

1 Persistent problems in the construction of matrix
2 population models

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8 **Abstract**

Matrix population models (MPMs) are powerful tools for translating demographic and life history information into a form that can be used to address a wide range of research topics, such as projecting population dynamics, evaluating stressor impacts on populations, and studying life history evolution. However, the reliability of such studies depends on the MPM being constructed in a way that accurately reflects the species' life history. We highlight three errors commonly encountered in published MPMs: (1) failing to include survival in the fertility coefficient; (2) introducing a one-year delay in age at first reproduction; and (3) incorrectly calculating the growth rate out of a stage with a mean development time greater than the model time step. We review the sources of such errors and provide new analyses revealing the impact of such errors on model predictions. These MPM construction issues are treated extensively in textbooks, so their existence in current literature is surprising. To quantify the prevalence of such errors we examined and scored the original publications underlying the models in the COMADRE Animal Matrix Database [add sample sizes (MPMs, species, studies)]. The first two errors were found in 41% and 42%, respectively, of the published studies; all were in models that used a "post-breeding census" representation of the life cycle (in which newborns [eggs, neonates, fledglings, etc.] are explicitly included). Of the studies where stages may last longer than one time step, 25% constructed the growth rate using inappropriate formulas, and 37% used methods that were either undescribed or would generate the correct development time only if the population remained at the observed within-stage age structure. These results suggest that further efforts may be required to educate biologists on the construction of MPMs, perhaps in concert with the development of new software tools. Furthermore,

the conclusions of many studies that are based on MPMs may need to be re-examined, and synthetic studies using the COMADRE Database need to be accompanied by careful examination of the underlying studies.

9 *Key words:* COMADRE animal matrix database, Lefkovitch matrix, Leslie
10 matrix, Matrix population models, model construction, model validity

11 1. Introduction

12 The use of matrix population models (MPMs) for the ecological study of
13 plants and animals has expanded rapidly in recent years (Salguero-Gómez,
14 Jones, Archer, et al., 2016; Salguero-Gómez et al., 2015). MPMs are a
15 convenient way to synthesize demographic information about a population,
16 and are useful in addressing ecological, evolutionary, and management
17 questions (Caswell, 2001; Morris and Doak, 2002). Most early MPMs were
18 developed by collaborations between biologists with expertise on the species
19 at hand and modelers with deep understanding of the subtleties of modeling.
20 MPMs now, having been long promoted as a straightforward translation of
21 life-history information into quantitative analysis, are often being created by
22 biologists who are primarily empirical ecologists or conservation biologists,
23 and may have had little training in modeling beyond a graduate course in
24 population ecology. One might then ask, do these MPMs reliably reproduce
25 the life histories they are meant to represent?

26 It is important to remember that MPMs are approximations, both because
27 they still abstract away much biological detail and because demographic
28 measurements are imprecise. There are a variety of subtleties in constructing
29 such models, especially when breeding is spread across a substantial fraction
30 of the year or when there is substantial heterogeneity among individuals
31 within an age or stage class; much theoretical work has gone into making
32 MPMs more complex to account for this, and we are starting to better
33 understand the effects of these phenomena on simple models (e.g., Fujiwara
34 and Diaz-Lopez, 2017). But even setting aside these issues, we might hope
35 that, if we assume that a simple life history description (e.g., as embodied in
36 an age- or stage-structured life table) is a useful description of the population,
37 then the constructed MPM should accurately reproduce that description.

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38 If it does not, then we are introducing additional errors into the analysis
39 above and beyond the inherent approximations of modeling.

40 One of us (BEK) has spent two decades teaching MPMs to applied
41 masters students. This experience has revealed several aspects of MPM
42 construction that are particularly challenging to novices, and thus might be
43 similarly challenging to biologists who do not regularly create mathematical
44 models of their system. Furthermore, the students, when sent to find a
45 published model to analyze, have uncovered a suprising number of published
46 MPMs that have failed to meet these challenges. At one level, the errors
47 made by these novices are “simply” accounting issues, but our interpretation
48 is that the difficulty that **students** have in overcoming them is rooted in
49 the contrast between the often-loose way that terms are used to describe
50 biological populations and the very precise meanings of terms in MPMs
51 (together with some genuine linguistic ambiguity that plagues the field).

52 Our goal in this paper is to highlight these challenges (which are described
53 in the next section), quantify their impacts on the outputs of MPMs, and
54 document their prevalence in the peer-reviewed literature. We conclude with
55 a discussion of why these MPM construction errors are so prevalent, and
56 how we might improve future practice.

57 **2. Challenges in MPM construction**

58 We focus on animal models in which the demography is structured by
59 age (Leslie matrices; Leslie, 1945) or developmental stage (Lefkovitch matri-
60 ces; Lefkovitch, 1965); plant and size structured models present additional
61 complications (although ~~two~~ the issues presented here still apply). We also
62 focus on species whose reproduction is concentrated in a short breeding
63 season (so-called “birth-pulse” populations; Caughley, 1967). Many animals
64 and plants satisfy this assumption, and this allows us to avoid the extra
65 complications associated with using discrete-time models to represent a
66 continuously breeding species. Note that by “breeding season” we mean the
67 season in which individuals are born or hatched, not the season during which
68 their parents mate. Age starts counting from birth, so that a reproductively
69 mature individual breeds on or about their birthday: e.g., if the age at first
70 reproduction is 5 years, then an individual has its first offspring on its fifth
71 birthday. 

72 MPMs project the population from one nominal census date to the next.
73 While the census could in principle be at any time, in practice nearly all

Table 1: A sample life table for a species that reaches sexual maturity at age 3 (adult stage). Adults continue to survive and reproduce indefinitely with the same survival and birth rates. Note that, because we are using the stage-structured convention for assigning age class names (see text), the table may look slightly different from those in many textbooks.

