Summary

The summary file contains summary information for all the raw files processed with a single MaxQuant run. The summary information consists of some MaxQuant parameters, information of the raw file contents, and statistics on the peak detection. Based on this file a quick overview can be gathered on the quality of the data in the raw file.

The last row in this file contains the summary information for each column on each of the processed files.

Name	Separator	Description
Raw file		The raw file processed.
Experiment		Experiment name assigned to this LC-MS run in the experimental design.
Enzyme		The protease used to digest the protein sample.
Enzyme mode		The protease used to digest the protein sample.
Enzyme first search		The protease used for the first search.
Enzyme mode first search		The protease used for the first search.
Use enzyme first search		Marked with '+' when a different protease setup was used for the first search.
Variable modifications		The variable modification(s) used during the identification of peptides.
Fixed modifications		The fixed modification(s) used during the identification of peptides.
Multi modifications		The multi modification(s) used during the identification of peptides.
Variable modifications first search		The variable modification(s) used during the first search.
Use variable modifications first search		Marked with '+' when different variable modifications were used for the first search.
Requantify		The number of labels used.
Multiplicity		The number of labels used.
Max. missed cleavages		The maximum allowed number of missed cleavages.
Labels0		The labels used in the labeling experiment. Allowed values for X: 0=light; 1=medium; 2=heavy label partner.
LC-MS run type		The type of LC-MS run. Usually it will be 'Standard' which refers to a conventional shotgun proteomics run with data-dependent MS/MS.
Time-dependent recalibration		When marked with '+', time-dependent recalibration was applied to improve the data quality.
MS		The number of MS spectra recorded in this raw file.
MS/MS		The number of MS/MS spectra recorded in this raw file.
MS3		The number of MS3 spectra recorded in this raw file.
MS/MS Submitted		The number of tandem MS spectra submitted for analysis.
MS/MS Submitted (SIL)		The number of tandem MS spectra submitted for analysis, where the precursor ion was detected as part of a labeling cluster.
MS/MS Submitted (ISO)		The number of tandem MS spectra submitted for analysis, where the precursor ion was detected as an isotopic pattern.
MS/MS Submitted (PEAK)		The number of tandem MS spectra submitted for analysis, where the precursor ion was detected as a single peak.
MS/MS Identified		The total number of identified tandem MS spectra.
MS/MS Identified (SIL)		The total number of identified tandem MS spectra, where the precursor ion was detected as part of a labeling cluster.
MS/MS Identified (ISO)		The total number of identified tandem MS spectra, where the precursor ion was detected as an isotopic pattern.
MS/MS Identified (PEAK)		The total number of identified tandem MS spectra, where the precursor ion was detected as a single peak.
MS/MS Identified [%]		The percentage of identified tandem MS spectra.
MS/MS Identified (SIL) [%]		The percentage of identified tandem MS spectra, where the precursor ion was detected as part of a labeling cluster.
MS/MS Identified (ISO) [%]		The percentage of identified tandem MS spectra, where the precursor ion was detected as an isotopic pattern.
MS/MS Identified (PEAK) [%]		The percentage of identified tandem MS spectra, where the precursor ion was detected as a single peak.
Peptide Sequences Identified		The total number of unique peptide amino acid sequences identified from the recorded tandem mass spectra.
Peaks		The total number of peaks detected in the full scans.
Peaks Sequenced		The total number of peaks sequenced by tandem MS.

Peaks Sequenced [%]	The percentage of peaks sequenced by tandem MS.
Peaks Repeatedly Sequenced	The total number of peaks repeatedly sequenced (i.e. 1 or more times) by tandem MS.
Peaks Repeatedly Sequenced [%]	The percentage of peaks repeatedly sequenced (i.e. 1 or more times) by tandem MS.
Isotope Patterns	The total number of detected isotope patterns.
Isotope Patterns Sequenced	The total number of isotope patterns sequenced by tandem MS.
Isotope Patterns Sequenced (z>1)	The total number of isotope patterns sequenced by tandem MS with a charge state of 2 or more.
Isotope Patterns Sequenced [%]	The percentage of isotope patterns sequenced by tandem MS.
Isotope Patterns Sequenced (z>1) [%]	The percentage of isotope patterns sequenced by tandem MS with a charge state of 2 or more.
Isotope Patterns Repeatedly Sequenced	The total number of isotope patterns repeatedly sequenced (i.e. 1 or more times) by tandem MS.
Isotope Patterns Repeatedly Sequenced [%]	The percentage of isotope patterns repeatedly sequenced (i.e. 1 or more times) by tandem MS.
Recalibrated	When marked with '+', the masses taken from the raw file were recalibrated.
Av. Absolute Mass Deviation [ppm]	The average absolute mass deviation found comparing to the identification mass in parts per million.
Mass Standard Deviation [ppm]	The standard deviation of the mass deviation found comparing to the identification mass in parts per million.
Av. Absolute Mass Deviation [mDa]	The average absolute mass deviation found comparing to the identification mass in milli-Dalton.
Mass Standard Deviation [mDa]	The standard deviation of the mass deviation found comparing to the identification mass in milli-Dalton.

Evidence

The evidence file combines all the information about the identified peptides and normally is the only file required for processing the results. Additional information about the peptides, modifications, proteins, etc. can be found in the other files by unique identifier linkage.

Name	Separator	Description
Sequence		The identified AA sequence of the peptide.
Length		The length of the sequence stored in the column 'Sequence'.
Modifications		Post-translational modifications contained within the identified peptide sequence.
Modified sequence		Sequence representation including the post-translational modifications (abbreviation of the modification in brackets before the modified AA). The sequence is always surrounded by underscore characters ('_').
Oxidation (M) Probabilities		Sequence representation of the peptide including PTM positioning probabilities ([01], where 1 is best match) for 'Oxidation (M)'.
Phospho (STY) Probabilities		Sequence representation of the peptide including PTM positioning probabilities ([01], where 1 is best match) for 'Phospho (STY)'.
Oxidation (M) Score Diffs		Sequence representation for each of the possible PTM positions in each possible configuration, the difference is calculated between the identification score with the PTM added to that position and the best scoring identification where no PTM is added to that position. When this value is negative, it is unlikely that the particular modification is located at this position.
Phospho (STY) Score Diffs		Sequence representation for each of the possible PTM positions in each possible configuration, the difference is calculated between the identification score with the PTM added to that position and the best scoring identification where no PTM is added to that position. When this value is negative, it is unlikely that the particular modification is located at this position.
Acetyl (Protein N-term)		The number of occurrences of the modification 'Acetyl (Protein N-term)'.
Oxidation (M)		The number of occurrences of the modification 'Oxidation (M)'.
Phospho (STY)		The number of occurrences of the modification 'Phospho (STY)'.
Missed cleavages		Number of missed enzymatic cleavages.
Proteins		The identifiers of the proteins this particular peptide is associated with.
Leading proteins		The identifiers of the proteins in the proteinGroups file, with this protein as best match, this particular peptide is associated with. When multiple matches are found here, the best scoring protein can be found in the 'Leading Razor Protein' column.
Leading razor protein		The identifier of the best scoring protein, from the proteinGroups file this, this peptide is associated to.
Туре		The type of the feature. 'MSMS' for an MS/MS spectrum without an MS1 isotope pattern assigned. 'ISO-MSMS' MS1 isotope cluster identified by MS/MS. 'MULTI-MSMS' MS1 labeling cluster identified by MS/MS. 'MULTI-SECPEP' MS1 labeling cluster identified by MS/MS as second peptide. 'MULTI-MATCH' MS1 labeling cluster identified by matching between runs. In case of label-free data there is no difference between 'MULTI' and 'ISO'.
Raw file		The name of the RAW-file the mass spectral data was derived from.
Experiment		
MS/MS m/z		The m/z used for fragmentation (not necessarily the monoisotopic m/z).
Charge		The charge-state of the precursor ion.
m/z		The recalibrated mass-over-charge value of the precursor ion.
Mass		The predicted monoisotopic mass of the identified peptide sequence.
Resolution		The resolution of precursor ion measured in Full Width at Half Maximum (FWHM).
Uncalibrated - Calibrated m/z [ppm]		The difference between the uncalibrated and recalibrated mass-over-charge value of the precursor ion measured in parts-per-million. This gives an indication of the mass drift in the original data, which was automatically corrected by MaxQuant.

