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Solution to Exercise 2 (Image Processing and Pattern Recognition)

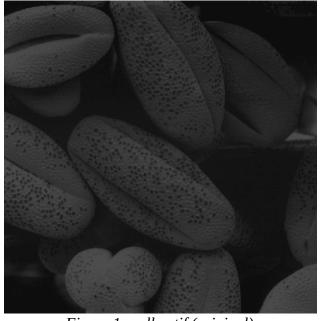
Q1. Load the images pollen.tif and bone-scan-GE.tif. Display them with imtool. Look at the pixel values in some area. Convert them into double. Display and look at the values again.

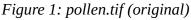
Solution

Matlab Script

```
pollen = imread('C:\Users\HP\Desktop\IPPLAssignment2\pollen.tif');
imtool(pollen)
pollen_double = double(pollen);
imtool(pollen_double)
bone = imread('C:\Users\HP\Desktop\IPPLAssignment2\bone-scan-GE.tif');
imtool(bone)
bone_double = double(bone);
imtool(bone double)
```

Matlab Output





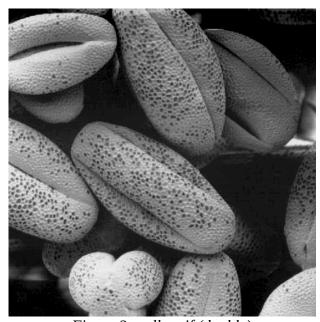


Figure 2: pollen.tif (double)

Figure 2 is contrast adjusted image (from [0,1] to [13,83]) using Adjust Constrast Tool of imtool. The pixel values in both original and double image is found out to be same for both images. Also contrast is adjusted from [0,1] to [0.92] in Figure 4 as well. It is clear that left bone scan is front side scan and right bone scan is the back side scan of a person



Figure 4: bone-scan-GE.tif (original)



Figure 3: bone-scan-GE.tif (double, adjusted)

Q2. Use thresholding and regionprops to count the nuclei in the following images:

nuclei1.tif nuclei2.tif

How many nuclei are there in the picture?

What is the average area of a nucleus?

Which image was easier? Why? How can the segmentation be improved?

Try to write your programs in a generic fashion.

Comment and maybe even include functions.

Solution

Matlab Script

```
function e2q2()
    clc;
    clear;
    imtool close all;
```

```
% Exercise 2 Question 2
function count_nuclei(file_path, file_name)
    if exist([file path, file name])
        disp(['For image: ', file name]);
        nuclei1 = imread(strcat(file path, file name));
        imtool(nuclei1)
        % imhist(nuclei1)
        nuclei1 gray = mat2gray(nuclei1);
        T nuclei1 gray = graythresh(nuclei1 gray);
        nuclei1_gray_binary = im2bw(nuclei1_gray, T_nuclei1_gray);
        imtool(nuclei1 gray binary)
        [L_nuclei1, num_nuclei1] = bwlabel(nuclei1_gray_binary);
        disp([' Number of nuclei: ', num2str(num nuclei1)]);
        D_nuclei1 = regionprops(L_nuclei1, 'area');
        area list nuclei1 = [D nuclei1.Area];
        avg_area_nuclei1 = mean(area_list_nuclei1);
        disp([' Avg Area of nuclei: ', num2str(avg area nuclei1)]);
        disp(' ');
        % hist(area_list_nuclei1)
    else
        disp(['For image: ', file name]);
        disp(' This file path is invalid');
        disp(' ');
    end
end
count nuclei('C:\Users\HP\Desktop\IPPLAssignment2\', 'nuclei1.tif')
count nuclei('C:\Users\HP\Desktop\IPPLAssignment2\', 'nuclei2.tif')
```

Matlab Output

end

For image: nuclei1.tif
Number of nuclei: 80

Avg Area of nuclei: 1113.125

For image: nuclei2.tif
Number of nuclei: 18359
Avg Area of nuclei: 4.8994

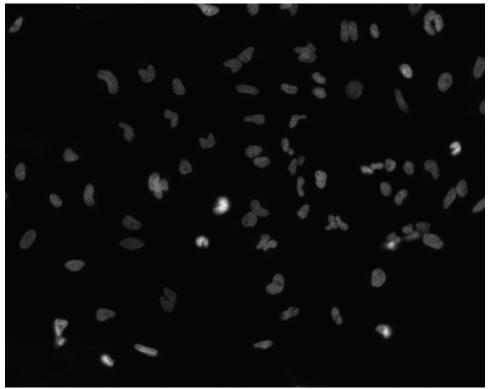


Figure 5: nuclei.tif (original)



Figure 6: nuclei1.tif (grayscaled, binary)



Figure 7: nuclei2.tif (original)

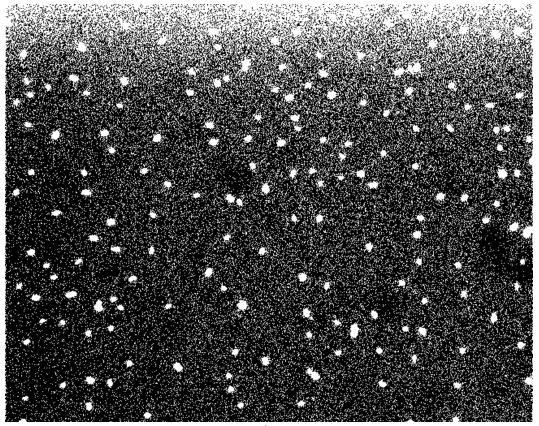


Figure 8: nuclei2.tif (grayscaled, binary)

Analysis

The total number of nuclei in *Figure 5* is **80** with average area of **1113.125** and the total number of nuclei in *Figure 7* is **18359** with average area of **4.8994**.

