**The Acromegaly Gene Expression Signature in Human Adipose Tissue Reveals Possible New Mechanisms for Enhanced Lipolysis and Insulin Resistance**

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**ABBREVIATED TITLE:** Analysis of Acromegalic Adipose Tissue

**KEY TERMS:** Acromegaly, Lipolysis, Insulin Resistance, Growth Hormone

**WORD COUNT:** 2736

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**GRANT SUPPORT**: This work was supported by Motor City Golf Classic (MCGC) Grant # G010640.

**DISCLOSURE STATEMENT:** The authors have nothing do disclose

**Structured Abstract:**

Context: GH affects several molecular pathways regulating proliferation and metabolism. Objective: To study tissue transcriptional changes induced by GH

Design. Prospective observational study from March 2011 to June 2012.

Setting: Tertiary referral center at the University of Michigan Pituitary clinic

Participants: Patients with acromegaly (n=9) or non functioning pituitary adenoma (n=11).

Intervention: The patients underwent clinical and metabolic profiling including assessment of HOMA-IR. Explants of adipose tissue were assayed ex-vivo for lipolysis. Adipose tissue was analysed by RNAseq.

Main Outcome: differences in adipose tissue mRNA expression between acromegaly patients and controls.

Results: There was a trend for enhanced adipose tissue lipolysis in acromegaly patients (p=0.09). When correcting for BMI there was a significant decrease in insulin sensitivity in the acromegaly patients (P=0.0027). We observed several expected transcriptional changes (*IGF1*, *IGFBP3*) as well as several novel transcriptional changes, some of which may be important for GH signal regulation (*PTPN3* and *PTPN4*) and the effect of GH on growth and proliferation. Several transcripts could potentially be important in GH-induced metabolic changes. Specifically induction of *LPL*, ,*ABHD5*, , *ACVR1C* could contribute to enhanced lipolysis. Higher expression of *SCD* and *TCF7L2* could contribute to insulin resistance. Expression of *HSD11B1* was reduced and *GR* was increased, predicting modified glucocorticoid activity in acromegaly.

Conclusions: We identified the acromegaly gene expression signature in human adipose tissue. The significance of altered expression of specific transcripts will enhance our understanding of the metabolic and proliferative changes associated with acromegaly

**Introduction**

Acromegaly, excess growth hormone (GH) production secondary to a pituitary adenoma, has important metabolic effects. The two most significant effects of GH on metabolism in adipose tissue are insulin resistance and lipolysis (1). Insulin resistance, presenting as diabetes or impaired glucose tolerance, is found in most acromegalic patients (2), and contributes to the enhanced morbidity (3). To study the effect of excess GH on adipose tissue, we used next generation RNA sequencing to study adipose tissue from acromegaly patients and controls. We found a distinctive pattern of changes in many transcripts that are highly associated with acromegaly. Many of these alterations may contribute to the metabolic effect of GH and reveal novel mechanisms of GH-induced insulin resistance and lipolysis in adipose tissue.

**Methods**

**Patient recruitment**

The study was approved by the institutional review board of the University of Michigan Medical System. Written informed consent was obtained from all patients. Patients were recruited consecutively from those undergoing transsphenoidal adenomectomy at the University of Michigan for acromegaly or non-functioning pituitary adenoma over a 12 month period. Exclusion criteria were age <18, current hormone treatment including glucocorticoids, malignancy, inflammatory disease, diabetes type 1 and established pituitary hormone deficiencies. For each patient, a data sheet was completed including, age, sex, anthropometric measurements, diagnosis of hypertension, diabetes, results of blood tests and medications. Fasting blood samples were assayed for glucose (Siemens Advia 1800) and insulin (Life Technologies) as instructed by the manufacturers.

**Subcutaneous fat biopsy**

During the course of pituitary surgery a routine subcutaneous fat graft is utilized to seal the surgical field upon completion of the procedure. 500 mg of this fat graft was used for the study. ~100 mg were utilized for ex vivo lipolysis assay ~200 mg was snap frozen in liquid nitrogen and stored at -80 degrees for RNA preparation.