Uncalibrated - Calibrated m/z [Da]	The difference between the uncalibrated and recalibrated mass-over-charge value of the precursor ion measured in parts-per-million. This gives an indication of the mass drift in the original data, which was automatically corrected by
Mass error [ppm]	MaxQuant. Mass error of the recalibrated mass-over-charge value of the precursor ion in comparison to the predicted monoisotopic mass of the identified peptide sequence in parts per million.
Mass error [Da]	Mass error of the recalibrated mass-over-charge value of the precursor ion in comparison to the predicted monoisotopic mass of the identified peptide sequence in milli-Dalton.
Uncalibrated mass error [ppm]	Mass error of the uncalibrated mass-over-charge value of the precursor ion in comparison to the predicted monoisotopic mass of the identified peptide sequence.
	Note: This column can contain missing values (denoted as NaN).
Uncalibrated mass error [Da]	Mass error of the uncalibrated mass-over-charge value of the precursor ion in comparison to the predicted monoisotopic mass of the identified peptide sequence.
	Note: This column can contain missing values (denoted as NaN).
Max intensity m/z 0	Summed up eXtracted Ion Current (XIC) of all isotopic clusters associated with the identified AA sequence. In case of a labeled experiment this is the total intensity of all the isotopic patterns in the label cluster.
Retention time	The uncalibrated retention time in minutes in the elution profile of the precursor ion.
Retention length	The total retention time length of the peak (last timepoint first timepoint).
Calibrated retention time	The recalibrated retention time in minutes in the elution profile of the precursor ion.
Calibrated retention time start	The recalibrated retention start in minutes in the elution profile of the precursor ion.
Calibrated retention time finish	The recalibrated retention finish in minutes in the elution profile of the precursor ion.
Retention time calibration	The difference in minutes between the uncalibrated and recalibrated retention time. This gives an indication of the retention time drift in the original data, which was automatically corrected by MaxQuant.
Match time difference	Note: This column can contain missing values (NaN). When the option match between runs is used in MaxQuant, this value indicates the time difference between the feature from the raw file it was taken from and the feature from the raw file it was matched to.
Match m/z difference	When the option match between runs is used in MaxQuant, this value indicates the m/z difference between the feature from the raw file it was taken from and the feature from the raw file it was matched to.
Match q-value	This is the q-value for features that have been identified by 'matching between runs'.
Match score	The andromeda score of the MS/MS identification that is the source of this identification by 'matching between runs'.
Number of data points	The number of data points (peak centroids) collected for this peptide feature.
Number of scans	The number of MS scans that the 3d peaks of this peptide feature are overlapping with.
Number of isotopic peaks	The number of isotopic peaks contained in this peptide feature.
PIF	Short for Parent Ion Fraction; indicates the fraction the target peak makes up of the total intensity in the inclusion window.
Fraction of total spectrum	The percentage the ion intensity makes up of the total intensity of the whole spectrum.
Base peak fraction	The percentage the parent ion intensity in comparison to the highest peak in the MS spectrum.
PEP	Posterior Error Probability of the identification. This value essentially operates as a p-value, where smaller is more significant.
MS/MS count	The number of sequencing events for this sequence, which matches the number of identifiers stored in the column MS/MS IDs.
MS/MS scan number	The RAW-file derived scan number of the MS/MS with the highest peptide identification score (the highest score is stored in the column 'Score').
Score	Andromeda score for the best associated MS/MS spectrum.
Delta score	Score difference to the second best identified peptide.
Combinatorics	Number of possible distributions of the modifications over the peptide sequence.

Intensity	Summed up extracted Ion Current (XIC) of all isotopic clusters associated with the identified AA sequence. In case of a labeled experiment this is the total intensity of all the isotopic patterns in the label cluster.
Reporter intensity corrected 1	
Reporter intensity corrected 2	
Reporter intensity corrected 3	
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Reporter intensity count 3	
Reporter intensity count 4	
Reporter intensity count 5	
Reporter intensity count 6	
Reporter intensity count 7	
Reporter intensity count 8	
Reporter intensity count 9	
Reporter intensity count 9	
Reporter PIF	
Reporter fraction	
Reverse	When marked with 't' this particular poptide was found to be
Reverse	When marked with '+', this particular peptide was found to be part of a protein derived from the reversed part of the decoy database. These should be removed for further data analysis.
Potential contaminant	When marked with '+', this particular peptide was found to be part of a commonly occurring contaminant. These should be removed for further data analysis.
id	A unique (consecutive) identifier for each row in the evidence table, which is used to cross-link the information in this file with the information stored in the other files.
Protein group IDs	The identifier of the protein-group this redundant peptide sequence is associated with, which can be used to look up the extended protein information in the file 'proteinGroups.txt'. As a single peptide can be linked to multiple proteins (e.g. in the case of razor-proteins), multiple ids can be stored here separated by a semicolon. As a protein can be identified by multiple peptides, the same id can be found in different rows.
Peptide ID	The identifier of the non-redundant peptide sequence.
Mod. peptide ID	Identifier of the associated modification summary stored in the file 'modificationSpecificPeptides.txt'.
MS/MS IDs	Identifier(s) of the associated MS/MS summary(s) stored in the file 'msms.txt'.
Best MS/MS	Identifier(s) of the best MS/MS associated spectrum stored in the file 'msms.txt'.
AIF MS/MS IDs	Identifier(s) of the associated All Ion Fragmentation MS/MS summary(s) stored in the file 'aifMsms.txt'.
Oxidation (M) site IDs	Identifier(s) of the modification summary stored in the file 'Oxidation (M)Sites.txt'.
Phospho (STY) site IDs	Identifier(s) of the modification summary stored in the file 'Phospho (STY)Sites.txt'.

Peptides

The peptides table contains information on the identified peptides in the processed raw-files.

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Unique (Groups)	When marked with '+', this particular peptide is unique to a single protein group in the proteinGroups file.
Unique (Proteins)	When marked with '+', this particular peptide is unique to a single protein sequence in the fasta file(s).
Charges	All charge states that have been observed.
PEP	Posterior Error Probability of the identification. This value essentially operates as a p-value, where smaller is more significant.
Score	Highest Andromeda score for the associated MS/MS spectra.
Experiment TMT1	Number of evidence entries for this 'Experiment'.
Experiment TMT1phos	Number of evidence entries for this 'Experiment'.
Experiment TMT2	Number of evidence entries for this 'Experiment'.
Experiment TMT2phos	Number of evidence entries for this 'Experiment'.
Reporter intensity corrected 1	<u> </u>
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Intensity	Summed up eXtracted Ion Current (XIC) of all isotopic clusters associated with the identified AA sequence. In case of a labeled experiment this is the total intensity of all the isotopic patterns in the label cluster.
Intensity TMT1	Summed up eXtracted Ion Current (XIC) of all isotopic clusters associated with the identified AA sequence. In case of a labeled experiment this is the total intensity of all the isotopic patterns in the label cluster.
Intensity TMT1phos	Summed up eXtracted Ion Current (XIC) of all isotopic clusters associated with the identified AA sequence. In case of a labeled experiment this is the total intensity of all the isotopic patterns in the label cluster.
Intensity TMT2	Summed up eXtracted Ion Current (XIC) of all isotopic clusters associated with the identified AA sequence. In case of a labeled experiment this is the total intensity of all the isotopic patterns in the label cluster.
Intensity TMT2phos	Summed up eXtracted Ion Current (XIC) of all isotopic clusters associated with the identified AA sequence. In case of a labeled experiment this is the total intensity of all the isotopic patterns in the label cluster.
Reverse	When marked with '+', this particular peptide was found to be part of a protein derived from the reversed part of the decoy database. These should be removed for further data analysis.
Potential contaminant	When marked with '+', this particular peptide was found to be part of a commonly occurring contaminant. These should be removed for further data analysis.
id	A unique (consecutive) identifier for each row in the peptides table, which is used to cross-link the information in this table with the information stored in the other tables.
Protein group IDs	The identifiers of the protein groups this peptide was linked to, referenced against the proteinGroups table.
Mod. peptide IDs	Identifier(s) for peptide sequence(s), associated with the peptide, referenced against the corresponding modified peptides table.
Evidence IDs	Identifier(s) for analyzed peptide evidence associated with the protein group referenced against the evidence table.
MS/MS IDs	The identifiers of the MS/MS scans identifying this peptide, referenced against the msms table.
Best MS/MS	The identifier of the best (in terms of quality) MS/MS scan identifying this peptide, referenced against the msms table.
Oxidation (M) site IDs	Identifier(s) for site(s) associated with the protein group, which show(s) evidence of the modification, referenced against the appropriate modification site file.
Phospho (STY) site IDs	Identifier(s) for site(s) associated with the protein group, which show(s) evidence of the modification, referenced against the appropriate modification site file.
MS/MS Count	

Modification-specific peptides

Name	Separator	Description
Sequence		The identified AA sequence of the peptide.
Modifications		Post-translational modifications contained within the sequence. When no modifications exist, this is set to 'unmodified'.
Mass		Charge corrected mass of the precursor ion.
Mass Fractional Part		The values after the decimal point (ie value - floor(value)).
Protein Groups		IDs of the protein groups to which this peptide belongs.
Proteins		The identifiers of the proteins this particular peptide is associated with.
Unique (Groups)		When marked with '+', this particular peptide is unique to a single protein group in the proteinGroups file.
Unique (Proteins)		When marked with '+', this particular peptide is unique to a single protein sequence in the fasta file(s).
Acetyl (Protein N-term)		Number of Acetyl (Protein N-term) on this peptide.
Oxidation (M)		Number of Oxidation (M) on this peptide.
Phospho (STY)		Number of Phospho (STY) on this peptide.
Missed cleavages		Number of missed enzymatic cleavages.
Experiment TMT1		Number of evidence entries for this 'Experiment'.
Experiment TMT1phos		Number of evidence entries for this 'Experiment'.
Experiment TMT2		Number of evidence entries for this 'Experiment'.
Experiment TMT2phos		Number of evidence entries for this 'Experiment'.
Retention time		Retention time in minutes averaged over the evidence entries belonging to this modification-specific peptide.
Calibrated retention time		Calibrated retention time averaged over the evidence entries belonging to this modification-specific peptide. Obviously this only makes sense if retention time recalibration has been performed which is the case when matching between run is selected.
Charges		All charge states that have been observed.
PEP		Posterior Error Probability of the identification. This value essentially operates as a p-value, where smaller is more significant.
MS/MS scan number		The RAW-file derived scan number of the MS/MS with the highest peptide identification score (the highest score is stored in the column 'Score').
Raw file		The name of the RAW-file the mass spectral data was derived from.
Score		Andromeda score for the best identified among the associated MS/MS spectra.
Delta score		Score difference to the second best identified peptide.
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MS/MS Count	

