# RESEARCH PAPER



# Continental-scale hotspots of pelagic fish abundance inferred from commercial catch records

Scott L. Nichol<sup>2</sup> | M. Julian Caley<sup>4,5</sup> | Reg A. Watson<sup>6,7</sup>

<sup>1</sup>Centre for Marine Futures & UWA Oceans Institute, School of Biological Sciences, University of Western Australia, Crawley, Western Australia, 6009, Australia

<sup>2</sup>Geoscience Australia, Canberra, Australian Capital Territory, 2601, Australia

<sup>3</sup>Institute of Zoology, Zoological Society of London, London, NW1 4RY, United Kingdom

<sup>4</sup>School of Mathematical Sciences, Oueensland University of Technology. Brisbane, Queensland, 4000, Australia

<sup>5</sup>Centre of Excellence for Mathematical and Statistical Frontiers, Australian Research Council, Parkville, Victoria, Australia

<sup>6</sup>Institute for Marine and Antarctic Studies, University of Tasmania, Taroona, Tasmania, 7001. Australia

<sup>7</sup>Centre for Marine Socioecology, Hobart, Tasmania, 7001, Australia

## Correspondence

Phil J. Bouchet, Centre for Marine Futures M092. The University of Western Australia. 35 Stirling Highway, Crawley, WA, 6009, Australia.

Email: phil.bouchet@uwa.edu.au

#### Funding information

Australian Government's Department of Sustainability, Environment, Water, Population and Communities; University of Western Australia; Australian Research Council, Grant/Award Number: DP140101377

Editor: Amanda Bates

## Abstract

Aim: Protected areas have become pivotal to the modern conservation planning toolbox, but a limited understanding of marine macroecology is hampering their efficient design and implementation in pelagic environments. We explored the respective contributions of environmental factors and human impacts in capturing the distribution of an assemblage of commercially valuable, largebodied, open-water predators (tunas, marlins and mackerels).

Location: Western Australia.

Time period: 1997-2006.

Major taxa studied: Pelagic fishes.

Methods: We compiled 10 years of commercial fishing records from the Sea Around Us Project and derived relative abundance indices from standardized catch rates while accounting for confounding effects of effort, year and gear type. We used these indices to map pelagic hotspots over a 0.5°-resolution grid and built random forests to estimate the importance of 33 geophysical, oceanographic and anthropogenic predictors in explaining their locations. We additionally examined the spatial congruence between these hotspots and an extensive network of marine reserves and determined whether patterns of co-occurrence deviated from random expectations using null model simulations.

Results: First, we identified several pelagic hotspots off the coast of Western Australia. Second, geomorphometrics explained up to 50% of the variance in relative abundance of pelagic fishes, and submarine canyon presence ranked as the most influential variable in the North bioregion. Seafloor complexity, geodiversity, salinity, temperature variability, primary production, ocean energy, current regimes and human impacts were also identified as important predictors. Third, spatial overlap between hotspots and marine reserves was limited, with most high-abundance areas primarily found in zones where anthropogenic activities are subject to few regulations.

Main conclusions: This study reveals geomorphometrics as valuable indicators of the distribution of mobile fish species and highlights the relevance of harnessing static topography as a key element in any blueprint for ocean zoning and spatial management.

# KEYWORDS

catch rate standardization, fisheries data, geomorphometrics, hotspot, marine reserves, pelagic fishes, random forest, spatial modelling, submarine canyons, topography

## 1 | INTRODUCTION

The past decades have seen unprecedented changes in the abundance of marine living resources. Despite divergent perspectives on the present status and future prospects of the world's fisheries (Daan, Gislason, Pope, & Rice, 2011; Froese, Zeller, Kleisner, & Pauly, 2012), overexploitation since the 1950s is widely accepted as a catalyst of modern declines in elasmobranchs (sharks, skates and rays) and teleosts globally (Letessier, Bouchet, & Meeuwig, 2017). Although efforts to rebuild fisheries are now underway in several ecosystems (Worm et al., 2009). their success to date remains relatively modest because insufficient control on current exploitation rates often precludes recovery, even for resilient stocks that may have adapted to moderate levels of extractive pressure (Neubauer, Jensen, Hutchings, & Baum, 2013).

In this context, a new generation of multilateral environmental treaties has emerged to reverse the large-scale erosion of biodiversity. The United Nations Convention on Biological Diversity (CBD; http://www.cbd.int/) is perhaps one of the best-known examples and presently binds 196 countries to take legislative and policy action to tackle the loss of threatened species. A key element of the convention's strategic plan is Aichi Target 11, which commits to expand the coverage of marine reserves (MRs) to at least 10% of the world's ocean by 2020. Although the ecological and socio-economic merits of MRs are well established in coastal systems (e.g., Angulo-Valdés & Hatcher, 2010; Kerwath, Winker, Götz, & Attwood, 2013), their utility in pelagic environments remains more contentious, in part owing to the perception that mobile species require protection over too large a geographical space to be logistically, politically or financially practical to implement or enforce (Kaplan, Chassot, Gruss, & Fonteneau, 2010). However, such assumptions rarely account for heterogeneous population structuring (Grewe et al., 2015), partial migration, residency, site fidelity, philopatry (Chapman, Feldheim, Papastamatiou, & Hueter, 2015), evolutionary reductions in mobility (Mee, Otto, & Pauly, 2017) and predictable aggregative behaviour in upper-trophic-level organisms (e.g., Kessel et al., 2014). There is now growing consensus that even mobile predators with extensive home ranges, such as seabirds (Young, Maxwell, Conners, & Shaffer, 2015), cetaceans (Gormley et al., 2012), turtles (Scott et al., 2012), fishes (Kerwath et al., 2009) and sharks (White et al., 2017), can benefit from spatial closures (Breen, Posen, & Righton, 2015), provided mortality rates do not rise disproportionately in adjacent unprotected waters as a result of effort displacement (Powers & Abeare, 2009). This is especially true where reserves strategically encompass core habitat areas or sites in which key life-cycle events perennially occur (Hooker, Hyrenbach, Corrigan, Polovina, & Reeves, 2011).

Sparse ecological data in remote offshore waters [i.e., beyond the territorial sea baseline, both within exclusive economic zones (EEZs) and beyond; Webb, Vanden Berghe, & O'Dor, 2010] make the direct identification of critical habitat difficult and call for robust surrogates of biological diversity to predict, delineate and prioritize candidate sites for zoning (McArthur et al., 2010). Complex topography has been recognized as a determinant of wildlife dynamics across numerous taxa

(Bouchet, Meeuwig, Salgado Kent, Letessier, & Jenner, 2015), which suggests that locating protected areas over sites of rugged terrain could yield a range of conservation gains (Harris & Whiteway, 2009; Michael, Jahncke, & Hyrenbach, 2014). Morato et al. (2008), Morato, Hoyle, Allain, and Nicol (2010) and Worm, Lotze, and Myers (2003) illustrated this possibility in the open ocean by showing that North Atlantic and Pacific seamounts were important centres of taxonomic richness of special interest for the management of threatened vertebrates. Nevertheless, whether static topography could be widely used as a key element in ocean planning remains to be investigated in other ocean basins and for different types of geomorphologies. Submarine canyons, for instance, are prominent and commonly occurring physical seabed features throughout the world's oceans. Although their role as drivers of primary productivity, plankton abundance and benthic biomass is well documented (Fernandez-Arcaya et al., 2017), our understanding of their importance to pelagic megafauna remains nascent and somewhat skewed towards marine mammals (e.g., Moors-Murphy, 2014).

Here, we combine long-term landings and fishing effort datasets from the Sea Around Us Project (http://www.seaaroundus.org) with a recent reclassification of Australian submarine canyons (Huang, Nichol, Harris, & Caley, 2014) to: (a) determine the location of abundance hotspots for a suite of commercially important predatory pelagic fishes within the western part of Australia's exclusive economic zone (hereafter 'wEEZ'); (b) examine associations between these hotspots and an array of abiotic variables, including seabed topography, on a continental scale; and (c) assess their spatial congruence with a subset of Australia's 3.1 million km<sup>2</sup> national network of Commonwealth Marine Reserves (http://www.environment.gov.au/topics/marine/marine-reserves).

# 2 | MATERIALS AND METHODS

## 2.1 | Fish and environmental data

Annual commercial fishing records for the wEEZ were extracted from the quality-checked databases compiled by Sea Around Us but did not include the recent catch reconstructions that capture estimates of recreational and illegal fishing (Pauly & Zeller, 2016). These data are global, readily accessible (http://www.seaaroundus.org/data/), transparent, vetted through extensive peer review and were adopted as the only spatially resolved dataset available that currently allowed analyses at macroecological scales. Landings (in tonnes) originated primarily from yearly catch reports (corrected for discarded bycatch) produced by the Food and Agriculture Organization (FAO) (Watson, Alder, Kitchingman, & Pauly, 2005; see Supporting Information Appendix S1). Effort statistics were obtained from an array of public domain sources (Watson et al., 2013) and standardized to a common unit of vessel engine power and operation time (kilowatt sea days; kwsd), following Anticamara, Watson, Gelchu, and Pauly (2011). Both catch and effort were disaggregated into a grid of  $0.5^{\circ}$  (latitude)  $\times$   $0.5^{\circ}$  (longitude) spatial cells using a rule-based algorithm, a technical description of which is provided elsewhere (Watson, Kitchingman, Gelchu, & Pauly, 2004; Watson et al., 2013). The full dataset used here spanned the period 1950WILEY

2006 and comprised 5,640,222 entries of 111 species and higher taxa, representing a total catch of 3.11 million tonnes extracted over  $3.35 \text{ million km}^2$  of the eastern Indian Ocean between 93 and 129° E and 8 and 39° S.

