A microscopic image showing several cells against a blue background. The cells are circular with a textured surface, resembling water droplets. Each cell contains a central purple sphere, likely representing the nucleus. The cells are scattered across the frame, with some in sharp focus and others blurred in the background.

# Analysis of a single-cell RNA-seq dataset

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San Francisco

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## GENERAL INFORMATION

- N = **28,700** cells from **4** murine mammary tumors.
- Information on the expression of > 20,000 genes per cell.
- Single cells were dissociated from mouse mammary gland tumors, immuno-stained and **sorted on the CD45+ marker**.
- In 2 samples, single cells were dissociated with **enzymatic digestion** and in 2 samples with the **SimpleFlowTM System**.
- Single cell RNA-seq was performed with 10xGenomics.



## THE GOALS



Prepare a pipeline to analyze single-cell RNA-seq data for current and future use in our lab



Test whether there are differences between samples dissociated with distinct methodologies  
(enzymatic digestion versus SimpleFlow)

# INDEX

1. Quality control & cell filtering
2. Normalization & dimensionality reduction
3. Clustering & artifact control
4. Marker selection
5. Cell type annotation
6. Coloring by genes
7. Comparison of the dissociation methods
8. The take aways



# 01

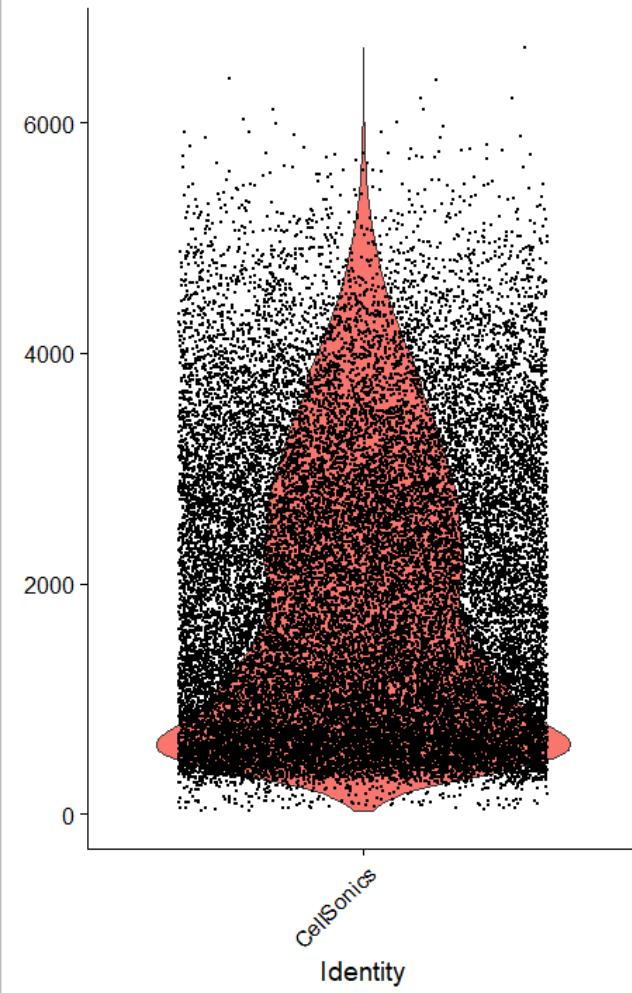
## Quality control & cell filtering



1. Import to R the output of 10X cell ranger (filtered\_feature\_bc\_matrix)
2. Create a **Seurat** object
3. Calculate quality control metrics:
  - nCount RNA (UMI reads/cell)
  - nFeature RNA (number of detected genes/cell)
  - % mitochondrial genes
  - % ribosomal genes

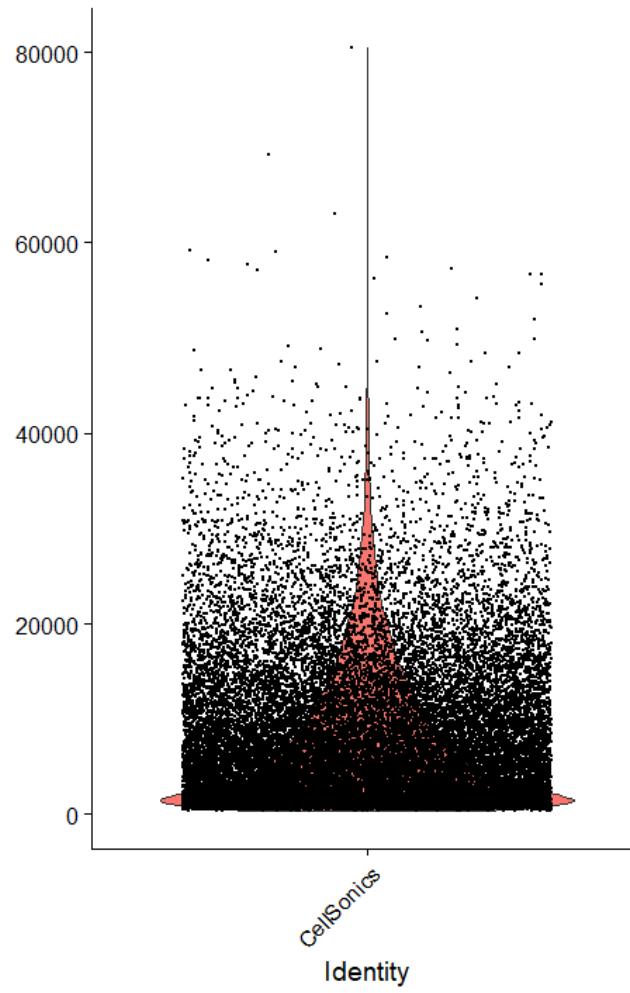
(# detected genes/cell)

nFeature\_RNA

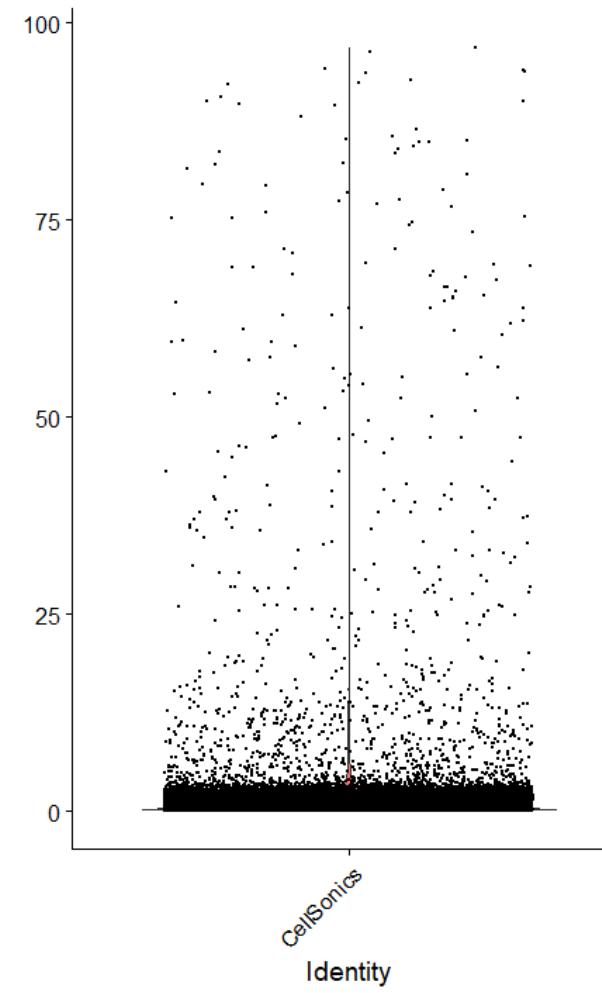


(UMI reads/cell)

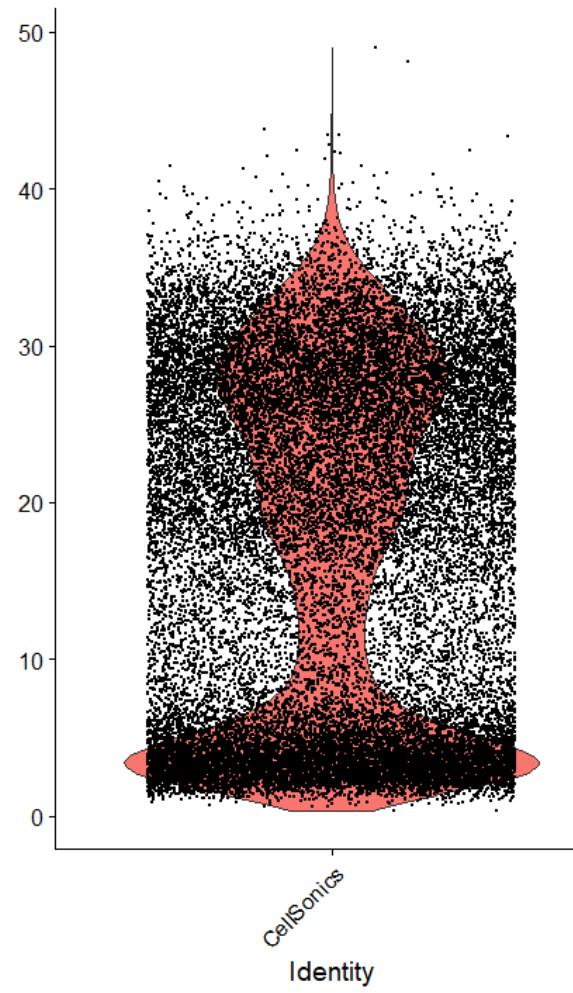
nCount\_RNA



percent.mt



percent.rb



4. Based on that, filter cells to remove dead cells / empty droplets / doublets

```
> table(srat[['QC']]) #Most cells (26.0007) pass the quality control.
```

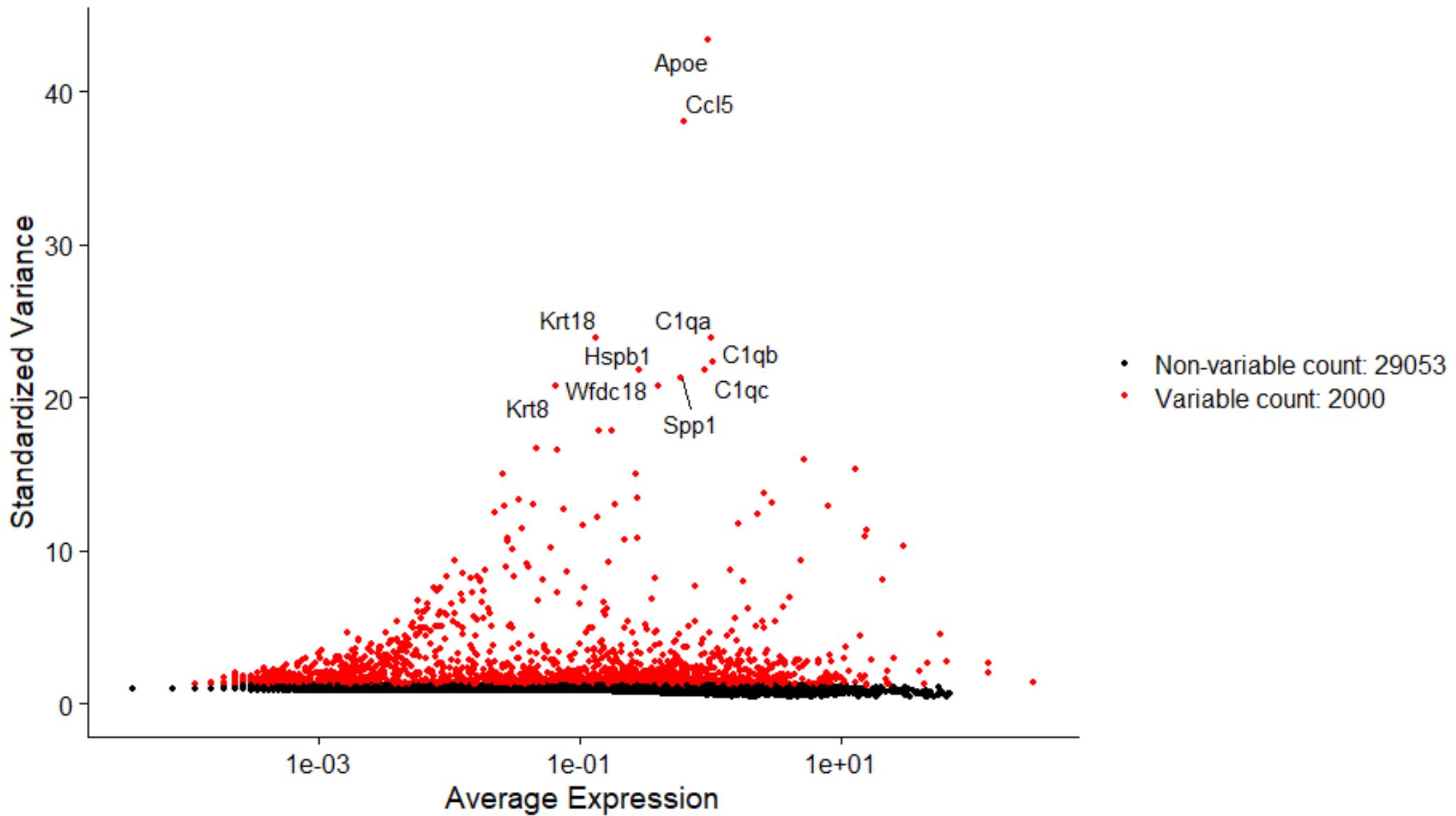
High_MT	High_MT,Low_nFeature	Pass
78	2615	26007