Age in years (x)	Annual survival (σ_x)	Birth rate (b_x)	Stage
0	0.2	0	Newborn
1	0.4	0	Juvenile
2	0.4	0	Juvenile
3	0.9	3	Adult
4	0.9	3	Adult
\vdots	\vdots	\vdots	\vdots

MPMs either census the population just before breeding (“prebreeding census”) or just after breeding (“postbreeding census”). In a prebreeding census, the youngest age class is made up of individuals all with age (nearly) one, which we designate as age one. In a postbreeding census, the first age class would comprise newborn individuals, all of age zero, the second class would comprise individuals of age exactly one, etc. We refer to newborn individuals with a subscript of zero; as a result, our lifetables and postbreeding Leslie models will look different from most textbooks, which assign newborns to “age class one” and use a subscript of one.

2.1. Ensuring that the fertility coefficient spans a full timestep

In a life table (e.g., Table 1), age specific survival and birth rates look as if they should make equivalent contributions to a population model. But there is an important difference: survival (which we denote σ_x for Leslie models and σ_i for Lefkovitch models, where x is the age and i is stage) represents the fraction of individuals in a class (x or i) that survive for a full timestep, from time t to time $t + 1$. In contrast, the birth rate (which we denote b_x for age-structured demography and b_i for stage-structured demography) is instantaneous; it is the average number of offspring produced at time t by an individual that is alive at time t . It is tempting to draw a life cycle graph like the one in Fig. 1a (and we commonly see that in the literature). While this works as a *conceptual* diagram, translating it directly into an MPM by coverting each arrow in the graph into a matrix element is incorrect: each

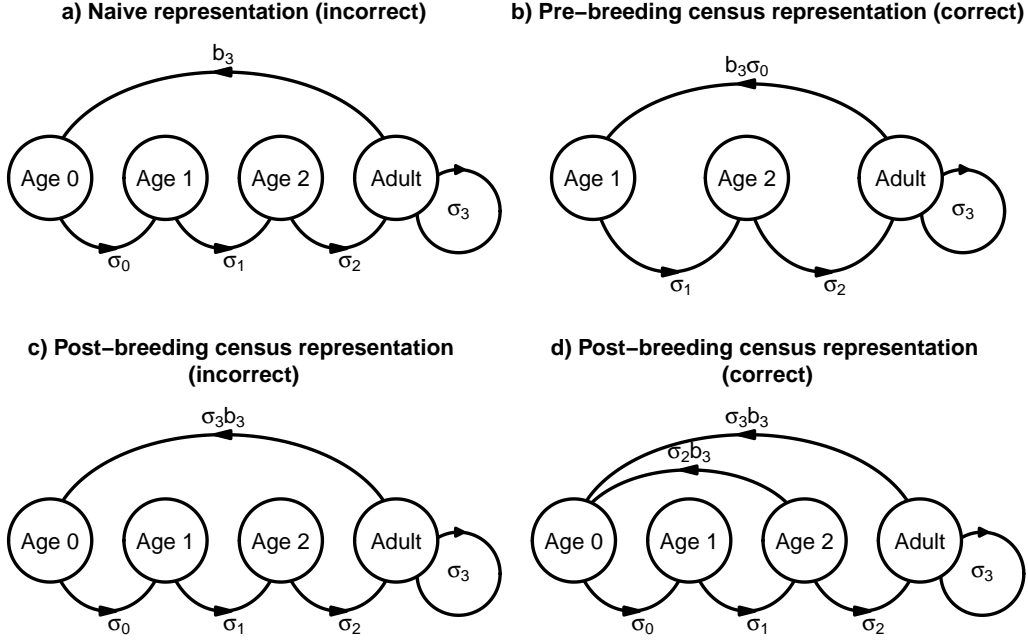


Figure 1: Four life cycle graphs that might (or might not) represent the life table in Table 1. (a) A “naive” representation that associates each arrow with a vital rate. This fails to account for the fact that the fertility coefficients must span a timestep and therefor include a survival term. (b) A pre-breeding census representation, in which the youngest censused class is Age 1 and the adult birth rate is multiplied by newborn survival. (c) A post-breeding census representation, in which the adult burth rate is correctly multiplied by adult survival, but which fails to account for the fact that Age 2 individuals will have reproduced (as adults) just prior to the next census. (d) A correct post-breeding census representation.

96 element in the matrix must span a timestep, and m_x does not accomplish
 97 that.

98 To get the timestep in there, b_x needs to be multiplied by a survival
 99 term—either that of the parent or of the offspring. For a pre-breeding census
 100 model, the fertility coefficient is $F_x = b_x \sigma_0$: the parent, in class x , produces
 101 b_x offspring immediately after the census, and then these offspring survive
 102 to the end of the timestep at rate σ_0 (Fig. 1b). For an annual timestep, F_x
 103 represents the number of one-year-olds next year produced by an individual
 104 of age x this year.

105 In a post-breeding census, the parent (which will have just reproduced if
 106 it is already an adult) must survive for a timestep, aging by a timestep and
 107 possibly maturing into a new class, and then reproduces with a birth rate

appropriate to its class at the end of the timestep. If we use x' to denote the parent's class at time $t + 1$, then the fertility coefficient is $F_x = \sigma_x b_{x'}$ (Fig. 1d). For an annual timestep, F_x is the number of zero-year-olds (newborns) next year produced ~~next year~~ by an individual that was in class x this year. Properly accounting for x' is a separate challenge that we address in the next subsection.

2.2. Matching the age at first reproduction to the species' life history

In an age-structured population, the lowest age with a non-zero birth rate represents the *age at first reproduction*; we will call that age x_m , for “age at [reproductive] maturity.” In a prebreeding census model (Fig. 1b), this does not present a conceptual challenge, as the newly matured, about-to-reproduce-for-the-first-time individuals are already classified as age x_m , matching intuition. However, in a post-breeding census model, the individuals who are age x_m at the end of the timestep, and have just reproduced for the first time, were age $x_m - 1$ at the beginning of the timestep. A model that has the first fertility coefficient associated with age x_m (Fig. 1c) results in the modelled age at first reproduction being $x_m + 1$, a year delay relative to the actual life history. The solution is to add a fertility coefficient linking age class $x_m - 1$ to age class zero—the lower of the two fertility arrows in Fig. 1d. Embracing this fertility coefficient requires overcoming cognitive dissonance—“juveniles” are reproducing!—and we have found that novices actively resist this.