We also collated a biophysical dataset composed of 51 variables computed over the same  $0.5^{\circ}$  cells as the fisheries data (Table 1, Supporting Information Appendix S3, Table S1, and Supporting Information Appendix S2, Figure S1). Most of these variables were geomorphometrics (measures of seabed complexity, n=20) assembled from existing archives curated by Geoscience Australia, including a newly revised digital catalogue of submarine canyons (Huang et al., 2014). Recognizing that mesoscale hydrographic features attract mobile megafauna (Scales et al., 2014) and that human activities can affect spatial patterns in biodiversity by driving changes in species composition and abundances (Navarro, Coll, Cardador, Fernández, & Bellido, 2015), the dataset also included a number of oceanographic factors (n=14; Table 1, Supporting Information Appendix S3, Table S1, and Supporting Information Appendix S4) as well as indices of cumulative anthropogenic threats (n=3; Table 1 and Supporting Information Appendix S3, Table S1).

# 2.2 Data processing

Our stepwise approach to data preparation and analysis is illustrated in Figure 1, and a brief description of each step follows below (see Supporting Information Appendix S1 for full details). Analyses were carried out in R 3.3.1 (R Foundation for Statistical Computing) and Matlab 2012a (MathWorks, Inc.).

## 2.2.1 | Gear matching

Catch and effort were estimated independently. Landing records were segregated by species and fishing method and were well resolved both spatially and temporally. In contrast, the effort data were incomplete and reported on the basis of gear rather than taxa. To guarantee compatibility between the two datasets, we re-allocated every fishing gear type to one of five discrete and mutually exclusive categories, namely gillnet (GIL), seine (SEI), trawl (TRW), line (LIN) and miscellaneous (OTH) (Supporting Information Appendix S2, Figure S2). We used this classification key as a common denominator to pair all landings with a corresponding value of effort for any combination of year × grid cell using a purpose-built Microsoft Access 2011 (Microsoft Corporation) query.

### 2.2.2 | Filtering

We filtered the data to generate a time series spanning the period 1997–2006, excluding all demersal or benthopelagic species caught and all grid cells situated outside the continental wEEZ (Supporting Information Appendix S1 and Appendix S3, Table S2). Catch records for unidentified species were discarded, and those reported only in families and genera were reapportioned to confirmed species in proportion to their relative contribution to the total family- or genusspecific catch. Finally, gears contributing < 5% of total landings were omitted, effectively removing all benthic fishing from our study area. Clear outliers (see Supporting Information Appendix S1 for definition), including particularly small coastal grid cells and exaggerated estimates

of effort, were also excluded (Supporting Information Appendix S3, Table S2).

# 2.2.3 | Geographical partitioning

To control for spatial non-stationarity, we subdivided the consolidated data into four contiguous bioregions: North, Gascoyne, West and South. These reflected broadly homogeneous environmental conditions and biological assets (Supporting Information Appendix S2, Figure S3), consistent with management boundaries recognized by the Western Australian Department of Fisheries (http://www.fish.wa.gov.au/).

#### 2.2.4 | Imputation of missing effort data

Although the filtered landings achieved full temporal and spatial coverage, with a positive catch value for each grid cell  $\times$  year combination, available effort estimates were incomplete and required reconstruction in locations where none existed (Supporting Information Appendix S2, Figure S4). We performed gap filling on the effort matrix using the *smoothn* package (https://au.mathworks.com/matlabcentral/fileex-change/25634-fast-n-easy-smoothing) introduced by Garcia (2010), which applies penalized regression to smooth evenly gridded data in multidimensional space. The algorithm was executed on a 3-D space-time cube (year  $\times$  latitude  $\times$  longitude) of effort values, summed across fishing gear types owing to data scarcity in some grid cells (Supporting Information Appendix S2, Figure S5). After imputation, effort predictions were re-allocated to all gears proportionally to their original usage rate in (a) each cell  $\times$  year when cell-level data existed or (b) the bioregion as a whole when cell-level data were missing.

# 2.2.5 | Catch rate standardization

The use of fisheries-dependent data for ecological inference hinges on standardizing the catch per unit effort (CPUE) to control for confounding elements that may affect catchability (Maunder & Punt, 2004). We constructed lognormal generalized linear models (GLMs) to standardize CPUE values for each bioregion (Supporting Information Appendix S2, Figure S6) across year, fishing gear category, grid cell and species body weight (as a proxy for size-mediated gear selectivity; Supporting Information Appendix S1). A subset of nine plausible models was assessed and competing formulations were ranked based on their second-order (corrected) Akaike information criterion scores (AICc; Table 2). We forced the inclusion of grid cell as a factor in all models, but its interaction with year was not considered owing to the paucity of records on a per year and cell basis. Following Francis (1999), the coefficients (β) of the grid cell term were scaled to canonical form to guarantee invariance to the choice of baseline level. Scaled values were taken as relative fish abundance indices and calculated for grid cell i as:  $\beta_i' = \frac{\beta_i}{\bar{\rho}}$ , where  $\bar{\beta}$  is the geometric mean of all  $\beta$ , defined as  $\bar{\beta} = \sqrt[n]{\prod_{i=1}^{n} \beta_i}$ .

## 2.2.6 | Random forests

We gauged quantitative relationships between predictor variables and abundance indices with random forests (Breiman, 2001), using the party R package (Strobl, Hothorn, & Zeileis, 2009) to construct 2,000 unbiased conditional inference trees of fish abundance (on the log scale) in each bioregion (Strobl, Boulesteix, Zeileis, & Hothorn, 2007).



Variable	Name	Unit	State	Ecological interpretation
Bathymetric/to	pographic			
CI	Contour index	d.u.	Static	Vertical circulation and mixing, eddy formation, prey refugia, prey entrapment
CRS	Cross-sectional curvature	rad/m	Static	Vertical circulation and mixing, eddy formation, prey refugia, prey entrapment
FRD	Fractal dimension	d.u.	Static	Vertical circulation and mixing, eddy formation, prey refugia, prey entrapment
LSRI	Land surface ruggedness index	d.u.	Static	Vertical circulation and mixing, eddy formation, prey refugia, prey entrapment
RUG	Rugosity	d.u.	Static	Vertical circulation and mixing, eddy formation, prey refugia, prey entrapment
Geological				
CAN	Presence/absence of one or more submarine canyons	d.u.	Static	Breeding/spawning habitat, migration cue, prey availability
CANpercent	Percentage of grid cell area occupied by submarine canyons	%	Static	Breeding/spawning habitat, migration cue, prey availability
CANhead	Presence/absence of one or more canyon heads	d.u.	Static	Productivity (upwelling), food availability, feeding ground
CANadj	Number of adjacent cells containing submarine canyons	d.u.	Static	Population connectivity, larval dispersal
CANdepth	Maximum canyon depth within a cell	m	Static	Prey availability, breeding/spawning habitat
FEATcount	Number of distinct geomorphic features within a cell	d.u.	Static	Prey and habitat diversity
FEATdom	Dominant geomorphic feature class	d.u.	Static	Prey availability, breeding/spawning habitat
Oceanographic				
CUREW	East-west current velocity	m/s	Dynamic	Nutrient inputs, oxygenation, enhanced productivity, larval drift and juvenile recruitment
CURNS	North-south current velocity	m/s	Dynamic	Nutrient inputs, oxygenation, enhanced productivity, larval drift and juvenile recruitment
FFD	Daily sea surface temperature frontal frequency	%	Dynamic	Food availability, migration cue
L2	Regional circulation regimes	d.u.	,	Eddy formation, enhanced primary and secondary production
L3	Ocean energy	d.u.	Dynamic	Prey availability, breeding habitat, feeding success, larval growth rates
MIX	Mixed layer depth	m		Prey availability, physiological tolerance, oxygen availability
PP	Annual mean primary production			Prey availability
PPstd SAL	SD of annual mean primary production Annual mean salinity at the surface	mg C/m²/day PSU		Prey availability Prey availability, physiological tolerance, hatching rate
SSTstd	SD of sea surface temperature	°C	,	Spawning cue, breeding habitat, metabolic stress
WAT	Water mass at surface	d.u.	,	Prey availability, physiological tolerance
Anthropogenic				
Hi	Mean cumulative human impact	d.u.	,	Mortality, sublethal disturbance, displacement
Hir Him	Range of cumulative human impacts  Maximum cumulative human impact	d.u. d.u.		Mortality, sublethal disturbance, displacement Mortality, sublethal disturbance, displacement
HIIII	Maximum cumulative numan impact	u.u.	Dynamic	iviortaiity, subjettiai disturbance, displacement

Note. Only the top 15 random forest predictors from each bioregion are shown (Figure 3). See Supporting Information Appendix S3, Table S1 for the full list and links to data sources. Geomorphometrics encompass both 'bathymetric/topographic' and 'geological' parameters. d.u. = dimensionless unit.