Oxidation (M)Sites

Name	Separator	Description
Proteins	_	Identifiers of proteins this site is associated with.
Positions within proteins		For each protein identifier in the 'Proteins' column you find here the position of the site in the respective protein sequence. The index of the first amino acid in the sequence is 1.
Leading proteins		
Protein		Identifier of the protein this peptide is associated with.
Fasta headers		Descriptions of proteins this peptide is associated with.
Localization prob		
Score diff		
PEP		The posterior error probability (PEP) of the best identified modified peptide containing this site.
Score		The Andromeda score of the best identified modified peptide containing this site.
Delta score		The Andromeda delta score of the best identified modified peptide containing this site.
Score for localization		The Andromeda score of the MS/MS spectrum used for calculating the localization score for this site.
Localization prob TMT1		
Score diff TMT1		
PEP TMT1		
Score TMT1		
Localization prob TMT1phos		
Score diff TMT1phos		
PEP TMT1phos		
Score TMT1phos		
Localization prob TMT2		
Score diff TMT2		
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Score TMT2		
Localization prob TMT2phos		
Score diff TMT2phos		
PEP TMT2phos		+
Score TMT2phos		
Diagnostic peak		
Number of Oxidation (M)		Different numbers of Oxidation (M) on peptides that this site is
. ,		involved in.
Amino acid		
Sequence window		
Modification window		
Peptide window coverage		
Oxidation (M) Probabilities		
Oxidation (M) Score diffs		
Position in peptide		
Charge		Charge state of the precursor ion.
Mass error [ppm]		Mass error of the recalibrated mass-over-charge value of the precursor ion in comparison to the predicted monoisotopic mass of the identified peptide sequence.
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Reporter intensity count 9 TMT2phos3	
Reporter intensity count 10 TMT2phos1	
Reporter intensity count 10 TMT2phos2	
Reporter intensity count 10 TMT2phos3	
Intensity	Summed up eXtracted Ion Current (XIC) of all isotopic clusters associated with the identified AA sequence. In case of a labeled experiment this is the total intensity of all the isotopic patterns in the label cluster.
Intensity1	Summed up eXtracted Ion Current (XIC) of all isotopic clusters associated with the identified AA sequence. In case of a labeled experiment this is the total intensity of all the isotopic patterns in the label cluster.
Intensity2	Summed up eXtracted Ion Current (XIC) of all isotopic clusters associated with the identified AA sequence. In case of a labeled experiment this is the total intensity of all the isotopic patterns in the label cluster.
Intensity3	Summed up eXtracted Ion Current (XIC) of all isotopic clusters associated with the identified AA sequence. In case of a labeled experiment this is the total intensity of all the isotopic patterns in the label cluster.
Ratio mod/base	
Intensity TMT1	Summed up eXtracted Ion Current (XIC) of all isotopic clusters associated with the identified AA sequence. In case of a labeled experiment this is the total intensity of all the isotopic patterns in the label cluster.
Intensity TMT1phos	Summed up eXtracted Ion Current (XIC) of all isotopic clusters associated with the identified AA sequence. In case of a labeled experiment this is the total intensity of all the isotopic patterns in the label cluster.
Intensity TMT2	Summed up extracted Ion Current (XIC) of all isotopic clusters associated with the identified AA sequence. In case of a labeled experiment this is the total intensity of all the isotopic patterns in the label cluster.
Intensity TMT2phos	Summed up eXtracted Ion Current (XIC) of all isotopic clusters associated with the identified AA sequence. In case of a labeled experiment this is the total intensity of all the isotopic patterns in the label cluster.
Ratio mod/base TMT1	
Ratio mod/base TMT1phos	
Ratio mod/base TMT2	
Ratio mod/base TMT2phos	
Intensity TMT11	Summed up eXtracted Ion Current (XIC) of all isotopic clusters associated with the identified AA sequence. In case of a labeled experiment this is the total intensity of all the isotopic patterns in the label cluster.

Intensity TMT12	Summed up eXtracted Ion Current (XIC) of all isotopic clusters associated with the identified AA sequence. In case of a labeled experiment this is the total intensity of all the isotopic patterns in the label cluster.
Intensity TMT13	Summed up eXtracted Ion Current (XIC) of all isotopic clusters associated with the identified AA sequence. In case of a labeled experiment this is the total intensity of all the isotopic patterns in the label cluster.
Intensity TMT1phos1	Summed up eXtracted Ion Current (XIC) of all isotopic clusters associated with the identified AA sequence. In case of a labeled experiment this is the total intensity of all the isotopic patterns in the label cluster.
Intensity TMT1phos2	Summed up eXtracted Ion Current (XIC) of all isotopic clusters associated with the identified AA sequence. In case of a labeled experiment this is the total intensity of all the isotopic patterns in the label cluster.
Intensity TMT1phos3	Summed up eXtracted Ion Current (XIC) of all isotopic clusters associated with the identified AA sequence. In case of a labeled experiment this is the total intensity of all the isotopic patterns in the label cluster.
Intensity TMT21	Summed up eXtracted Ion Current (XIC) of all isotopic clusters associated with the identified AA sequence. In case of a labeled experiment this is the total intensity of all the isotopic patterns in the label cluster.
Intensity TMT22	Summed up eXtracted Ion Current (XIC) of all isotopic clusters associated with the identified AA sequence. In case of a labeled experiment this is the total intensity of all the isotopic patterns in the label cluster.
Intensity TMT23	Summed up eXtracted Ion Current (XIC) of all isotopic clusters associated with the identified AA sequence. In case of a labeled experiment this is the total intensity of all the isotopic patterns in the label cluster.
Intensity TMT2phos1	Summed up eXtracted Ion Current (XIC) of all isotopic clusters associated with the identified AA sequence. In case of a labeled experiment this is the total intensity of all the isotopic patterns in the label cluster.
Intensity TMT2phos2	Summed up eXtracted Ion Current (XIC) of all isotopic clusters associated with the identified AA sequence. In case of a labeled experiment this is the total intensity of all the isotopic patterns in the label cluster.
Intensity TMT2phos3	Summed up eXtracted Ion Current (XIC) of all isotopic clusters associated with the identified AA sequence. In case of a labeled experiment this is the total intensity of all the isotopic patterns in the label cluster.
Reverse	When marked with '+', this particular peptide was found to be part of a protein derived from the reversed part of the protein sequence database. These should be removed for further data analysis.
Potential contaminant	When marked with '+', this particular peptide was found to be part of a commonly occurring contaminant. These should be removed for further data analysis.
id	A unique (consecutive) identifier for each row in the site table, which is used to cross-link the information in this file with the information stored in the other files.
Protein group IDs	The identifier of the protein-group this peptide sequence is associated with, which can be used to look up the extended protein information in the file 'proteinGroups.txt'. As a single peptide can be linked to multiple proteins (e.g. in the case of razor-proteins), multiple id's can be stored here separated by a semicolon. As a protein can be identified by multiple peptides, the same id can be found in different rows.
Positions	The positions of the modifications in the protein amino acid sequence.
Position	The position of the modification in the protein amino acid sequence.
Peptide IDs	Identifier(s) of the associated peptide sequence(s) summary, which can be found in the file 'peptides.txt'.
Mod. peptide IDs	Identifier(s) of the associated peptide sequence(s) summary, which can be found in the file 'modificationSpecificPeptides.txt'.
Evidence IDs	Identifier(s) for analyzed peptide evidence associated with the protein group referenced against the evidence table.
MS/MS IDs	The identifiers of the MS/MS scans identifying this peptide, referenced against the msms table.
Best localization evidence ID	
Best localization MS/MS ID	
Best localization raw file	
Best localization scan number	
Best score evidence ID	
Best score MS/MS ID	

Best score scan number	
Best PEP evidence ID	
Best PEP MS/MS ID	
Best PEP raw file	
Best PEP scan number	

Phospho (STY)Sites

Name	Separator	Description
Proteins		Identifiers of proteins this site is associated with.
Positions within proteins		For each protein identifier in the 'Proteins' column you find here the position of the site in the respective protein sequence. The index of the first amino acid in the sequence is 1.
Leading proteins		
Protein		Identifier of the protein this peptide is associated with.
Fasta headers		Descriptions of proteins this peptide is associated with.
Localization prob		
Score diff		
PEP		The posterior error probability (PEP) of the best identified modified peptide containing this site.
Score		The Andromeda score of the best identified modified peptide containing this site.
Delta score		The Andromeda delta score of the best identified modified peptide containing this site.
Score for localization		The Andromeda score of the MS/MS spectrum used for calculating the localization score for this site.
Localization prob TMT1		
Score diff TMT1		
PEP TMT1		
Score TMT1		
Localization prob TMT1phos		
Score diff TMT1phos		
PEP TMT1phos		
Score TMT1phos		
Localization prob TMT2		
Score diff TMT2		
PEP TMT2		
Score TMT2		
Localization prob TMT2phos		
Score diff TMT2phos		
PEP TMT2phos		
Score TMT2phos		
Diagnostic peak		
Number of Phospho (STY)		Different numbers of Phospho (STY) on peptides that this site is involved in.
Amino acid		
Sequence window		
Modification window		
Peptide window coverage		
Phospho (STY) Probabilities		
Phospho (STY) Score diffs		
Position in peptide		1
Charge Mass error [ppm]		Charge state of the precursor ion. Mass error of the recalibrated mass-over-charge value of the precursor ion in comparison to the predicted monoisotopic mass of the identified peptide sequence.
Reporter intensity corrected 11	+	mass of the luchtimed peptide sequence.
Reporter intensity corrected 11 Reporter intensity corrected 12	+	+
Reporter intensity corrected 12	+	+
Reporter intensity corrected 13		+
Reporter intensity corrected 21 Reporter intensity corrected 22	+	+
Reporter intensity corrected 23		
Reporter intensity corrected 31		
Reporter intensity corrected 32		
Reporter intensity corrected 33		
Reporter intensity corrected 41		
Reporter intensity corrected 42		
Reporter intensity corrected 43		