An alternative approach for postbreeding census Leslie models is to change the indexing scheme, associating x with the individual's age at the *end* of the timestep (rather than the age at the beginning of the timestep, as we have done above). Then the individuals who will mature and reproduce before the next census are called age x_m , and their fertility is $F_{x_m} = \hat{\sigma}_{x_m} b_{x_m}$ (we use $\hat{\sigma}$ for survival to clarify that it is a differently indexed parameter from σ as used above). Thus, the cognitive dissonance is finessed by “hiding” the fact that these individuals started the timestep as juveniles. Indeed, many textbooks use this indexing convention. However, this alternative indexing convention cannot be maintained when moving from age-structured to stage-structured models, as not all individuals that start the timestep as juveniles will end it as adults.

So let us look at stage-structured models. The simplest stage-structured model has newborns, a nonreproductive juvenile class that spans multiple timesteps, and reproductive adults (Fig. 2). Within the juvenile class, some individuals remain juveniles in the next timestep (should they survive),

146 whereas others mature into adults. If γ_J is the fraction surviving individuals
 147 that mature (“grow”) at the end of the timestep, then the Juvenile–Juvenile
 148 transition is given by $P_J = \sigma_J(1 - \gamma_J)$ and the Juvenile–Adult transition
 149 is given by $G_J = \sigma_J\gamma_J$. We will treat the estimation of γ_J in the next
 150 section. As in age-structured models, the fertility coefficient needs to include
 151 a survival term, so Fig. 2a is incorrect. In a prebreeding census model,
 152 the adult birth rate must be multiplied by newborn survival, and newborns
 153 removed from the stage vector (Fig. 2b), whereas in a postbreeding census
 154 model, the birth rate must be multiplied by the parent’s survival (Figs. 2c-d).

155 If the criterion for being classed as “adult” is that the individual can
 156 reproduce, then just-matured adults should have non-zero fertility. In the
 157 prebreeding census model (Fig. 2b), the individuals that have just matured
 158 (made the transition from Juvenile to Adult) will reproduce at the beginning
 159 of the next timestep. However, in the ~~“intuitive”~~ postbreeding census model
 160 (Fig. 2c), the individuals that just matured won’t reproduce until the end
 161 of the next timestep, a full timestep after their transition to adulthood. In
 162 reality, the individuals that just made the Juvenile–Adult transition were
 163 already adults at the just-passed breeding season, and hence have had their
 164 first opportunity to reproduce. To represent this we need a fertility coefficient
 165 leading out of the juvenile class— $F_J = G_Jb_A$ (Fig. 2d)—which again seems
 166 to defy common sense. Although this is the textbook solution, there a number
 167 of other ways one might finesse this issue, such as classifying individuals as
 168 adults at the beginning of the timestep in which they mature, or creating a
 169 special “pre-adult” class between juveniles and adults. However, ~~none of these~~
 170 ~~are as straightforward as the re-indexing gambit for age structured models;~~
 171 ~~they introduce changes~~ such as giving the maturing juveniles adult survival
 172 rather than juvenile survival (which might or might not be desirable), or
 173 shortening the “juvenile” stage duration to account for the extra stage.

174 In both stage- and age-structured models, failing to have the fertility
 175 coefficient out of the last juvenile class in postbreeding census models (or
 176 having another workaround to the problem) will lead to a one-year delay in
 177 the age at first breeding, relative to the life history that the model is meant
 178 to represent.

179 2.3. Ensuring that the mean time in each developmental stage matches the 180 species’ life history

181 The final challenge is associated with estimating γ_x , the fraction of
 182 individuals maturing out of stage x . With repeated observations of marked
 183 individuals, one can simply derive this from the observations, for example,

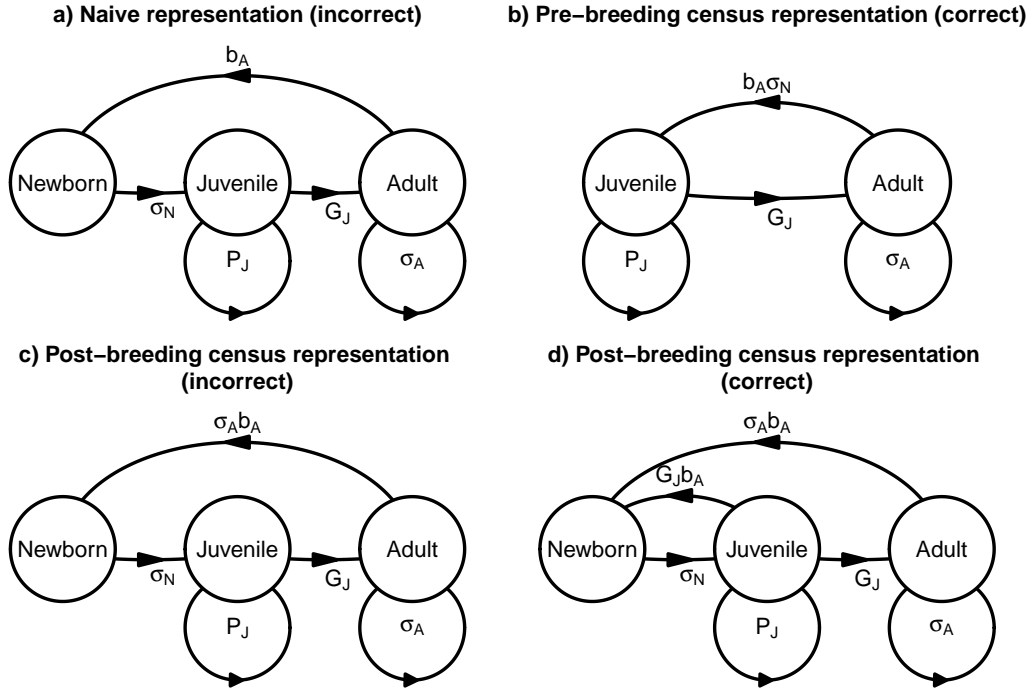


Figure 2: Four life cycle graphs that might (or might not) represent stage structured life table in Table 1. (a) A “naive” representation that associates each arrow with a vital rate. This fails to account for the fact that the fertility coefficients must span a timestep and therefor include a survival term. (b) A pre-breeding census representation, in which the youngest censused class is the juvenile stage and the adult birth rate is multiplied by newborn survival. (c) A post-breeding census representation, in which the adult burth rate is correctly multiplied by adult survival, but which fails to account for the fact that the maturing juveniles will have reproduced (as adults) just prior to the next census. (d) A correct post-breeding census representation.

