Importing and combining data sets

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Importing data into R

The readr package (found in the tidyverse collection) contains a number of useful functions of the form read_* to import data. For example, if you have a .csv file, you would use the read_csv function

Download a file from uniprot.org

After selecting some columns of interest, click the Download button and download as a compressed Text file

Rename the file to something simple (yet informative!), like uniprot and make sure the extension is .tsv

To import into RStudio, run the following

```
uniprot <- read_tsv("uniprot.tsv")

## Parsed with column specification:
## cols(
## Entry = col_character(),
## `Gene names` = col_character(),
## Length = col_double()</pre>
```

You can also use the readr package to import data from a URL

For example, to load a dataset from the (very useful) Tidy Tuesday series, run the following

pizza <- read_csv("https://raw.githubusercontent.com/rfordatascience/tidytuesday/master/data/2019/2019-

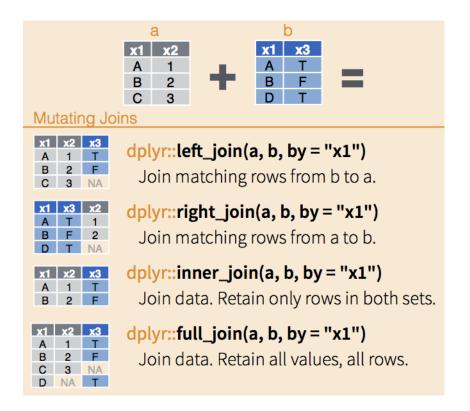
This data set contains ratings of various pizzerias in Manhattan

Combining datasets

)

There are many times when you have two or more overlapping datasets that you would like to combine

The dplyr package has a number of *_join functions for this purpose



left_join

Returns all rows from a, and all columns from a and b

Rows in a with no match in b will have NA values in the new columns

If there are multiple matches between a and b, all combinations of the matches are returned

First, load the two datasets needed for this example - proteins and mitocarta

```
library(proteins)
library(mitocarta)
```

Take a look at the variables in each dataset

gene name in proteins and symbol in mitocarta contain the gene IDs for each gene. They can therefore serve as a common variable

Let's join on this

left_join example

left_join proteins with mitocarta and assign the output to a new object called pm_left

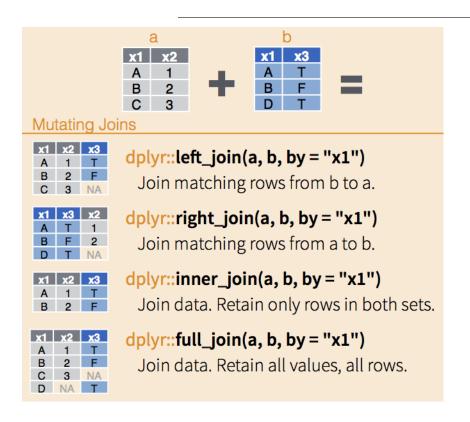
```
pm_left <- left_join(proteins, mitocarta, by = c("gene_name" = "symbol"))</pre>
pm_left %>% head(1)
## # A tibble: 1 x 50
     uniprot_id gene_name gene_name_alt protein_name protein_name_alt sequence
```

```
##
     <chr>
                <chr>>
                           <chr>
                                         <chr>
                                                       <chr>>
## 1 P04217
                A1BG
                           <NA>
                                         "Alpha-1B-g~ Alpha-1-B glyco~ MSMLVVF~
## # ... with 44 more variables: length <dbl>, mass <dbl>, training_dataset <chr>,
      human_gene_id <dbl>, mouse_ortholog_gene_id <dbl>, synonyms <chr>,
```