***Ex vivo* lipolysis**

25 mg pieces of adipose tissue were pre-incubated for 15 minutes in KRBH buffer (sigma) at 37°C and then incubated for 1 hour at 37°C in 300 μl KRBH in the presence or absence of isoproterenol 30nM in duplicate. Glycerol was assayed in supernatants using a glycerol assay kit (sigma) as instructed by manufacturer.

**Transcriptomic Analysis**

Total RNA was extracted from adipose tissue using the RNEasy kit (Qiagen)and its quality was verified using the Agilent 2100 Bioanalyzer (Agilent Technologies). At the University of Michigan DNA Sequencing Core, cDNA libraries from polyA mRNA were prepared using TruSeq cDNA synthesis kit and sequenced using a HiSeq 2000 (Illumina). Samples were run on 2 lanes of a HiSeq 2000 (Illumina) generating 8 612 682 to 16 469 501 single-ended 50bp reads per sample. These were aligned to the human genome (Enembl GRCh37.69, Genbank Assembly ID GCA\_000001405.9) using TopHat version 2.0.6 (4), Bowtie version 2.0.2 (5) and Samtools version 0.1.18 . Only the highest expressing transcript was analysed. Gene expression was analyzed using DESeq version 1.14.0 (6). All p-values were adjusted by the method of Benjamini-Hochberg (7). These subjects corresponded to the patients described in Table 1, with the exception of subjects 32 and 35 (both acromegaly patients), which had clinical data but no RNAseq data.

For re-analysis of the Huo dataset (8), we downloaded these data from the Gene Expression Omnibus (GSE2120) and compared the 48h treated control data to the 48h GH treated data. Analysis was performed using limma (version 3.16.7; (9)), with all p-values adjusted by the method of Benjamini-Hochberg (7).

**Statistics**

Statistical significance in this study was defined as a p-value of less than 0.05. All statistical tests were performed using the R package (version 3.0.2,(10)). Unless otherwise indicated, between group comparisons were analyzed using a Welch’s two sample t-test. All p-values were corrected for multiple observations by the method of Benjamini and Hochberg (7). To test for correlations we calculated Pearson's product-moment correlation. To test for gene set enrichment, either for gene ontology or KEGG categories, we used the GOseq package (version 1.14.0 using GO.db version 2.10.1 and KEGG.db version 2.10.1), taking sequence length bias into account and using the Wallenius approximation for the null distribution (11). Resulting p-values were adjusted by the method of Benjamini and Hochberg (7). Redundant GO categories were manually removed. To test for enrichment of genes identified in the Huo *et al.* dataset (8), we used Fisher’s exact test. To correct for the effects of BMI on insulin sensitivity, we generated a linear model using the natural logarithm of the HOMA-IR score as the dependent variable, and the BMI and the diagnosis as the independent variables. We observed no evidence of an interaction between BMI and the diagnosis in this model (p=0.617). This model had an adjusted R2 of 0.567.

**Results**

**Patient characteristics**

Clinical and metabolic measurements were obtained for 9 acromegaly patients and 11 controls. Patient characteristics are shown in Table 1. There was no statistically significant difference in body mass index (BMI), abdominal circumference or weight. Acromegaly patients trended to be younger and taller than their controls.

**Acromegaly patients were more insulin resistant and had higher lipolysis**

Acromegaly patients trended to have higher fasting glucose levels (p=0.086) and higher fasted insulin (p=0.11, Figure 1A-B). When combined, we observed a trend towards higher HOMA-IR scores in the acromegalic patients (p=0.11, Figure 1C). We observed a significant association between BMI and insulin sensitivity (HOMA-IR score) across our subjects (p=0.022, R2=0.286), we therefore corrected for the wide range of BMI’s in this study and detected a significant decrease in insulin sensitivity in the acromegaly patients (a HOMA-IR score increase of 2.407, with a 95% confidence interval of 1.45-4.25; p= 0.012; Figure 1D).