# 02

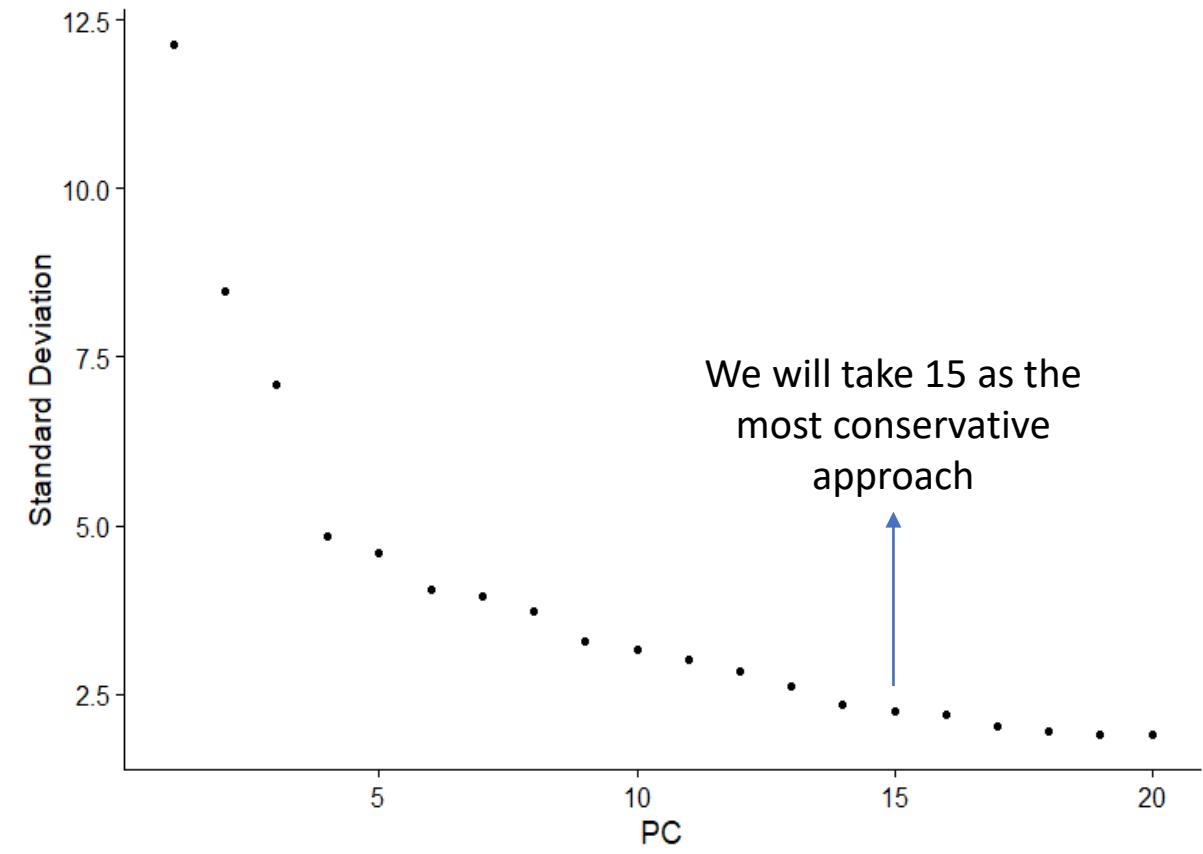
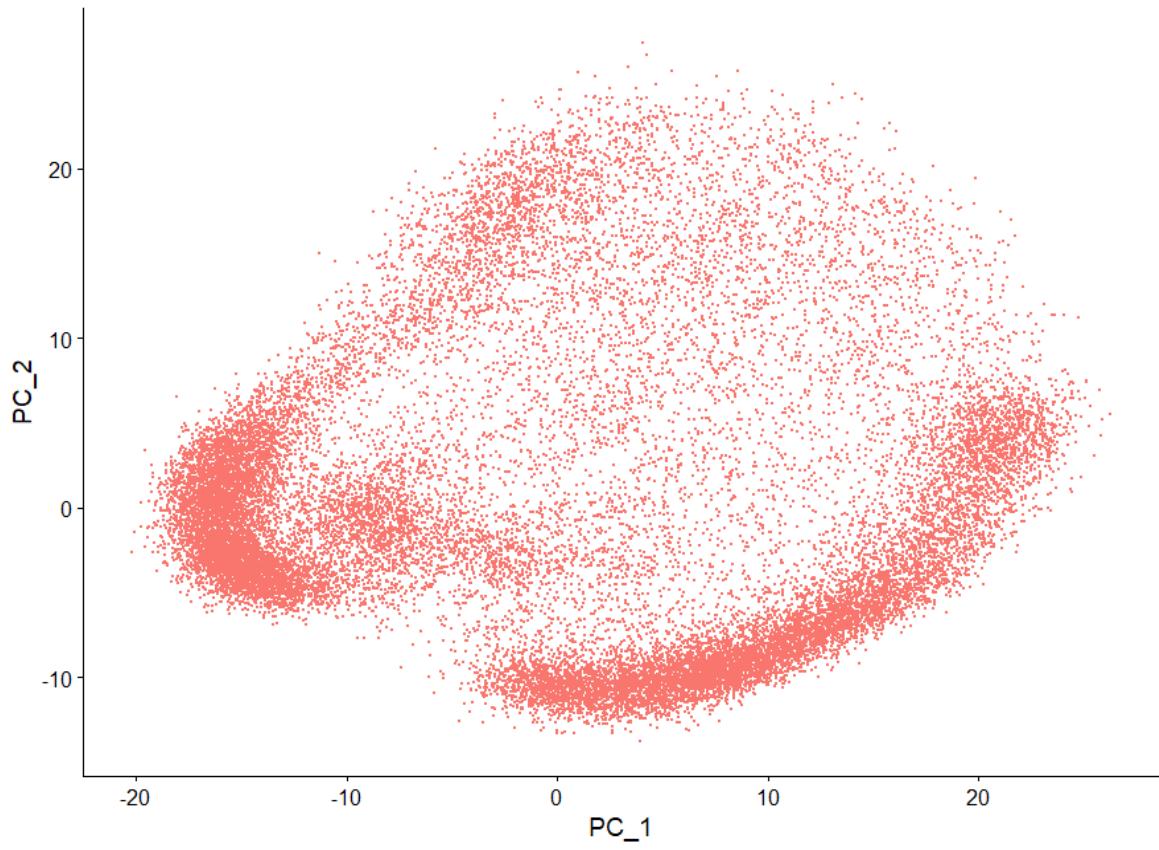
Normalization &  
dimensionality  
reduction



5. Normalize the data to account for sequencing depth using the `NormalizeData()` R function
  - Method: Feature counts for each cell are divided by the total counts for that cell and multiplied by the `scale.factor` (10.000 default). This is then log transformed.
6. Identify the 2.000 genes with the highest variance between samples using the `FindVariableFeatures()` R function



7. Scale the data by Z-score transforming the gene expression
8. Perform a Principal Component Analysis (PCA) dimensionality reduction using the 2.000 genes with the highest variance
9. Identify how many Principal Components (PCs) we should use in downstream analyses by ranking the % of variance explained by each of them using the ElbowPlot() R function

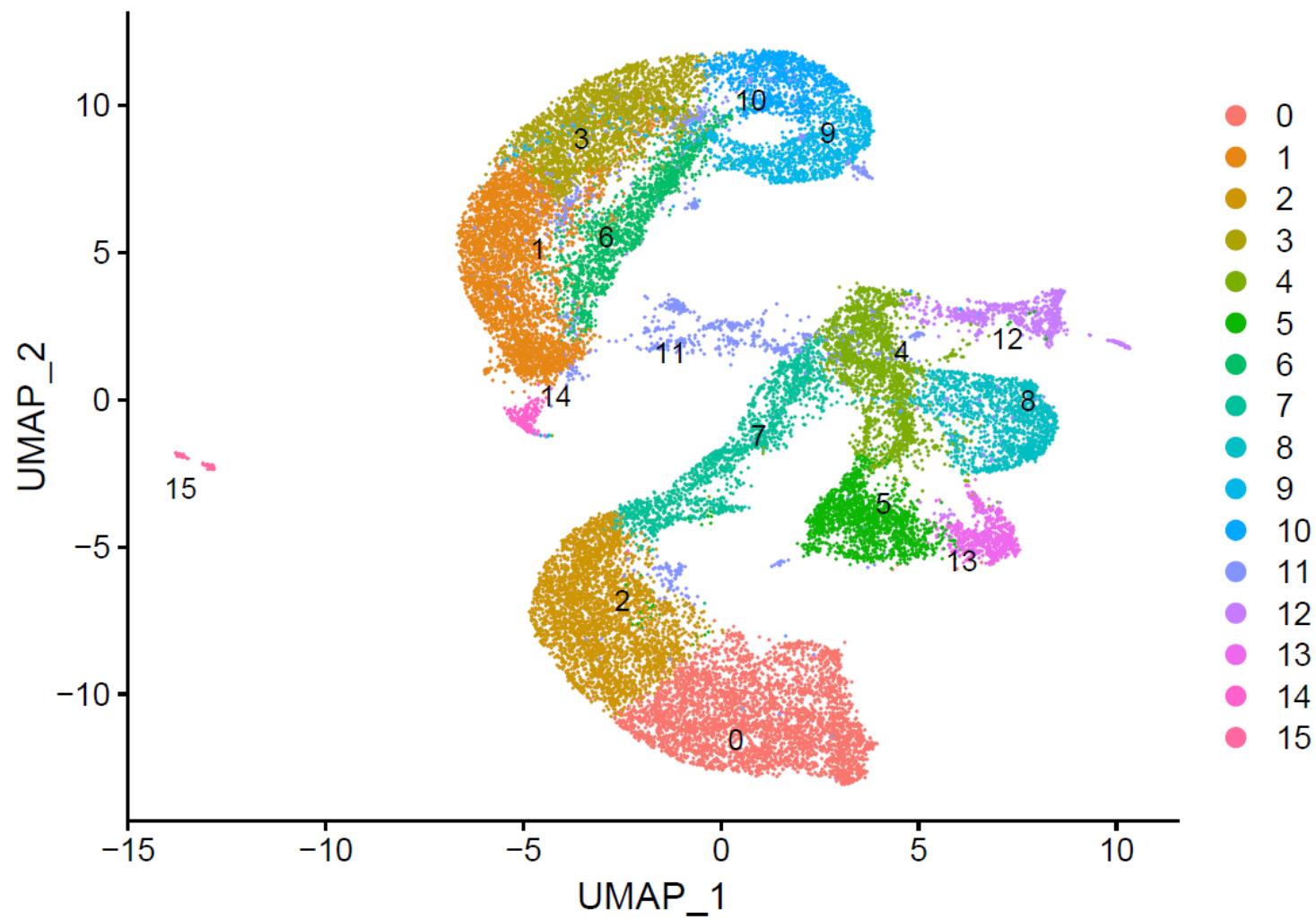


# 03

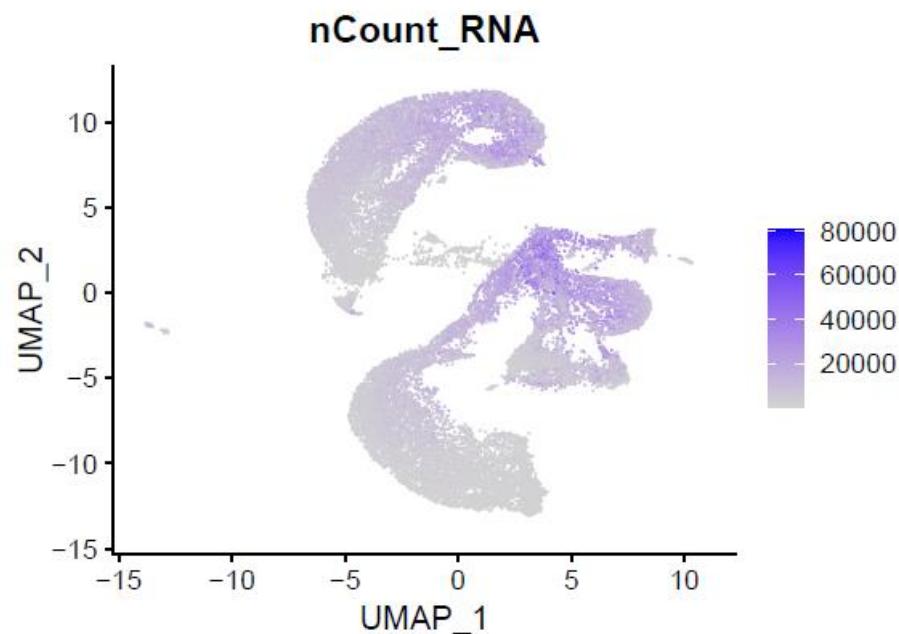
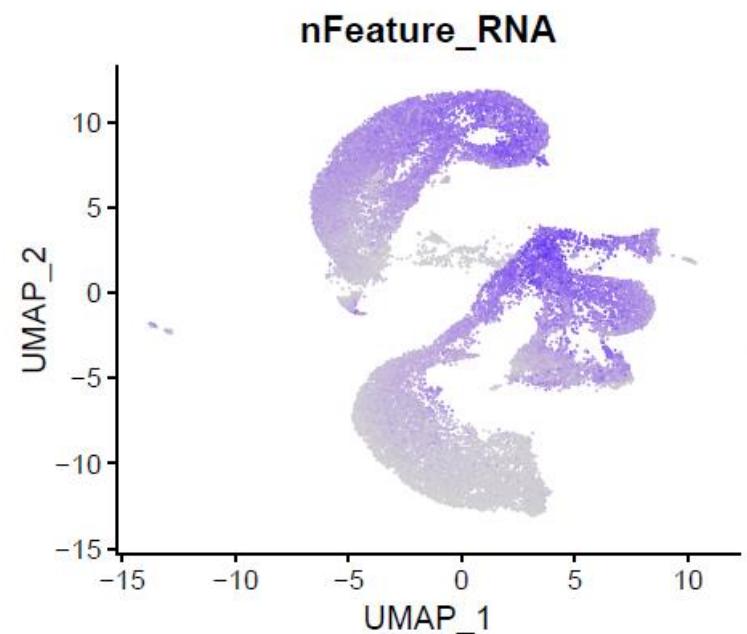
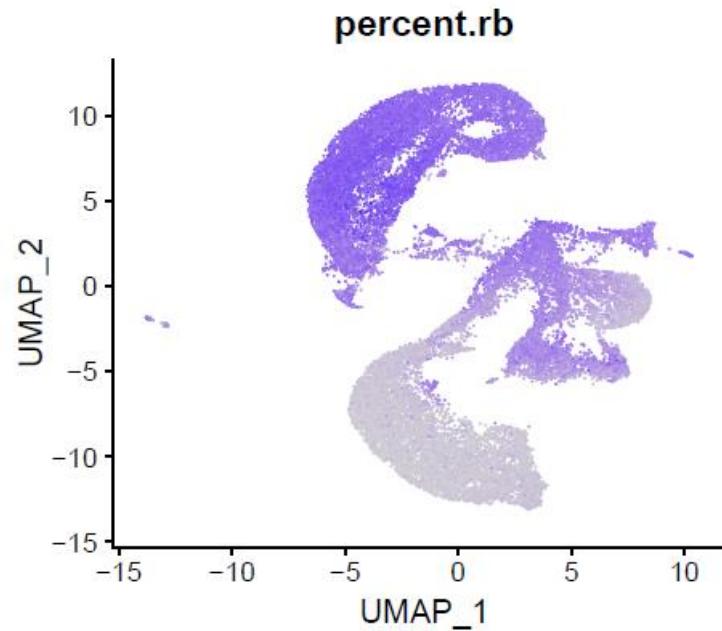
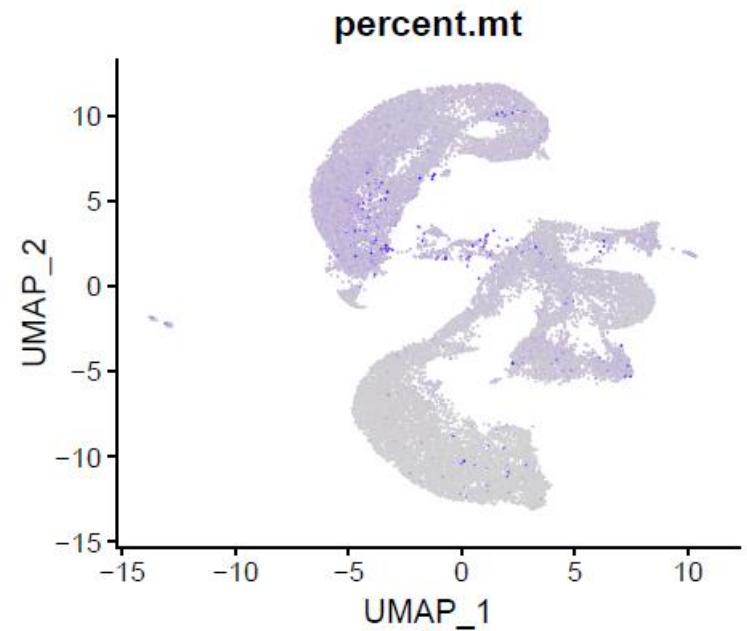
## Clustering & artifact control