Reporter intensity corrected 52	
Reporter intensity corrected 53	
Reporter intensity corrected 61	
Reporter intensity corrected 62	
Reporter intensity corrected 63	
Reporter intensity corrected 72	
Reporter intensity corrected 73	
Reporter intensity corrected 81	
Reporter intensity corrected 82	
Reporter intensity corrected 83	
Reporter intensity corrected 91	
Reporter intensity corrected 92	
Reporter intensity corrected 93	
Reporter intensity corrected 101	
Reporter intensity corrected 102	
Reporter intensity corrected 103	
Reporter intensity derrected 166	
Reporter intensity 13	
Reporter intensity 21	
Reporter intensity 22	
Reporter intensity 23	
Reporter intensity 31	
Reporter intensity 32	
Reporter intensity 33	
Reporter intensity 41	
Reporter intensity 42	
Reporter intensity 43	
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TMT2phos3 Reporter intensity count 2 TMT2phos1 Reporter intensity count 2 TMT2phos2 Reporter intensity count 2 TMT2phos3 Reporter intensity count 3 TMT2phos1 Reporter intensity count 3 TMT2phos2 Reporter intensity count 3 TMT2phos2 Reporter intensity count 3 TMT2phos3 Reporter intensity count 4 TMT2phos1 Reporter intensity count 4 TMT2phos2 Reporter intensity count 4	Reporter intensity count 1 TMT2phos2	
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TMT2phos2 Reporter intensity count 4	Reporter intensity count 4 TMT2phos1	
Reporter intensity count 4 TMT2phos3		
	Reporter intensity count 4 TMT2phos3	

Reporter intensity count 5 TMT2phos1	
Reporter intensity count 5	
Reporter intensity count 5 TMT2phos3	
Reporter intensity count 6 TMT2phos1	
Reporter intensity count 6 TMT2phos2	
Reporter intensity count 6 TMT2phos3	
Reporter intensity count 7 TMT2phos1	
Reporter intensity count 7 TMT2phos2	
Reporter intensity count 7 TMT2phos3	
Reporter intensity count 8 TMT2phos1	
Reporter intensity count 8 TMT2phos2	
Reporter intensity count 8 TMT2phos3	
Reporter intensity count 9 TMT2phos1	
Reporter intensity count 9 TMT2phos2	
Reporter intensity count 9 TMT2phos3	
Reporter intensity count 10 TMT2phos1	
Reporter intensity count 10 TMT2phos2	
Reporter intensity count 10 TMT2phos3	
Intensity	Summed up eXtracted Ion Current (XIC) of all isotopic clusters associated with the identified AA sequence. In case of a labeled experiment this is the total intensity of all the isotopic patterns in the label cluster.
Intensity1	Summed up eXtracted Ion Current (XIC) of all isotopic clusters associated with the identified AA sequence. In case of a labeled experiment this is the total intensity of all the isotopic patterns in the label cluster.
Intensity2	Summed up eXtracted Ion Current (XIC) of all isotopic clusters associated with the identified AA sequence. In case of a labeled experiment this is the total intensity of all the isotopic patterns in the label cluster.
Intensity3	Summed up eXtracted Ion Current (XIC) of all isotopic clusters associated with the identified AA sequence. In case of a labeled experiment this is the total intensity of all the isotopic patterns in the label cluster.
Ratio mod/base	
Intensity TMT1	Summed up eXtracted Ion Current (XIC) of all isotopic clusters associated with the identified AA sequence. In case of a labeled experiment this is the total intensity of all the isotopic patterns in the label cluster.
Intensity TMT1phos	Summed up eXtracted Ion Current (XIC) of all isotopic clusters associated with the identified AA sequence. In case of a labeled experiment this is the total intensity of all the isotopic patterns in the label cluster.
Intensity TMT2	Summed up extracted Ion Current (XIC) of all isotopic clusters associated with the identified AA sequence. In case of a labeled experiment this is the total intensity of all the isotopic patterns in the label cluster.
Intensity TMT2phos	Summed up eXtracted Ion Current (XIC) of all isotopic clusters associated with the identified AA sequence. In case of a labeled experiment this is the total intensity of all the isotopic patterns in the label cluster.
Ratio mod/base TMT1	
Ratio mod/base TMT1phos	
Ratio mod/base TMT2	
Ratio mod/base TMT2phos	
Intensity TMT11	Summed up eXtracted Ion Current (XIC) of all isotopic clusters associated with the identified AA sequence. In case of a labeled experiment this is the total intensity of all the isotopic patterns in the label cluster.

Intensity TMT12	Summed up extracted Ion Current (XIC) of all isotopic clusters associated with the identified AA sequence. In case of a labeled experiment this is the total intensity of all the isotopic patterns in the label cluster.
Intensity TMT13	Summed up extracted Ion Current (XIC) of all isotopic clusters associated with the identified AA sequence. In case of a labeled experiment this is the total intensity of all the isotopic patterns in the label cluster.
Intensity TMT1phos1	Summed up eXtracted Ion Current (XIC) of all isotopic clusters associated with the identified AA sequence. In case of a labeled experiment this is the total intensity of all the isotopic patterns in the label cluster.
Intensity TMT1phos2	Summed up eXtracted Ion Current (XIC) of all isotopic clusters associated with the identified AA sequence. In case of a labeled experiment this is the total intensity of all the isotopic patterns in the label cluster.
Intensity TMT1phos3	Summed up eXtracted Ion Current (XIC) of all isotopic clusters associated with the identified AA sequence. In case of a labeled experiment this is the total intensity of all the isotopic patterns in the label cluster.
Intensity TMT21	Summed up extracted lon Current (XIC) of all isotopic clusters associated with the identified AA sequence. In case of a labeled experiment this is the total intensity of all the isotopic patterns in the label cluster.
Intensity TMT22	Summed up eXtracted Ion Current (XIC) of all isotopic clusters associated with the identified AA sequence. In case of a labeled experiment this is the total intensity of all the isotopic patterns in the label cluster.
Intensity TMT23	Summed up eXtracted Ion Current (XIC) of all isotopic clusters associated with the identified AA sequence. In case of a labeled experiment this is the total intensity of all the isotopic patterns in the label cluster.
Intensity TMT2phos1	Summed up eXtracted Ion Current (XIC) of all isotopic clusters associated with the identified AA sequence. In case of a labeled experiment this is the total intensity of all the isotopic patterns in the label cluster.
Intensity TMT2phos2	Summed up eXtracted Ion Current (XIC) of all isotopic clusters associated with the identified AA sequence. In case of a labeled experiment this is the total intensity of all the isotopic patterns in the label cluster.
Intensity TMT2phos3	Summed up eXtracted Ion Current (XIC) of all isotopic clusters associated with the identified AA sequence. In case of a labeled experiment this is the total intensity of all the isotopic patterns in the label cluster.
Occupancy TMT1	
Occupancy ratioTMT1	
Occupancy error scale TMT1	
Occupancy TMT1phos	
Occupancy ratioTMT1phos	
Occupancy error scale TMT1phos	
Occupancy TMT2	
Occupancy ratioTMT2	
Occupancy error scale TMT2	
Occupancy TMT2phos	
Occupancy ratioTMT2phos	
Occupancy error scale TMT2phos	
Reverse	When marked with '+', this particular peptide was found to be part of a protein derived from the reversed part of the protein sequence database. These should be removed for further data analysis.
Potential contaminant	When marked with '+', this particular peptide was found to be part of a commonly occurring contaminant. These should be removed for further data analysis.
id	A unique (consecutive) identifier for each row in the site table, which is used to cross-link the information in this file with the information stored in the other files.
Protein group IDs	The identifier of the protein-group this peptide sequence is associated with, which can be used to look up the extended protein information in the file 'proteinGroups.txt'. As a single peptide can be linked to multiple proteins (e.g. in the case of razor-proteins), multiple id's can be stored here separated by a semicolon. As a protein can be identified by multiple peptides, the same id can be found in different rows.
Positions Position	The positions of the modifications in the protein amino acid sequence.
I OSIUOII	The position of the modification in the protein amino acid sequence.
Peptide IDs	Identifier(s) of the associated peptide sequence(s) summary, which can be found in the file 'peptides.txt'.