Subcutaneous adipose tissue chunks for lipolysis assay were available from 6 acromegaly patients and 9 controls. As shown in Figure 1E, acromegaly patients trended towards higher basal lipolysis (p=0.11), and higher lipolysis in the presence of isoproterenol (p=0.14). These data are consistent with previous reports linking GH signaling with increased lipolysis (12).

**Transcriptomic Analysis**

To determine which genes are altered in adipose tissue in acromegaly subjects, we performed a transcriptomic analysis of subcutaneous adipose tissue mRNA from 7 acromegalic patients and 11 controls. We identified 103 genes that had significantly different expression in acromegaly, of these 25 genes were downregulated and 78 were upregulated in adipose tissue from the acromegalic patients. These transcripts form a signature identifying transcriptional differences in adipose tissue in response to long-term exposure to GH (Figure 2 and Supplementary Table 1).

Gene set enrichment analysis using gene ontology terms (11,13) showed enrichment of genes in the gene ontology categories involved in development, signaling and lipid biosynthetic processes (Table 2). These included several categories specifically related to steroid biogenesis, indicating a novel alteration in sterol metabolism in acromegaly patients. We also tested for enrichment of KEGG pathways and found enrichment in genes involved in TGF beta signaling and prostate cancer (Table 2). The negative relationship between GH signaling and TGF beta signaling has been previously reported in cardiomyocytes (14), as has the increased incidence some cancers, including prostate cancer, and acromegaly (reviewed in (15)).

Previous work by Huo *et al*. (8) examined an analogous *in vitro* system, where 3T3-F442A adipocytes were treated with GH for a variety of time points. The longest time point (48h) is potentially analogous to the chronic GH exposure that occurs in adipose tissue from acromegaly patients. We re-analyzed that data set, looking at only the 48h GH treatment time point. Out of the 103 genes identified in our acromegaly analysis, 34 have mouse homologs, corresponding to 48 probes in that dataset. From examination of these probesets, we found 12 probes altered in the same direction in a statistically significant manner (or 15 %). Therefore the genes from the acromegaly dataset are significantly enriched in the GH treated dataset (p=1.61 × 10-4). The genes that reached statistical significance in both datasets were *Fmo1, Phldb2, Igsf10, Igfbp3, Igf1, Scd2, Scp2, Ptger3 and Capn6.*

**Established GH responsive genes are upregulated in adipose tissue from acromegaly patients**

Since acromegaly is caused by an overproduction of GH, we first analyzed known GH responsive genes. We found that expression of GH responsive genes, including *IGF1* and *IGFBP3*, are elevated in acromegalic patients (Figure 3A-B). IGF-1 has been shown to be induced in adipocytes exposed to GH (16), while there is no previous reports regarding IGF-BP3 induction. The confirmation of these previously reported acromegaly or GH dependent transcriptional changes strengthens our interpretation of other transcriptional changes.

Within the acromegalic subjects, there was a correlation between *IGF1* mRNA and levels of IGF-1 in serum in the acromegaly patients (R2=0.51, p=0.043; Figure 3C), reflecting that increased induction of *IGF1* in adipose tissue in is similar in its extent to serum IGF1 induction. Serum IGF1 is primarily thought to be derived from liver tissue (17). Our data demonstrates expression of the adipose tissue *IGF1* gene correlates well with that of serum IGF-1, suggesting that adipose tissue may also be a source of IGF-1 in acromegalic subjects.

**Negative regulators of GH signaling are upregulated in acromegaly patients**

*SOCS2* AND *CISH*, both suppressors of cytokine signaling previously reported to be important in downregulating GH signaling We found that both these genes are upregulated in acromegaly (Figure 3D-E). These genes have been previously reported to be induced in liver and muscle by GH (18), while *SOCS2* has also been reported to be induced in adipocytes by GH (16). The induction in adipose tissue has not been described previously. These genes likely reflect activation of a negative feedback loop in response to chronic high growth hormone levels.