For comparison, we developed models from both the full environmental dataset and a reduced set comprising geomorphometrics only. We also assessed the importance of each predictor through unconditional permutation (Strobl, Boulesteix, Kneib, Augustin, & Zeileis, 2008) and determined how many top-ranking variables should be designated as truly 'important' by implementing a modified version of the recursive selection protocol described by Sabatia and Burkhart (2014) (see Supporting Information Appendix S1 for full details).

#### 2.2.7 | Hotspot detection

Bartolino, Maiorano, and Colloca (2011b) proposed cumulative relative frequency distributions (CRFDs) as a simple way of delineating spatial hotspots in a continuous parameter of interest (e.g., density, diversity).

The method is objective and preferable to the more traditional Getis-Ord Gi\* or Moran's I, which rely heavily on a priori user-selected settings (Bartolino, Maiorano, & Colloca, 2011a). We obtained CRFD curves by plotting the relative values of fish abundance indices in each bioregion against their own frequency distribution. The majority of CRFDs derived from biological data should increase monotonically and approach an upper asymptote, such that tangents to the curve can then be interpreted as rates of accumulation, with high-density areas (hotspots) characterized by tangent slopes  $<45^\circ$ . Instead of empirically approximating tangents from pairs of adjacent points, we fitted local polynomial regressions (LOESS smoothers, obtained from the R package fANCOVA; Wang, 2010) to the CRFDs (Supporting Information Appendix S2, Figure S7), with smoothing span widths chosen according

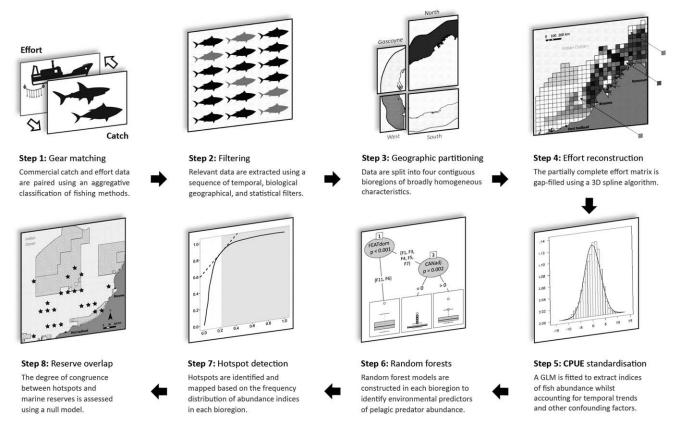


FIGURE 1 Graphical representation of the grooming and analysis of the Sea Around Us data

TABLE 2 Model selection summary from the generalized linear model standardization of pelagic fish catch rates

	North Gascoyne		West		South			
Model formulation	ΔAICc	Adjusted R <sup>2</sup>	ΔAICc	Adjusted R <sup>2</sup>	ΔAICc	Adjusted R <sup>2</sup>	ΔAICc	Adjusted R <sup>2</sup>
$\alpha + CeII + Year + log(E) + log(A) + \epsilon$	23,976	.355	9,372.1	.204	10,611	.235	9,911	.203
$\alpha + CeII + Year + Gear + log(E) + log(A) + \epsilon$	10,541	.456	4,687.1	.347	6,445	.356	8,411	.256
$\alpha + \text{Cell} + \text{Year} + \text{Gear} + \text{Weight} + \text{log(E)} + \text{log(A)} + \epsilon$	4,928	.459	3,063.7	.347	3,997	.369	1,936.6	.384
$\begin{array}{l} \alpha + \text{Cell} + \text{Year} + \text{Gear} + \text{Weight} + \text{Dport} \\ + \log(\text{E}) + \log(\text{A}) + \epsilon \end{array}$	4,928	.459	3,063.7	.347	3,997	.369	1,936.6	.384
$\alpha + \text{Cell} + \text{Year} + \text{Gear} \times \text{Weight} + \text{log(E)} + \text{log(A)} + \epsilon$	3,739	.468	2,838.1	.354	3,604.9	.380	1,249.3	.404
$\alpha + Cell + Year + Gear \times Dport + log(E) + log(A) + \epsilon$	10,389	.457	3,940.2	.368	5,326	.385	8,281	.261
$\begin{array}{l} \alpha + \text{Cell} + \text{Year} \times \text{Dport} + \text{Gear} + \text{Weight} \\ + \log(\text{E}) + \log(\text{A}) + \epsilon \end{array}$	4,478	.463	2,425.5	.365	3,613.2	.380	1,785.1	.389
$\begin{array}{l} \alpha + Cell + Year \times Gear + Weight + Dport \\ + log(E) + log(A) + \epsilon \end{array}$	3,182	.472	767.8	.409	956.3	.445	988.3	.412
$\begin{array}{l} \alpha + Cell + Year \times Gear \times Weight + log(E) \\ + log(A) + \epsilon \end{array}$	0	.493	0	.428	0	.467	0	.440

Note. The best model [with minimal second-order corrected Akaike information criterion (AICc) and maximal adjusted  $R^2$ ] is shown in bold.  $\Delta$ AICc quantifies the difference in AICc score between the current and best models. The response variable was the log of catches (in tonnes).  $\alpha$  is the intercept,  $\epsilon$  is the residual variation, log(E) and log(A) are offset terms for fishing effort and water surface area respectively, and the standard notation '×' represents all covariate main effects and their associated interactions. Weights are the species-specific values reported in Supporting Information, Appendix 3, Table S3.

to the bias-corrected AICc. This approach is less sensitive to noise and enables a more robust detection of fish hotspots that reflects fundamental ecological signals rather than data idiosyncrasies.

## 2.2.8 | Spatial overlap between MRs and hotspots

We matched hotspots with MRs and measured their spatial congruence according to Jaccard's similarity coefficient *J* (Real & Vargas, 1996). We developed null models to determine the probability of obtaining these patterns by chance, with the null expectation that hotspots could occur anywhere within each bioregion. Following Leroux, Schmiegelow, and Nagy (2007), this was achieved by randomly selecting without replacement the same number of grid cells as identified hotspots, calculating *J*, and reiterating this permutation 10,000 times. We then compared the simulated distribution of Jaccard indices to the corresponding observed values via two-sample Mann–Whitney–Wilcoxon tests. The same steps were followed to quantify overlap with Marine National Parks (MNPs).

## 2.2.9 | Uncertainty propagation

Estimates of variance for relative abundance indices, predictor rankings, hotspot thresholds and Jaccard values were acquired by applying a non-parametric bootstrapping procedure, which resampled the CPUE data randomly and with replacement 100 times.

## 3 | RESULTS

# 3.1 | Fish landings and abundance indices

The catch data consisted of 23 pelagic species (22 teleosts, one elasmobranch), of which 12 are highly migratory (Annex I of the 1982 Convention on the Law of the Sea; Supporting Information Appendix S3, Table S3). Mackerels and tunas dominated, with six species (*Katsuwonus pelamis*, skipjack tuna; *Thunnus maccoyii*, southern bluefin tuna; *Thunnus albacares*, yellowfin tuna; *Thunnus obesus*, bigeye tuna; *Scomberomorus commerson*, narrow-banded Spanish mackerel; and *Trachurus declivis*, greenback horse mackerel) making up nearly 75% of all landings over the 10 years of the study. Northern fisheries contributed nearly twice as much to total catches (37%) as those operating in other bioregions (18% Gascoyne, 23% West and 22% South).

The GLM formulation that minimized the AICc contained a three-way interaction between year, fishing gear category and species body weight (Table 2). Bootstrap catch rate standardization models explained between 45 and 50% of the deviance [mean adjusted  $R^2$  and 95% percentile confidence interval, North: 0.5 (0.49–0.5); Gascoyne: 0.44 (0.43–0.45); West: 0.48 (0.46–0.49); South: 0.45 (0.44–0.47)]. Fish abundance was highest in the North bioregion (spatial regional mean  $\pm$  SD:  $15.7 \pm 54.1$ ), followed by the South (2.6  $\pm$  4.7), Gascoyne (1.7  $\pm$  1.4) and West (1.6  $\pm$  2.5) in that order (Supporting Information Appendix S2, Figure S8). Model prediction uncertainty, expressed as the coefficient of variation (CV) of bootstrap values, showed a reverse pattern, being largest in the South (mean regional CV  $\pm$  SD: 0.36  $\pm$  0.07), relative to the West (0.31  $\pm$  0.05), Gascoyne (0.27  $\pm$  0.05) and North (0.23  $\pm$  0.06) (Supporting Information Appendix S2, Figure S9).