Mod. peptide IDs	Identifier(s) of the associated peptide sequence(s) summary, which can be found in the file 'modificationSpecificPeptides.txt'.
Evidence IDs	Identifier(s) for analyzed peptide evidence associated with the protein group referenced against the evidence table.
MS/MS IDs	The identifiers of the MS/MS scans identifying this peptide, referenced against the msms table.
Best localization evidence ID	
Best localization MS/MS ID	
Best localization raw file	
Best localization scan number	
Best score evidence ID	
Best score MS/MS ID	
Best score raw file	
Best score scan number	
Best PEP evidence ID	
Best PEP MS/MS ID	
Best PEP raw file	
Best PEP scan number	

Protein groups

The Protein Groups table contains information on the identified proteins in the processed raw-files. Each single row contains the group of proteins that could be reconstructed from a set of peptides.

Name	Separator	Description
Protein IDs		Identifier(s) of protein(s) contained in the protein group. They are sorted by number of identified peptides in descending order.
Majority protein IDs		These are the IDs of those proteins that have at least half of the peptides that the leading protein has.
Peptide counts (all)		Number of peptides associated with each protein in protein group, occurring in the order as the protein IDs occur in the 'Protein IDs' column. Here distinct peptide sequences are counted. Modified forms or different charges are counted as one peptide.
Peptide counts (razor+unique)		Number of peptides associated with each protein in protein group, occurring in the order as the protein IDs occur in the 'Protein IDs' column. Here distinct peptide sequences are counted. Modified forms or different charges are counted as one peptide.
Peptide counts (unique)		Number of peptides associated with each protein in protein group, occurring in the order as the protein IDs occur in the 'Protein IDs' column. Here distinct peptide sequences are counted. Modified forms or different charges are counted as one peptide.
Fasta headers		Fasta headers(s) of protein(s) contained within the group.
Number of proteins		Number of proteins contained within the group. This corresponds to the number of entries in the column 'Protein IDs'.
Peptides		The total number of peptide sequences associated with the protein group (i.e. for all the proteins in the group).
Razor + unique peptides		The total number of razor + unique peptides associated with the protein group (i.e. these peptides are shared with another protein group).
Unique peptides		The total number of unique peptides associated with the protein group (i.e. these peptides are not shared with another protein group).
Peptides TMT1		Number of peptides (distinct peptide sequences) in experiment TMT1
Peptides TMT1phos		Number of peptides (distinct peptide sequences) in experiment TMT1phos
Peptides TMT2		Number of peptides (distinct peptide sequences) in experiment TMT2
Peptides TMT2phos		Number of peptides (distinct peptide sequences) in experiment TMT2phos
Razor + unique peptides TMT1		Number of razor + unique peptides (distinct peptide sequences) in experiment TMT1
Razor + unique peptides TMT1phos	3	Number of razor + unique peptides (distinct peptide sequences) in experiment TMT1phos
Razor + unique peptides TMT2		Number of razor + unique peptides (distinct peptide sequences) in experiment TMT2
Razor + unique peptides TMT2phos	5	Number of razor + unique peptides (distinct peptide sequences) in experiment TMT2phos
Unique peptides TMT1		Number of unique peptides (distinct peptide sequences) in experiment TMT1
Unique peptides TMT1phos		Number of unique peptides (distinct peptide sequences) in experiment TMT1phos
Unique peptides TMT2		Number of unique peptides (distinct peptide sequences) in experiment TMT2
Unique peptides TMT2phos		Number of unique peptides (distinct peptide sequences) in experiment TMT2phos
Sequence coverage [%]		Percentage of the sequence that is covered by the identified peptides of the best protein sequence contained in the group.
Unique + razor sequence coverage [%]		Percentage of the sequence that is covered by the identified unique and razor peptides of the best protein sequence contained in the group.
Unique sequence coverage [%]		Percentage of the sequence that is covered by the identified unique peptides of the best protein sequence contained in the group.
Mol. weight [kDa]		Molecular weight of the leading protein sequence contained in the protein group.
Sequence length		The length of the leading protein sequence contained in the group.

Sequence lengths	The length of all sequences of the proteins contained in the group.
Q-value	This is the ratio of reverse to forward protein groups.
Score	Protein score which is derived from peptide posterior error probabilities.
Reporter intensity corrected 1	
Reporter intensity corrected 2	
Reporter intensity corrected 3	
Reporter intensity corrected 4	
Reporter intensity corrected 5	
Reporter intensity corrected 6	
Reporter intensity corrected 7	
Reporter intensity corrected 8	
Reporter intensity corrected 9	
Reporter intensity corrected 10	
Reporter intensity 1	
Reporter intensity 2	
Reporter intensity 3	
Reporter intensity 4	
Reporter intensity 5	
Reporter intensity 6	
Reporter intensity 7	
Reporter intensity 8	
Reporter intensity 9	
Reporter intensity 10	
Reporter intensity count 1	
Reporter intensity count 2	
Reporter intensity count 3	
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Reporter intensity count 4	
Reporter intensity count 5	
Reporter intensity count 6	
Reporter intensity count 7	
Reporter intensity count 8	
Reporter intensity count 9	
Reporter intensity count 10	
Reporter intensity corrected 1 TMT1	
Reporter intensity corrected 2 TMT1	
Reporter intensity corrected 3 TMT1	
Reporter intensity corrected 4 TMT1	
Reporter intensity corrected 5 TMT1	
Reporter intensity corrected 6 TMT1	
Reporter intensity corrected 7 TMT1	
Reporter intensity corrected 8 TMT1	
Reporter intensity corrected 9 TMT1	
Reporter intensity corrected 10 TMT1	
Reporter intensity corrected 1 TMT1phos	
Reporter intensity corrected 2 TMT1phos	
Reporter intensity corrected 3	
TMT1phos Reporter intensity corrected 4	
TMT1phos Reporter intensity corrected 5 TMT1phos	
Reporter intensity corrected 6	
TMT1phos Reporter intensity corrected 7	
TMT1phos Reporter intensity corrected 8 TMT1phos	
TMT1phos Reporter intensity corrected 9	
TMT1phos Reporter intensity corrected 10	
TMT1phos Reporter intensity corrected 1 TMT2	
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Reporter intensity corrected 3 TMT2	
Reporter intensity corrected 4 TMT2	
Reporter intensity corrected 5 TMT2	
Reporter intensity corrected 6 TMT2	
Reporter intensity corrected 7 TMT2	
Reporter intensity corrected 8 TMT2	
Reporter intensity corrected 9 TMT2	
Reporter intensity corrected 10	
TMT2	
Reporter intensity corrected 1 TMT2phos	
Reporter intensity corrected 2 TMT2phos	
Reporter intensity corrected 3 TMT2phos	
Reporter intensity corrected 4 TMT2phos	
Reporter intensity corrected 5 TMT2phos	
Reporter intensity corrected 6 TMT2phos	
Reporter intensity corrected 7 TMT2phos	
Reporter intensity corrected 8 TMT2phos	
Reporter intensity corrected 9 TMT2phos	
Reporter intensity corrected 10 TMT2phos	
Reporter intensity 1 TMT1	
Reporter intensity 2 TMT1	
Reporter intensity 3 TMT1	
Reporter intensity 4 TMT1	
Reporter intensity 4 TMT1	
Reporter intensity 6 TMT1	
Reporter intensity 7 TMT1	
Reporter intensity 8 TMT1	
Reporter intensity 9 TMT1	
Reporter intensity 10 TMT1	
Reporter intensity 1 TMT1phos	
Reporter intensity 2 TMT1phos	
Reporter intensity 3 TMT1phos	
Reporter intensity 4 TMT1phos	
Reporter intensity 5 TMT1phos	
Reporter intensity 6 TMT1phos	
Reporter intensity 7 TMT1phos	
Reporter intensity 8 TMT1phos	
Reporter intensity 9 TMT1phos	
Reporter intensity 10 TMT1phos	
Reporter intensity 1 TMT2	
Reporter intensity 2 TMT2	
Reporter intensity 3 TMT2	
Reporter intensity 4 TMT2	
Reporter intensity 5 TMT2	
Reporter intensity 6 TMT2	
Reporter intensity 7 TMT2	
Reporter intensity 8 TMT2	
Reporter intensity 9 TMT2	
Reporter intensity 10 TMT2	
Reporter intensity 1 TMT2phos	
Reporter intensity 2 TMT2phos	
Reporter intensity 3 TMT2phos	
Reporter intensity 4 TMT2phos	
Reporter intensity 5 TMT2phos	
Reporter intensity 6 TMT2phos	
Reporter intensity 7 TMT2phos	
Reporter intensity 8 TMT2phos	

Reporter intensity 9 TMT2phos	
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Reporter intensity 10 TMT2phos	
Reporter intensity count 1 TMT1	
Reporter intensity count 2 TMT1	
Reporter intensity count 3 TMT1	
Reporter intensity count 4 TMT1	
Reporter intensity count 5 TMT1	
Reporter intensity count 6 TMT1	
Reporter intensity count 7 TMT1	
Reporter intensity count 8 TMT1	
Reporter intensity count 9 TMT1	
Reporter intensity count 10 TMT1	
Reporter intensity count 1 TMT1phos	
Reporter intensity count 2 TMT1phos	
Reporter intensity count 3 TMT1phos	
Reporter intensity count 4 TMT1phos	
Reporter intensity count 5 TMT1phos	
Reporter intensity count 6 TMT1phos	
Reporter intensity count 7 TMT1phos	
Reporter intensity count 8 TMT1phos	
Reporter intensity count 9 TMT1phos	
Reporter intensity count 10 TMT1phos	
Reporter intensity count 1 TMT2	
Reporter intensity count 2 TMT2	
Reporter intensity count 3 TMT2	
Reporter intensity count 4 TMT2	
Reporter intensity count 5 TMT2	
Reporter intensity count 6 TMT2	
Reporter intensity count 7 TMT2	
Reporter intensity count 8 TMT2	
Reporter intensity count 9 TMT2	
Reporter intensity count 10 TMT2	
Reporter intensity count 1 TMT2phos	
Reporter intensity count 2 TMT2phos	
Reporter intensity count 3 TMT2phos	
Reporter intensity count 4 TMT2phos	
Reporter intensity count 5 TMT2phos	
Reporter intensity count 6 TMT2phos	
Reporter intensity count 7 TMT2phos	
Reporter intensity count 8 TMT2phos	
Reporter intensity count 9 TMT2phos	
Reporter intensity count 10 TMT2phos	
Sequence coverage TMT1 [%]	Percentage of the sequence that is covered by the identified peptides in this sample of the longest protein sequence contained within the group.
Sequence coverage TMT1phos [%]	Percentage of the sequence that is covered by the identified peptides in this sample of the longest protein sequence contained within the group.
Sequence coverage TMT2 [%]	Percentage of the sequence that is covered by the identified peptides in this sample of the longest protein sequence contained within the group.

