DNA detection techniques:
Quantitative polymerase chain reaction (PCR) + Electrophoresis:
○ Reagents required:
- DNA
→ 2* Primers (antiparallel structure)
Taq Polymerase Taq
Polynerase buffer
· dNTP
· Water
○ PCR cycle:
DNA double strand separation (95°C) 1min
▶ Primer annealing (~ 65°C) 30sec
DNA elongation (~ 75°C) 30sec + 5min
Electrophoresis separate DNA fragments based on relative molecular mass.
Electric current applied to agarose gel
 Fragments placed in wells at negative end, attracted by positive side.
Speed of movement inversely correlated to fragment size.
Visualised using intercalating agent stains.
Sanger sequencing - Low throughput sequencing
○ Components:
Template ssDNA
- Primers
DNA polymerase I
• 4 dNTPs
4 ddNTP (small amounts labelled in different colours)
ddNTP terminates sequence generation results in different length fragment
○ Radioactive labelling (all dNTP and ddNTP are radioactive to improve intensity on polyacrylamide gel)
Different termination length separated by electrophoresis. Deduce sequence from last nucleotide
○ Fluorescence labelling (only ddNTP are fluoresent tagged)
Colour of fragment shows last nucleotide, capillary gel electrophoresis allow drawing of electropherogram.
Next Generation Sequencing (NGS)
Characteristics: Lower costs, faster, higher accuracy, more sensitive.
Illumina sequencing (sequencing by synthesis):
○ Library preparation: Fragmentation of DNA, adaptors are ligated onto the ends, complementary to immobilised
adaptors on solid surface
○ Cluster generation: bridge amplification of DNA, create clusters of identical immobilised DNA segments
○ Sequencing by synthesis:
A, T, C, G dNTP are added with an attached fluorescent terminator, DNA segments elongate by one
nucleotide.
 Imaging: laser illumination causes each ATCG dNTP to emit different colour fluorescence, image recorded.
Peset: fluorescent terminator sequence cleaved off, cycle continues.
○ Analysis: Fluorescent images of each cluster sequenced, sequences aligned and assembled into longer sequence

Genetic engineering Restriction endonuclease: Three types of endonuclease (I II III), type I and III have different recognition site and cleavage site, type II have same. Recognise short palindromic sequence (~6bp) Catalyse hydrolysis of phosphodiester bonds (Type II RE does not require ATP for hydrolysis) Can create staggered cuts, blunt cuts Cloning Cleavage of plasmid Segment of desirable gene cleaved ou tusing RE Incorporation of segment into the plasmid, sealed by ligase Transformation of recombinant vector into the host cell, allow replication Selection of cells that successfully uptaken the vector. Allow multiply Clustered Regularly Interspaced Short Palindromic Repeats (CRISPR) CRISPR locus contains: Transactivating CRISPR RNA (TracrRNA) Cas gene AT-rich leader sequence (promotor) Short semi-palindromic sequences (~20bp), with spacers in between, spacers are specific foreign DNA sequences acquired by the bacteria, incorporated into the array using cas9. Upon viral invasion: RNA polymerase bind to leader sequence, transcription of CRISPR RNA (crRNA) precursors Transcription and translation of cas genes into cas proteins. Trans-activating crRNA (tracrRNA) transcribed, bind to crRNA precursor, cleaved by RNase III, form gRNA gRNA incorporate with cas nuclease protein - form CRISPR complex Spacer RNA in crRNA recognise foreign sequence, cas induce double stranded breaks Before cutting, cas9 check for protospacer adjacent motif (PAM), a sequence downstream of complementary part (XGG for cas9). Applications: can be used to create sticky ends, allow deletion, insertion of segments Prime editing Cas9 nuclease replaced by cas9 nickase, which only induce single stranded breaks, bound to reverse transcriptase gRNA replaced by prime editing gRNA(pegRNA), also contains a primer binding sequence and a edited desired RNA template crRNA recognise sequence, nickase induce ssb, nicked ssDNA bind with primer binding sequence, RT transcribe desired DNA. Desired DNA sequence incorporated back into gap created by nickase, unedited strand fixed with cellular mechanism Means of delivery Ex-vivo delivery: Cell extracted from the host, edited in culture solution then replaced In-vivo delivery: Editing complex introduced into host via viral factor; nanoparticle; microinjection; lipofection etc

6 PCR components
3 PCR cycle
Electrophoresis
Sanger sequencing (2 types)
4 NGS characteristic
4 stages Illumina sequencing
Type Restriction endonuclease, actions
Cloning
4 components of the CRISPR locus
Viral infection
Prime editing
Delivery