We also observed induction of the tyrosine phosphatases, *PTPN3* (also called PTP-H1) and *PTPN4* in acromegaly (Supplementary Figure 1A-B). *PTPN3* has been reported to bind GH receptor *in vitro* in the presence of GH (19), and its overexpression reduces STAT5 signaling in response to GH (20). *Ptpn3* Knockout mice have excessive GH activity, as demonstrated by excessive growth accompanied by a strong induction of liver *IGF1* mRNA and serum IGF-1 (20). This is the first report of enhanced abundance of PTPN3 mRNA in response to GH exposure. The increased expression of *PTPN3* that we have observed in acromegaly suggests that this may be an additional negative feedback pathway induced by GH and reducing GH signaling.

**Genes controlling DNA replication, proliferation and apoptosis**

We observed difference in expression of several different genes regulating cellular proliferation in acromegalic subjects. Cyclin E (*CCNE1)*, which is important for transfer from G1 to S, was increased in acromegalic patients, and cyclin dependent kinase inhibitor B (*CDKN2B*) was decreased (Supplementary Figures 1C-D). Of note, *CDKN2B* is also a diabetes susceptibility gene identified repeatedly in GWAS studies (21,22). Nucleosome assembly protein 1-like 1 (*NAP1L1*) and origin recognition complex, subunit 2(*ORC2*), which are important for DNA replication, and the antiapoptotic regulators *BAG4*(BCL2-associated athanogene 4) and *CAPN6* (calpain 6) were also induced (Supplementary Figures 1E-H). Apoptosis signal-regulating kinase 1 (*MAP3K5*) expression is higher in acromegaly, and there is also a trend for higher expression of its downstream substrate p38*δ*  (*MAPK13,* p=0.071, Supplementary Figure 1I-J). The pathway is thought to regulate apoptosis in response to stress. The effect of GH on these transcripts has not been reported before, and they could account for the effects of enhanced cell proliferation and apoptosis in response to GH.

**Transcripts regulating lipid metabolism and localization that may contribute to enhanced lipolysis**

To determine the potential causes of the increased lipolysis observed in Figure 1E and F, we examined the expression of human lipase in these adipose tissue lysates. We observed no significant difference in expression of the three classical triglyceride lipolysis enzymes hormone sensitive lipase (*LIPE*), adipose triglyceride lipase (*PNPLA*) or monoglycerol lipase (*MGLL*; Figure 4A). Lipoprotein lipase (*LPL*), the lipase important for lipolysis of triglycerides in apolipoproteins, was significantly more highly expressed in acromegaly patients (Figure 4A). A strong induction of LPL expression in response to GH and absence of change in HSL was demonstrated before in a preadipocyte cell line (23) and in adipose tissue biopsies from GH deficient patients after treatment with GH (24). Notably, studies that addressed LPL enzymatic activity and not expression have found no change or even a reduction in LPL activity in response to GH treatment (25,26).

Although neither Hormone Sensitive Lipase (*LIPE*) or ATGL (*PNPLA2*) were altered, a direct regulator of hormone sensitive lipase and adipose triglyceride lipase activity in adipocytes, abhydrolase domain containing 5 (*ABHD5*, also called CGI58, see Figure 4B) (27), was expressed at higher levels in adipose tissue from acromegaly patients. This suggests that these activators are candidates for the induction of lipolysis by GH.

We also examined the expression of G-protein coupled receptors that induce lipolysis. The 3 adrenergic (*ADRB3*) and Oxytocin receptors (*ACVR1C*), were more highly expressed in acromegaly patients compared to the controls, though at relatively low levels (Figure 4C-D).

Angiopoietin like 4 (*ANGPTL4*), which has been recently shown to be induced in muscle in response to acute GH treatment, and suggested as a possible regulator of lipolysis (28) was not observed to be significantly different between the patients and the controls (p=0.77). As far as glycogen synthesis is concerned, expression of two glycogenolysis enzymes, muscle glycogen phosphorylase (*PYGM*) and phosphohexomutase (*PGM3*), was significantly higher in the acromegaly patients. The significance of these in adipose tissue, or changes in glycogen content in acromegalic adipose tissue has not been characterized.