184 using multistate capture-mark-recapture models (CITE, although this is not
 185 guaranteed to be robust; see Discussion). ~~However, in~~ many cases, the goal
 186 is to combine information on stage-specific survival, often variously collected,
 187 with knowledge about the stage duration (the number of timesteps that
 188 individuals remain in a stage before maturing). If stage durations are fixed
 189 (e.g., all individuals mature after two years as juveniles), then the goal is to
 190 create a “stage-for-age” model (e.g., Ebert) where the mean stage duration
 191 in the model equals the fixed stage duration in the life history. ~~Even where~~
 192 real stage durations are ~~somewhat~~ variable, if there is a well-defined mean
 193 stage duration, then ~~we can still aspire to have the modeled~~ mean stage
 194 duration match the actual mean stage duration.

195 Caswell (2001) has a section of his book describing various ways of doing
 196 this. ~~It’s not clear that there is a~~ single “best” approach, as the stage-
 197 structured model will never match the age-structured model ~~in all ways~~ (see
 198 Discussion). Nevertheless, the primary analyses performed on most published
 199 MPMs are to calculate the asymptotic growth rate (λ_1) and **the sensitivity**
 200 **of λ_1** to underlying demographic parameters. For these calculations, there
 201 is one (and only one) recipe to create a stage-structured model that will
 202 generate the same results as the equivalent age-structured model with fixed
 203 stage durations. **As best we can tell, it was first introduced by Caswell**
 204 **(1989), although its first application to an ecological populaton may have**
 205 **been by Crowder et al. (1994); it is covered in Caswell’s (2001) section on**
 206 **stage-structured models.**

207 The basic idea is that γ represents the fraction of individuals in the stage
 208 that have spent enough time in the stage to mature. This, in turn, depends
 209 on the **(virtual)** age structure within the stage. In general, this age structure
 210 could have any form, depending on the recent history of the population; the
 211 one case in it is well defined is when the population is at the stable age/stage
 212 distribution. This is exactly the condition under which λ_1 is the population
 213 growth rate. Under these conditions, there are two factors that affect the
 214 age distribution within the stage, and hence the fraction of individuals of an
 215 appropriate age to mature. The first is the stage-specific mortality, which
 216 determines how a cohort shrinks as it ages. The second is the population
 217 growth rate, which determines the degree to which one year’s entering cohort
 218 is larger or smaller than the previous one. This makes γ non-trivial to
 219 calculate, as the resulting formula involves λ_1 , which can only be calculated
 220 once the MPM has been constructed and calibrated on data! The solution
 221 is an iterative approach: take an initial guess of λ_1 , calculate γ from the

222 formula; calculate the dominant eigenvalue of the resulting matrix; use that
 223 as a new guess for λ_1 ; and repeat until the value of λ_1 stops changing. For
 224 someone used to programming, this is straightforward to implement; but
 225 non-modellers find this daunting, and we do not know of any programs that
 226 implement this algorithm, aside from some difficult-to-generalize Matlab
 227 code in Morris and Doak (2002). **Check this out** We will refer to this as
 228 the “asymptotic age-within-stage structure” (AAS) model.

229 Thus, non-modellers will be tempted by easier-to-calculate formulas.
 230 Some discover a predecessor to the above Crowder et al. approach that
 231 appears in Crouse et al. (1987). This formula is complex-looking but
 232 straightforward to calculate, as it doesn’t require iteration (it incorporates
 233 the within-cohort dynamics, but *assumes* that $\lambda_1 = 1$). We will refer to
 234 this as the “stationary age-within-stage structure” (SAS) model (where
 235 “stationary” means that the population is neither growing nor declining).

236 An even simpler approach ~~is to simply~~ say that, if the desired duration
 237 of stage i is T_i timesteps, then the fraction maturing is $1/T_i$. ~~This effectively~~
 238 ~~assumes that the age distribution is perfectly flat, which will only be true~~
 239 ~~under asymptotic conditions if $\sigma_i/\lambda_1 = 1$. Its popularity probably stems not~~
 240 ~~just from the fact that it is simple to calculate and explain, but because~~
 241 ~~it is the first solution to the problem listed by Caswell (2001), who calls~~
 242 ~~it “intuitive.”~~ We will refer to this as the “flat age-within-stage structure”
 243 (FAS) model.

244 A rather different approach to estimating γ_i does not explicitly attempt
 245 to match the mean stage duration, but instead merely estimates γ_i as the
 246 observed fraction of surviving individuals in the stage that mature to the
 247 next stage. As long as the modelled age-within-stage distribution remains
 248 the same as it was during the observation period, then the model will get
 249 the mean stage duration correct. However, whereas the observed fraction
 250 maturing will change as the age-within-stage distribution changes in the
 251 actual population, changes in the modeled age-within-stage distribution will
 252 not be reflected in changing maturation fraction. We will refer to this as
 253 the “observed age-within-stage structure” (OAS) model.

254 As mentioned above, only the AAS model will replicate λ_1 and its
 255 sensitivities from the fully age-structured model. Of course, a final approach,
 256 if the stage durations really are fixed, is to “unroll” the stage, replacing
 257 the single stage class with T_i age classes with identical survival coefficients
 258 (or, if the assumption of homogenous survival within the stage was an
 259 approximation, with actual age-specific survival coefficients). After analysis,

the age classes can be collapsed back to their stage to generate a stage structure or sensitivity analyses that are stage-specific rather than age-specific.