# 3.2 | Pelagic hotspots

Numerous hotspots of fish abundance, and clusters thereof, were consistently identified, with a high frequency of inclusion in bootstrap resamples (Figure 2). These included areas adjacent to the Scott reefs (14.1° S, 121.8° E), the edge of the Exmouth Plateau and the Argo-Rowley terrace (17° S, 117° E), offshore waters northwest of Barrow Island (20.8° S, 115.4° E), the length of Ningaloo Reef peninsula (22.5° S, 113.5° E) south to Shark Bay (26° S, 114° E), Rottnest Island and the Perth canyon (31.9° S, 115.1° E), Geographe Bay (33.6° S, 115.3° E) and the vicinity of Cape Leeuwin (35° S, 115.1° E), as well as along a significant stretch of the southern coastline from the Bremer Basin (35.5° S, 119.5° E), east to the western half of the Great Australian Bight (32° S, 129° E).

Random forest models built on the full set of biophysical predictors explained between 24 and 70% of the out-of-bag variance (R2) in fish abundance (Table 3 and Supporting Information Appendix S1). Geomorphometrics accounted for more than half of this percentage in all cases, and although reduced models suffered a loss of predictive power, performance remained satisfactory, particularly in the North (R<sup>2</sup> > 50%). The number and order of predictors selected as important also varied spatially (Figure 3 and Supporting Information Appendix S3, Table S4). For example, canyon presence in neighbouring cells (CANadj) was the top-ranked parameter in the North (bootstrap mean rank  $\pm$ SD: 1.06  $\pm$  0.28), closely followed by fractal dimension (FRD; 2.28  $\pm$ 0.55). In contrast, the Gascoyne bioregion was clearly driven by oceanography, with sea surface temperature variability (SSTstd;  $1.0 \pm 0.0$ ) and annual mean primary production (PP;  $2.27 \pm 0.51$ ) emerging as the most influential pair. Likewise, the West was dominated by PP (1.79  $\pm$ 0.89), ocean energy (L3; 2.01  $\pm$  0.98) and regional circulation regimes (L2;  $2.45 \pm 0.92$ ), whereas in the South, L3 ( $1.0 \pm 0.0$ ), dominant geomorphic features (FEATdom;  $2.0 \pm 0.0$ ) and east-west current velocity (CUREW;  $3.62 \pm 0.59$ ) were most important (Figure 3). Despite this variation in importance of variables, measures of static topographic complexity, including canyon attributes, made up an average of 34% of the top 10 splitting variables and 43% of all predictors retained after variable selection. Despite lower mean ranks, some metrics exhibited particularly high selection frequencies; for instance, geodiversity (FEATcount; 100% in the North and 75% in the West), the contour index (CI; 91% in the West and the South) and, to a lesser extent, canyon orientation (CANorient; 58% in the South) or canyon distribution (CANadj; 56% in the West). A small number of predictors, including CANadj, SAL, PP or SSTstd, were recurrently important throughout the entire wEEZ. In contrast, the frequency of chlorophyll peak index (FCPI) was of relatively trivial importance, as retained in 17% of bootstrap runs on average in only three of the four bioregions. Two human presence indicators (Hi and Him) were especially dominant (mean rank < 10 and frequency > 80%) in the North and the South bioregions.

### 3.3 | Spatial overlap

Hotspots occupied a mean area of 198,017 [95% percentile CI: (153,398-239,884)] km<sup>2</sup> across bioregions [n = 77 (60-94) cells; i.e.,

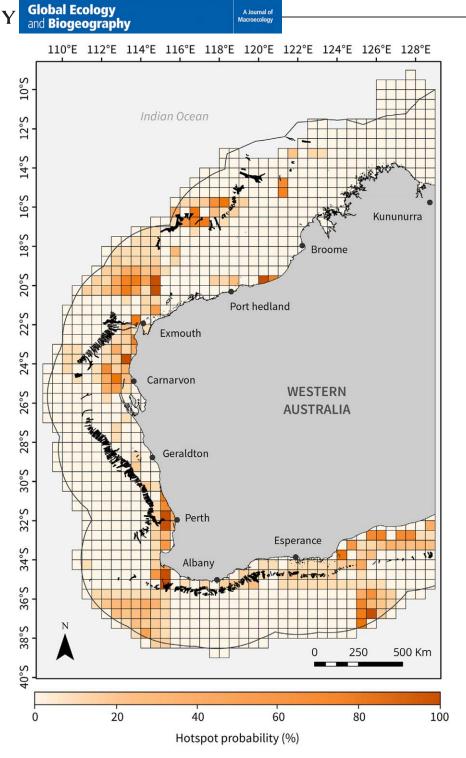


FIGURE 2 Pelagic fish hotspots derived from the Sea Around Us data. Hotspot probability was derived as the frequency with which each grid cell was selected as a hotspot across n = 100 bootstrap iterations, with darker tones denoting higher values. Submarine canyons (Huang et al., 2014) are overlaid in black

8.4% of total] and were most abundant in the North [30% (18-40) of the cumulative number of hotspots in the entire wEEZ] but most prevalent in Gascoyne [12% (7-17) of the number of grid cells in the bioregion]. In comparison, MR coverage was 796,110 km<sup>2</sup> (n = 316 cells) and was more prominent in the North (32% of the cumulative number of reserve cells in the entire wEEZ) but widest in the West (39% of the number of grid cells in the bioregion). Congruence was highest in the

North bioregion, although Jaccard scores remained generally very low everywhere (< 0.2), suggesting that a substantial proportion of the MR network as a whole does not intersect hotspots (Figure 4). All Mann-Whitney-Wilcoxon tests were significant (null permutation models, p < .01), and hotspots coincided with reserves less frequently than expected by chance alone in all bioregions but the North. Overlap with MNPs was substantially lower than with the wider MRs in both the



TABLE 3 Predictive accuracy of the random forest models of pelagic fish abundance

		Bioregions				
Input	Performance metric	North	Gascoyne	West	South	
All variables	RMSE	1.58 (1.54-1.61)	1.0 (0.93-1.05)	0.77 (0.73-0.83)	1.07 (1.02-1.11)	
All variables	$R^2$	.70 (.6971)	.44 (.3948)	0.24 (0.19-0.29)	.5 (.4654)	
Geomorphometrics	RMSE	1.93 (1.9-1.96)	1.16 (1.1-1.21)	0.81 (0.76-0.87)	1.23 (1.17-1.27)	
Geomorphometrics	$R^2$	.51 (.552)	.19 (.1624)	.17 (.1223)	.33 (.337)	

Note. Values represent bootstrap means (with associated 95% percentile confidence intervals). Performance is evaluated on the out-of-bag data (see Supporting Information Appendix S1). R<sup>2</sup> represents the percentage of variance explained. RMSE = root mean squared error.

Gascoyne and West bioregions. This trend was only marginally reversed in the North and South.

### 4 | DISCUSSION

Our study offers quantitative insights into the occurrence patterns of an assemblage of highly mobile, pelagic predatory fishes in the eastern Indian Ocean. Prediction maps revealed several fish hotspots along the northwestern, southwestern and southern continental shelves of Western Australia, broadly consistent with findings from previous research. For example, tuna and billfish species richness peaks in similar parts of the North and Gascoyne bioregions (Trebilco et al., 2011), and

analyses of movement behaviour in tiger sharks (*Galeocerdo cuvier*; Ferreira et al., 2015) and pygmy blue whales (*Balaenoptera musculus brevicauda*; Double et al., 2014) indicate that pelagic hotspots coincide with home range cores and areas of distinctly higher occupancy times in both species (Supporting Information Appendix S2, Figure S11). These congruent spatial patterns suggest a potential common basis to hotspot formation across multiple taxa (Bouchet et al., 2015). Higher relative abundance was also inferred at a number of discrete sites, confirming their importance to marine megafauna (Supporting Information Appendix S2, Figure S8); these included the seasonally productive Bremer basin, a foraging ground for white sharks (*Carcharodon carcharias*) and killer whales (*Orcinus orca*), the Albany canyon group, a

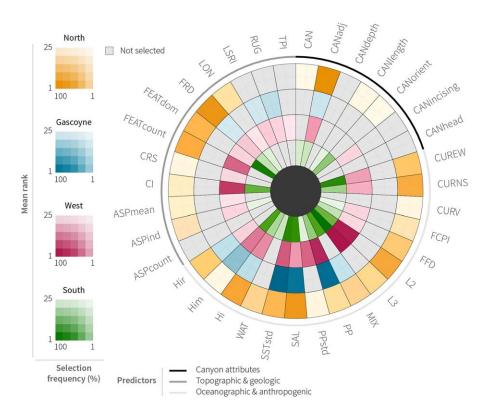


FIGURE 3 Summary of predictor importance in the random forest models. Each bioregion is assigned a different colour scheme and position on the wheel, from the outer (North) to the inner rings (South). Darker tones indicate predictors that ranked both highly and consistently (across bootstrap resamples), according to the bivariate key. Predictors that were not retained after variable selection (see Supporting Information Appendix S1 for details) are shown in grey. Canyon attributes are identified in black, topographic variables in dark grey and all remaining predictors in light grey (left). A full list of variable abbreviations is provided in Supporting Information Appendix S3, Table S1