**Transcripts altered in acromegaly that may contribute to insulin resistance**

We observed no difference in expression of canonical transcripts important for insulin signaling and response to insulin in adipocytes, including insulin receptor (*INSR*), *IRS1, IRS2*, *AKT1-3*, or *SLC2A4* (GLUT4; see Supplementary Figure 2A-G). This indicates that the observed insulin resistance is not caused by transcriptional changes in these genes. In fact, most of these genes trended to be more highly expressed in the adipose tissue from these insulin resistant patients, potentially underlying an upregulation that compensates for an alternative insulin resistant mechanism.

One previously identified candidate is the phosphoinositide-3-kinase, regulatory subunit 1 (*PIK3R1*, also called p85α), which was induced by GH in mouse adipose tissue (29) and thought to contribute to GH-induced insulin resistance (29). In our study it trended to be more highly expressed in the acromegaly patients, but the difference was not statistically significant (P=0.95). *TNF* transcript levels are significantly reduced in acromegalic patients while the proinflammatory cytokines *IL1B*, *IL6* and *MCP1* and the proinflammatory protein kinases *IKKB*, *IKBKE* all trend towards lower expression (data not shown). The JNK and PKC family members were upregulated by more than 15% between controls and acromegaly patients. These data support the hypothesis that insulin resistance in these patients is not due to enhanced inflammatory signaling.

A possible link between acromegaly and insulin resistance may be the increased expression of Stearoyl-CoA desaturase (*SCD*, delta-9-desaturase), a fatty acid desaturase whose activity is associated with metabolic syndrome 2, 3, in acromegaly patients. SCD Expression of has recently been shown to be induced by GH in mice 4,5, Figure 4E).

*TCF7L2*, a transcription factor regulating many metabolism genes known as a diabetes susceptibility gene1 is upregulated in the acromegaly patients. Mice with liver specific knockout of TCF7L2 are hypoglycemic, while transgenic mice overexpressing liver TCF7L2 are hyperglycemic. TCF7L2 in subcutaneous fat is higher and expression of splice isoforms is reduced in subcutaneous fat and in liver following bariatric surgery. Higher expression of TCF7L2 could also also be linked to insulin resistance in acromegaly.

**Glucocorticoid regulation**

11βHydroxysteroid dehydrogenase 1 (*HSD11B1*), the enzyme that activates cortisol, was reduced 4 fold in acromegaly patients (Supplementary Figure 2H). The downregulation of expression and activity of this enzyme by GH/IGF1 has been confirmed both *in vitro* (30), in GH deficient patients treated with GH (31) and in acromegaly patients (32,33). In addition, we found higher expression of the glucocorticoid receptor (*NR3C1*) in acromegaly patients (Supplementary Figure 2I).

**Summary**

In this study we have described a transcriptional signature in adipose tissue from subjects with acromegaly. We identified 103 adipose tissue transcripts altered in acromegaly. Some of these genes may be direct targets of increased GH or IGF-1 signaling in adipose tissue, whereas others may be secondary adaptations to this condition.

The main limitation of our study is the small sample size. The fact that the patients consistently had a relatively uniform change of expression of these genes suggests that we are able to draw valid conclusions about adipose tissue in acromegalic patients even from this small cohort. Furthermore, as mentioned throughout, our data agrees with several previous studies in animal models and patients. The confirmation of these previously reported GH-dependent transcriptional changes strengthens our interpretation of other transcriptional changes.

These data provide a variety of novel transcriptional changes that may be causative of the co-morbidities associated with acromegaly. Further studies in animals and cells using knockout or overexpression of specific transcripts may verify which of the changes is crucial in metabolic effects of GH in adipose tissue.

**Acknowledgements**

We thank Charlotte Gunden, Elizabeth Walkowiak and Eric Vasbinder for their valuable help in the study. We would also like to thank Solomon S. Solomon (UTHSC) for helpful suggestions.