#

```
## #
       target_p_score <dbl>, mito_domain_score <chr>,
## #
       coexpression_gnf_n50_score <dbl>, pgc_induction_score <dbl>,
## #
       yeast_mito_homolog_score <chr>, rickettsia_homolog_score <chr>,
## #
       msms_score <chr>, mcarta2_score <dbl>, mcarta2_fdr <dbl>,
## #
       mcarta2_list <dbl>, mcarta2_evidence <chr>, hg19_chromosome <fct>,
## #
       hg19 start <dbl>, hg19 stop <dbl>, msms num tissues <dbl>,
       msms num peptides unique <dbl>, msms num spectra <dbl>,
## #
       msms_total_intensity <dbl>, msms_percent_coverage <dbl>, tissues <chr>,
## #
## #
       cerebrum_total_peak_intensity_log10 <dbl>,
## #
       cerebellum_total_peak_intensity_log10 <dbl>,
## #
       brainstem_total_peak_intensity_log10 <dbl>,
       spinalcord_total_peak_intensity_log10 <dbl>,
## #
## #
       kidney_total_peak_intensity_log10 <dbl>,
## #
       liver_total_peak_intensity_log10 <dbl>,
## #
       heart_total_peak_intensity_log10 <dbl>,
## #
       skeletalmuscle_total_peak_intensity_log10 <dbl>,
## #
       adipose_total_peak_intensity_log10 <dbl>,
## #
       smallintestine total peak intensity log10 <dbl>,
## #
       largeintestine_total_peak_intensity_log10 <dbl>,
## #
       stomach_total_peak_intensity_log10 <dbl>,
## #
       placenta_total_peak_intensity_log10 <dbl>,
## #
       testis_total_peak_intensity_log10 <dbl>,
## #
       hpa_primary_subcellular_localization_2015 <chr>
```

Now you have one dataset with additional useful information



right_join

Returns all rows from b, and all columns from a and b

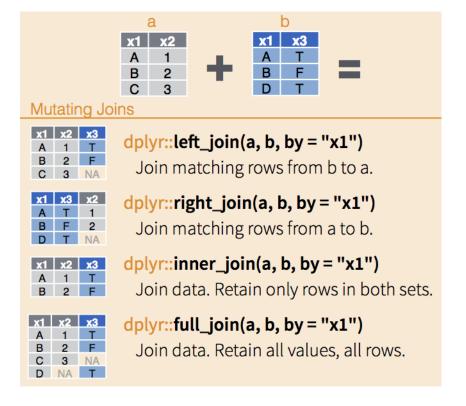
Rows in b with no match in a will have NA values in the new columns

If there are multiple matches between a and b, all combinations of the matches are returned

right_join example

right_join proteins with mitocarta and assign the output to a new object called pm_right

```
pm_right <- right_join(proteins, mitocarta, by = c("gene_name" = "symbol"))</pre>
pm_right %>% head(1)
## # A tibble: 1 x 50
##
     uniprot_id gene_name gene_name_alt protein_name protein_name_alt sequence
##
     <chr>>
                                                      <chr>
                                                                        <chr>
                <chr>
                          <chr>>
                                         <chr>>
## 1 P04217
                A1BG
                          <NA>
                                         "Alpha-1B-g~ Alpha-1-B glyco~ MSMLVVF~
## # ... with 44 more variables: length <dbl>, mass <dbl>, training_dataset <chr>,
       human_gene_id <dbl>, mouse_ortholog_gene_id <dbl>, synonyms <chr>,
## #
## #
       description <chr>, ensembl_gene_id <chr>, protein_length <dbl>,
## #
       target_p_score <dbl>, mito_domain_score <chr>,
## #
       coexpression_gnf_n50_score <dbl>, pgc_induction_score <dbl>,
## #
       yeast_mito_homolog_score <chr>, rickettsia_homolog_score <chr>,
## #
       msms_score <chr>, mcarta2_score <dbl>, mcarta2_fdr <dbl>,
## #
       mcarta2_list <dbl>, mcarta2_evidence <chr>, hg19_chromosome <fct>,
## #
       hg19 start <dbl>, hg19 stop <dbl>, msms num tissues <dbl>,
## #
       msms_num_peptides_unique <dbl>, msms_num_spectra <dbl>,
## #
       msms total intensity <dbl>, msms percent coverage <dbl>, tissues <chr>,
## #
       cerebrum_total_peak_intensity_log10 <dbl>,
## #
       cerebellum_total_peak_intensity_log10 <dbl>,
## #
       brainstem total peak intensity log10 <dbl>,
## #
       spinalcord total peak intensity log10 <dbl>,
## #
       kidney_total_peak_intensity_log10 <dbl>,
       liver_total_peak_intensity_log10 <dbl>,
## #
## #
       heart_total_peak_intensity_log10 <dbl>,
## #
       skeletalmuscle_total_peak_intensity_log10 <dbl>,
       adipose_total_peak_intensity_log10 <dbl>,
## #
## #
       smallintestine_total_peak_intensity_log10 <dbl>,
## #
       largeintestine_total_peak_intensity_log10 <dbl>,
## #
       stomach_total_peak_intensity_log10 <dbl>,
## #
       placenta_total_peak_intensity_log10 <dbl>,
## #
       testis_total_peak_intensity_log10 <dbl>,
       hpa primary subcellular localization 2015 <chr>
## #
```