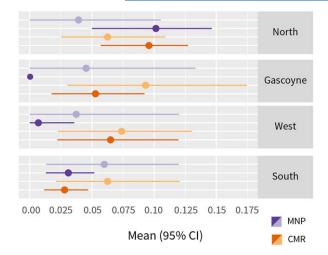


FIGURE 4 Congruence between pelagic fish hotspots and marine reserves. Overlap is measured as the Jaccard similarity coefficient J, which ranges from 0 (no overlap) to 1 (complete overlap). Lighter and darker colours capture the distribution of values under random null models (n=10,000) and empirical bootstrap resamples (n=100), respectively. Mean values are shown as filled circles, and their associated 95% percentile confidence intervals (CIs) as continuous lines. CMR = Commonwealth Marine Reserves (all zones); MNP = Marine National Parks (no-take zones only)

favourable habitat for sperm whales (*Physeter macrocephalus*; Johnson et al., 2016), the Recherche Archipelago, the Rowley Shoals, the Ashmore, Scott, Cartier and Ningaloo reefs (Sleeman et al., 2007) and, to a lesser degree, the waters off Dirk Hartog Island (Letessier et al., 2013).

Importantly, we add to a growing body of literature demonstrating that mobile top predators congregate at discrete and perennial sites (Graham et al., 2016). As international support for expanding the world's marine reserve coverage continues to rise (Singleton & Roberts, 2014), the hotspot concept may become particularly appropriate in guiding long-term MR placement and focusing research attention and resources on regions of persistently high ecological value for mobile species, whilst conferring maximal conservation benefits per dollar invested. Such information will be crucial because designing reserves is notoriously difficult and constrained by the costs of sampling vast volumes of ocean (Letessier et al., 2017) as well as by the necessity to accommodate a broad gamut of socio-economic and geopolitical interests. Thus far, most extant pelagic MRs have been established opportunistically without reliance on well-defined scientific criteria (Roberts, 2000) and/or residually where there is little perceived conflict with resource users (Devillers et al., 2015). In many information-poor settings, such as offshore waters, surrogate-based approaches may thus be the only viable option for improvement (McArthur et al., 2010). If so, the main difficulty will then lie in identifying not only a universally accepted operational definition of what constitutes a hotspot, an exercise so far thwarted by mixed interpretations (Bouchet et al., 2015), but also reliable proxies that can predict hotspot locations and possibly their change through time. Both will require the creation of standardized operating procedures that reconcile the numerous ways in which hotspots are measured (Marchese, 2015), importance scores calculated and predictors pruned (Szymczak et al., 2016).

Methodological biases notwithstanding, conflicting results from empirical studies have fuelled controversy about the application of abiotic surrogates to conservation planning scenarios (cf. Rees, Jordan, Price, Coleman, & Davis, 2014 with Stevens & Connolly, 2004). Such disputes are likely to stem from unresolved questions regarding the spatial and temporal stability of surrogate relationships or the effects of data quality and availability on indicator performance (Mellin, 2015). However, as efforts to test the validity of explanatory variables continue to be made, it will become easier to draw generalizations and identify those that perform consistently better across ecosystems, regions and scales (Sutcliffe, Klein, Pitcher, & Possingham, 2015). Geomorphometrics have generally remained broadly unvalidated proxies of oceanic biodiversity (but see Morato et al., 2010; Worm et al., 2003) in part because large portions of the seafloor are yet to be mapped and the majority of seamounts, submarine canyons and other prominent features around the world are still poorly explored (Huvenne & Davies, 2014). This suggests that weak correlations between static topography and predator hotspots may, at least historically, be more likely to reflect sporadic and uneven sampling than the absence of genuine wildlifehabitat relationships. Syntheses are also beginning to emerge that highlight the value of using geodiversity to prioritize areas for biological conservation (Beier, Hunter, & Anderson, 2015). Here, we have confirmed the value of geomorphometry by showing that it can be a good predictor of fish abundance on a macroecological scale and highlighting several indices of topographic complexity that have consistent relationships with pelagic biodiversity. That said, not all geomorphometrics were equally important. For instance, reflecting the observations of Huang et al. (2014) that the Australian margin is both physically and morphologically heterogeneous, submarine canyon metrics were outperformed in some bioregions but not others. We see two possible explanations for this. First, the formation and maintenance of openocean hotspots may demand a suite of interacting oceanographic and biophysical forces that are not associated with all canyons or topographic features (e.g., upwellings, fronts, eddies, physical retention of prey; Hazen et al., 2013). Second, some canyons may provide favourable conditions for pelagic fish species only episodically. If the latter is true, the relatively coarse temporal and spatial resolution of our data might not have been sufficient to reveal such variable relationships. This could be the case for canyon heads, which were not retained as important predictors of hotspots in our analysis but which are frequently reported to be sustained by cyclical upwelling events (Rennie et al., 2009). Likewise, the Oceanic Shoals Commonwealth Marine Reserve (11.5° S, 128.5° E) was identified here as a cold spot of fish abundance despite records of seasonally elevated pelagic diversity (Nichol et al., 2013).

In order to fully explore the hotspot spectrum and prioritize candidate areas for protection more robustly, a conceptual shift is warranted whereby hotspots are no longer defined in purely geographical terms, but are rather mapped in the four dimensions of latitude, longitude, time and depth, with explicit evaluations of their levels of intra/inter-

annual variability (Santora & Veit, 2013). Despite being essential to long-term resource management, such evaluations of hotspot persistence are seldom attempted in practice (Santora & Sydeman, 2015). A natural way of quantifying the consistency of hotspots through time is to determine the frequency with which a given site exceeds a chosen hotness level. Piacenza et al. (2015) proposed a 'universal' threshold in the form of a mean value across years, whereas Santora and Veit (2013) recommended that one standard deviation above a grand regional mean (> 1SD) be used as a common cut-off to align with previous work on remotely sensed patterns in ocean colour (Suryan, Santora, & Sydeman, 2012) and krill and whale distributions (Santora & Reiss, 2011). Although pragmatic, neither approach was feasible in the present study, and future efforts should be made to characterize hotspot variability based on finer-scale time-series data.

Assessing the processes underpinning the environmental preferences of wildlife species is a major challenge in the pelagic realm. The trophodynamics and habitat usage of tunas, billfishes and their relatives are complex, dynamic and niche-dependent such that species occupying temperate or tropical eco-regions may exhibit contrasting tolerance for, and responses to, similar environmental signals (Arrizabalaga et al., 2015). Such non-stationarity (illustrated in Supporting Information Appendix S2, Figure S10) may be mediated, and further complicated by, biotic interactions of varying intensity and direction across latitudes (Schemske, Mittelbach, Cornell, Sobel, & Roy, 2009). This may explain why temperature, kinetic energy, oxygen concentration and salinity are often seen as important predictors of biogeographical range but a mechanistic understanding of their influence is often missing in the literature. For example, the role of salinity, a consistently high-scoring variable in all bioregions (Supporting Information Appendix S3, Table S4), in determining the occurrence of pelagic species, is particularly obscure albeit some evidence exists that haline fronts may be indirectly linked with reproductive success (e.g., Alvarez-Berastegui et al., 2014), or prey density and therefore favourable foraging areas (e.g., Maury, Gascuel, Marsac, Fonteneau, & Rosa, 2001). Remotely sensed measurements of ocean colour and their derivatives, such as FCPI, are more readily interpretable, but may lack explanatory power (Supporting Information Appendix S3, Table S4) if the target organisms sit several trophic levels above primary producers (Grémillet et al., 2008), track productivity at depth by following the deep scattering layer or chlorophyll maxima and/or time lags occur between chlorophyll peaks and resource availability for consumers (Navarro et al., 2015).

Few studies have investigated the relative contribution of anthropogenic factors in controlling the distribution of ocean wildlife (Navarro, Cardador, Fernández, Bellido, & Coll, 2016). Distributional shifts caused by climate change or overfishing have been documented in some species (Fromentin, Reygondeau, Bonhommeau, & Beaugrand, 2014), but relationships with cumulative, distal factors can prove complex and taxon-specific (Navarro et al., 2015). As such, disentangling the respective effects of oceanographic conditions, migratory behaviour, density dependence, exploitation history and population structure on habitat selection remains a significant challenge. Here, the prevalence of human impact measures (Hi and Him), especially in the north

of the wEEZ, is unsurprising because this region boasts a diverse array of active and productive industries (e.g., petroleum, tourism, shipping, defence) earmarked for accelerated growth in coming decades. Further research is, however, needed to define the strength, directionality and persistence of their impacts on individual species at both population and organism levels. Comprehensive syntheses of contemporary pressures and their trends should greatly facilitate this process.