**References**

1. **Vijayakumar A, Novosyadlyy R, Wu Y, Yakar S, LeRoith D** 2010 Biological effects of growth hormone on carbohydrate and lipid metabolism. Growth Horm. IGF Res. 20:1–7

2. **Ezzat S, Forster MJ, Berchtold P, Redelmeier DA, Boerlin V, Harris AG** 1994 Acromegaly. Clinical and biochemical features in 500 patients. Medicine (Baltimore). 73:233–240

3. **Colao A, Baldelli R, Marzullo P, Ferretti E, Ferone D, Gargiulo P, et al.** 2000 Systemic hypertension and impaired glucose tolerance are independently correlated to the severity of the acromegalic cardiomyopathy. J. Clin. Endocrinol. Metab. 85:193–199

4. **Kim D, Pertea G, Trapnell C, Pimentel H, Kelley R, Salzberg SL** 2013 TopHat2: accurate alignment of transcriptomes in the presence of insertions, deletions and gene fusions. Genome Biol. BioMed Central Ltd; 14:R36

5. **Langmead B, Trapnell C, Pop M, Salzberg SL** 2009 Ultrafast and memory-efficient alignment of short DNA sequences to the human genome. Genome Biol. 10:R25

6. **Anders S, Huber W** 2010 Differential expression analysis for sequence count data. Genome Biol. BioMed Central Ltd; 11:R106

7. **Benjamini Y, Hochberg Y** 1995 Controlling the False Discovery Rate: A Practical and Powerful Approach to Multiple Testing. J. R. Stat. Soc. Ser. B 57:289–300

8. **Huo JS, McEachin RC, Cui TX, Duggal NK, Hai T, States DJ, et al.** 2006 Profiles of growth hormone (GH)-regulated genes reveal time-dependent responses and identify a mechanism for regulation of activating transcription factor 3 by GH. J. Biol. Chem. 281:4132–4141

9. **Smyth GK** 2005 Limma: linear models for microarray data. In: Gentleman R, Carey V, Dudoit S, Irizarry R, Huber W, editors. Bioinforma. Comput. Biol. Solut. Using R Bioconductor New York: Springer; p. 397–420

10. **R Development Core Team** 2011 R: A language and environment for statistical computing.

11. **Young MD, Wakefield MJ, Smyth GK, Oshlack A** 2010 Gene ontology analysis for RNA-seq: accounting for selection bias. Genome Biol. 11:R14

12. **Moller L, Norrelund H, Jessen N, Flyvbjerg A, Pedersen SB, Gaylinn BD, et al.** 2009 Impact of growth hormone receptor blockade on substrate metabolism during fasting in healthy subjects. J. Clin. Endocrinol. Metab. 94:4524–4532

13. **Ashburner M, Ball CA, Blake JA, Botstein D, Butler H, Cherry JM, et al.** 2000 Gene ontology: tool for the unification of biology. The Gene Ontology Consortium. Nat. Genet. 25:25–29

14. **Imanishi R, Ashizawa N, Ohtsuru A, Seto S, Akiyama-Uchida Y, Kawano H, et al.** 2004 GH suppresses TGF-beta-mediated fibrosis and retains cardiac diastolic function. Mol. Cell. Endocrinol. 218:137–146

15. **Jenkins PJ, Besser M** 2001 Clinical perspective: acromegaly and cancer: a problem. J. Clin. Endocrinol. Metab. 86:2935–2941

16. **Fleenor D, Arumugam R, Freemark M** 2006 Growth hormone and prolactin receptors in adipogenesis: STAT-5 activation, suppressors of cytokine signaling, and regulation of insulin-like growth factor I. Horm. Res. 66:101–110

17. **Haluzik M, Yakar S, Gavrilova O, Setser J, Boisclair Y, LeRoith D** 2003 Insulin resistance in the liver-specific IGF-1 gene-deleted mouse is abrogated by deletion of the acid-labile subunit of the IGF-binding protein-3 complex: relative roles of growth hormone and IGF-1 in insulin resistance. Diabetes 52:2483–2489