3. Consequences of incorrect MPM construction

We evaluate the impacts of these errors in MPM construction by examining several endpoints that many analyses focus on: the asymptotic population growth rate (λ_1), sensitivity analysis of λ_1 to changes in underlying vital rates, and life history statistics such as generation time. We approach this evaluation through theoretical analysis (where feasible and informative) and by examining two case studies: a lionfish (*Pterois* sp.) model with very high population growth (Morris et al., 2011); and a pair of American alligator (*Alligator mississippiensis*) models that project rapidly declining and nearly constant population dynamics, respectively (Dunham et al., 2014). These studies made all three of the errors described above; we singled them out not because they are particularly egregious (many other studies make these errors) but because the authors did an exceptional job of describing the species' life history, allowing us to infer the model they meant to construct. All three of these models had other minor errors, which we corrected (while retaining the three focal errors) to create "baseline" models. We then constructed models that fixed one, two, or three of the focal errors to see how these errors affect the model projections and conclusions.

A matrix population model for lionfish was constructed by Morris et al. (2011) to investigate the potential approaches for controlling the invasive species. The modeled life history consists of three stages (larvae, juvenile, and adult), and time step of the model is one month. The original model was a post-breeding census model, but it did not include the survival of adults in the fertility rate. The average duration of the juvenile stage was assumed to be 12 months in the model. This meant they take 14 months for the first reproduction (one month in larvae stage, 12 months in juvenile stage, and one additional month in fertility rate) even though they start reproducing in 12 months. The authors used FAS model for calculating the transition rates for juveniles. We developed four stage-structured population models that gradually correct for some or all of these problems and one Leslie matrix (Table 2). Along with the original models, we used the six models to calculate λ_1 , stable stage distribution, reproductive value, sensitivity and elasticity of λ_1 to stage-specific survival rate and fecundity, damping ratio, and generation time (Appendix 1).

297 For American alligator populations, Dunham et al. (2014) developed
 298 two stage-structured matrix population models to compare the status of
 299 northern and southern populations. The original models consist of five
 300 stages (eggs, larvae, juvenile, subadults, and adults), and the time step of
 301 the model is one year. Similarly to the lionfish model, the alligator models
 302 are post-breeding census models, but the authors did not include the survival
 303 of adults in the fertility rate. The first stage was egg stage, but it only lasted
 304 for three months; therefore, there was clear inconsistency in the time steps
 305 among stages. Dunham et al. (2014) used AAS model to calculate transition
 306 rates for juvenile and subadult stages. We developed two additional stage-
 307 structured models that corrects some or all of these problems and one Leslie
 308 matrix for each population (Table 2). The four population models for each
 309 population were used to calculate the same quantities that we calculated
 310 with the lionfish models (Appendix 1).

Table 2: List of models and description of changes made to the original models.

Population	Model Description	
Lionfish	L1	- Original three-stage model in Morris et al., 2011
	L2	- Correction of juvenile duration - Incorporation of adult survival in fertility rate
	L3	- All of the corrections in L2 - Incorporation of fertility rate of juvenile stage
	L4	- All of the corrections in L3 - Use of SAS model for calculating juvenile transition rate
	L5	- All of the corrections in L3 - Use of AAS model for calculating juvenile transition rate
	L6	- Leslie matrix
American Alligator (Northern)	A1	- Original five-stage model in Dunham et al. 2014
	A2	- Reduction in the number of stage to four (hatchling, juvenile, subadult, and adult) - Incorporation of survival rate into fertility rate of adult stage

Population	Model Description
American Alligator (Southern)	- Addition of fertility rate to subadult stage
	A3 - All of the corrections made in A2
	- Calculation of juvenile transition rates using AAS model
	A4 - Leslie matrix
	A5 - Original five-stage model in Dunham et al. 2014
	A6 - Reduction in the number of stage to four (hatchling, juvenile, subadult, and adult)
	- Incorporation of survival rate into fertility rate of adult stage
	- Addition of fertility rate to subadult stage
A7	- All of the corrections made in A6
	- Calculation of juvenile transition rates using AAS model
A8	- Leslie matrix

3.1. Effects on asymptotic population growth rate (λ_1)

The original lionfish model (L1) had λ_1 of 1.125; this meant they grew by 12.5% each month (Figure S1). Even though they are invasive species and rapidly expanding. The population growth rate is unrealistically high. By reducing the average duration in juvenile stage (L2) and incorporating fertility rate on juvenile stage, the population growth rate increased further (L3). However, the use of SAS model to calculate the juvenile transition rate (L4) reduced λ_1 substantially. This is because for the duration of the juvenile stage (11 time steps on average), the survival rate is low. Incorporation of λ_1 in calculation of the transition rates (AAS models; L5) reduced λ_1 because $\lambda_1 \gg 1$. This meant that there was an increase in number of individuals recruited into juvenile stage from larval stage; therefore, the actual age-distribution declines faster with age than the stable age distribution. The Leslie matrix (L6, which is expected to be least prone to the errors associated with the manipulations of parameters, and model L5 had the same λ_1 .

The original alligator models had λ_1 of 0.87 for the northern population and 1.02 for the northern population (Figure S8). In contrast to lionfish example, λ_1 was affected very little with the corrections. Changes in the fertility rate had very little effect because λ_1 was not sensitive to fertility

rate. Changes in the transition rates for juvenile and subadult stages had little effect because, for the short durations in the stages (7 years in each stage and 3 years in each stage for northern and southern populations, respectively), they had relatively high survival rates (0.78 and 0.73 for juvenile and subadult stages of both populations).

3.2. Effects on sensitivity analysis of λ_1

Similarly to the results on λ_1 , sensitivity and elasticity of λ_1 to stage-specific survival and fecundity were affected more for lionfish than alligator. For lionfish population, the rank order of both sensitivity and elasticity among stages changed from L3 to L4 as we changed the way we calculate the transition rates (Figures S4 & S5). This implies that conservation strategy can be affected by the errors in developing a population matrix.

For lionfish models, changes in sensitivity and elasticity are less clear (Figures S11 & S12), but stable stage distribution (Figure S9) and reproductive values (Figure S10) are affected by the errors. These quantities are used for calculating sensitivity and elasticity of λ_1 . For example, the original model substantially under-estimated the stable stage distribution and over-estimated the reproductive value of adults.