Against a backdrop of limited global marine protection (Costello & Ballantine, 2015) and a failure of conventional management to halt declines in ocean health, support for the implementation of placebased conservation strategies, such as marine reserves, is rapidly growing. In the past, MRs have largely mirrored the static frameworks that proved successful on land, yet recognition of the more fluid and threedimensional nature of the pelagic realm has prompted calls for more complex dynamic ocean management (DOM) approaches that can harness near-real-time data to provide adaptive and flexible responses to changes in the distribution and behaviour of species, habitats and resource users alike (Maxwell et al., 2015). Although a valid and pragmatic concept, successful examples of its execution remain few (but see Dunn, Maxwell, Boustany, & Halpin, 2016) and restricted to developed countries with sufficient logistical capacity and financial means to make DOM a viable option. Nonetheless, with access to technology rapidly expanding, DOM should become increasingly feasible in the future.

This does not imply that static reserves are ill-suited to mobile organisms. Indeed, mounting evidence suggests that even migratory species can benefit from static closures, for the following reasons: (a) protecting part of an animal's range or life cycle contributes to reducing overall mortality; (b) pelagic species are not necessarily as far-ranging as previously believed (White et al., 2017) and tend to aggregate around predictable bathymetric and hydrographic features; (c) their rates of residency, philopatry or site fidelity have generally been underestimated; (d) static MRs ought to be easier to enforce and therefore more likely to bear tangible benefits in species conservation; and (e) neglecting the potential for evolution of individual movement rates has compromised expectations of MR effectiveness (Mee et al., 2017). In this context, Australia recently declared a national network of MRs that occupies more than a third of its entire jurisdiction (c. 3.1 million km<sup>2</sup>). This areal coverage is exceptional by international standards; however, the reserve system in its current form provides low levels of protection equality across habitats and bathymetric classes (Barr & Possingham, 2013). Our analysis demonstrates (despite some caveats, see Supporting Information Appendix S1) that other natural assets, namely hotspots of mobile predatory wildlife, are also significantly under-represented. Whilst the declaration of the network has been a milestone in Australia's ecosystem-based approach to conservation, work remains to be done to ensure that the framework in place is ecologically coherent and enables rapid progress towards the new target set by the IUCN 2014 Sydney World Parks Congress to have at least 30% of ocean environments afforded strict protection within the next 15 years. Of course, marine reserves are only one conservation tool and are not a blanket solution to the problem of declining fish

A Journal

populations and biodiversity loss (Allison, Lubchenco, & Carr, 1998). They can, however, complement management efforts focused on setting and enforcing sustainable exploitation levels, controlling illegal fishing, mitigating pollution, capping fleet capacities, decreasing reliance on destructive gear and reducing bycatch rates. We also recognize that the mission statement of the world's protected area portfolio has expanded far beyond the original objectives the first reserves were created to fulfil in the early 1900s. Today, MRs are not only promoted as a means of preserving iconic wildlife and seascapes, but also to help bolster national economies, increase tourism, support the livelihood of local communities, alleviate poverty, replenish depleted stocks and provide resilience in the face of environmental change. The relevance of MRs for pelagic species will therefore need to be balanced against these and numerous other goals.

Here, we have provided empirical evidence that geomorphometrics can be meaningful proxies of macroecological patterns in pelagic marine species, a notion long suspected to be true but seldom comprehensively tested (with some exceptions; e.g., Morato et al., 2010; Worm et al., 2003). We also reinforced the notion that landing statistics can be instructive in a biogeographical context (Reygondeau et al., 2012), provided they are handled with care, transparency and a thorough understanding of their limitations in addressing specific questions. Appropriate use of these data is crucial because they provide some of the most spatially and temporally extensive information available for marine organisms, making them relevant as inputs in spatial planning within the data-limited pelagic ocean. Moreover, we demonstrate that significant opportunities to advance existing conservation frameworks await within offshore waters. The establishment of a global 'hotspot repository' (Hazen et al., 2013), to which this study contributes, is an essential step in developing an effective and flexible system of ocean management.

#### **ACKNOWLEDGMENTS**

We are grateful to the Sea Around Us team and to Daniel Pauly in particular for making the fisheries landing data available for this study. We would like to acknowledge Scott Foster, Damien Garcia, Johnathan Kool, Timothy Leslie, Jarod Santora and Carolin Strobl for their guidance at various stages of the data assembly and analysis, and we also thank two anonymous reviewers whose constructive comments considerably improved the quality of this manuscript. P.J. B. was supported by the Marine Biodiversity Hub through the Australian Government's National Environmental Research Program (NERP), administered by the Department of the Environment. NERP Marine Biodiversity Hub partners include the Institute for Marine and Antarctic Studies, University of Tasmania; CSIRO, Geoscience Australia, Australian Institute of Marine Science, Museum Victoria, Charles Darwin University and the University of Western Australia. P.J.B. was also the recipient of a scholarship for international research fees (SIRF) during the course of this work. R.A.W. acknowledges funding support from the Australian Research Council Discovery project support (DP140101377). Z.H. and S.L.N. publish with permission of the Chief Executive Officer, Geoscience Australia.

#### **ORCID**

Phil J. Bouchet http://orcid.org/0000-0002-2144-2049

#### REFERENCES

- Allison, G. W., Lubchenco, J., & Carr, M. H. (1998). Marine reserves are necessary but not sufficient for marine conservation. *Ecological Appli*cations, 8, S79–S92.
- Alvarez-Berastegui, D., Ciannelli, L., Aparicio-Gonzalez, A., Reglero, P., Hidalgo, M., López-Jurado, J. L., . . . Alemany, F. (2014). Spatial scale, means and gradients of hydrographic variables define pelagic seascapes of bluefin and bullet tuna spawning distribution. PLoS One, 9, e109338.
- Angulo-Valdés, J. A., & Hatcher, B. G. (2010). A new typology of benefits derived from marine protected areas. *Marine Policy*, 34, 635–644.
- Anticamara, J. A., Watson, R., Gelchu, A., & Pauly, D. (2011). Global fishing effort (1950–2010): Trends, gaps, and implications. Fisheries Research, 107, 131–136.
- Arrizabalaga, H., Dufour, F., Kell, L., Merino, G., Ibaibarriaga, L., Chust, G., ... Fraile, I. (2015). Global habitat preferences of commercially valuable tuna. Deep Sea Research Part II: Topical Studies in Oceanography, 113. 102–112.
- Barr, L. M., & Possingham, H. P. (2013). Are outcomes matching policy commitments in Australian marine conservation planning?. *Marine Policy*, 42, 39–48.
- Bartolino, V., Maiorano, L., & Colloca, F. (2011a). Frequency distribution curves and the identification of hotspots: Response to comments. *Population Ecology*, *53*, 603–604.
- Bartolino, V., Maiorano, L., & Colloca, F. (2011b). A frequency distribution approach to hotspot identification. *Population Ecology*, 53, 351–359.
- Beier, P., Hunter, M. L., & Anderson, M. (2015). Special section: Conserving nature's stage. Conservation Biology, 29, 613-617.
- Bouchet, P. J., Meeuwig, J. J., Salgado Kent, C. P., Letessier, T. B., & Jenner, C. K. (2015). Topographic determinants of mobile vertebrate predator hotspots: Current knowledge and future directions. *Biological Reviews*, 90, 699–728.
- Breen, P., Posen, P., & Righton, D. (2015). Temperate marine protected areas and highly mobile fish: A review. *Ocean & Coastal Management*, 105, 75–83.
- Breiman, L. (2001). Random forests. Machine Learning, 45, 5-32.
- Chapman, D. D., Feldheim, K. A., Papastamatiou, Y. P., & Hueter, R. E. (2015). There and back again: A review of residency and return migrations in sharks, with implications for population structure and management. *Annual Review of Marine Science*, 7, 547–570.
- Costello, M. J., & Ballantine, B. (2015). Biodiversity conservation should focus on no-take marine reserves: 94% of marine protected areas allow fishing. *Trends in Ecology and Evolution*, 30, 507–509.
- Daan, N., Gislason, H., Pope, J. G., & Rice, J. C. (2011). Apocalypse in world fisheries? The reports of their death are greatly exaggerated. ICES Journal of Marine Science: Journal du Conseil, 68, 1375–1378.
- Devillers, R., Pressey, R. L., Grech, A., Kittinger, J. N., Edgar, G. J., Ward, T., & Watson, R. (2015). Reinventing residual reserves in the sea: Are we favouring ease of establishment over need for protection?. Aquatic Conservation: Marine and Freshwater Ecosystems, 25, 480–504.
- Double, M. C., Andrews-Goff, V., Jenner, K. C. S., Jenner, M.-N., Laverick, S. M., Branch, T. A., & Gales, N. J. (2014). Migratory movements of pygmy blue whales (*Balaenoptera musculus brevicauda*) between Australia and Indonesia as revealed by satellite telemetry. *PLoS One*, 9, e93578.

