

## Sources of DNA damage

- Intrinsic

- Metabolic byproduct (ROS, uric acid)
- Replication error (nucleotide mismatch, topoisomerase not repaired)
- rNTP instead of dNTP
- Spontaneous damage (deamination)

- Extrinsic

- Ionising radiation (U.V.)
- Intercalating agent (Disrupt base stacking)
- Alkylating agent (Disrupt sugar phosphate backbone EtBr)
- Genotoxic agents (Phthalates)
- Base analogues (Replace base nucleotides) (5-bromouracil replaces thymine)

## Repair mechanisms

### Single strand DNA repair

- Post replication mismatch repair (MMR)

- Mismatched dNTP identified in the polymerase
- Replication stops, 3' end of daughter strand transferred to exonuclease site
- Mismatched nucleotide cleaved off, 3' end transferred back to catalysing site

- Mutator protein (Mut) dependent (In E.Coli)

- Mismatch identified immediately after leaving polymerase
- Dam methylase hemimethylate the dsDNA GATC sequence, parent strand methylated, not the daughter strand.
- MutS and MutL bind to mismatch site, read bidirectionally, form loop, until hemimethylated GATC
- MutS MutL recruit MutH, cleaves unmethylated daughter strand from GATC.
- Helicase II + Pol I unwind and cleave daughter strand to mismatch
- Pol III add nucleotide to gap, sealed by ligase

- Base Excision Repair (BER)

- Glycosylase bind to mismatch, flip out the base, cleave glycosidic bond of nitrogenous base
- AP endonuclease cleave phosphodiester bond of mismatched nucleotide
- Repaired by Pol I, sealed by ligase

- Nucleotide excision repair (NER), bulkier than BER

- Four Uvr proteins (UvrA~D)
- UvrA and UvrB scan genome, UvrB stays at site of mismatch
- UvrA leaves, B recruits C to unwind and excise ~15 nucleotide fragment
- C leaves, D removes the fragment, all leaves
- Pol I repair, ligase seal

- Direct reversal of modifications

- Photolyase can reverse UV induced thymine dimers
- Methyltransferase can reverse methylated nucleotides
- No DNA excision, no template needed, but costly as enzyme commit suicide afterwards

- Models of translesion synthesis (TLS) (replication across damaged nucleotide)

- Polymerase switching: High fidelity pol stops at lesion, switch to TLS Pol, then switch back
- Gap filling: High fidelity Pol does not pair dNTP at lesion, filled by TLS Pol

## Double strand DNA repair

- Non-homologous end joining
  - Characteristic: Always available, common in non-dividing cells (G0, G1, S), prone to insertion & deletion
  - Ku70/80 recognise and bind to double stranded breaks
  - Recruit DNA-protein catalytic subunit (DNA-PKcs), form DNA-PK complex
  - Binding of nuclease Artemis, phosphorylated, activated and trims the end of DSB (prone to deletion)
  - Modification of ends by other enzymes (fixing gaps, trimming, prone to insertion)
  - Joining of blunt ends, ligase seal.
- Homology directed repair
  - Characteristic: Available in dividing cells (G2/S stage), less error prone
  - MRN recognise the error sequence
  - MRN recruit ATM to activate array of downstream proteins. exonuclease resect the end and create 3' overhangs.
  - RPA binds to ssDNA, stabilisation, BRCA2 and RAD51 replace RPA ssDNA, forming nucleofilament
  - RAD51 scans for homologous sequence in sister chromatid, RAD54 direct invasion
  - RAD51 dissociate, Polymerase synthesis of invading strand.
  - Invading strand dissociate, bind to 3' overhang other damaged strand, polymerase + ligase seal.
- Cell cycle checkpoints
  - G1: before S phase, check for nutrient availability and cell size
  - G2: Before mitosis check for complete replication and DNA damage
  - M (Spindle): before anaphase, check if spindles attached to kinetochores
  - S: Check for DNA fidelity and genome stability
- Proteins involved in cell cycle regulation:
  - Cyclin: activate CDK, drive division. Different cyclin signal different cycle events
    - Cyclin dependent kinase (CDK), upregulate cell cycle
    - Cyclin dependent kinase Inhibitor (CDKI), downregulate cell cycle



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Cell cycle checkpoints, what is assessed