3.3. Effects on estimated generation time

A variety of life history statistics, such as generation time (T) and net reproductive rate (R_0), are key quantities in comparative studies of population demography. Since these quantities are commonly estimated by analyzing MPMs (e.g., Salguero-Gómez, Jones, Jongejans, et al., 2016), it is critical to understand how sensitive these estimates are to errors in MPM construction. Here, we focus on generation time. Previous work suggests that age-structured models provide more reliable generation time estimates than do stage-structured models (Fujiwara and Diaz-Lopez, 2017), but how this depends on potential errors in matrix construction has not been examined.

There are a variety of ways to define generation time in demographic studies (Coale, 1972). In this study, we used the mean age of mothers to represent the generation time, applying formulas developed by Bienvenu and Legendre (2015). We found substantial variations among the stage-structured models. For the lionfish population, all of the stage-structured models substantially underestimated (by as much as 50%) the generation time relative to the age-structured model (Fig. S7). However, the bias was greatest under models L1-L3, which contain the most errors. For the alligator

populations, the deviations of generation time estimates between the stage-structured models and the age-structured models were more modest (Fig. S14). Although the same types of errors/approximations were incorporated in the models for northern (from A1 to A3) and southern populations (from A5 to A7), the changes in estimated generation time was not consistent between the two populations. For example, it increased from A1 to A2 whereas it decreased from A5 to A6, suggesting simple predictions of how various errors introduced in constructing population matrix would affect the generation time are challenging.

3.4. *Effects on transient dynamics*

The asymptotic growth rate and its sensitivity to underlying parameters are relevant to a population's dynamics when the population is at the stable age or stage distribution. However, in real populations subject to environmental variability or perturbations of abundance, many applications depend on knowing about the population's transient dynamics when it is far from the stable distribution. There are a range of approaches to quantifying transient dynamics (Stott, 2016); here we focus on the one of the simplest, the damping ratio, which measures how quickly transient dynamics dissipate over time. The damping ratio is given by $\lambda_1/|\lambda_2|$, where λ_1 is the eigenvalue of the largest magnitude and λ_2 is that of the second largest.

For lionfish, λ_1 remained almost the same between L1 and L2, but the damping ratio changed between them indicating the reduced juvenile duration also reduced the magnitude of λ_2 . Between L5 and L6, λ_1 and stable stage distribution were almost the same, but there was reduction in damping ratio, indicating the conversion from age-structured (L6) to stage-structured (L5) models influenced the damping ratio. Nevertheless, among all stage-structured model, L5 approximated damping ratio closest to that under L6.

The damping ratio of the alligator models between northern (A1-A4) and southern (A5-A8) populations changed differently. For example, there were larger changes for southern population than northern population, indicating it is difficult to predict how the damping ratio is affected among stage-structured models. Interestingly, the models with the most errors (A1 and A5) had the damping ratios closest to that of age structured models (A4 and A8, respectively). However, this is probably because stage-structured model generally overestimate damping ratio because the number of types (age and stage classes) is reduced from age- to stage-structured models, allowing a population to converge quickly to the asymptotic dynamics. However,

the errors were introduced to A1 and A5 to increase the duration of pre-adult stages, which caused the damping ratio to be underestimated. The bias associated with stage-structured models and the introduced errors in constructing the stage-structured models probably cancelled each other.

4. Prevalence of construction errors in published MPMs

4.1. Methods

To evaluate the prevalence of these errors in published MPMs, we examined a sample of the studies contained in the COMADRE animal matrix model database (Salguero-Gómez, Jones, Archer, et al., 2016). Using version 2.01 of the database, we subset the data to studies of nonhuman animals that had a DOI (as a simple filter to eliminate non-peer-reviewed studies). This left 65 studies published prior to the year 2000, and roughly twice that number published from 2000 to 2018. We retained all of the 20th century studies and took a random sample of 60 of the 21st century studies. Although many studies publish multiple models, representing different sites or species, we take the study as the unit of observation, as a similar approach was usually taken in all the models within a publication.

Using a haphazard subset of studies, we developed a protocol to systematically assess each study (Appendix 2). This protocol was applied by NNN members of the COMADRE digitization team, all graduate students in demography at the Max Planck Institute for Demographic Research. After initial training, consistency was ensured by having all members of the team, as well as the lead author of this paper, independently apply the protocol to the same set of papers until all were getting consistent results. If a question didn't apply (e.g., if it was not a stage-structure model), the answer was coded "NA;" if the answer could not be determined from the information in the publication, it was coded as "unknown."

- Questions related to fertility
- Questions related to first reproduction
- Questions related to maturation
- Statistical analysis: means, trends through time

4.2. Results

Note: The data collection by the Compadrinos is ongoing. Here we report an analysis based on data collected for the initial feasibility study for the project. The publications are recent and haphazardly chosen, but for this

preliminary review we have analyzed and presented these data in the same form that we plan for the final dataset.

We were able to unambiguously identify the components of the fecundity term in 24 studies. 50% of these studies failed to include an appropriate survival component in the fertility coefficients; 100% of these errors were in post-breeding census models. The frequency of these errors had no detectable trend over time ($P = 0.539$; Fig. 3a).

The potential for missing the reproductive event associated with first reaching reproductive age is only a feature of post-breeding census models. Of the 18 studies in which we could unambiguously determine both the last pre-reproductive stage or age class and in which we could identify fertility coefficients, 67% made this error. The frequency of these errors had no detectable trend over time ($P = 0.495$; Fig. 3b).

16 studies included models having at least one stage class that was meant to last for multiple timesteps; we were able to unambiguously classify the rule defining the maturation rate out of the stage(s) in 11 of them. Of these, 55% did not use a rule that would generate a value of λ_1 that would match an age-structured model with the target mean stage duration (Fig. 4). The frequency of these errors had no detectable trend over time ($P = 0.842$; Fig. 3c).

5. Discussion

[This outline will be fleshed out in the final version!]