Sequence coverage TMT2phos [%]	Percentage of the sequence that is covered by the identified peptides in this sample of the longest protein sequence contained within the group.
Intensity	Summed up eXtracted Ion Current (XIC) of all isotopic clusters associated with the identified AA sequence. In case of a labeled experiment this is the total intensity of all the isotopic patterns in the label cluster.
Intensity TMT1	Summed up eXtracted Ion Current (XIC) of all isotopic clusters associated with the identified AA sequence. In case of a labeled experiment this is the total intensity of all the isotopic patterns in the label cluster.
Intensity TMT1phos	Summed up eXtracted Ion Current (XIC) of all isotopic clusters associated with the identified AA sequence. In case of a labeled experiment this is the total intensity of all the isotopic patterns in the label cluster.
Intensity TMT2	Summed up eXtracted Ion Current (XIC) of all isotopic clusters associated with the identified AA sequence. In case of a labeled experiment this is the total intensity of all the isotopic patterns in the label cluster.
Intensity TMT2phos	Summed up eXtracted Ion Current (XIC) of all isotopic clusters associated with the identified AA sequence. In case of a labeled experiment this is the total intensity of all the isotopic patterns in the label cluster.
MS/MS count	
Only identified by site	When marked with '+', this particular protein group was identified only by a modification site.
Reverse	When marked with '+', this particular protein group contains no protein, made up of at least 50% of the peptides of the leading protein, with a peptide derived from the reversed part of the decoy database. These should be removed for further data analysis. The 50% rule is in place to prevent spurious protein hits to erroneously flag the protein group as reverse.
Potential contaminant	When marked with '+', this particular protein group was found to be a commonly occurring contaminant. These should be removed for further data analysis.
id	A unique (consecutive) identifier for each row in the proteinGroups table, which is used to cross-link the information in this file with the information stored in the other files.
Peptide IDs	Identifier(s) of the associated peptide sequence(s) summary, which can be found in the file 'peptides.txt'.
Peptide is razor	Indicates for each peptide ID if it is a razor or group unique peptide (true) or a non unique non razor peptide (false).
Mod. peptide IDs	
Evidence IDs	
MS/MS IDs	
Best MS/MS	The identifier of the best (in terms of quality) MS/MS scans identifying the peptides of this protein, referenced against the msms table.
Oxidation (M) site IDs	Identifier(s) for site(s) associated with the protein group, which show(s) evidence of the modification, referenced against the appropriate modification site file.
Phospho (STY) site IDs	Identifier(s) for site(s) associated with the protein group, which show(s) evidence of the modification, referenced against the appropriate modification site file.
Oxidation (M) site positions	Positions of the sites in the leading protein of this group.
Phospho (STY) site positions	Positions of the sites in the leading protein of this group.

All peptides

Name	Separator	Description
Raw file		Name of the raw file the spectral data was extracted from.
Туре		The type of detection for the peptide. MULTI – A labeling multiplet was detected. ISO – An isotope pattern was detected.
Charge		The charge state of the peptide.
m/z		The mass divided by the charge of the charged peptide.
Mass		The mass of the neutral peptide ((m/z-proton) * charge).
Uncalibrated m/z		m/z before recalibrations have been applied.
Resolution		The resolution of the peak detected for the peptide measured in Full Width at Half Maximum (FWHM).
Number of data points		The number of data points (peak centroids) collected for this peptide feature.
Number of scans		The number of MS scans that the 3d peaks of this peptide feature are overlapping with.
Number of isotopic peaks		The number of isotopic peaks contained in this peptide feature.
PIF		Short for Parent Ion Fraction; indicates the fraction the target peak makes up of the total intensity in the inclusion window.
Mass fractional part		The values after the radix point (ie value - floor(value)).
Mass deficit		Empirically derived deviation measure to the next nearest integer scaled to center around 0. Can be used to visually detect contaminants in a plot setting Mass against this value. m*a+b - round(m*a+b) m: the peptide mass a: 0.99954
Mass precision [ppm]		b: -0.04 The precision of the mass detection of the peptide in parts-permillion.
Max intensity m/z 0		Summed up extracted Ion Current (XIC) of all isotopic clusters associated with the identified AA sequence. In case of a labeled experiment this is the total intensity of all the isotopic patterns in the label cluster.
Retention time		The retention time of the peak detected for the peptide measured in minutes.
Retention length		The total retention time width of the peak (last timepoint – first timepoint) in seconds.
Retention length (FWHM)		The full width at half maximum value retention time width of the peak in seconds.
Min scan number		The first scan number at which the peak was encountered.
Max scan number		The last scan number at which the peak was encountered.
Identified		When marked with '+' this particular MS/MS scan was identified as a peptide; when marked with '-' no identification was made.
MS/MS IDs		Unique identifier linking this identification to the MS/MS scans.
Sequence		The identified AA sequence of the peptide.
Length		The length of the sequence stored in the column "Sequence".
Modifications		Post-translational modifications contained within the sequence. When no modifications exist, this is set to 'unmodified'.
Madron de anno ano		Note: This column only set when this MS/MS spectrum has been identified.
Modified sequence		Sequence representation of the peptide including location(s) of modified AAs. Note: This column only set when this MS/MS spectrum has
Proteins		been identified. Identifiers of proteins this peptide is associated with.
		Note: This column only set when this MS/MS spectrum has been identified.
Score		The score of the identification (higher is better).
		Note: This column only set when this MS/MS spectrum has been identified.
Intensity		Summed up eXtracted Ion Current (XIC) of all isotopic clusters associated with the identified AA sequence. In case of a labeled experiment this is the total intensity of all the isotopic patterns in the label cluster.
Intensities		Elution profile.
Isotope pattern		Isotope pattern.

MS/MS Count	The number of MS/MS spectra recorded for the peptide.
MSMS Scan Numbers	The scan numbers where the MS/MS spectra were recorded.
MSMS Isotope Indices	Indices of the isotopic peaks that the MS/MS spectra reside on. A value of 0 corresponds to the monoisotopic peak.

MS scans

The msScans table contains information about the full scans, which can be used to verify data quality and generated useful statistics about the interaction between the samples and LC.

Name	Separator	Description
Raw file		The name of the RAW-file the mass spectral data originates from.
Scan number		The scan number (defined in the raw-file) at which the full scan was made.
Scan index		The consecutive index of this full scan.
Retention time		The retention time at which the full scan was made.
Cycle time		The total time (full scan including the tandem MS scans) this full scan has taken up.
Ion injection time		The total injection time that was required to capture the specified amount of ions. This value is limited by a maximum, which can be used to determine whether the time has maxed out (indicative of a bad acquisition).
Base peak intensity		The intensity of the most intense ion in the spectrum.
Total ion current		The total intensity acquired in the full scan.
MS/MS count		The number of tandem MS scans that were made based on this full scan (e.g. a top 10 method selects the top 10 most intense ions in the scan and fragments those).
Mass calibration		The applied mass correction in Th to the full scan.
Experiment		
Peak length		The average time between the start and the end of the peaks detected in the full scan.
Isotope pattern length		The average time between the start and the end of the isotope patterns detected in the full scan.
Multiplet length		The average time between the start and the end of the isotope patterns of the labeling multiplets detected in the full scan.
Peaks / s		The average number of peaks detected per second of chromatography.
Single peaks / s		The average number of single peaks detected per second of chromatography.
Isotope patterns / s		The average number of isotope patterns detected per second of chromatography.
Single isotope patterns / s		The average number of single isotope patterns detected per of second chromatography.
Multiplets / s		The average number of labeling multiplets detected per of second chromatography.
Identified multiplets / s		The percentage of labeling multiplets actually identified.
Multiplet identification rate [%]		The percentage of the detected labeling multiplets that were identified.
Intens Comp Factor		Taken from the Thermo RAW file.
CTCD Comp		Taken from the Thermo RAW file.
RawOvFtT		For Thermo Fisher only. TIC estimation done with the orbitrap cell.
AGC Fill		Taken from the Thermo RAW file.