- Dunn, D. C., Maxwell, S. M., Boustany, A. M., & Halpin, P. N. (2016). Dynamic ocean management increases the efficiency and efficacy of fisheries management. *Proceedings of the National Academy of Sciences USA*, 113, 668–673.
- Fernandez-Arcaya, U., Ramirez-Llodra, E., Aguzzi, J., Allcock, A. L., Davies, J. S., Dissanayake, A., . . . Van den Beld, I. M. J. (2017). Ecological role of submarine canyons and need for canyon conservation: A review. *Frontiers in Marine Science*, 4, 1–26.
- Ferreira, L. C., Thums, M., Meeuwig, J. J., Vianna, G. M. S., Stevens, J., McAuley, R., & Meekan, M. G. (2015). Crossing latitudes—Longdistance tracking of an apex predator. PLoS One, 10, e0116916.
- Francis, R. I. C. C. (1999). The impact of correlations in standardised CPUE indices. In *New Zealand fisheries assessment research document* 99/42 (p. 31). New Zealand Ministry of Fisheries, Wellington, New Zealand
- Froese, R., Zeller, D., Kleisner, K., & Pauly, D. (2012). What catch data can tell us about the status of global fisheries. *Marine Biology*, 159, 1283–1292.
- Fromentin, J. M., Reygondeau, G., Bonhommeau, S., & Beaugrand, G. (2014). Oceanographic changes and exploitation drive the spatio-temporal dynamics of Atlantic bluefin tuna (Thunnus thynnus). Fisheries Oceanography, 23, 147–156.
- Garcia, D. (2010). Robust smoothing of gridded data in one and higher dimensions with missing values. Computational Statistics & Data Analysis, 54, 1167–1178.
- Gormley, A. M., Slooten, E., Dawson, S., Barker, R. J., Rayment, W., Du Fresne, S., & Bräger, S. (2012). First evidence that marine protected areas can work for marine mammals. *Journal of Applied Ecology*, 49, 474–480.
- Graham, F., Rynne, P., Estevanez, M., Luo, J., Ault, J. S., & Hammerschlag, N. (2016). Use of marine protected areas and exclusive economic zones in the subtropical western North Atlantic Ocean by large highly mobile sharks. *Diversity and Distributions*, 22, 534–546.
- Grémillet, D., Lewis, S., Drapeau, L., Van Der Lingen, C. D., Huggett, J. A., Coetzee, J. C., ... Ryan, P. G. (2008). Spatial match-mismatch in the Benguela upwelling zone: Should we expect chlorophyll and seasurface temperature to predict marine predator distributions?. *Journal of Applied Ecology*, 45, 610-621.
- Grewe, P. M., Feutry, P., Hill, P. L., Gunasekera, R. M., Schaefer, K. M., Itano, D. G., . . . Davies, C. R. (2015). Evidence of discrete yellowfin tuna (*Thunnus albacares*) populations demands rethink of management for this globally important resource. *Scientific Reports*, 5, 16916.
- Harris, P. T., & Whiteway, T. (2009). High seas marine protected areas: Benthic environmental conservation priorities from a GIS analysis of global ocean biophysical data. Ocean & Coastal Management, 52, 22–38.
- Hazen, E. L., Suryan, R. M., Santora, J. A., Bograd, S. J., Watanuki, Y., & Wilson, R. P. (2013). Scales and mechanisms of marine hotspot formation. *Marine Ecology Progress Series*, 487, 177–183.
- Hooker, S. K., Hyrenbach, K. D., Corrigan, C., Polovina, J. J., & Reeves, R. R. (2011). Making protected area networks effective for marine top predators. *Endangered Species Research*, 13, 203–218.
- Huang, Z., Nichol, S. L., Harris, P. T., & Caley, M. J. (2014). Classification of submarine canyons of the Australian continental margin. *Marine Geology*, 357, 362–383.
- Huvenne, V. A. I., & Davies, J. S. (2014). Towards a new and integrated approach to submarine canyon research. Deep Sea Research Part II: Topical Studies in Oceanography, 104, 1–5.
- Johnson, C. M., Beckley, L. E., Kobryn, H., Johnson, G. E., Kerr, I., & Payne, R. (2016). Crowdsourcing modern and historical data identifies

- sperm whale (*Physeter macrocephalus*) habitat offshore of southwestern Australia. Frontiers in Marine Science. 3, 1–16.
- Kaplan, D. M., Chassot, E., Gruss, A., & Fonteneau, A. (2010). Pelagic MPAs: The devil is in the details. *Trends in Ecology and Evolution*, 25, 62–63
- Kerwath, S. E., Thorstad, E. B., Naesje, T. F., Cowley, P. D., Økland, F., Wilke, C., & Attwood, C. G. (2009). Crossing invisible boundaries: The effectiveness of the Langebaan Lagoon Marine Protected Area as a harvest refuge for a migratory fish species in South Africa. Conservation Biology, 23, 653–661.
- Kerwath, S. E., Winker, H., Götz, A., & Attwood, C. G. (2013). Marine protected area improves yield without disadvantaging fishers. *Nature Communications*, 4, 2347.
- Kessel, S. T., Chapman, D. D., Franks, B. R., Gedamke, T., Gruber, S. H., Newman, J. M., ... Perkins, R. G. (2014). Predictable temperatureregulated residency, movement and migration in a large, highly mobile marine predator (Negaprion brevirostris). *Marine Ecology Pro*gress Series, 514, 175–190.
- Leroux, S. J., Schmiegelow, F. K., & Nagy, J. A. (2007). Potential spatial overlap of heritage sites and protected areas in a boreal region of northern Canada. Conservation Biology, 21, 376–386.
- Letessier, T. B., Bouchet, P. J., & Meeuwig, J. J. (2017). Sampling mobile oceanic fishes and sharks: Implications for fisheries and conservation planning. *Biological Reviews*, 92, 627–646.
- Letessier, T. B., Meeuwig, J. J., Gollock, M., Groves, L., Bouchet, P., Chapuis, L., ... Koldewey, H. J. (2013). Assessing pelagic fish and shark populations: The application of demersal techniques to the mid-water. *Methods in Oceanography*, 8, 41–55.
- Marchese, C. (2015). Biodiversity hotspots: A shortcut for a more complicated concept. *Global Ecology and Conservation*, *3*, 297–309.
- Maunder, M. N., & Punt, A. E. (2004). Standardizing catch and effort data: A review of recent approaches. *Fisheries Research*, 70, 141–159.
- Maury, O., Gascuel, D., Marsac, F., Fonteneau, A., & Rosa, A.-L. D. (2001). Hierarchical interpretation of nonlinear relationships linking yellowfin tuna (Thunnus albacares) distribution to the environment in the Atlantic Ocean. Canadian Journal of Fisheries and Aquatic Sciences, 58, 458-469.
- Maxwell, S. M., Hazen, E. L., Lewison, R. L., Dunn, D. C., Bailey, H., Bograd, S. J., ... Crowder, L. B. (2015). Dynamic ocean management: Defining and conceptualizing real-time management of the ocean. *Marine Policy*, 58, 42–50.
- McArthur, M. A., Brooke, B. P., Przeslawski, R., Ryan, D. A., Lucieer, V. L., Nichol, S., . . . Radke, L. C. (2010). On the use of abiotic surrogates to describe marine benthic biodiversity. *Estuarine, Coastal and Shelf Science*, 88, 21–32.
- Mee, J. A., Otto, S. P., & Pauly, D. (2017). Evolution of movement rate increases the effectiveness of marine reserves for the conservation of pelagic fishes. Evolutionary Applications, 10, 444–461.
- Mellin, C. (2015). Abiotic surrogates in support of marine biodiversity conservation. In D. Lindenmayer, P. Barton, & J. Pierson (Eds.), Indicators and surrogates of biodiversity and environmental change (p. 216). Melbourne: CSIRO Publishing.
- Michael, P. E., Jahncke, J., & Hyrenbach, K. D. (2014). Relative influence of static and dynamic features on black-footed albatross (*Phoebastria* nigripes) habitat use in central California Sanctuaries. Fisheries Oceanography, 23, 18–31.
- Moors-Murphy, H. B. (2014). Submarine canyons as important habitat for cetaceans, with special reference to the gully: A review. *Deep Sea Research Part II: Topical Studies in Oceanography*, 104, 6–19.