- A. Summary of impacts
- B. Summary of prevalence
- C. Why does this happen? Most texts are incomplete (especially for stage-structured models) and/or have inconsistent or confusing notation. Talk about Caswell (2001) as example. Also, these are “threshold concepts.”
- D. Recommendations for practitioners:
 1. Use prebreeding census whenever possible
 2. Unroll developmental stages (address sensitivity analysis challenges raised by Fujiwara and Diaz-Lopez, p. 2: “The inclusion of a large number of age-classes for long-lived organisms can make the interpretation of the sensitivity and elasticity analyses complicated because individuals in multiple age classes are often practically identical but separated in an age-structured

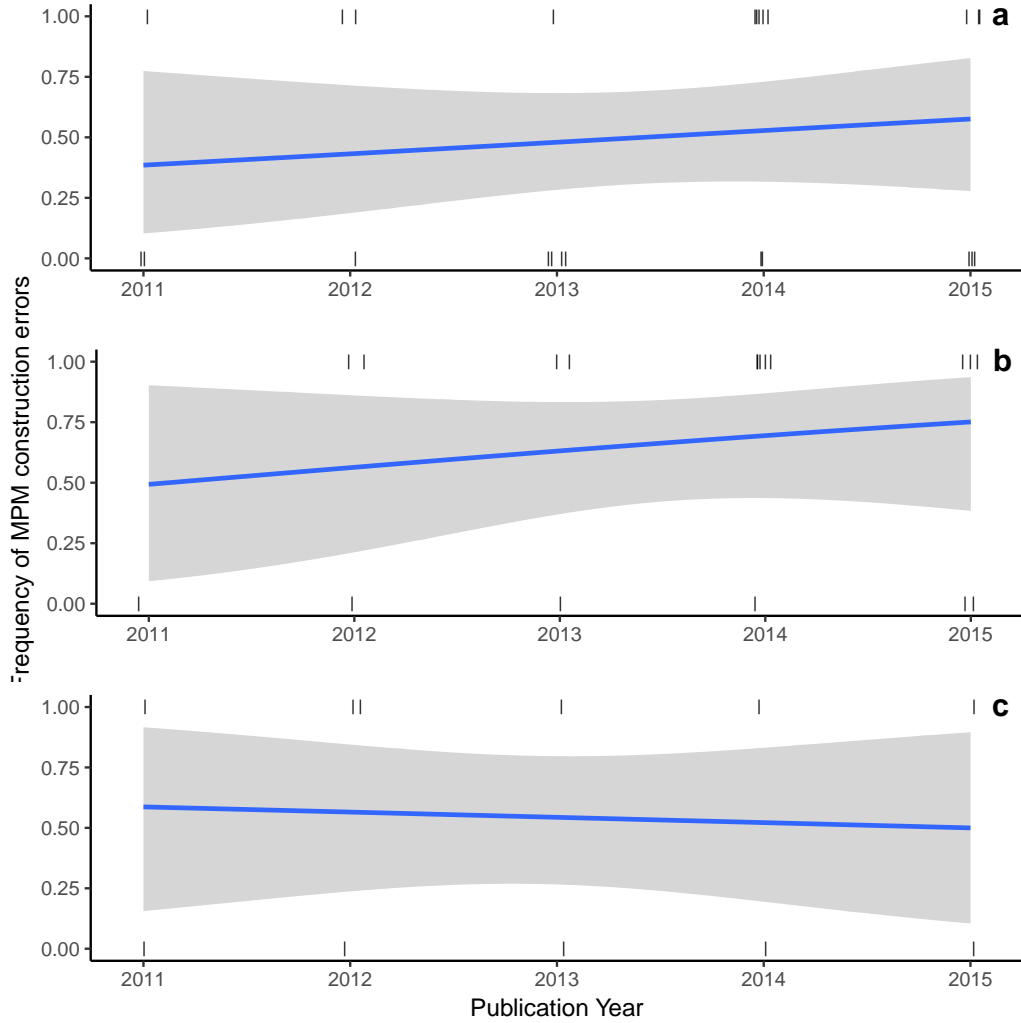


Figure 3: Trends in matrix population model construction errors through time. Trend line is logistic regression; vertical lines represent the data (jittered horizontally to prevent overlap). (a) Frequency of errors in the fertility coefficient, among all studies. (b) Frequency of errors in timing of first reproduction, among studies with post-breeding census models. (c) Frequency of errors in mean stage duration, among studies with stage classes having mean stage durations > 1 timestep.