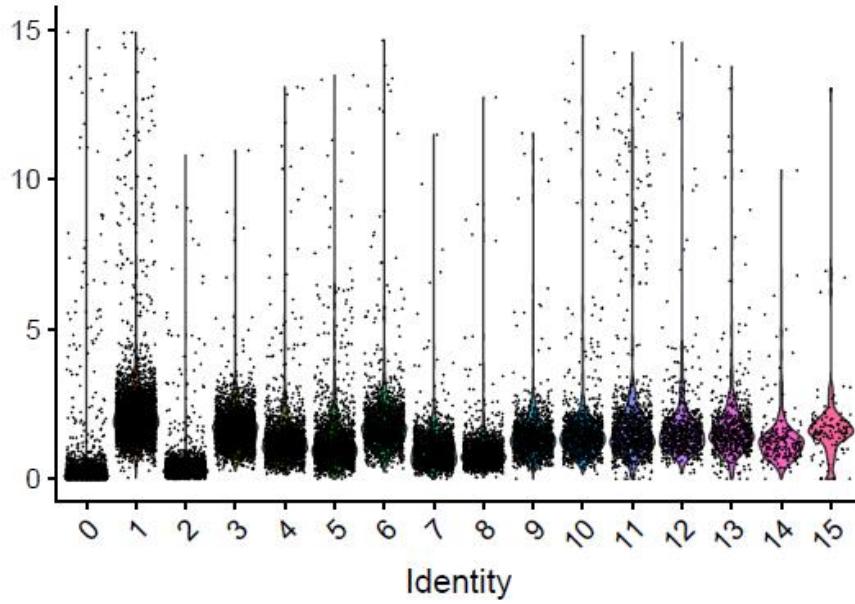
10. Using these number of PCs, perform clustering of the cells using the K-Nearest Neighbors (KNN) algorithm based on the euclidean distance in PCA space with the FindNeighbors() and FindClusters() R functions
11. For visualization purposes, generate UMAP reduced dimensionality representation of the results



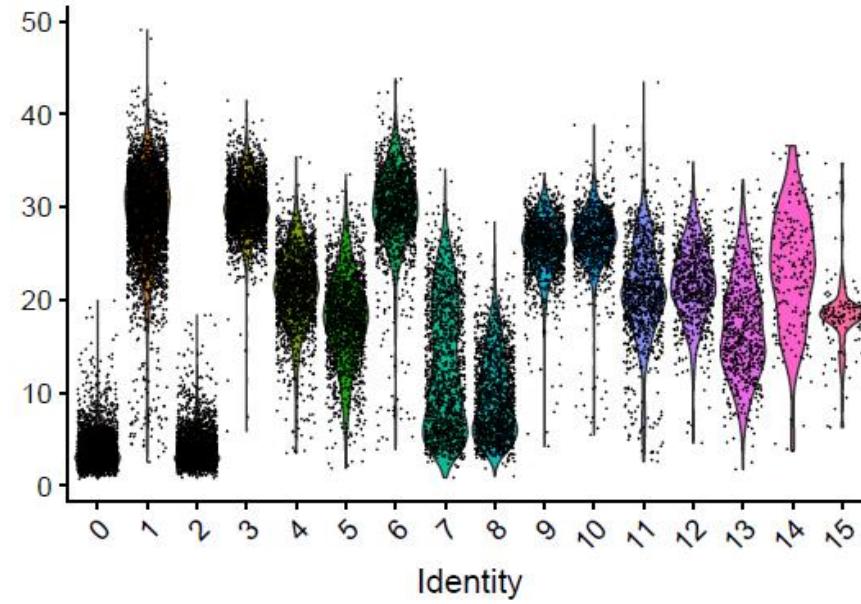
12. In order to control for possible artifacts, compare the distribution within clusters of the quality control (QC) metrics (nCount RNA, nFeature RNA, % mitochondrial genes, % ribosomal genes, +/- cell cycle score)



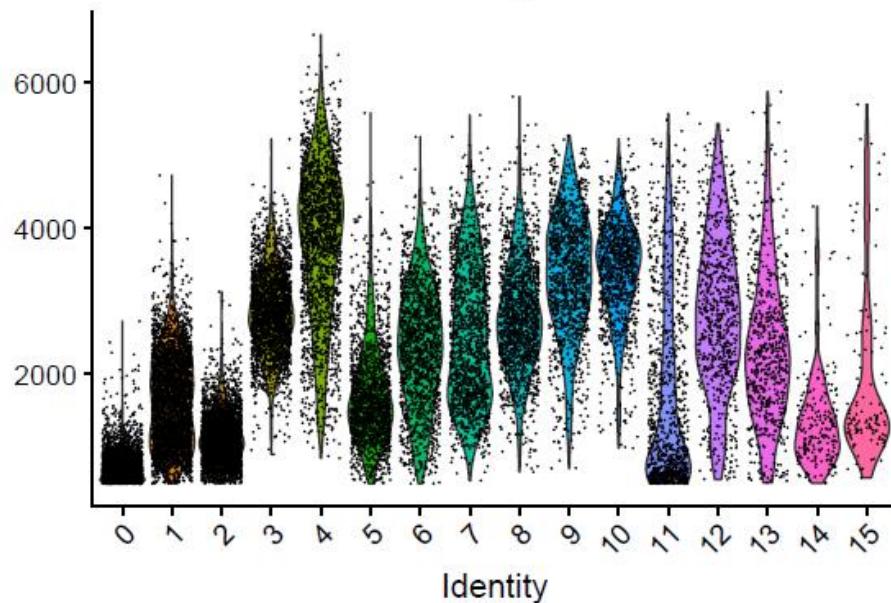
**percent.mt**



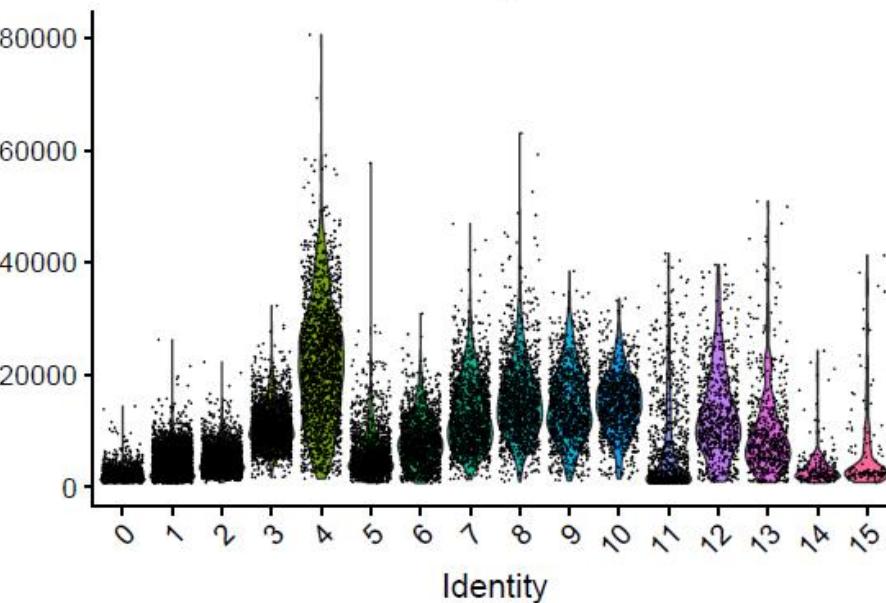
**percent.rb**



**nFeature\_RNA**



**nCount\_RNA**



13. Correct for the identified confounding factors that change dramatically between clusters using using vars.to.regres from the SCTransform() R function

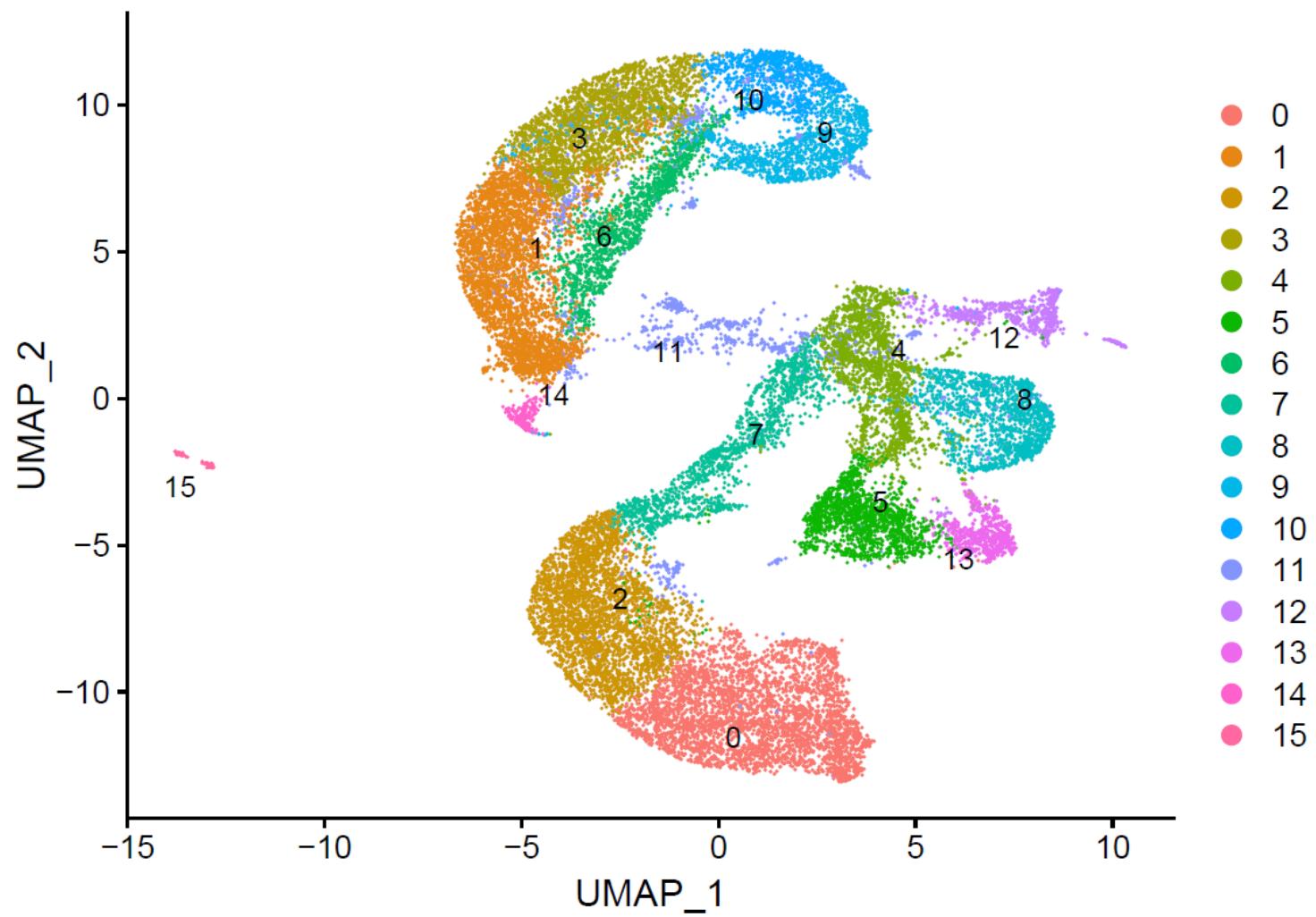
In this case, however, the distribution of none of the metrics seems to be an artifact after consultation with the experts (Ye lab from UCSF), so no correction is needed

# 04

## Marker selection



14. Find markers for every cluster by comparing it to all remaining cells using the Wilcoxon Rank Sum test with the `FindAllMarkers()` R function



### Top 5 markers of each cluster

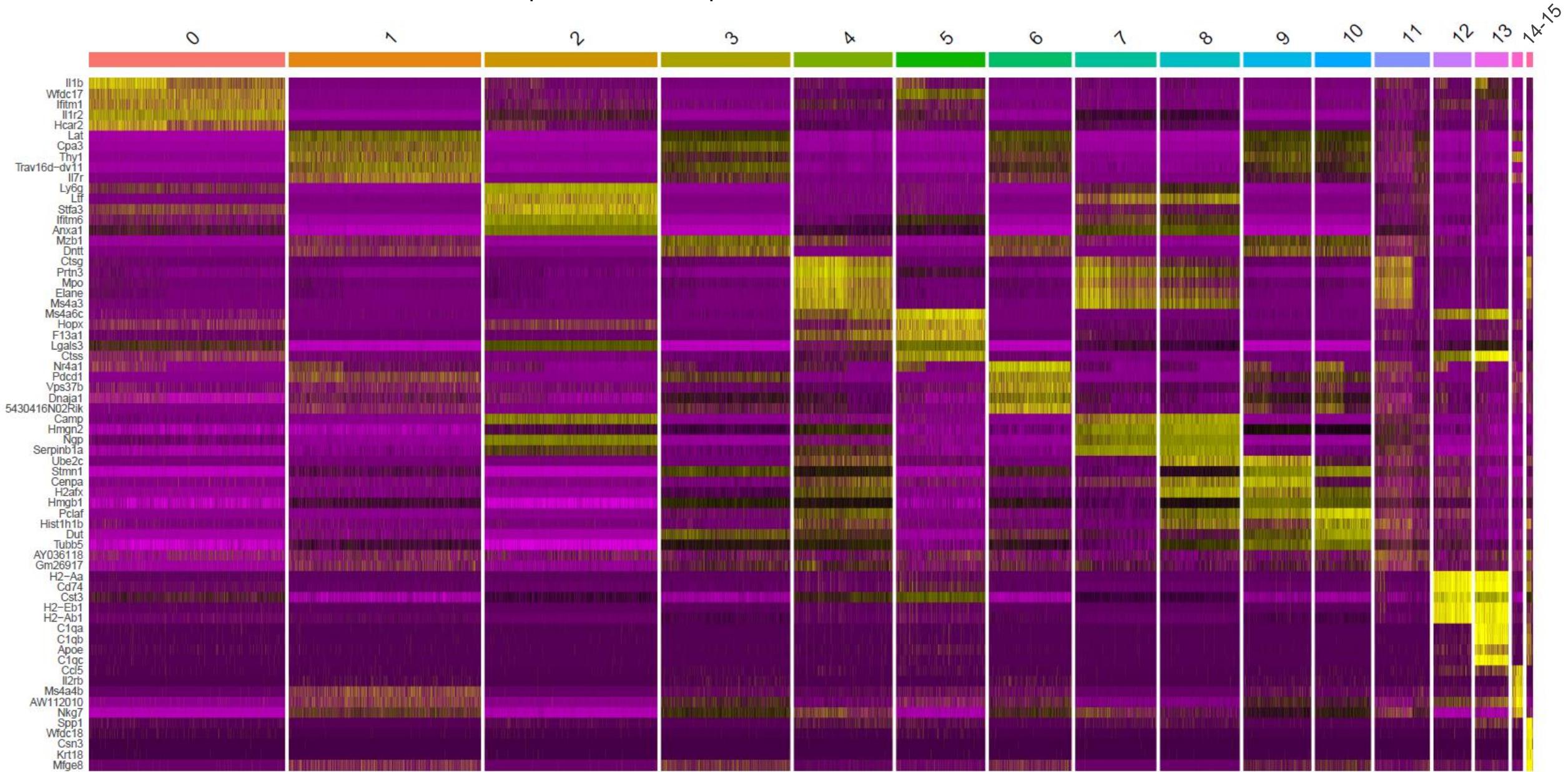
cluster	gene
0	Il1b
0	Wfdc17
0	Ifitm1
0	Il1r2
0	Hcar2
1	Lat
1	Cpa3
1	Thy1
1	Trav16d-dv11
1	Il7r
2	Ly6g
2	Ltf
2	Stfa3
2	Ifitm6
2	Anxa1
3	Lgals1
3	Cpa3
3	Mzb1
3	Dntt
3	Pebp1

cluster	gene
4	Ctsg
4	Prtn3
4	Mpo
4	Elane
4	Ms4a3
5	Ms4a6c
5	Hopx
5	F13a1
5	Lgals3
5	Ctss
6	Nr4a1
6	Pdcd1
6	Vps37b
6	Dnaja1
6	5430416N02Rik
7	Elane
7	Mpo
7	Prtn3
7	Camp
7	Hmgn2