- Morato, T., Hoyle, S. D., Allain, V., & Nicol, S. J. (2010). Seamounts are hotspots of pelagic biodiversity in the open ocean. *Proceedings of the National Academy of Sciences USA*, 107, 9707–9711.
- Morato, T., Varkey, D. A., Damaso, C., Machete, M., Santos, M., Prieto, R., . . . Pitcher, T. J. (2008). Evidence of a seamount effect on aggregating visitors. *Marine Ecology Progress Series*, 357, 23–32.
- Navarro, J., Cardador, L., Fernández, Á. M., Bellido, J. M., & Coll, M. (2016). Differences in the relative roles of environment, prey availability and human activity in the spatial distribution of two marine mesopredators living in highly exploited ecosystems. *Journal of Biogeography*, 43, 440–450.
- Navarro, J., Coll, M., Cardador, L., Fernández, Á. M., & Bellido, J. M. (2015). The relative roles of the environment, human activities and spatial factors in the spatial distribution of marine biodiversity in the Western Mediterranean Sea. Progress in Oceanography, 131, 126–137.
- Neubauer, P., Jensen, O. P., Hutchings, J. A., & Baum, J. K. (2013). Resilience and recovery of overexploited marine populations. *Science*, 340, 347–349.
- Nichol, S. L., Howard, F. J. F., Kool, J., Stowar, M., Bouchet, P., Radke, L., ... Heyward, A. (2013). Oceanic shoals commonwealth marine reserve (timor sea) biodiversity survey. In *Post-survey report GA0339/* SOL5650 (p. 112). Geoscience Australia, Canberra.
- Pauly, D., & Zeller, D. (2016). Catch reconstructions reveal that global marine fisheries catches are higher than reported and declining. *Nature Communications*, 7, 10244.
- Piacenza, S. E., Thurman, L. L., Barner, A. K., Benkwitt, C. E., Boersma, K. S., Cerny-Chipman, E. B., ... Heppell, S. S. (2015). Evaluating temporal consistency in marine biodiversity hotspots. *PLoS One*, 10, e0133301
- Powers, J. E., & Abeare, S. M. (2009). Fishing effort redistribution in response to area closures. *Fisheries Research*, 99, 216–225.
- Real, R., & Vargas, J. M. (1996). The probabilistic basis of Jaccard's index of similarity. *Systematic Biology*, 45, 380–385.
- Rees, M. J., Jordan, A., Price, O. F., Coleman, M. A., & Davis, A. R. (2014). Abiotic surrogates for temperate rocky reef biodiversity: Implications for marine protected areas. *Diversity and Distributions*, 20, 284–296.
- Rennie, S., Hanson, C. E., McCauley, R. D., Pattiaratchi, C., Burton, C., Bannister, J., ... Jenner, M. N. (2009). Physical properties and processes in the Perth Canyon, Western Australia: Links to water column production and seasonal pygmy blue whale abundance. *Journal of Marine Systems*, 77, 21–44.
- Reygondeau, G., Maury, O., Beaugrand, G., Fromentin, J. M., Fonteneau, A., & Cury, P. (2012). Biogeography of tuna and billfish communities. Journal of Biogeography, 39, 114–129.
- Roberts, C. M. (2000). Selecting marine reserve locations: Optimality versus opportunism. *Bulletin of Marine Science*, *66*, 581–592.
- Sabatia, C. O., & Burkhart, H. E. (2014). Predicting site index of plantation loblolly pine from biophysical variables. Forest Ecology and Management, 326, 142–156.
- Santora, J. A., & Reiss, C. S. (2011). Geospatial variability of krill and top predators within an Antarctic submarine canyon system. *Marine Biology*, 158, 2527–2540.
- Santora, J. A., & Sydeman, W. J. (2015). Persistence of hotspots and variability of seabird species richness and abundance in the southern California Current. *Ecosphere*, *6*, 1–19.
- Santora, J.A., & Veit, R. (2013). Spatio-temporal persistence of top predator hotspots near the Antarctic Peninsula. Marine Ecology Progress Series, 487, 287–304.

- Scales, K. L., Miller, P. I., Hawkes, L. A., Ingram, S. N., Sims, D. W., & Votier, S. C. (2014). On the front line: Frontal zones as priority at-sea conservation areas for mobile marine vertebrates. *Journal of Applied Ecology*, 51, 1575–1583.
- Schemske, D. W., Mittelbach, G. G., Cornell, H. V., Sobel, J. M., & Roy, K. (2009). Is there a latitudinal gradient in the importance of biotic interactions?. Annual Review of Ecology, Evolution, and Systematics, 40, 245–269.
- Scott, R., Hodgson, D. J., Witt, M. J., Coyne, M. S., Adnyana, W., Blumenthal, J. M., ... Godley, B. J. (2012). Global analysis of satellite tracking data shows that adult green turtles are significantly aggregated in Marine Protected Areas. Global Ecology and Biogeography, 21, 1053–1061.
- Singleton, R. L., & Roberts, C. M. (2014). The contribution of very large marine protected areas to marine conservation: Giant leaps or smoke and mirrors?. *Marine Pollution Bulletin*, 87, 7–10.
- Sleeman, J., Meekan, M., Wilson, S., Jenner, C., Jenner, M., Boggs, G., ... Bradshaw, C. (2007). Biophysical correlates of relative abundances of marine megafauna at Ningaloo Reef, Western Australia. *Marine and Freshwater Research*, 58, 608–623.
- Stevens, T., & Connolly, R. M. (2004). Testing the utility of abiotic surrogates for marine habitat mapping at scales relevant to management. Biological Conservation, 119, 351–362.
- Strobl, C., Boulesteix, A.-L., Kneib, T., Augustin, T., & Zeileis, A. (2008). Conditional variable importance for random forests. BMC Bioinformatics, 9(1), 11.
- Strobl, C., Boulesteix, A.-L., Zeileis, A., & Hothorn, T. (2007). Bias in random forest variable importance measures: Illustrations, sources and a solution. *BMC Bioinformatics*, 8(1), 21.
- Strobl, C., Hothorn, T., & Zeileis, A. (2009). Party on! A new, conditional variable importance measure for random forests available in the party package. *The R Journal*, 1, 14–17.
- Suryan, R. M., Santora, J. A., & Sydeman, W. J. (2012). New approach for using remotely sensed chlorophyll a to identify seabird hotspots. Marine Ecology Progress Series, 451, 213–225.
- Sutcliffe, P., Klein, C., Pitcher, C., & Possingham, H. (2015). The effectiveness of marine reserve systems constructed using different surrogates of biodiversity. Conservation Biology, 29, 657–667.
- Szymczak, S., Holzinger, E., Dasgupta, A., Malley, J. D., Molloy, A. M., Mills, J. L., ... Bailey-Wilson, J. E. (2016). r2VIM: A new variable selection method for random forests in genome-wide association studies. *BioData Mining*, 9, 7.
- Trebilco, R., Halpern, B. S., Flemming, J. M., Field, C., Blanchard, W., & Worm, B. (2011). Mapping species richness and human impact drivers to inform global pelagic conservation prioritisation. *Biological Conservation*, 144, 1758–1766.
- Wang, X.-F. (2010). fANCOVA: Nonparametric analysis of covariance. R package version 0.5-1.
- Watson, R. A., Alder, J., Kitchingman, A., & Pauly, D. (2005). Catching some needed attention. *Marine Policy*, 29, 281–284.
- Watson, R. A., Cheung, W. W. L., Anticamara, J. A., Sumaila, R. U., Zeller, D., & Pauly, D. (2013). Global marine yield halved as fishing intensity redoubles. Fish and Fisheries, 14, 493–503.
- Watson, R. A., Kitchingman, A., Gelchu, A., & Pauly, D. (2004). Mapping global fisheries: Sharpening our focus. Fish and Fisheries, 5, 168-177.
- Webb, T. J., Vanden Berghe, E., & O'Dor, R. (2010). Biodiversity's big wet secret: The global distribution of marine biological records reveals chronic under-exploration of the deep pelagic ocean. PLoS One, 5, e10223.





White, T. D., Carlisle, A. B., Kroodsma, D. A., Block, B. A., Casagrandi, R., De Leo, G. A., . . . McCauley, D. J. (2017). Assessing the effectiveness of a large marine protected area for reef shark conservation. *Biological Conservation*, 207, 64–71.

Worm, B., Hilborn, R., Baum, J. K., Branch, T. A., Collie, J. S., Costello, C., ... Zeller, D. (2009). Rebuilding global fisheries. *Science*, 325, 578–585.

Worm, B., Lotze, H., & Myers, R. (2003). Predator diversity hotspots in the blue ocean. Proceedings of the National Academy of Sciences USA, 100, 9884–9888.

Young, H. S., Maxwell, S. M., Conners, M. G., & Shaffer, S. A. (2015). Pelagic marine protected areas protect foraging habitat for multiple breeding seabirds in the central Pacific. *Biological Conservation*, 181, 226–235.

## **BIOSKETCH**

PHIL BOUCHET is a postdoctoral researcher interested in the use of statistical models to support conservation planning decisions

throughout the pelagic ocean. He works on a range of vertebrate species (cetaceans, sharks and fishes), at different spatial scales (local, regional and global) and across a number of biogeographical areas (temperate to tropical).

## SUPPORTING INFORMATION

Additional Supporting Information may be found online in the supporting information tab for this article.

**How to cite this article:** Bouchet PJ, Meeuwig JJ, Huang Z, et al. Continental-scale hotspots of pelagic fish abundance inferred from commercial catch records. *Global Ecol Biogeogr.* 2017;26:1098–1111. https://doi.org/10.1111/geb.12619