

ABCB1_proximal_seq_tfs

charvi khanna

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```
# Load necessary libraries
library(TFBSTools)
library(Biostrings)

## Loading required package: BiocGenerics

## Loading required package: generics

##
## Attaching package: 'generics'

## The following objects are masked from 'package:base':
##       as.difftime, as.factor, as.ordered, intersect, is.element, setdiff,
##       setequal, union

##
## Attaching package: 'BiocGenerics'

## The following objects are masked from 'package:stats':
##       IQR, mad, sd, var, xtabs

## The following objects are masked from 'package:base':
##       anyDuplicated, aperm, append, as.data.frame, basename, cbind,
##       colnames, dirname, do.call, duplicated, eval, evalq, Filter, Find,
##       get, grep, grepl, is.unsorted, lapply, Map, mapply, match, mget,
##       order, paste, pmax, pmax.int, pmin, pmin.int, Position, rank,
##       rbind, Reduce, rownames, sapply, saveRDS, table, tapply, unique,
##       unsplit, which.max, which.min

## Loading required package: S4Vectors

## Loading required package: stats4

##
## Attaching package: 'S4Vectors'
```

```

## The following object is masked from 'package:utils':
##
##     findMatches

## The following objects are masked from 'package:base':
##
##     expand.grid, I, uname

## Loading required package: IRanges

##
## Attaching package: 'IRanges'

## The following object is masked from 'package:grDevices':
##
##     windows

## Loading required package: XVector

## Loading required package: GenomeInfoDb

##
## Attaching package: 'Biostrings'

## The following object is masked from 'package:base':
##
##     strsplit

library(JASPAR2022)

## Loading required package: BiocFileCache

## Loading required package: dbplyr

library(dplyr)

##
## Attaching package: 'dplyr'

## The following objects are masked from 'package:dbplyr':
##
##     ident, sql

## The following objects are masked from 'package:Biostrings':
##
##     collapse, intersect, setdiff, setequal, union

## The following object is masked from 'package:GenomeInfoDb':
##
##     intersect

```

```

## The following object is masked from 'package:XVector':
##
##      slice

## The following objects are masked from 'package:IRanges':
##
##      collapse, desc, intersect, setdiff, slice, union

## The following objects are masked from 'package:S4Vectors':
##
##      first, intersect, rename, setdiff, setequal, union

## The following objects are masked from 'package:BiocGenerics':
##
##      combine, intersect, setdiff, setequal, union

## The following object is masked from 'package:generics':
##
##      explain

## The following objects are masked from 'package:stats':
##
##      filter, lag

```

```

## The following objects are masked from 'package:base':
##
##      intersect, setdiff, setequal, union

```

```

library(writexl)
library(knitr)
library(kableExtra)

```

```

## Warning: package 'kableExtra' was built under R version 4.5.2

```

```

##
## Attaching package: 'kableExtra'

```

```

## The following object is masked from 'package:dplyr':
##
##      group_rows

```

```

# Global chunk options
knitr::opts_chunk$set(echo = TRUE, message = FALSE, warning = FALSE)

```

```

ABCB1_proximal <- DNAString("TCACAGGTAAAGAGTAATGATAGCCTTAAAATGGTAATACAAGTGTATCCAGTACCAAGAGGAGGAGCTAC")

```

```

# Get human motifs

```

```

opts <- list(species = 9606)
motifs <- getMatrixSet(JASPAR2022, opts)

```

```

# Convert motifs to PWMs using log2 probability ratios

pwm <- toPWM(motifs, type = "log2probratio")

# Scan sequence for all motifs

hits <- searchSeq(
pwm,
ABCB1_proximal,
seqname = "ABCB1_proximal",
min.score = "95%", # high threshold
strand = "*" # both strands
)

# Keep only motifs with at least one hit

hits_filtered <- hits[sapply(hits, length) > 0]

# Convert to data.frame and add TF names

hits_df <- do.call(rbind, lapply(hits_filtered, as.data.frame))
hits_df$TF <- rep(names(hits_filtered), sapply(hits_filtered, length))

# Keep only desired columns

hits_df_small <- hits_df %>%
select(class, start, end, siteSeqs, absScore, relScore)