inner_join

Returns all rows from a where there are matching values in b, and all columns from a and b If there are multiple matches between a and b, all combination of the matches are returned

inner join example

```
inner_join proteins with mitocarta and assign the output to a new object called pm_inner
```

```
pm_inner <- inner_join(proteins, mitocarta, by = c("gene_name" = "symbol"))
pm_inner %>% head(1)

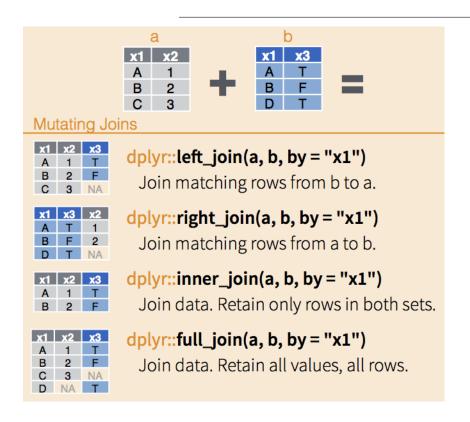
## # A tibble: 1 x 50

## uniprot_id gene_name gene_name_alt protein_name protein_name_alt sequence
```

```
##
##
     <chr>
                <chr>
                          <chr>>
                                                      <chr>
                                                                        <chr>
## 1 P04217
                A1BG
                          <NA>
                                         "Alpha-1B-g~ Alpha-1-B glyco~ MSMLVVF~
## # ... with 44 more variables: length <dbl>, mass <dbl>, training_dataset <chr>,
       human_gene_id <dbl>, mouse_ortholog_gene_id <dbl>, synonyms <chr>,
## #
## #
       description <chr>, ensembl_gene_id <chr>, protein_length <dbl>,
## #
       target_p_score <dbl>, mito_domain_score <chr>,
## #
       coexpression_gnf_n50_score <dbl>, pgc_induction_score <dbl>,
## #
       yeast_mito_homolog_score <chr>, rickettsia_homolog_score <chr>,
## #
       msms_score <chr>, mcarta2_score <dbl>, mcarta2_fdr <dbl>,
## #
       mcarta2_list <dbl>, mcarta2_evidence <chr>, hg19_chromosome <fct>,
       hg19_start <dbl>, hg19_stop <dbl>, msms_num_tissues <dbl>,
## #
## #
       msms num peptides unique <dbl>, msms num spectra <dbl>,
       msms_total_intensity <dbl>, msms_percent_coverage <dbl>, tissues <chr>,
## #
## #
       cerebrum_total_peak_intensity_log10 <dbl>,
## #
       cerebellum_total_peak_intensity_log10 <dbl>,
## #
       brainstem_total_peak_intensity_log10 <dbl>,
```

```
## #
       spinalcord_total_peak_intensity_log10 <dbl>,
## #
       kidney_total_peak_intensity_log10 <dbl>,
## #
       liver total peak intensity log10 <dbl>,
## #
       heart_total_peak_intensity_log10 <dbl>,
## #
       skeletalmuscle_total_peak_intensity_log10 <dbl>,
## #
       adipose total peak intensity log10 <dbl>,
       smallintestine_total_peak_intensity_log10 <dbl>,
## #
## #
       largeintestine_total_peak_intensity_log10 <dbl>,
## #
       stomach_total_peak_intensity_log10 <dbl>,
## #
       placenta_total_peak_intensity_log10 <dbl>,
       testis_total_peak_intensity_log10 <dbl>,
       hpa_primary_subcellular_localization_2015 <chr>
## #
```