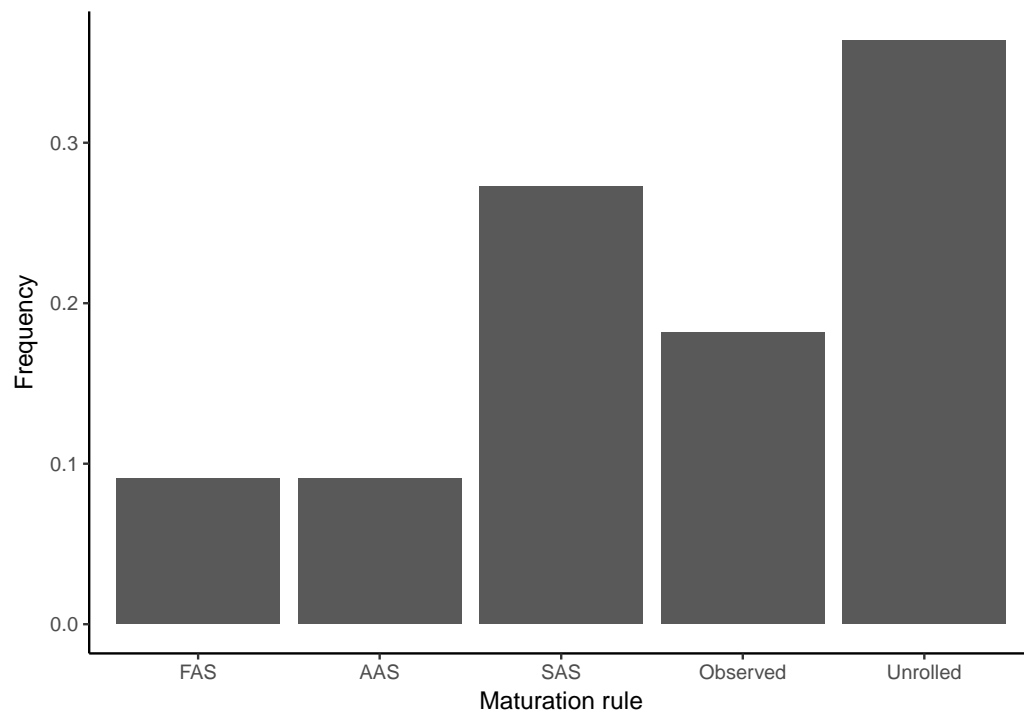


Figure 4: Frequency of approaches for setting maturation rates from stages with mean duration exceeding one timestep. [LGF: use white bars] [RSG: need to explain the categories on x axis]

476 model. Consequently, when long-lived organisms are studied, it
 477 is common to convert age-specific vital rates into stage-specific
 478 vital rates, and to use stage-structured population matrices for
 479 calculating λ_1 and generation time.”)

480 E. Recommendations for experts:

- 481 1. Develop handbooks and training materials that are both compre-
 482 hensive and clear to novices (reference threshold concepts)
- 483 2. Develop software tools to help guide practitioners through the
 484 process of translating demographic information in to MPMs

485 F. Revisiting prior results

- 486 1. Individual studies
- 487 2. Syntheses

488 5.1. Text about causes of confusion

489 This nomenclatural ambiguity plays a role in all three of the “accounting
 490 errors” that follow. An additional challenge is that most textbooks use incon-
 491 sistent notation and subscripting conventions for age- and stage-structured
 492 models. Age-structured models follow a convention that makes sense for
 493 a continuously breeding species: an individual is assigned to an age class
 494 that is numbered with the individual’s age at its *next* birthday (but note
 495 that this differs from colloquial use in most languages, where we identify a
 496 person’s age with their *previous* birthday). When looking at a species with
 497 discrete breeding seasons, this convention is perfectly fine for a pre-breeding
 498 census model, as all individuals are just a tiny fraction of a timestep away
 499 from their next birthday, and an individual that is almost three years old
 500 will have characteristics almost indistinguishable from one that is exactly
 501 three years old. [RSG: “a bit too verbose”] In contrast, for a post-breeding
 502 census model, this convention would identify an individual that is just a tiny
 503 bit past its third birthday as a four-year-old. This means that “ x -year-old
 504 survival” (often denoted P_x) refers to survival from age x to age $x + 1$ in a
 505 pre-breeding census model, but from age $x - 1$ to age x in a post-breeding
 506 census model.

507 This is potentially confusing, but most practitioners do not move between
 508 these models so it is probably not a big issue. However, transitioning from
 509 an age-structured model (which is what all textbooks use for a detailed
 510 explanation of MPMs) to a stage-structured model creates major challenges,
 511 at least for post-breeding census models. This is because, whereas an
 512 individual that was x years old at its recent birthday will be exactly $x + 1$

513 years old at its next birthday, and can be unambiguously identified as
 514 “age class $x + 1$,” an individual that was in stage x at its recent birthday
 515 might be in any of a number of stages at its next birthday, preventing the
 516 application of the age-structure convention. The solution is for post-breeding
 517 census stage-structured models to assign individuals to the stage class they
 518 were in at their recent birthday (bearing in mind that the age is measured
 519 in time steps). ; since most animal MPMs are at least partially stage-
 520 structured, we suspect that textbooks’ failure to force a confrontation with
 521 this cognitive dissonance in the conceptually simpler age-structured models
 522 leaves practitioners unprepared to face it (or even recognize its necessity) in
 523 stage-structured models. This is perfectly sensible, but *most textbooks make*
 524 *this transition without comment*. [CHECK WHAT MILLS DOES HERE]
 525 Thus it is not obvious to a novice that the carefully explained conventions
 526 of age-structured models no longer apply; we suspect that this contributes
 527 to some of the common errors described below. ; since most animal MPMs
 528 are at least partially stage-structured, we suspect that textbooks’ failure
 529 to force a confrontation with this cognitive dissonance in the conceptually
 530 simpler age-structured models leaves practitioners unprepared to face it (or
 531 even recognize its necessity) in stage-structured models.

532 5.2. Text about birth-flow models

533 However, much of the theoretical development of MPMs was motivated
 534 by human demography, in which reproduction occurs throughout the year
 535 (“birth-flow” populations). This may require a variety of mathematical ap-
 536 proximations to suitably transform continuous-time processes into a discrete-
 537 time representation, and gives a different view of the identity of individuals
 538 in a particular age or stage class. We will not treat the former here, but the
 539 latter is relevant.

540 The discretization of age into classes means that in a birth-flow popu-
 541 lation, the first age class represents all individuals between zero and one
 542 timestep old at time t , and thus we would describe it as “Age 0–1.”

543 But we often see birth-pulse models in which the age classes are given as
 544 a range—this is incorrect, and leads to ambiguity (does the class “Age 0–1”
 545 refer to newborns or to one-year-olds?) that can confuse the practitioner
 546 on how best to construct the model. This nomenclature problem likely
 547 has two sources. First, many texts introduce birth-flow models first (e.g.,
 548 the very first figure in Caswell, 2001, illustrating the notion of discrete age
 549 classes, shows the classes as ages 0–1, 1–2, etc.). Second, in colloquial use,
 550 age-associated terms often span an age range: for example, in ungulates the

term “yearling” refers to any individual between the ages of one and two. When that same term is applied to a class in a birth-pulse MPM, nontrivial cognitive effort is required to mentally redefine it (so that “Yearling” means age exactly one).

5.3. Misc discussion items

- Paragraph about how AAS, SAS, and FAS models are related to each other [MF].
- Mention other types of errors in MPM construction
- Inappropriate fecundity term (e.g., using birth-pulse for continuously breeding species)
- Issues with size, esp. using observed transitions between size classes
- Issues with seeds/seed banks

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