MZ range

Name	Separator	Description
Raw file		The name of the RAW-file the mass spectral data was derived from.
m/z		The mass-over-charge value.
Peaks / Da		The average number of peaks detected per Dalton.
Single peaks / Da		The average number of single peaks detected per Dalton.
Isotope patterns / Da		The average number of isotope patterns detected per Dalton.
Single isotope patterns / Da		The average number of single isotope patterns detected per Dalton.
SILAC pairs / Da		The average number of SILAC pairs detected per Dalton.
Identified SILAC pairs / Da		The percentage of SILAC pairs actually identified.
SILAC identification rate [%]		The percentage of the detected SILAC pairs that were identified.

MS/MS scans

Name	Separator	Description
Raw file		Name of the RAW file the spectral MS/MS data was extracted
		from.
Scan number		RAW file derived scan number for the MS/MS spectrum.
Retention time		Time point along the elution profile at which the MS/MS data was recorded.
Ion injection time		The ion inject time for the MS/MS scan. This can be used to determine if this time equals to the maximum ion inject time, general indicative of a lower quality spectrum.
Total ion current		The total ion current of the MS/MS scan. For Thermo data this value is calculated by summing all the intensity values found in the mass spectral data, which is different from the Xcalibur reported TIC (Xcalibur TIC is about 25% of the value reported here).
Collision energy		The collision energy used for the fragmentation that resulted in this MS/MS scan.
Summations		For time of flight instruments only.
Base peak intensity		The intensity of the most intense ion in the spectrum.
Elapsed time		The time the MS/MS scan took to complete.
Identified		When marked with '+' this particular MS/MS scan was identified as a peptide; when marked with '-' no identification was made.
MS/MS IDs		Unique identifier linking this identification to the MS/MS scans.
Sequence		The identified AA sequence of the peptide.
Length		The length of the sequence stored in the column "Sequence".
Filtered peaks		Number of peaks after the 'top X per 100 Da' filtering.
m/z		Recalibrated m/z of the precursor ion.
Mass		Charge corrected mass of the precursor ion.
Charge		Charge state of the precursor ion.
Туре		The type of precursor ion as identified by MaxQuant. ISO – isotopic cluster. PEAK – single peak. MULTI – labeling cluster.
Fragmentation		The type of fragmentation used to create the MS/MS spectrum. CID – Collision Induced Dissociation. HCD – High energy Collision induced Dissociation. ETD – Electron Transfer Dissociation.
Mass analyzer		The mass analyzer used to record the MS/MS spectrum. ITMS – Ion trap. FTMS – Fourier transform ICR or orbitrap cell. TOF – Time of flight.
Parent intensity fraction		The percentage the parent ion intensity makes up of the total intensity in the selection window.
Fraction of total spectrum		The percentage the parent ion intensity makes up of the total intensity of the whole MS spectrum.
Base peak fraction		The percentage the parent ion intensity in comparison to the highest peak in he MS spectrum.
Precursor full scan number		The full scan number where the precursor ion was selected for fragmentation.
Precursor intensity		The intensity of the precursor ion at the scannumber it was selected.
Precursor apex fraction		The fraction the intensity of the precursor ion makes up of the peak (apex) intensity.
Precursor apex offset		How many full scans the precursor ion is offset from the peak (apex) position.
Precursor apex offset time		How much time the precursor ion is offset from the peak (apex) position.
Scan event number		This number indicates which MS/MS scan this one is in the consecutive order of the MS/MS scans that are acquired after an MS scan.
Modifications		Post-translational modifications contained within the sequence. When no modifications exist, this is set to 'unmodified'.
		Note: This column only set when this MS/MS spectrum has been identified.
Modified sequence		Sequence representation of the peptide including location(s) of modified AAs.
		Note: This column only set when this MS/MS spectrum has been identified.

Proteins	Identifiers of proteins this peptide is associated with.
	Note: This column only set when this MS/MS spectrum has been identified.
Score	The score of the identification (higher is better).
	Note: This column only set when this MS/MS spectrum has been identified.
Experiment	
Reporter intensity corrected 1	
Reporter intensity corrected 2	
Reporter intensity corrected 3	
Reporter intensity corrected 4	
Reporter intensity corrected 5	
Reporter intensity corrected 6	
Reporter intensity corrected 7	
Reporter intensity corrected 8	
Reporter intensity corrected 9	
Reporter intensity corrected 10	
Reporter intensity 1	
Reporter intensity 2	
Reporter intensity 3	
Reporter intensity 4	
Reporter intensity 5	
Reporter intensity 6	
Reporter intensity 7	
Reporter intensity 8	
Reporter intensity 9	
Reporter intensity 10	
Reporter PIF	
Reporter fraction	
Intens Comp Factor	Taken from the Thermo RAW file.
CTCD Comp	Taken from the Thermo RAW file.
RawOvFtT	For Thermo Fisher only. TIC estimation done with the orbitrap cell.
AGC Fill	Taken from the Thermo RAW file.
Scan index	Consecutive index of the MS/MS spectrum.
MS scan index	Consecutive index of the MS spectrum prior to this MS/MS spectrum.
MS scan number	Scan number of the MS spectrum prior to this MS/MS spectrum.

MS/MS

Name	Separator	Description
Raw file		The name of the RAW file the mass spectral data was read from.
Scan number		The RAW-file derived scan number of the MS/MS spectrum.
Scan index		The consecutive index of the MS/MS spectrum.
Sequence		The identified AA sequence of the peptide.
Length		The length of the sequence stored in the column "Sequence".
Missed cleavages		Number of missed enzymatic cleavages.
Modifications		Post-translational modifications contained within the identified peptide sequence.
Modified sequence		Sequence representation including the post-translational modifications (abbreviation of the modification in brackets before the modified AA). The sequence is always surrounded by underscore characters ('_').
Oxidation (M) Probabilities		Sequence representation of the peptide including PTM positioning probabilities ([01], where 1 is best match) for 'Oxidation (M)'.
Phospho (STY) Probabilities		Sequence representation of the peptide including PTM positioning probabilities ([01], where 1 is best match) for 'Phospho (STY)'.
Oxidation (M) Score Diffs		
Phospho (STY) Score Diffs		
Acetyl (Protein N-term)		
Oxidation (M)		
Phospho (STY)		
Proteins		The identifiers of the proteins the identified peptide is associated with.
Charge		The charge state of the precursor ion.
Fragmentation		The type of fragmentation used to create the MS/MS spectrum. CID – Collision Induced Dissociation. HCD – High energy Collision induced Dissociation. ETD – Electron Transfer Dissociation.
Mass analyzer		The mass analyzer used to record the MS/MS spectrum. ITMS – Ion trap. FTMS – Fourier transform ICR or orbitrap cell. TOF – Time of flight.
Туре		The type of precursor ion as identified by MaxQuant. ISO – isotopic cluster. PEAK – single peak. MULTI – labeling cluster.
Scan event number		
Isotope index		
m/z		The mass-over-charge of the precursor ion.
Mass		The charge corrected mass of the precursor ion.
Mass error [ppm]		Mass error of the recalibrated mass-over-charge value of the precursor ion in comparison to the predicted monoisotopic mass of the identified peptide sequence expressed in parts per million.
Mass error [Da]		Mass error of the recalibrated mass-over-charge value of the precursor ion in comparison to the predicted monoisotopic mass of the identified peptide sequence expressed in atomic mass units.
Simple mass error [ppm]		
Retention time		The uncalibrated retention time in minutes where the MS/MS spectrum has been acquired.
PEP		Posterior Error Probability of the identification. This value essentially operates as a p-value, where smaller is more significant.
Score		Andromeda score for the best associated MS/MS spectrum.
Delta score		Score difference to the second best identified peptide with a different amino acid sequence.
Score diff		Score difference to the second best positioning of modifications identified peptide with the same amino acid sequence.
Localization prob		
Combinatorics		Number of possible distributions of the modifications over the peptide sequence.
PIF		Short for Parent Ion Fraction; indicates the fraction the target peak makes up of the total intensity in the inclusion window.