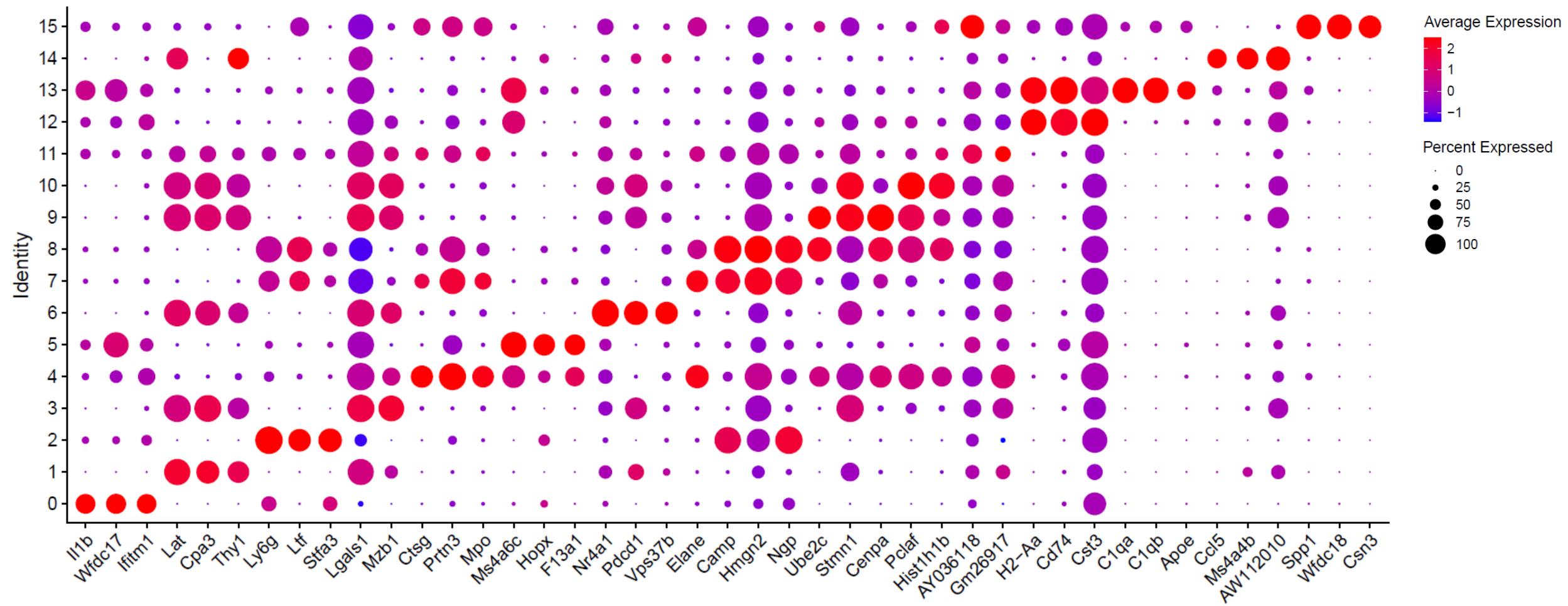
cluster	gene
8	Camp
8	Hmgn2
8	Ngp
8	Serpina1a
8	Ube2c
9	Ube2c
9	Stmn1
9	Cenpa
9	H2afx
9	Hmgb1
10	Pclf
10	Stmn1
10	Hist1h1b
10	Dut
10	Tubb5
11	mt-Cytb
11	Elane
11	Prtn3
11	AY036118
11	Gm26917

cluster	gene
12	H2-Aa
12	Cd74
12	Cst3
12	H2-Eb1
12	H2-Ab1
13	C1qa
13	C1qb
13	Apoe
13	C1qc
13	Cd74
14	Ccl5
14	Il2rb
14	Ms4a4b
14	AW112010
14	Nkg7
15	Spp1
15	Wfdc18
15	Csn3
15	Krt18
15	Mfge8

### Gene expression heatmap of the main markers across clusters



Gene expression bubble chart of the main markers across clusters



# 05

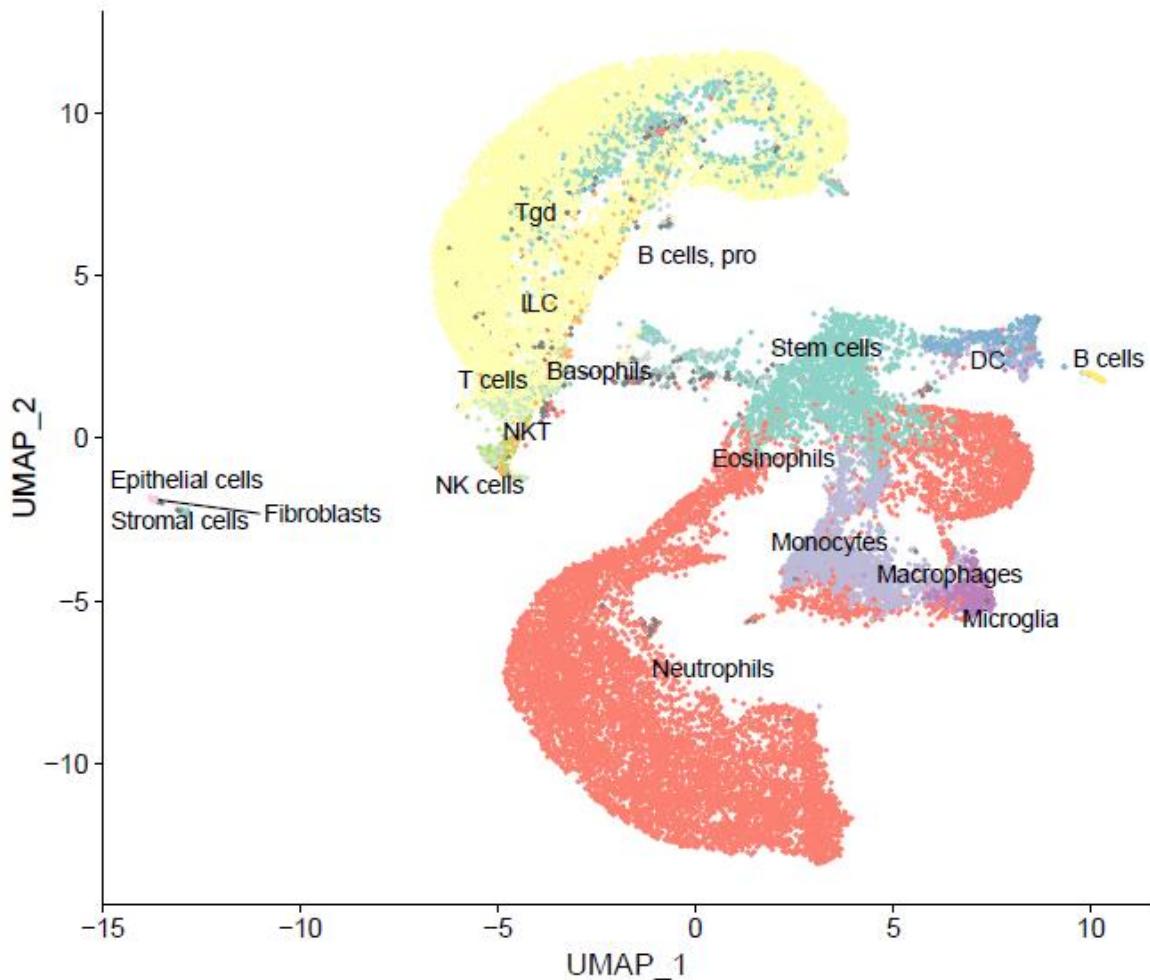
## Cell type annotation



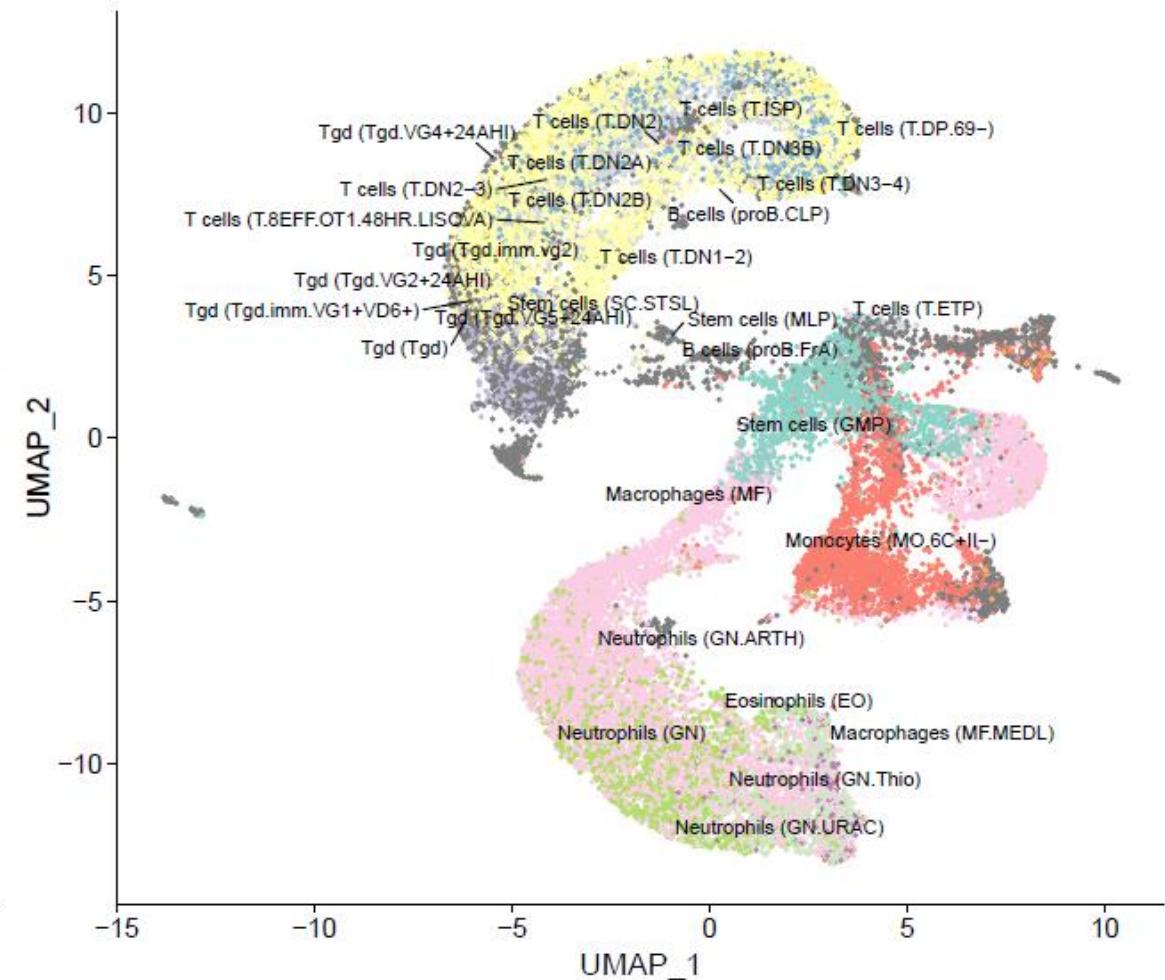
## 15. Assign cell types to the clusters using the ImmGen database as reference

- Convert the Seurat object into a single cell experiment object
- Get the ImmGen reference database from celldex package
- Assign cell types to the clusters using SingleR

Cell types: ImmGen (main labels)



Cell types: ImmGen (fine labels)



16. Check whether the cell types assigned with the main labels (general) and the fine labels (detailed subgroups) match well
17. Check what are the cell types from each cluster

ImmGen (main labels)