18. **Jørgensen JOL, Jessen N, Pedersen SB, Vestergaard E, Gormsen L, Lund SA, et al.** 2006 GH receptor signaling in skeletal muscle and adipose tissue in human subjects following exposure to an intravenous GH bolus. Am. J. Physiol. Endocrinol. Metab. 291:E899–905

19. **Pasquali C, Curchod M-L, Wälchli S, Espanel X, Guerrier M, Arigoni F, et al.** 2003 Identification of protein tyrosine phosphatases with specificity for the ligand-activated growth hormone receptor. Mol. Endocrinol. 17:2228–2239

20. **Pilecka I, Patrignani C, Pescini R, Curchod M-L, Perrin D, Xue Y, et al.** 2007 Protein-tyrosine phosphatase H1 controls growth hormone receptor signaling and systemic growth. J. Biol. Chem. 282:35405–35415

21. **Sladek R, Rocheleau G, Rung J, Dina C, Shen L, Serre D, et al.** 2007 A genome-wide association study identifies novel risk loci for type 2 diabetes. Nature 445:881–885

22. **Saxena R, Voight BF, Lyssenko V, Burtt NP, de Bakker PIW, Chen H, et al.** 2007 Genome-wide association analysis identifies loci for type 2 diabetes and triglyceride levels. Science 316:1331–1336

23. **Pradines-Figueres A, Barcellini-Couget S, Dani C, Baudoin C, Ailhaud G** 1990 Inhibition by serum components of the expression of lipoprotein lipase gene upon stimulation by growth hormone. Biochem. Biophys. Res. Commun. 166:1118–1125

24. **Khalfallah Y, Sassolas G, Borson-Chazot F, Vega N, Vidal H** 2001 Expression of insulin target genes in skeletal muscle and adipose tissue in adult patients with growth hormone deficiency: effect of one year recombinant human growth hormone therapy. J. Endocrinol. 171:285–292

25. **Simsolo RB** 1995 Effects of acromegaly treatment and growth hormone on adipose tissue lipoprotein lipase. J. Clin. Endocrinol. Metab. 80:3233–3238

26. **Richelsen B, Pedersen SB, Kristensen K, Børglum JD, Nørrelund H, Christiansen JS, et al.** 2000 Regulation of Lipoprotein Lipase and Hormone-Sensitive Lipase Activity and Gene Expression in Adipose and Muscle Tissue by Growth Hormone Treatment During Weight Loss in Obese Patients. 49:906–911

27. **Lass A, Zimmermann R, Haemmerle G, Riederer M, Schoiswohl G, Schweiger M, et al.** 2006 Adipose triglyceride lipase-mediated lipolysis of cellular fat stores is activated by CGI-58 and defective in Chanarin-Dorfman Syndrome. Cell Metab. 3:309–319

28. **Clasen BFF, Krusenstjerna-Hafstrøm T, Vendelbo MH, Thorsen K, Escande C, Møller N, et al.** 2013 Gene expression in skeletal muscle after an acute intravenous GH bolus in human subjects: identification of a mechanism regulating ANGPTL4. J. Lipid Res. 54:1988–1997

29. **Del Rincon J-P, Iida K, Gaylinn BD, McCurdy CE, Leitner JW, Barbour LA, et al.** 2007 Growth hormone regulation of p85alpha expression and phosphoinositide 3-kinase activity in adipose tissue: mechanism for growth hormone-mediated insulin resistance. Diabetes 56:1638–1646

30. **Napolitano a, Voice MW, Edwards CR, Seckl JR, Chapman KE** 1998 11Beta-hydroxysteroid dehydrogenase 1 in adipocytes: expression is differentiation-dependent and hormonally regulated. J. Steroid Biochem. Mol. Biol. 64:251–260