Fraction of total spectrum	The percentage the parent ion intensity makes up of the total
Base peak fraction	intensity of the whole spectrum. The percentage the parent ion intensity in comparison to the
Precursor Full ScanNumber	highest peak in he MS spectrum. The full scannumber where the precursor ion was selected for
	fragmentation.
Precursor Intensity	The intensity of the precursor ion at the scannumber it was selected.
Precursor Apex Fraction	The fraction the intensity of the precursor ion makes up of the peak (apex) intensity.
Precursor Apex Offset	How many full scans the precursor ion is offset from the peak (apex) position.
Precursor Apex Offset Time	How much time the precursor ion is offset from the peak (apex) position.
Diagnostic peak Phospho (STY) Y	
Matches	The species of the peaks in the fragmentation spectrum after TopN filtering.
Intensities	The intensities of the peaks in the fragmentation spectrum after TopN filtering.
Mass Deviations [Da]	The mass deviation of each peak in the fragmentation spectrum in absolute mass units.
Mass Deviations [ppm]	The mass deviation of each peak in the fragmentation spectrum in parts per million.
Masses	The masses-over-charge of the peaks in the fragmentation spectrum.
Number of Matches	The number of peaks matching to the predicted fragmentation
Intensity coverage	spectrum. The fraction of intensity in the MS/MS spectrum that is
Peak coverage	annotated. The fraction of peaks in the MS/MS spectrum that are
Neutral loss level	annotated. How many neutral losses were applied to each fragment in the
ETD identification type	Andromeda scoring. For ETD spectra several different combinations of ion series
Reverse	are scored. Here the highest scoring combination is indicated When marked with '+', this particular peptide was found to be
Reverse	part of a protein derived from the reversed part of the decoy database. These should be removed for further data analysis.
All scores	
All sequences	
All modified sequences	
Reporter intensity corrected 1	
Reporter intensity corrected 2	
Reporter intensity corrected 3	
Reporter intensity corrected 4	
Reporter intensity corrected 5	
Reporter intensity corrected 6	
Reporter intensity corrected 6 Reporter intensity corrected 7	
Reporter intensity corrected 6 Reporter intensity corrected 7 Reporter intensity corrected 8	
Reporter intensity corrected 6 Reporter intensity corrected 7 Reporter intensity corrected 8 Reporter intensity corrected 9	
Reporter intensity corrected 6 Reporter intensity corrected 7 Reporter intensity corrected 8 Reporter intensity corrected 9 Reporter intensity corrected 10	
Reporter intensity corrected 6 Reporter intensity corrected 7 Reporter intensity corrected 8 Reporter intensity corrected 9 Reporter intensity corrected 10 Reporter intensity 1	
Reporter intensity corrected 6 Reporter intensity corrected 7 Reporter intensity corrected 8 Reporter intensity corrected 9 Reporter intensity corrected 10 Reporter intensity 1 Reporter intensity 2	
Reporter intensity corrected 6 Reporter intensity corrected 7 Reporter intensity corrected 8 Reporter intensity corrected 9 Reporter intensity corrected 10 Reporter intensity 1 Reporter intensity 2 Reporter intensity 3	
Reporter intensity corrected 6 Reporter intensity corrected 7 Reporter intensity corrected 8 Reporter intensity corrected 9 Reporter intensity corrected 10 Reporter intensity 1 Reporter intensity 2 Reporter intensity 3 Reporter intensity 4	
Reporter intensity corrected 6 Reporter intensity corrected 7 Reporter intensity corrected 8 Reporter intensity corrected 9 Reporter intensity corrected 10 Reporter intensity 1 Reporter intensity 2 Reporter intensity 3 Reporter intensity 4 Reporter intensity 5	
Reporter intensity corrected 6 Reporter intensity corrected 7 Reporter intensity corrected 8 Reporter intensity corrected 9 Reporter intensity corrected 10 Reporter intensity 1 Reporter intensity 2 Reporter intensity 3 Reporter intensity 4 Reporter intensity 5 Reporter intensity 6	
Reporter intensity corrected 6 Reporter intensity corrected 7 Reporter intensity corrected 8 Reporter intensity corrected 9 Reporter intensity corrected 10 Reporter intensity 1 Reporter intensity 2 Reporter intensity 3 Reporter intensity 4 Reporter intensity 5 Reporter intensity 6 Reporter intensity 7	
Reporter intensity corrected 6 Reporter intensity corrected 7 Reporter intensity corrected 8 Reporter intensity corrected 9 Reporter intensity corrected 10 Reporter intensity 1 Reporter intensity 2 Reporter intensity 3 Reporter intensity 4 Reporter intensity 5 Reporter intensity 6 Reporter intensity 7 Reporter intensity 8	
Reporter intensity corrected 6 Reporter intensity corrected 7 Reporter intensity corrected 8 Reporter intensity corrected 9 Reporter intensity corrected 10 Reporter intensity 1 Reporter intensity 2 Reporter intensity 3 Reporter intensity 4 Reporter intensity 5 Reporter intensity 5 Reporter intensity 6 Reporter intensity 7 Reporter intensity 8 Reporter intensity 9	
Reporter intensity corrected 6 Reporter intensity corrected 7 Reporter intensity corrected 8 Reporter intensity corrected 9 Reporter intensity corrected 10 Reporter intensity 1 Reporter intensity 2 Reporter intensity 3 Reporter intensity 4 Reporter intensity 5 Reporter intensity 5 Reporter intensity 6 Reporter intensity 7 Reporter intensity 8 Reporter intensity 9 Reporter intensity 10	
Reporter intensity corrected 6 Reporter intensity corrected 7 Reporter intensity corrected 8 Reporter intensity corrected 9 Reporter intensity corrected 10 Reporter intensity 1 Reporter intensity 2 Reporter intensity 3 Reporter intensity 4 Reporter intensity 5 Reporter intensity 5 Reporter intensity 7 Reporter intensity 7 Reporter intensity 8 Reporter intensity 9 Reporter intensity 10 Reporter mass deviation [mDa] 1	
Reporter intensity corrected 6 Reporter intensity corrected 7 Reporter intensity corrected 8 Reporter intensity corrected 9 Reporter intensity corrected 10 Reporter intensity 1 Reporter intensity 2 Reporter intensity 3 Reporter intensity 4 Reporter intensity 5 Reporter intensity 5 Reporter intensity 7 Reporter intensity 7 Reporter intensity 8 Reporter intensity 9 Reporter intensity 10 Reporter mass deviation [mDa] 1 Reporter mass deviation [mDa] 2	
Reporter intensity corrected 6 Reporter intensity corrected 7 Reporter intensity corrected 8 Reporter intensity corrected 9 Reporter intensity corrected 10 Reporter intensity 1 Reporter intensity 2 Reporter intensity 3 Reporter intensity 4 Reporter intensity 5 Reporter intensity 5 Reporter intensity 7 Reporter intensity 8 Reporter intensity 9 Reporter intensity 10 Reporter mass deviation [mDa] 1 Reporter mass deviation [mDa] 3	
Reporter intensity corrected 6 Reporter intensity corrected 7 Reporter intensity corrected 8 Reporter intensity corrected 9 Reporter intensity corrected 10 Reporter intensity 1 Reporter intensity 2 Reporter intensity 3 Reporter intensity 4 Reporter intensity 5 Reporter intensity 5 Reporter intensity 7 Reporter intensity 8 Reporter intensity 9 Reporter intensity 10 Reporter mass deviation [mDa] 1 Reporter mass deviation [mDa] 3 Reporter mass deviation [mDa] 4	
Reporter intensity corrected 6 Reporter intensity corrected 7 Reporter intensity corrected 8 Reporter intensity corrected 9 Reporter intensity corrected 10 Reporter intensity 1 Reporter intensity 2 Reporter intensity 3 Reporter intensity 4 Reporter intensity 5 Reporter intensity 5 Reporter intensity 7 Reporter intensity 8 Reporter intensity 9 Reporter intensity 10 Reporter mass deviation [mDa] 1 Reporter mass deviation [mDa] 3 Reporter mass deviation [mDa] 4 Reporter mass deviation [mDa] 5	
Reporter intensity corrected 6 Reporter intensity corrected 7 Reporter intensity corrected 8 Reporter intensity corrected 9 Reporter intensity corrected 10 Reporter intensity 1 Reporter intensity 2 Reporter intensity 3 Reporter intensity 4 Reporter intensity 5 Reporter intensity 5 Reporter intensity 7 Reporter intensity 7 Reporter intensity 9 Reporter intensity 10 Reporter mass deviation [mDa] 1 Reporter mass deviation [mDa] 3 Reporter mass deviation [mDa] 4 Reporter mass deviation [mDa] 5 Reporter mass deviation [mDa] 6	
Reporter intensity corrected 6 Reporter intensity corrected 7 Reporter intensity corrected 8 Reporter intensity corrected 9 Reporter intensity corrected 10 Reporter intensity 1 Reporter intensity 2 Reporter intensity 3 Reporter intensity 4 Reporter intensity 5 Reporter intensity 5 Reporter intensity 7 Reporter intensity 7 Reporter intensity 8 Reporter intensity 9 Reporter intensity 10 Reporter mass deviation [mDa] 1 Reporter mass deviation [mDa] 2 Reporter mass deviation [mDa] 3 Reporter mass deviation [mDa] 4 Reporter mass deviation [mDa] 5 Reporter mass deviation [mDa] 6 Reporter mass deviation [mDa] 7	
Reporter intensity corrected 6 Reporter intensity corrected 7 Reporter intensity corrected 8 Reporter intensity corrected 9 Reporter intensity corrected 10 Reporter intensity 1 Reporter intensity 2 Reporter intensity 3 Reporter intensity 4 Reporter intensity 5 Reporter intensity 5 Reporter intensity 7 Reporter intensity 7 Reporter intensity 9 Reporter intensity 10 Reporter mass deviation [mDa] 1 Reporter mass deviation [mDa] 3 Reporter mass deviation [mDa] 4 Reporter mass deviation [mDa] 5 Reporter mass deviation [mDa] 6	

Reporter mass deviation [mDa] 10	
Reporter PIF	
Reporter fraction	
id	A unique (consecutive) identifier for each row in the msms table, which is used to cross-link the information in this file with the information stored in the other files.
Protein group IDs	The identifier of the protein-group this redundant peptide sequence is associated with, which can be used to look up the extended protein information in the file 'proteinGroups.txt'. As a single peptide can be linked to multiple proteins (e.g. in the case of razor-proteins), multiple id's can be stored here separated by a semicolon. As a protein can be identified by multiple peptides, the same id can be found in different rows.
Peptide ID	The identifier of the non-redundant peptide sequence.
Mod. peptide ID	Identifier of the associated modification summary stored in the file 'modificationSpecificPeptides.txt'.
Evidence ID	Identifier of the associated evidence stored in the file 'evidence.txt'.
Oxidation (M) site IDs	Identifier of the oxidation summary stored in the file 'Oxidation (M)Sites.txt'.
Phospho (STY) site IDs	Identifier of the oxidation summary stored in the file 'Phospho (STY)Sites.txt'.