DC  
Macrophages

Monocytes

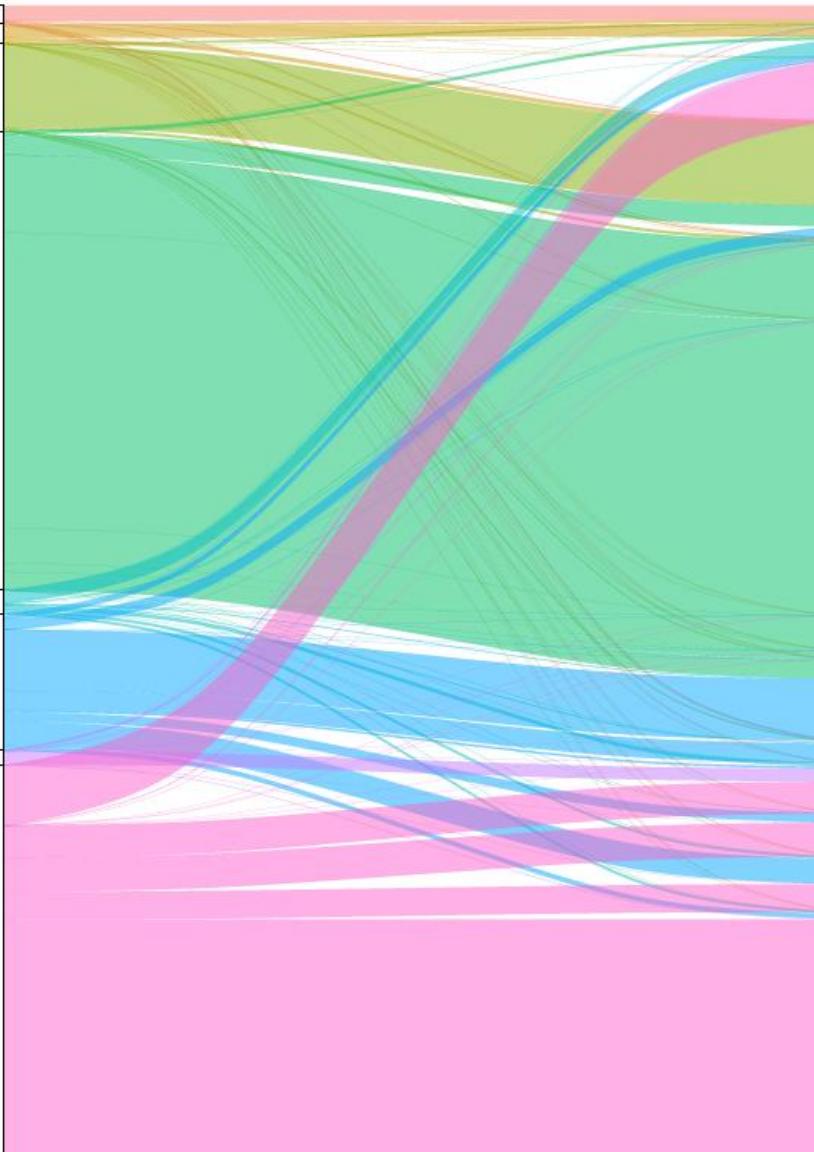
Neutrophils

Other

Stem cells

T cells

Tgd



ImmGen (fine labels)

DC  
Other

Tgd

Monocytes (MO.6C+II-)

Monocytes (other)

Neutrophils (GN)

Neutrophils (GN.ARTH)

Neutrophils (GN.URAC)

Neutrophils (other)

Stem cells (GMP)

Stem cells (other)

T cells (other)

T cells (T.DN2)

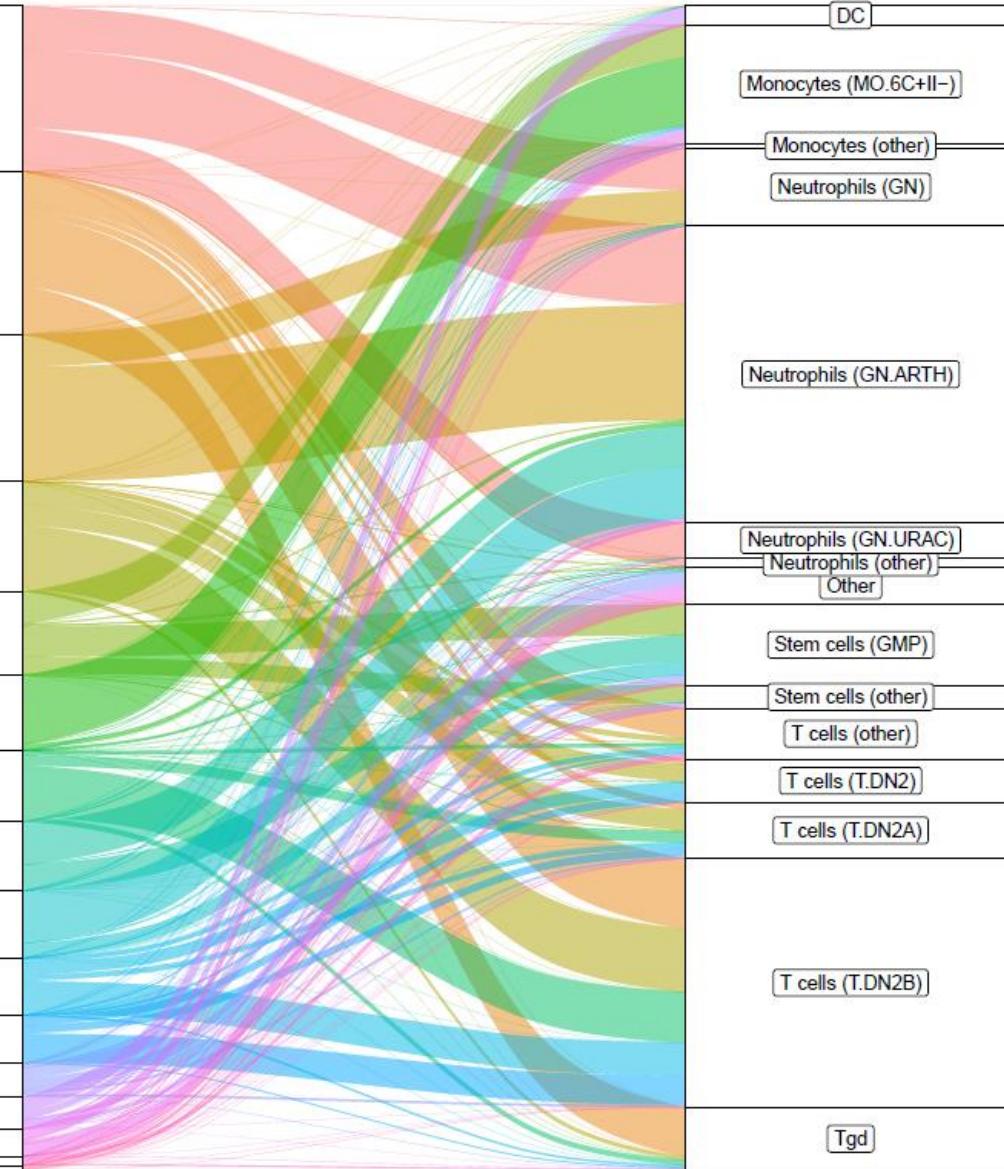
T cells (T.DN2A)

T cells (T.DN2B)

Clusters

0
1
2
3
4
5
6
7
8
9
10
11
12
13
14

ImmGen (fine labels)



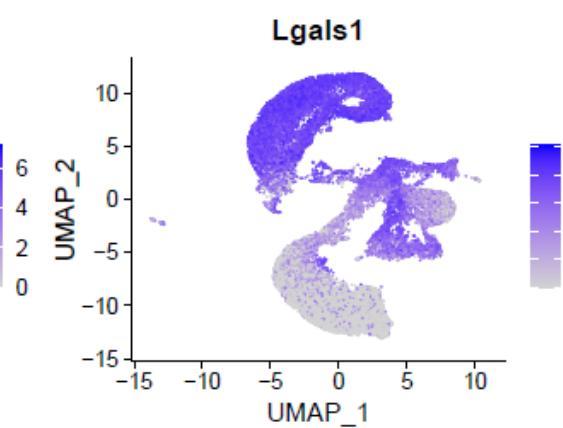
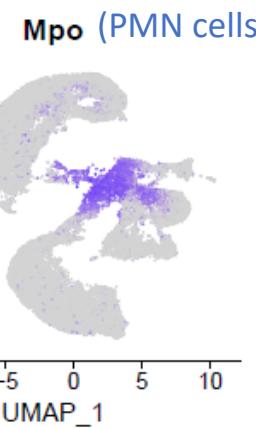
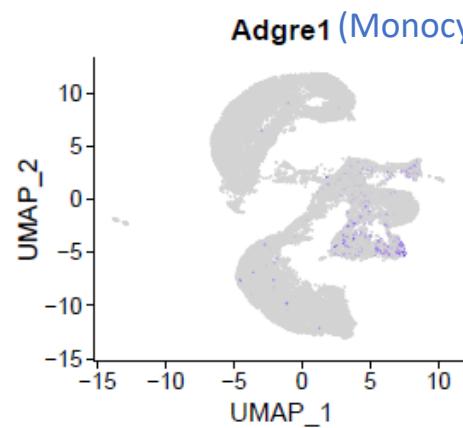
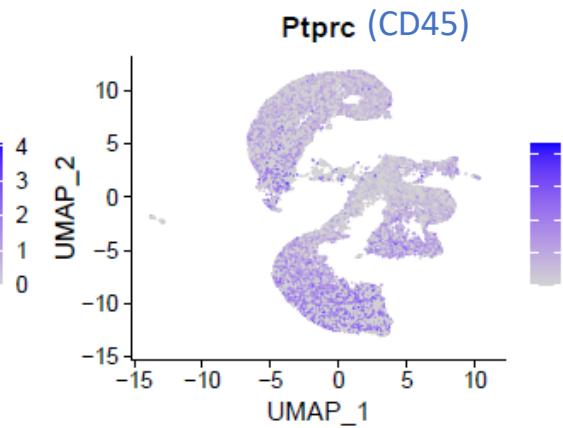
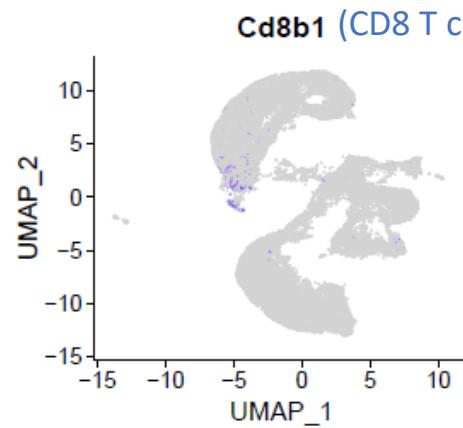
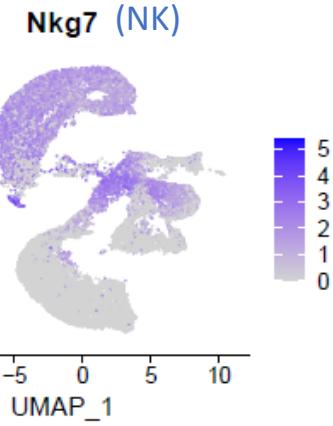
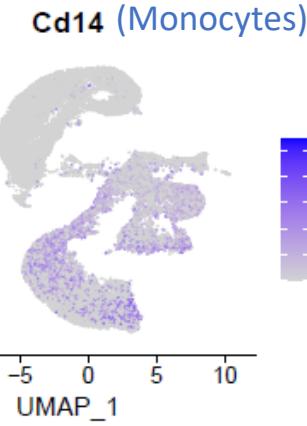
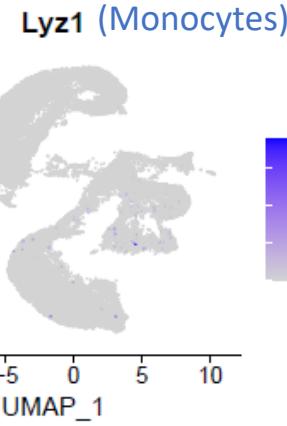
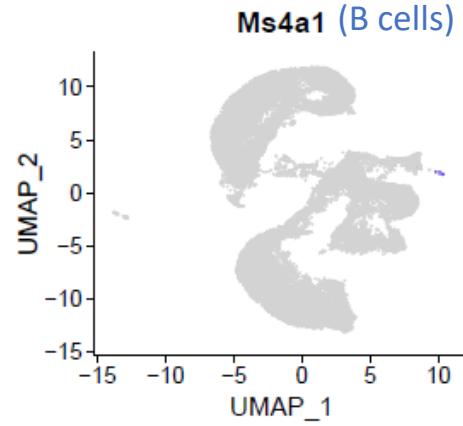
# 06