31. **Paulsen SK, Pedersen SB, Jørgensen JOL, Fisker S, Christiansen JS, Flyvbjerg A, et al.** 2006 Growth hormone (GH) substitution in GH-deficient patients inhibits 11beta-hydroxysteroid dehydrogenase type 1 messenger ribonucleic acid expression in adipose tissue. J. Clin. Endocrinol. Metab. 91:1093–1098

32. **Frajese G V, Taylor NF, Jenkins PJ, Besser GM, Monson JP** 2004 Modulation of cortisol metabolism during treatment of acromegaly is independent of body composition and insulin sensitivity. Horm. Res. 61:246–251

33. **Moore JS, Monson JP, Kaltsas G, Putignano P, Wood PJ, Sheppard MC, et al.** 1999 Modulation of 11beta-hydroxysteroid dehydrogenase isozymes by growth hormone and insulin-like growth factor: in vivo and in vitro studies. J. Clin. Endocrinol. Metab. 84:4172–4177

**Table Legends:**

**Table 1: Clinical characteristics.** Data represents mean +/- standard error.

**Table 2:** **Gene set enrichment of transcripts from acromegalic white adipose tissue.** Gene ontology biological process categories enriched by in transcripts that differ between acromegaly and control subjects. Type is the GO or KEGG classification of the term. DE indicates the number of differentially expressed genes in this category, Total is the total number of genes in that category. Lists were filtered to remove redundant categories.

**Figure Legends**

**Figure 1: Acromegalic patients tended to have reduced insulin sensitivity and higher lipolytic activity than their controls.** A) Fasting blood glucose levels. B) Fasting insulin levels. C) HOMA-IR score from Control or Acromegaly subjects. D) Plot of each subject’s BMI compared with the natural logarithm of their HOMA-IR score. A best fit line was drawn for each subject pool. E) *ex vivo* lipolysis as measured by glycerol release from excised white adipose tissue from control or acromegaly patients left untreated (Basal) or after stimulation with 30 nM isoproterenol (Iso). Data is presented as mean +/- standard error of the mean.

**Figure 2: Differential expression of genes in white adipose tissue from subject with compared to controls.** A) Heatmap of the differentially expressed genes in white adipose tissue. In where several transcripts derived from a single were identified as differentially expressed, only the most significant was shown. Individual values are colored as the log fold change for a particular gene in a particular subject compared to the average expression of that gene across all cohorts, with brown indicating less expression and green indicating more expression (designated in the key as Row Z-score). The bar across the top indicates the subject’s diagnosis, red for acromegaly and blue for controls.

**Figure 3: Previously reported GH targets are differentially expressed in acromegaly subjects.** A) mRNA Expression of A) *IGF1* and B) *IGFBP3* transcript levels in adipose tissue from control and acromegalic (Acro.) patients. Only the highest expressing transcript is shown for each gene. C) Comparason between *IGF1* mRNA and IGF-1 serum levels in patients with acromegaly. mRNA Expression of *CISH* and *SOCS2* in adipose tissue from acromegaly patients and controls. Asterisks indicate p<0.05. Barplots are presented as mean +/- standard error of the mean.

**Figure 4: Expression changes of selected genes, potentially responsible for lipolytic or insulin sensitivity alterations in acromegaly patients.** mRNA Expression profile of genes potentially involved in insulin sensitivity (A-B) or lipid metabolism (C-E). C) Lipases in human tissue were examined, only lipases with >100 FPKM are shown. Asterisks indicate p<0.05. Data indicates mean +/- standard error of the mean.

**Supplementary Data**

**Supplementary Table 1: Expression changes between control and acromegaly subjects.** Calculated expression, and expression changes for each transcript are shown along with raw, and adjusted p-values and the fold change for acromegaly data.

**Supplementary Figure 1: Expression changes of selected other transcripts.** mRNA Expression profile. Data indicates mean +/- standard error of the mean. Asterisk indicates p<0.05

**Supplementary Figure 2: Expression changes of selected insulin signal transduction genes.** mRNA Expression profile of genes involved in insulin signaling. Data indicates mean +/- standard error of the mean.

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