Figure 5 is easier than Figure 7 to interpret. In Figure 7, the nuclei were hidden due to image contrast and by using *thresholding operation* in Matlab, we can see the hidden nuclei as shown in figure 8. Finally, the number of nuclei were counted using regionprops command with Area property

Q3. Eliminate "bad" rice granules from the rice.png picture. Use regionprops to identify the rice you want to eliminate. Use the find function to change eliminate these grains from the labeled image. (You will probably want to write a loop that goes through each object in the labeled file)

Compare the original image to your new image (imshow and hist). How well did you do? Qualitatively, how many real grain got eliminated/how many bad grains remain?

Solution

Matlab Script

```
clc;
clear;
close all;

figure('Name', 'Question 3', 'NumberTitle', 'off');
font_size = 10;

% read image: rice.png
rice = imread('C:\Users\HP\Desktop\IPPLAssignment2\rice.png');

% show actual image: rice.png
subplot(2, 3, 1);
imshow(rice);
title('Rice (Actual Image)', 'FontSize', font_size, 'Interpreter', 'None');

% change png image to grayscale and calculate the threshold value,
% then create a new binary image using this calculated threshold value
rice_gray = mat2gray(rice);
T_rice_gray = graythresh(rice_gray);
```

```
rice gray binary = im2bw(rice gray, T rice gray);
% plot gray image and binary image
subplot(2, 3, 2);
imshow(rice gray);
title('Rice (Grayscale)', 'FontSize', font size, 'Interpreter', 'None');
% imtool(rice gray)
subplot(2, 3, 3);
imshow(rice gray binary);
title('Rice (Binary)', 'FontSize', font size, 'Interpreter', 'None');
% imtool(rice gray binary)
% enumerate all the objects in the figure
[L rice, num rice] = bwlabel(rice gray binary);
% then display no of object (this gives number of rice grain
disp(['Number of rice grain: ', num2str(num rice)]);
% num rice
% regionprops command is used to obtain quantitative information about the objects
% gray image is needed for 'MeanIntensity' for 'table'
D_rice = regionprops('table', L_rice, rice_gray, 'Area', 'MeanIntensity',
'SubarrayIdx');
% D rice = regionprops(L rice, 'area');
% making copy of binary image for two different analysis
rice gray binary good area = rice gray binary;
rice gray binary good mi = rice gray binary;
% sorting Area prop for analysis
area list rice = [D rice.Area];
sort(area list rice, 'descend');
% specify threshold value from sorted list
% then remove all those element that do not match threshold value
thres = 120;
D rice.isGreater = D rice.Area > thres;
objRemoveIdxA = find(~D rice.isGreater);
```

```
for i = find(~D rice.isGreater).'
     rice_gray_binary_good_area(D_rice.SubarrayIdx{i,1},D_rice.SubarrayIdx{i,2}) =
false;
end
% imtool(rice gray binary);
% plot this binary image (gives only good rice grain using Area prop)
subplot(2, 3, 4);
imshow(rice gray binary good area);
title('Rice (Binary, Good Area)', 'FontSize', font size, 'Interpreter', 'None');
% display number of eliminated rice grains
eliminated rice grains = length(objRemoveIdxA);
disp(['Number
               of
                      eliminated
                                     rice
                                            grains
                                                        (prop: Area):
                                                                              ٠,
num2str(eliminated rice grains)]);
% sorting MeanIntensity prop for analysis
mi list rice = [D rice.MeanIntensity];
sort(mi_list_rice, 'descend');
% specify threshold value from sorted list
% then remove all those element that do not match threshold value
thres = 0.80;
D rice.isGreater = D rice.MeanIntensity > thres;
objRemoveIdxMI = find(~D rice.isGreater);
for i = find(~D rice.isGreater).'
       rice gray binary good mi(D rice.SubarrayIdx{i,1},D rice.SubarrayIdx{i,2}) =
false;
end
% plot this binary image (gives only good rice grain using MeanIntensity prop)
subplot(2, 3, 5);
imshow(rice gray binary good area);
title('Rice (Binary, Good MI)', 'FontSize', font size, 'Interpreter', 'None');
% display number of eliminated rice grains
eliminated rice grains = length(objRemoveIdxMI);
disp(['Number
                of
                       eliminated
                                     rice
                                                grains
                                                        (prop: MI):
num2str(eliminated rice grains)]);
```

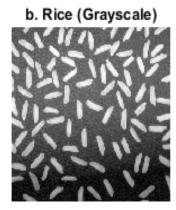
Matlab Output

Number of rice grain: 151

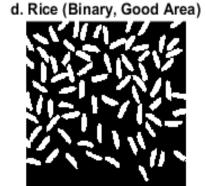
Number of eliminated rice grains (prop: Area): 78

Number of eliminated rice grains (prop: MI): 93

a. Rice (Actual Image)







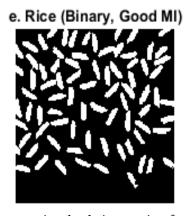


Figure 9: Separating bad rice grains from rice.png

<u>Analysis</u>

The original image (Figure 9a.) has darker side at the bottom of the image. So, when applying grayscaling function (Figure 9b.), some of the good rice grains were not detected properly in binary image (Figure 9c.) and thus around *eight* (8) good grains were eliminated in our analysis. Also the rice grains around the edges of the image is connected so that the area of those half and quarter rice grains are detected big. So we can see some half and quarter sized rice grains around the edges (Figure 9d and Figure 9e.). As such around *seven* (7) bad grains remained after the analysis. But looking at the overall detection, we can say that almost all of the bad grains are completely removed from the original image.