## Coloring by genes

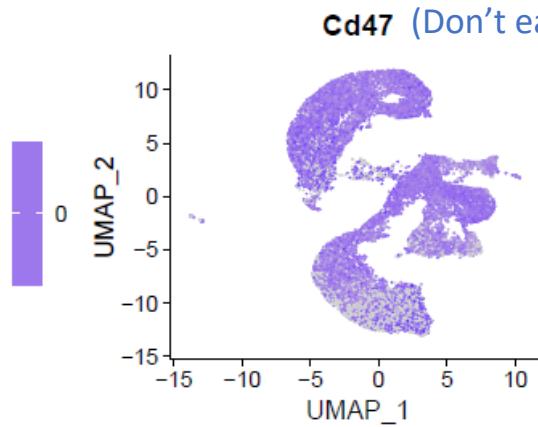
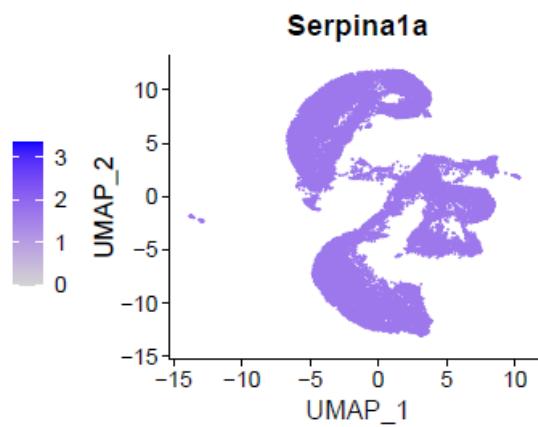
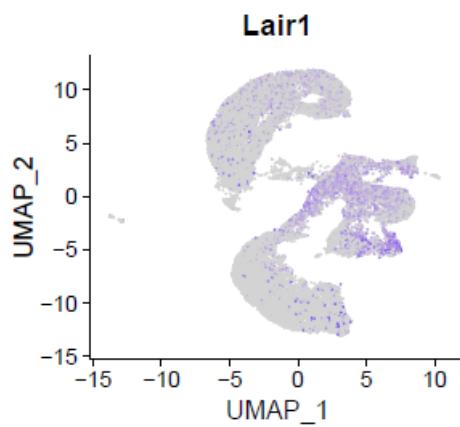
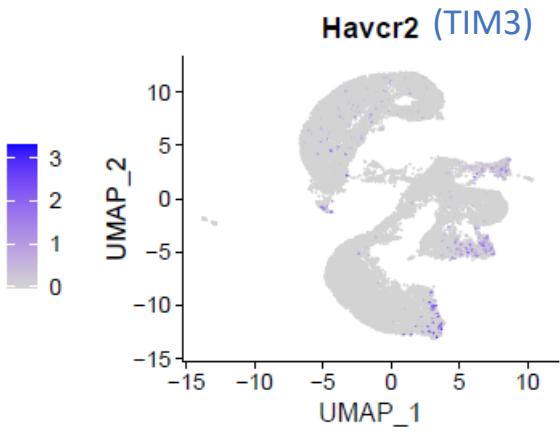
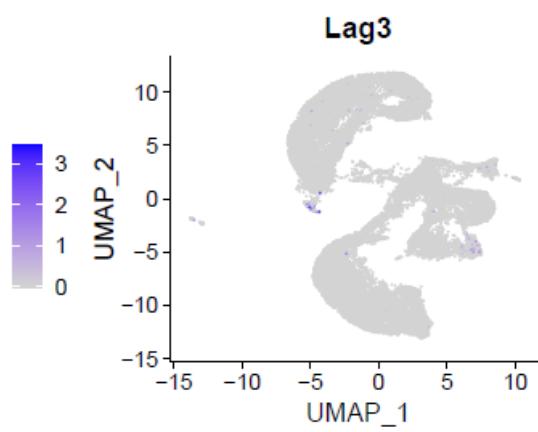
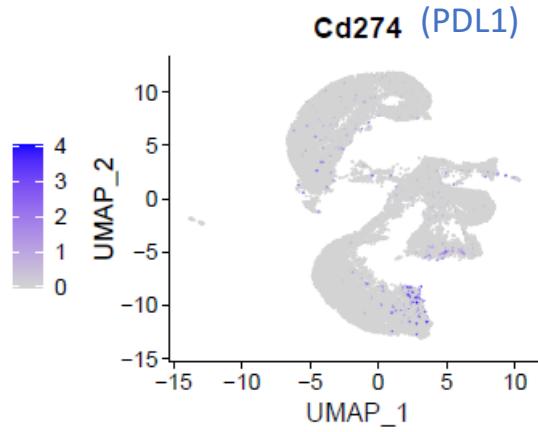
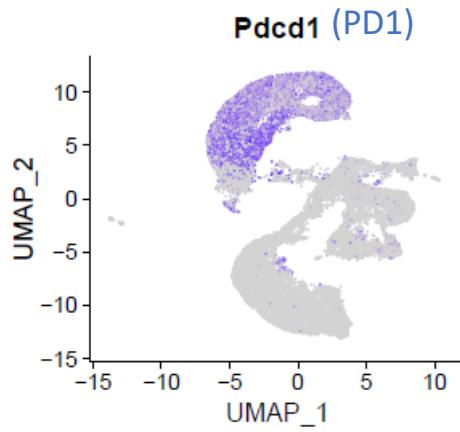
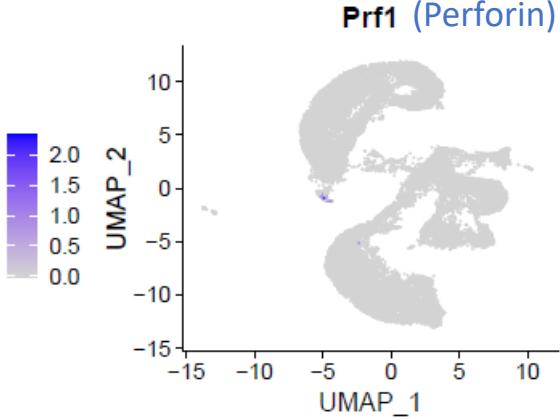
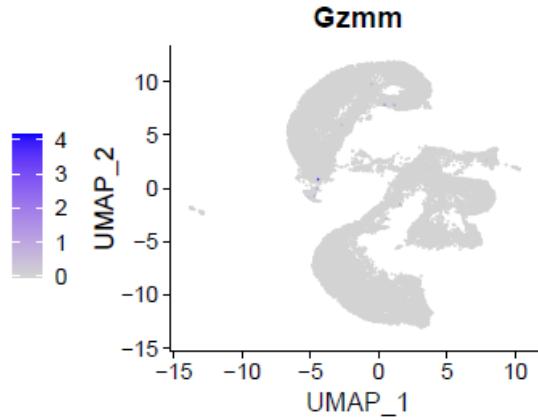
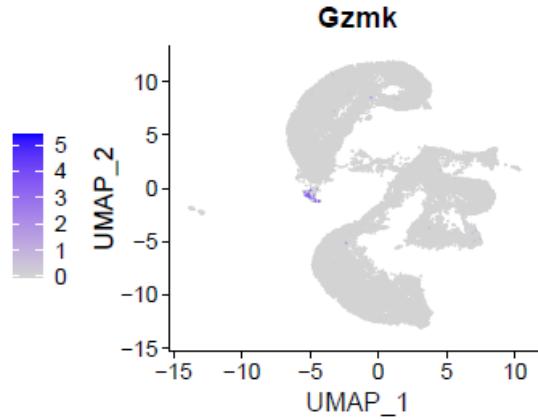
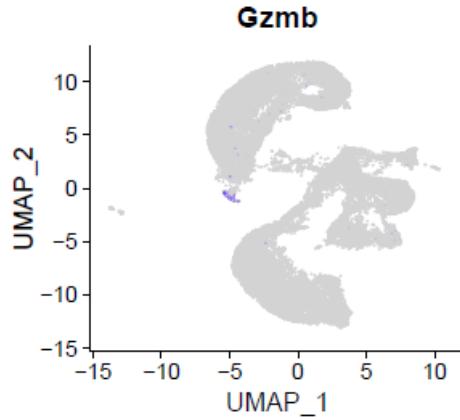


18. Visualize the distribution of the expression of some interesting genes across cells

Distribution of some famous markers

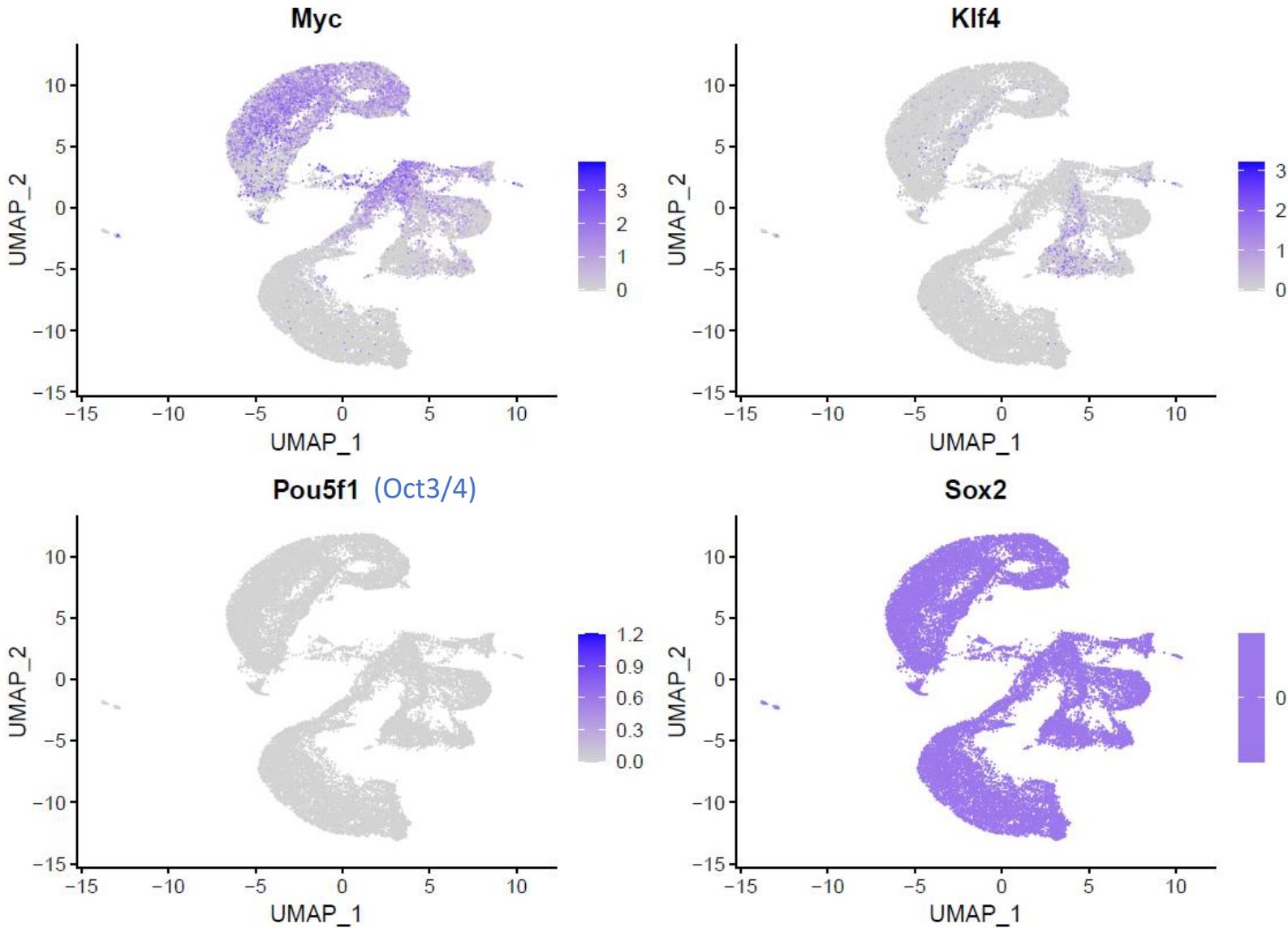


Distribution of some markers we were curious about



Granzyme h,  
Lair2 and  
Serpine A  
not available

### Distribution of the Yamanaka factors

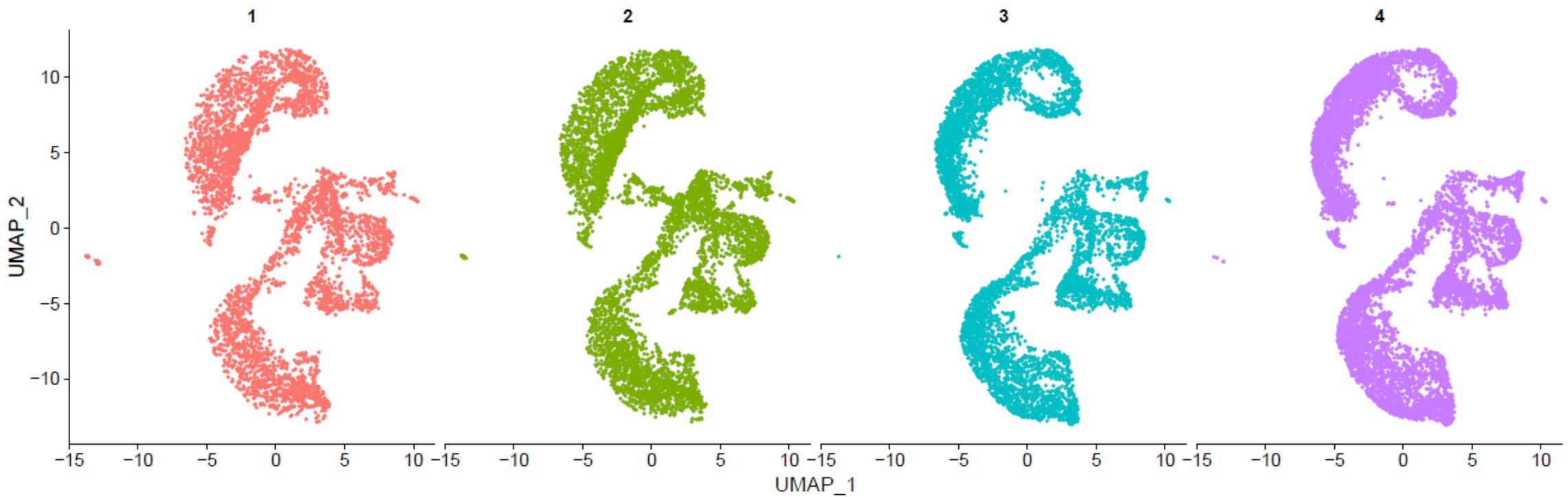


07

Comparison of  
the dissociation  
methods

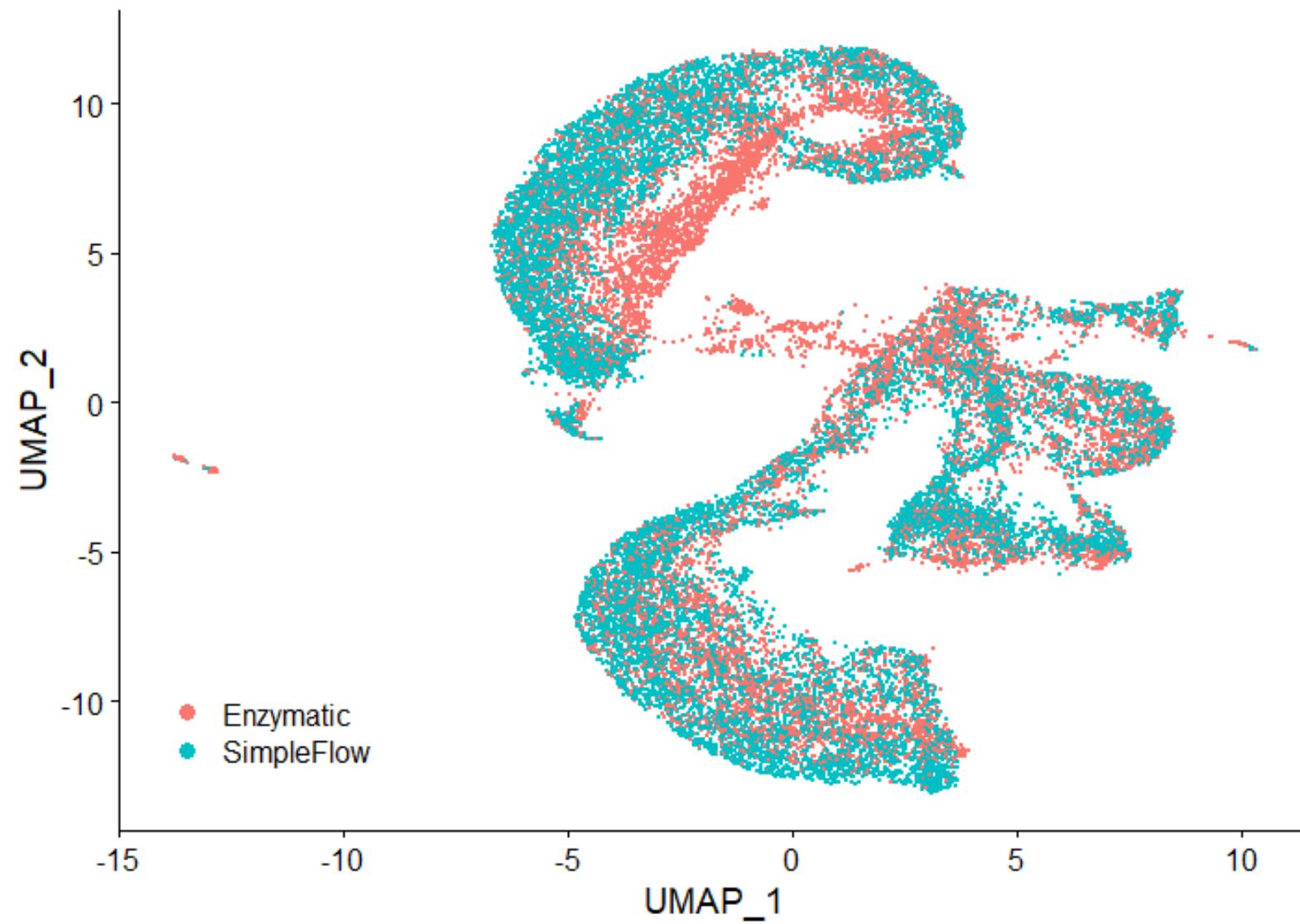


1. Visualize the distribution of cells from each murine mammary tumor and from the two dissociation groups



Samples 1 & 2: Single cells were dissociated with Enzymatic digestion

Samples 3 & 4: Single cells were dissociated with SimpleFlow™ System



2. Perform a differential gene expression (DGE) analysis between the two dissociation groups to identify the expression of what genes changes for cells of the same type under different dissociation techniques

