 **Ms. Firstname2 Lastname2**

897 Role2

898 Affiliation2

899 emailaddr2@domain.net

900