Why might this type of join be useful?



full_join

Returns all rows and all columns from both a and b

Where there are no matching values, returns NA for the one missing

full_join example

 $\verb|full_join| \ proteins \ with \ \verb|mitocarta| \ and \ assign \ the \ output \ to \ a \ new \ object \ called \ \verb|pm_full|$

```
pm_full <- full_join(proteins, mitocarta, by = c("gene_name" = "symbol"))
pm_full %>% head(1)
```

```
## # A tibble: 1 x 50
## uniprot_id gene_name gene_name_alt protein_name protein_name_alt sequence
```

```
##
     <chr>>
                <chr>
                          <chr>
                                         <chr>
                                                      <chr>>
                                                                        <chr>
## 1 P04217
                A1BG
                          <NA>
                                         "Alpha-1B-g~ Alpha-1-B glyco~ MSMLVVF~
## # ... with 44 more variables: length <dbl>, mass <dbl>, training dataset <chr>,
       human_gene_id <dbl>, mouse_ortholog_gene_id <dbl>, synonyms <chr>,
## #
       description <chr>, ensembl_gene_id <chr>, protein_length <dbl>,
## #
       target p score <dbl>, mito domain score <chr>,
## #
       coexpression gnf n50 score <dbl>, pgc induction score <dbl>,
       yeast_mito_homolog_score <chr>, rickettsia_homolog_score <chr>,
## #
## #
       msms score <chr>, mcarta2 score <dbl>, mcarta2 fdr <dbl>,
## #
       mcarta2_list <dbl>, mcarta2_evidence <chr>, hg19_chromosome <fct>,
## #
       hg19_start <dbl>, hg19_stop <dbl>, msms_num_tissues <dbl>,
## #
       msms_num_peptides_unique <dbl>, msms_num_spectra <dbl>,
## #
       msms_total_intensity <dbl>, msms_percent_coverage <dbl>, tissues <chr>,
## #
       cerebrum_total_peak_intensity_log10 <dbl>,
## #
       cerebellum_total_peak_intensity_log10 <dbl>,
## #
       brainstem_total_peak_intensity_log10 <dbl>,
## #
       spinalcord_total_peak_intensity_log10 <dbl>,
## #
       kidney total peak intensity log10 <dbl>,
## #
       liver_total_peak_intensity_log10 <dbl>,
## #
       heart total peak intensity log10 <dbl>,
## #
       skeletalmuscle_total_peak_intensity_log10 <dbl>,
## #
       adipose_total_peak_intensity_log10 <dbl>,
       smallintestine_total_peak_intensity_log10 <dbl>,
## #
## #
       largeintestine total peak intensity log10 <dbl>,
       stomach_total_peak_intensity_log10 <dbl>,
## #
       placenta_total_peak_intensity_log10 <dbl>,
## #
## #
       testis_total_peak_intensity_log10 <dbl>,
## #
       hpa_primary_subcellular_localization_2015 <chr>
```