For that, we will use the `FindAllMarkers()` R function that uses the Wilcoxon Rank Sum test

For simplicity, we will focus on the largest cell types

## Top 15 differentially expressed genes between dissociation methods by cell type

```
> head(Monocytes, n = 15)
```

		p_val	avg_log2FC	pct.1	pct.2	p_val_adj
●	Hspa1b	1.040215e-155	1.5454332	0.538	0.031	3.230179e-151
●	Nr4a1	5.232288e-130	2.4541140	0.743	0.241	1.624782e-125
●	Hspa1a	4.230262e-110	1.5763296	0.497	0.068	1.313623e-105
●	Dnajb1	1.186188e-75	1.6965480	0.659	0.305	3.683470e-71
Atf3		9.978361e-59	2.0161393	0.790	0.593	3.098580e-54
Ccl4		4.264070e-51	2.0004175	0.622	0.319	1.324122e-46
Hsp90aa1		5.711892e-46	1.0097817	0.786	0.552	1.773714e-41
Cdkn1a		7.336598e-39	1.0691153	0.688	0.447	2.278234e-34
H3f3b		1.700543e-37	0.5771917	1.000	0.999	5.280697e-33
Btg2		4.847031e-35	0.8355704	0.874	0.785	1.505149e-30
Srgn		8.298998e-34	0.8085828	0.903	0.772	2.577088e-29
Osm		4.413170e-32	1.0151987	0.428	0.199	1.370422e-27
Ier3		1.714669e-31	1.1314937	0.852	0.734	5.324563e-27
Ccl3		2.459093e-31	0.8946007	0.772	0.589	7.636220e-27
Il1b		1.501253e-30	2.0214160	0.450	0.237	4.661841e-26

```
> head(Tgd, n = 15)
```

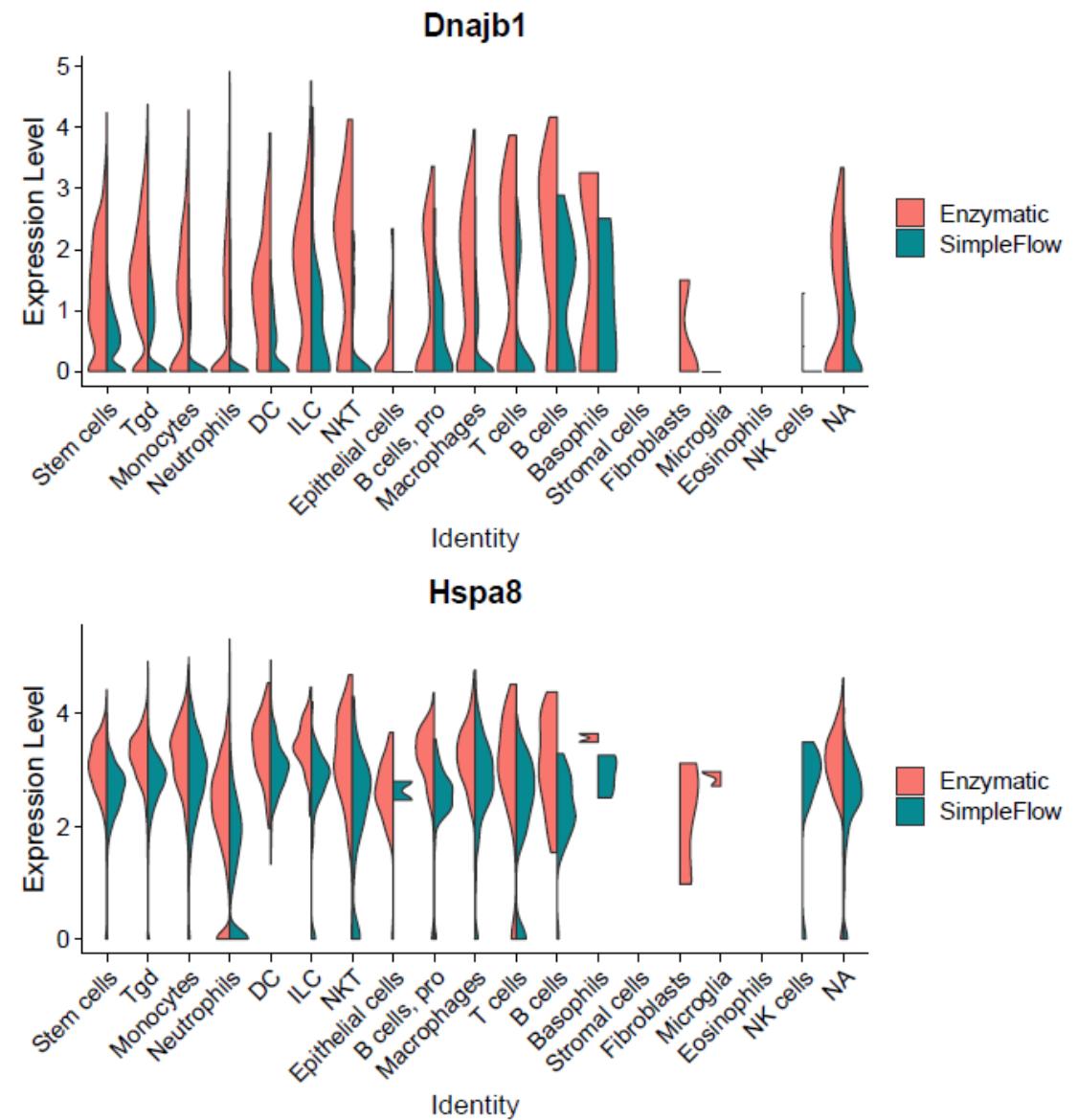
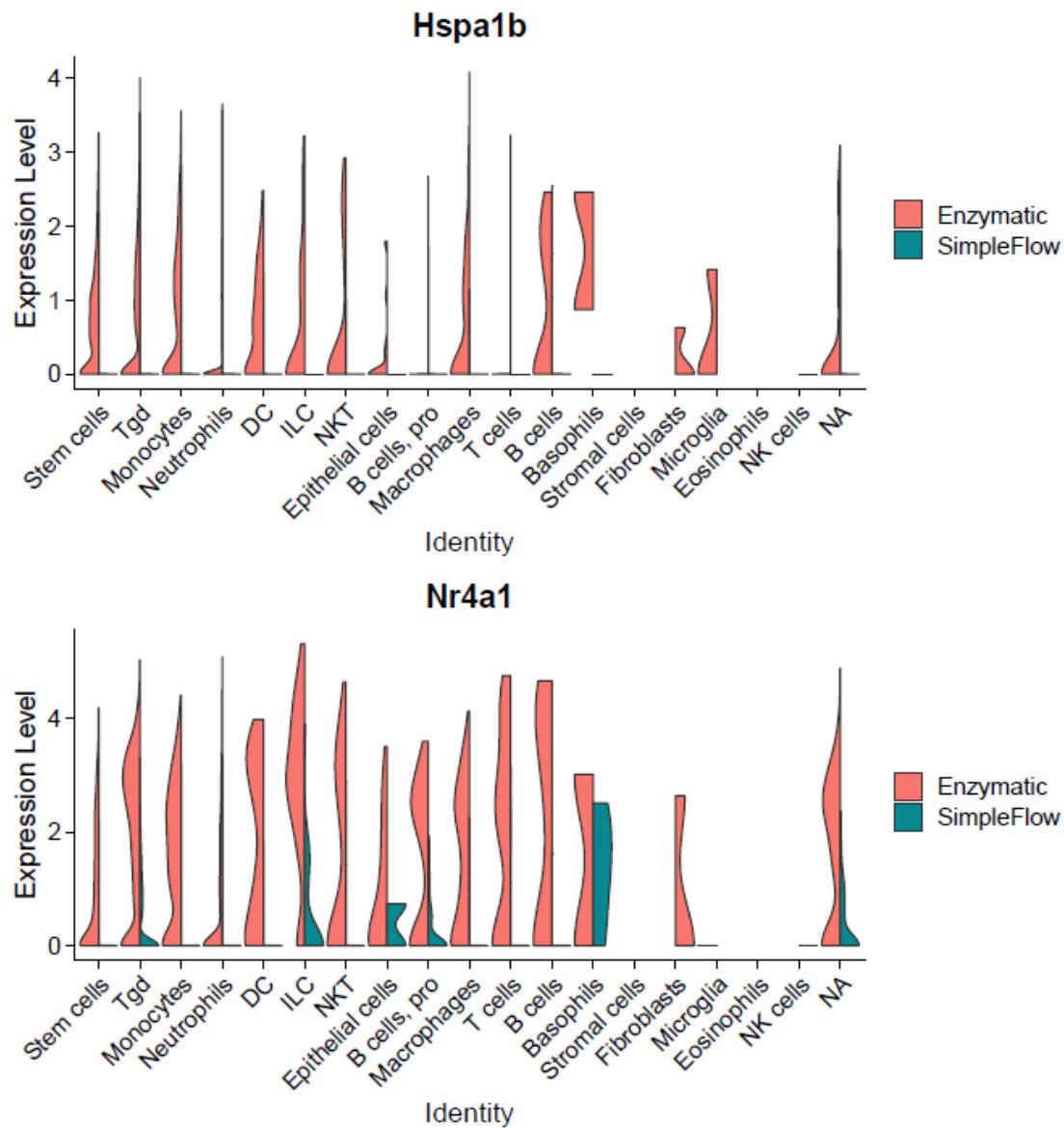
		p_val	avg_log2FC	pct.1	pct.2	p_val_adj
●	Dnajb1	0.000000e+00	1.4599982	0.854	0.613	0.000000e+00
●	Hspa8	0.000000e+00	0.6472637	0.996	0.985	0.000000e+00
●	Nr4a1	0.000000e+00	2.6383861	0.785	0.420	0.000000e+00
●	Hspa1b	0.000000e+00	1.5541028	0.560	0.016	0.000000e+00
●	Hspa1a	0.000000e+00	1.6776488	0.446	0.014	0.000000e+00
Hsp90aa1		2.583884e-275	0.6212419	0.969	0.909	8.023735e-271
Hsp90ab1		3.865601e-230	0.3527013	0.999	0.997	1.200385e-225
Cdkn1a		2.824298e-223	1.1446326	0.448	0.169	8.770294e-219
Ppp1r15a		3.299994e-209	0.8235387	0.765	0.499	1.024747e-204
Plaur		9.011602e-207	1.0946360	0.645	0.361	2.798373e-202
Atf3		4.832174e-199	0.9064369	0.207	0.015	1.500535e-194
Tnfsf11		2.023240e-198	0.6370298	0.195	0.011	6.282768e-194
Csrnp1		5.380092e-192	0.8110122	0.475	0.203	1.670680e-187
Nr4a2		5.067422e-191	0.8392308	0.403	0.147	1.573587e-186
Jun		1.030369e-186	0.8721499	0.897	0.748	3.199606e-182

```
> head(Neutrophils, n = 15)
```

		p_val	avg_log2FC	pct.1	pct.2	p_val_adj
G0s2		0.000000e+00	2.2647916	0.762	0.531	0.000000e+00
●	Hspa8	0.000000e+00	1.1134147	0.879	0.711	0.000000e+00
●	Nr4a1	0.000000e+00	1.9639366	0.435	0.071	0.000000e+00
●	Hspa1b	0.000000e+00	0.7527038	0.256	0.011	0.000000e+00
Csrnp1		1.446207e-240	1.0648900	0.448	0.153	4.490907e-236
●	Dnajb1	2.659682e-220	1.1938296	0.607	0.334	8.259110e-216
Klf6		1.515432e-217	0.8794987	0.892	0.740	4.705870e-213
Trem1		6.158587e-211	1.0356728	0.686	0.403	1.912426e-206
Plaur		1.202167e-208	1.0528873	0.825	0.656	3.733089e-204
Hsp90aa1		1.559493e-204	0.9325669	0.580	0.320	4.842694e-200
Fosl1		1.995955e-200	0.8110221	0.302	0.076	6.198039e-196
Nfkbid		6.757802e-194	0.8075579	0.298	0.078	2.098500e-189
Tnfaip6		5.856903e-184	0.6785843	0.175	0.016	1.818744e-179
●	Hspa1a	1.105735e-183	0.5400613	0.163	0.011	3.433638e-179
Hsp90ab1		2.126125e-175	0.8851453	0.778	0.622	6.602257e-171

```
> head(STEM.CELLS, n = 15)
```

		p_val	avg_log2FC	pct.1	pct.2	p_val_adj
●	Hspa1b	2.711773e-247	1.2752226	0.660	0.047	8.420869e-243
●	Hspa1a	1.161467e-155	1.2118726	0.489	0.044	3.606703e-151
●	Dnajb1	7.792747e-132	1.5208896	0.839	0.710	2.419882e-127
●	Nr4a1	3.926794e-123	1.9038705	0.587	0.220	1.219387e-118
●	Hspa8	2.028165e-112	0.6117943	0.987	0.995	6.298060e-108
Hsp90aa1		2.567353e-91	0.5895800	0.962	0.991	7.972401e-87
Jun		7.146602e-67	0.8336549	0.907	0.888	2.219234e-62
Hspf1		7.508380e-61	0.4787178	0.433	0.167	2.331577e-56
Hnrnpab		8.344769e-60	-0.4266460	0.874	0.980	2.591301e-55
Rps18		2.037699e-56	0.3285766	0.948	0.988	6.327668e-52
Bag3		2.962035e-56	0.5430598	0.403	0.160	9.198009e-52
Srsf2		4.995195e-56	-0.3655303	0.882	0.973	1.551158e-51
Gnai2		9.259230e-50	-0.4248351	0.819	0.947	2.875269e-45
Ppp1cc		1.249587e-49	-0.3403015	0.826	0.946	3.880342e-45
Gm10260		1.705010e-48	-0.2793919	0.980	1.000	5.294567e-44



3. Perform pathway analysis of the top 15 differentially expressed genes per cell type between dissociation groups using the DAVID platform ([link](#))

## Main differentially expressed pathways between dissociation methods by cell type

Sublist	Category	Term	RT	Genes	Count	%	P-Value	Fold Enrichment	Benjamini
□	UP_KEYWORDS	<a href="#">Stress response</a>	RT		6	40.0	6.8E-10	121.0	3.0E-8
□	UP_KEYWORDS	<a href="#">Chaperone</a>	RT		6	40.0	7.6E-8	47.5	1.7E-6
□	GOTERM_MF_DIRECT	<a href="#">unfolded protein binding</a>	RT		5	33.3	2.2E-7	83.1	1.6E-5
□	KEGG_PATHWAY	<a href="#">Protein processing in endoplasmic reticulum</a>	RT		6	40.0	2.5E-7	30.5	5.2E-6
□	KEGG_PATHWAY	<a href="#">Antigen processing and presentation</a>	RT		5	33.3	8.1E-7	52.1	8.5E-6

Neutrophils (n=10,202):