# Display top hits

kable(head(hits_df_small, 10), booktabs = TRUE, longtable = TRUE) %>%
kable_styling(latex_options = c("striped", "scale_down"))

```

```

## Warning in styling_latex_scale(out, table_info, "down"): Longtable cannot be
## resized.

```

| | class | start | end | siteSeqs | absScore | relScore |
|------------|---|-------|-----|-------------|-----------|-----------|
| MA0130.1.1 | C2H2 zinc finger factors | 486 | 491 | ATCCAC | 9.890853 | 1.0000000 |
| MA0130.1.2 | C2H2 zinc finger factors | 135 | 140 | CTCCAC | 9.675124 | 0.9940480 |
| MA0144.2 | STAT domain factors | 413 | 423 | TTACTGGGAAG | 10.125801 | 0.9534647 |
| MA0599.1.1 | C2H2 zinc finger factors | 132 | 141 | CCTCCACCCA | 10.182292 | 0.9625844 |
| MA0599.1.2 | C2H2 zinc finger factors | 143 | 152 | CTCCCACCCCT | 10.356340 | 0.9637991 |
| MA0599.1.3 | C2H2 zinc finger factors | 147 | 156 | TCTACTCCCA | 8.496201 | 0.9508171 |
| MA0610.1 | DM-type intertwined zinc finger factors | 556 | 566 | AATGTAACAAG | 13.616380 | 0.9700553 |
| MA0613.1 | Fork head/winged helix factors | 44 | 51 | ATAAACAC | 11.969470 | 0.9692009 |
| MA0618.1.1 | Homeo domain factors | 241 | 248 | TTAATGAG | 10.576243 | 0.9666860 |
| MA0618.1.2 | Homeo domain factors | 502 | 509 | ATAATTAG | 12.068281 | 0.9855810 |

```

# Filter hits with absolute score greater than 10

hits_df_filtered_small <- hits_df_small %>% filter(absScore > 10)

```

```
# Show filtered hits

kable(head(hits_df_filtered_small, 10), booktabs = TRUE, longtable = TRUE) %>%
kable_styling(latex_options = c("striped", "scale_down"))
```

```
## Warning in styling_latex_scale(out, table_info, "down"): Longtable cannot be
## resized.
```

| | class | | start | end | siteSeqs | absScore | relScore |
|------------|---|--|-------|-----|-------------|----------|-----------|
| MA0144.2 | STAT domain factors | | 413 | 423 | TTACTGGGAAG | 10.12580 | 0.9534647 |
| MA0599.1.1 | C2H2 zinc finger factors | | 132 | 141 | CCTCCACCCA | 10.18229 | 0.9625844 |
| MA0599.1.2 | C2H2 zinc finger factors | | 143 | 152 | CTCCCACCCCT | 10.35634 | 0.9637991 |
| MA0610.1 | DM-type intertwined zinc finger factors | | 556 | 566 | AATGTAACAAG | 13.61638 | 0.9700553 |
| MA0613.1 | Fork head/winged helix factors | | 44 | 51 | ATAAACAC | 11.96947 | 0.9692009 |
| MA0618.1.1 | Homeo domain factors | | 241 | 248 | TTAATGAG | 10.57624 | 0.9666860 |
| MA0618.1.2 | Homeo domain factors | | 502 | 509 | ATAATTAG | 12.06828 | 0.9855810 |
| MA0628.1.1 | Homeo domain factors | | 501 | 510 | AATAATTAGT | 13.17289 | 0.9737588 |
| MA0628.1.2 | Homeo domain factors | | 501 | 510 | ACTAATTATT | 13.17289 | 0.9737588 |
| MA0634.1 | Homeo domain factors | | 303 | 312 | TTCAATTATA | 10.59824 | 0.9542863 |

```
write_xlsx(hits_df_filtered_small, path = "ABCB1_proximal_hits_filtered.xlsx")
```

```
# Number of motifs scanned
```

```
length(pwm)
```

```
## [1] 692
```

```
# Number of motifs with hits
```

```
length(hits_filtered)
```

```
## [1] 98
```

```
# Number of filtered hits
```

```
nrow(hits_df_filtered_small)
```

```
## [1] 118
```