1) **Stress response**

Sublist	Category	Term	RT	Genes	Count	%	P-Value	Fold Enrichment	Benjamini
□	UP_KEYWORDS	<a href="#">Stress response</a>	RT		7	46.7	3.1E-12	141.1	1.6E-10
□	UP_KEYWORDS	<a href="#">Chaperone</a>	RT		6	40.0	7.6E-8	47.5	1.9E-6
□	KEGG_PATHWAY	<a href="#">Protein processing in endoplasmic reticulum</a>	RT		7	46.7	1.5E-7	22.9	9.2E-6
□	GOTERM_MF_DIRECT	<a href="#">unfolded protein binding</a>	RT		5	33.3	3.1E-7	77.5	2.9E-5
□	KEGG_PATHWAY	<a href="#">Estrogen signaling pathway</a>	RT		6	40.0	3.6E-7	33.6	1.1E-5

Tgd (n=9,706):

1) **Stress response**

Sublist	Category	Term	RT	Genes	Count	%	P-Value	Fold Enrichment	Benjamini
□	UP_KEYWORDS	<a href="#">Stress response</a>	RT		6	40.0	6.8E-10	121.0	3.8E-8
□	INTERPRO	<a href="#">Heat shock protein 70 family</a>	RT		4	26.7	7.1E-8	422.4	1.4E-6
□	INTERPRO	<a href="#">Heat shock protein 70, conserved site</a>	RT		4	26.7	7.1E-8	422.4	1.4E-6
□	UP_KEYWORDS	<a href="#">Chaperone</a>	RT		6	40.0	7.6E-8	47.5	1.5E-6
□	UP_KEYWORDS	<a href="#">Acetylation</a>	RT		12	80.0	8.1E-8	5.8	1.5E-6

Stem cells (n=3,109):

1) **Stress response**

2) **Heat shock protein 70 family**

Sublist	Category	Term	RT	Genes	Count	%	P-Value	Fold Enrichment	Benjamini
□	GOTERM_BP_DIRECT	<a href="#">response to heat</a>	RT		4	26.7	1.1E-5	3.2E-3	
□	UP_KEYWORDS	<a href="#">Stress response</a>	RT		4	26.7	1.2E-5	6.2E-4	
□	GOTERM_BP_DIRECT	<a href="#">negative regulation of cell proliferation</a>	RT		5	33.3	1.7E-4	2.5E-2	
□	UP_KEYWORDS	<a href="#">Chaperone</a>	RT		4	26.7	2.0E-4	3.8E-3	
□	UP_KEYWORDS	<a href="#">Cytokine</a>	RT		4	26.7	2.3E-4	3.8E-3	

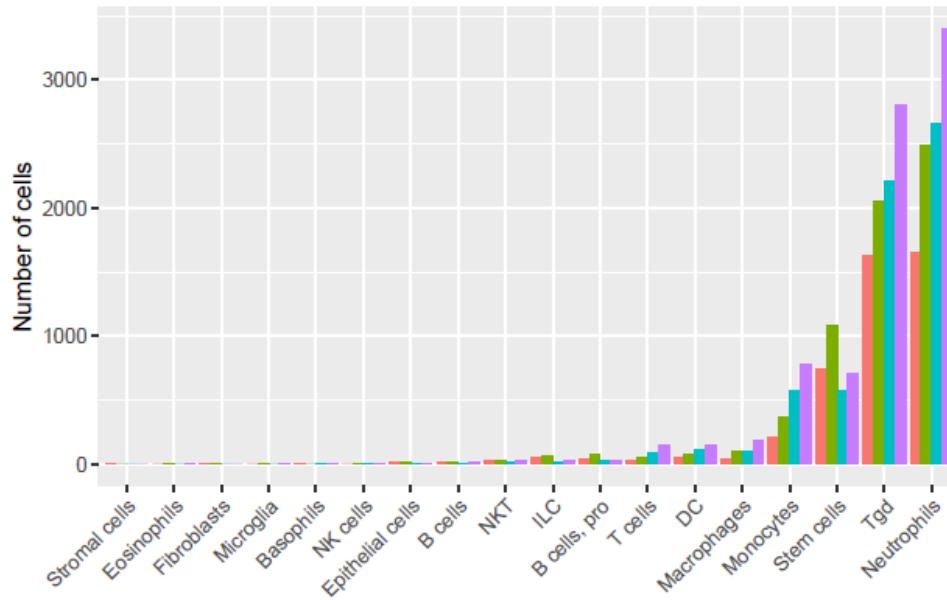
Monocytes (n=1,939):

1) **Response to heat**

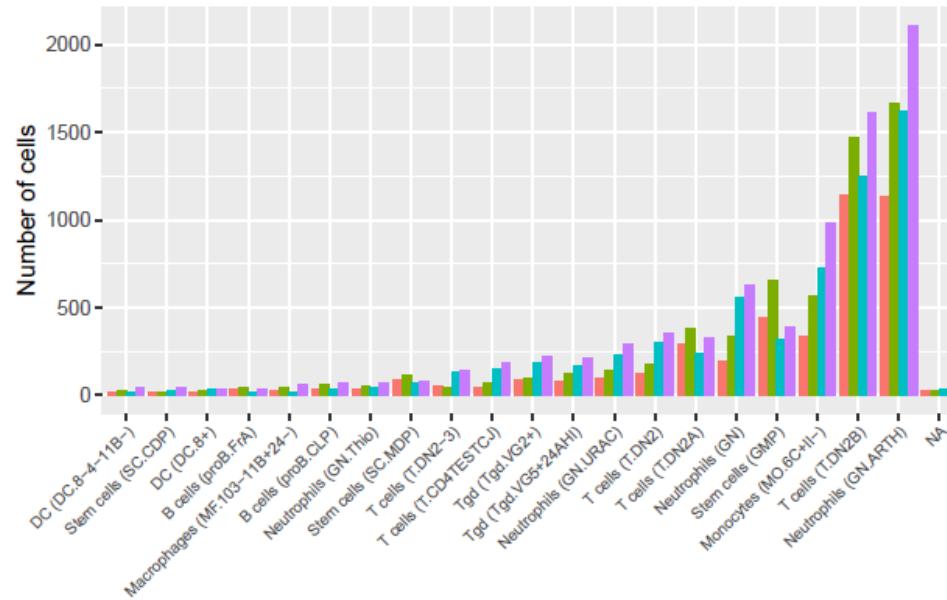
2) **Stress response**

4. Compare the cell type composition of samples from the two dissociation groups

Cell types: ImmGen (main labels)

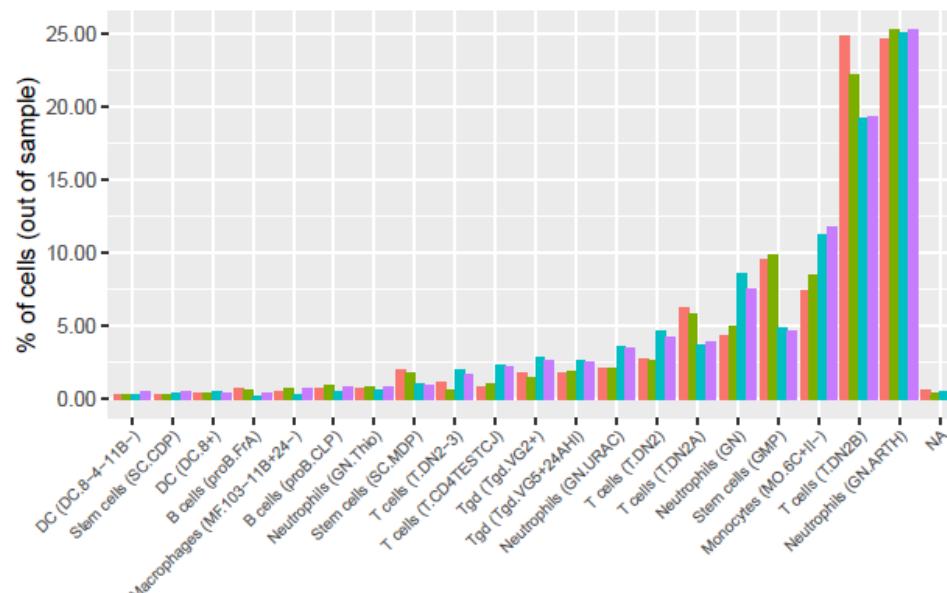
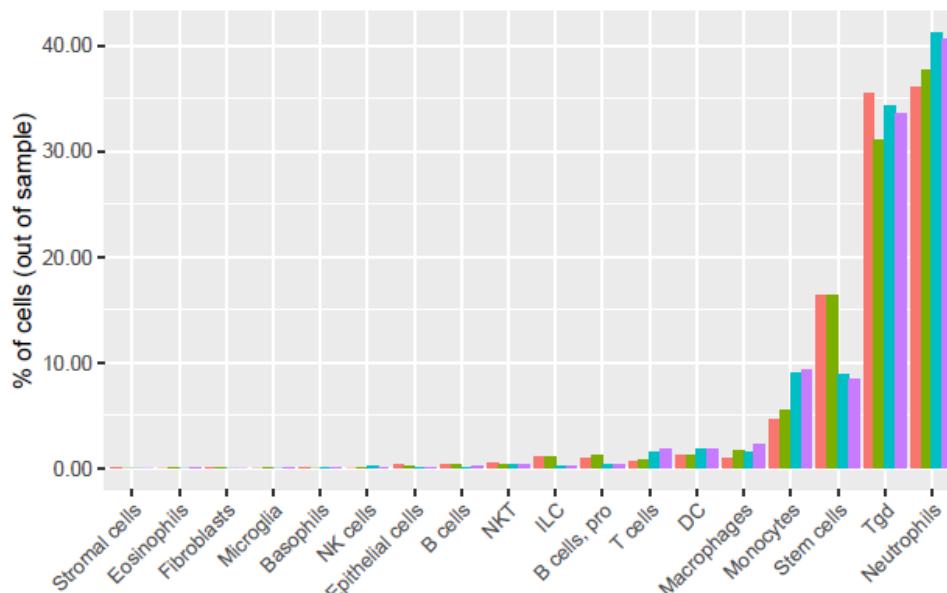


Cell types: ImmGen (fine labels)



Sample ID  
1  
2  
3  
4

Samples 1 & 2:  
Single cells were  
dissociated with  
enzymatic digestion

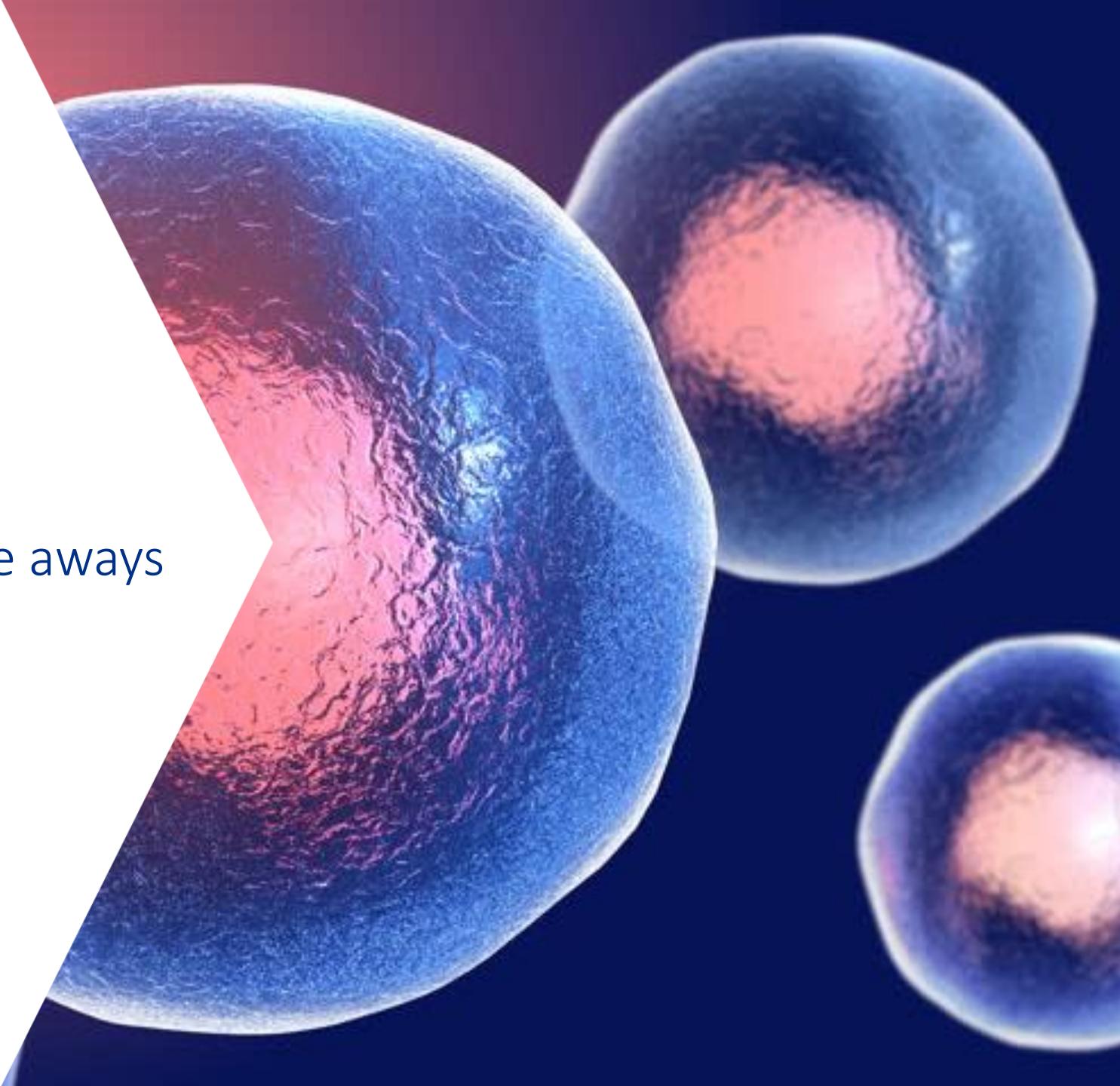


Total # cells per sample:

1 6596  
2 6610  
3 6458  
4 8343

08

The take aways



## Summary of key insights:



The most frequent cell types in the samples are neutrophils (GN.ARTH), T cells (T.DN2B), and monocytes (MO.6C+II-).



The cell type compositions of samples dissociated with enzymatic digestion and the SimpleFlowTM System are very similar, though samples dissociated with enzymatic digestion contain a slightly higher proportion of T cells (T.DN2B).



Cells dissociated with enzymatic digestion have a substantial increase in expression of genes from the “stress response” and the “response to heat” pathways. Overexpression of these pathways is indicative of cellular stress conditions imposed by the circumstances of tissue digestion and the high temperature of enzymatic digestion (it is performed at 37°, whereas SimpleFlowTM is performed at 4°).



Overall, the SimpleFlowTM system has demonstrated to be an excellent dissociation methodology of single cells because: 1) it generates similar results to those obtained with the conventional methodology of enzymatic digestion, and 2) reduces the cellular stress imposed by the procedures.



# THANKS

Does anyone have any questions?